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(54) **NOVEL STRAIN OF SARS-ASSOCIATED CORONAVIRUS AND APPLICATIONS THEREOF**

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(52) **U.S. Cl.** **424/221.1**; 435/5; 435/69.3;
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530/388.3; 536/23.72; 977/802

(57) **ABSTRACT**

The invention relates to a novel strain of severe acute respiratory syndrome (SARS)-associated coronavirus, resulting from a sample collected in Hanoi (Vietnam), reference number 031589, nucleic acid molecules originating from the genome of same, proteins and peptides coded by said nucleic acid molecules and, more specifically, protein N and the applications thereof, for example, as diagnostic reagents and/or as a vaccine.

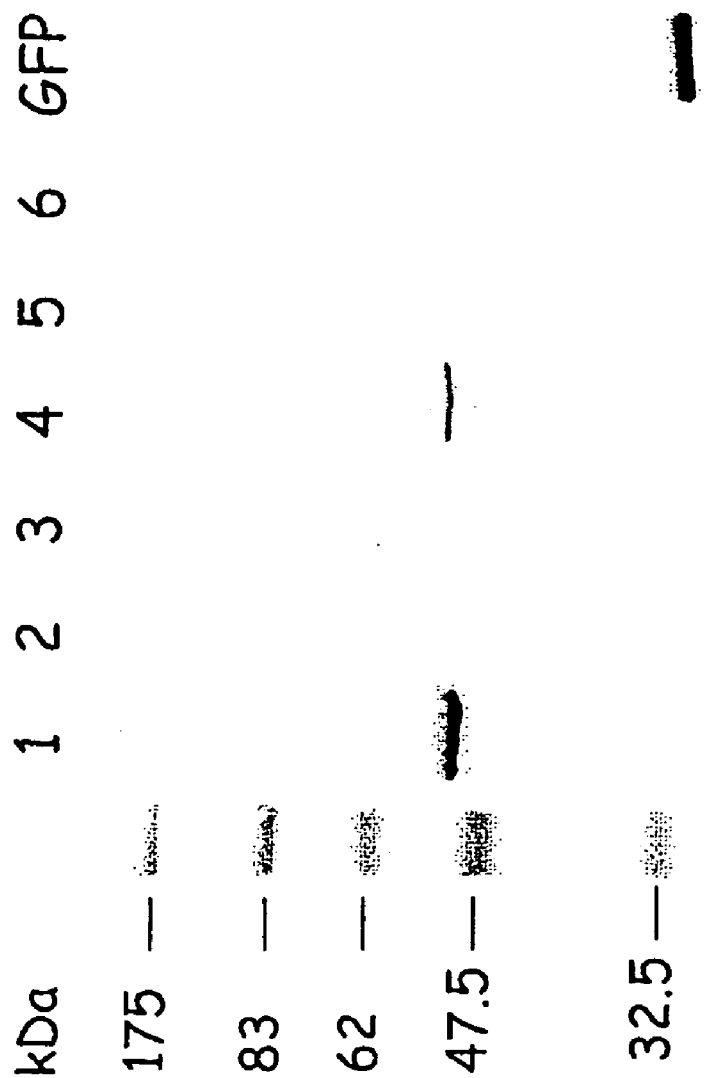


FIGURE 1

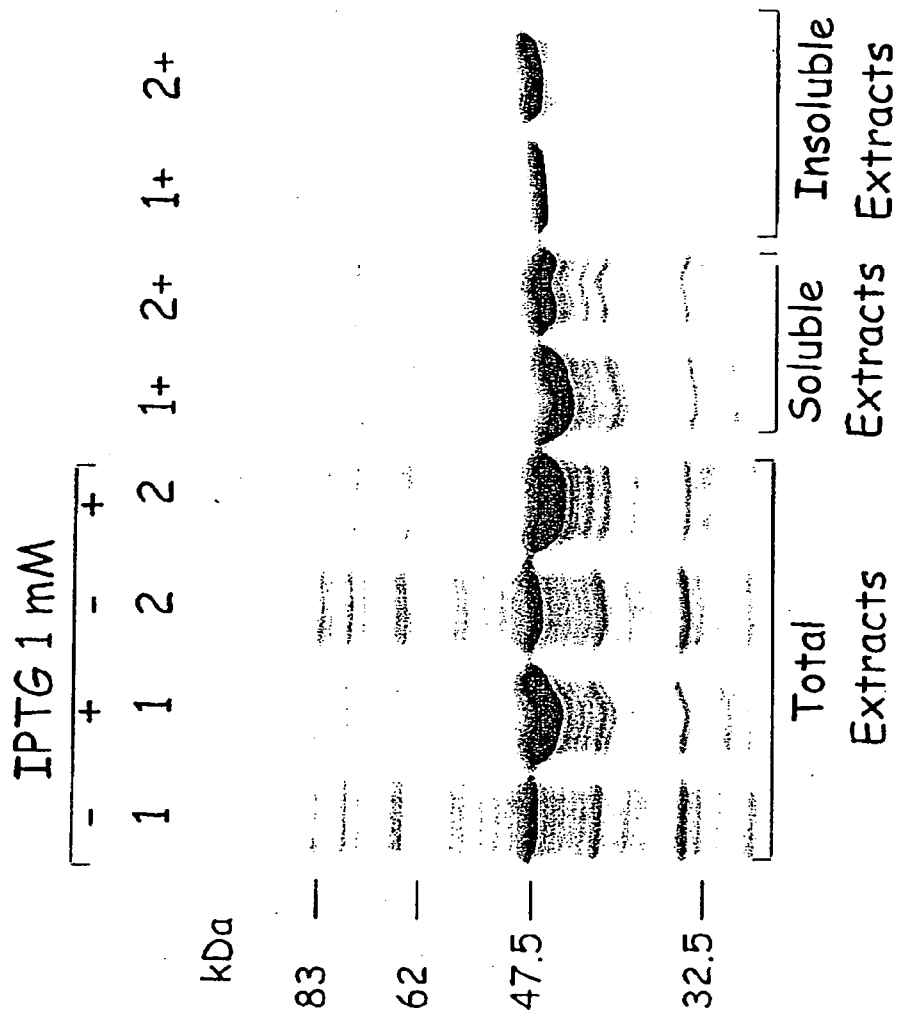


FIGURE 2

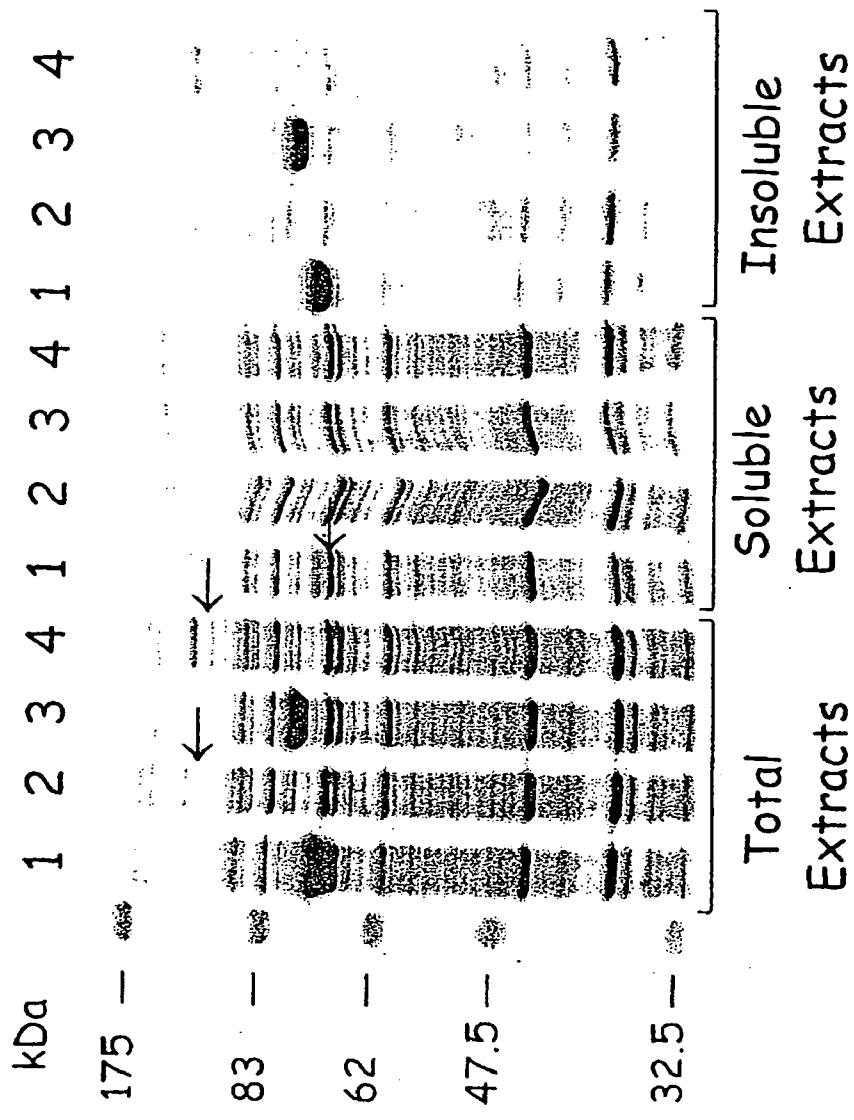


FIGURE 3

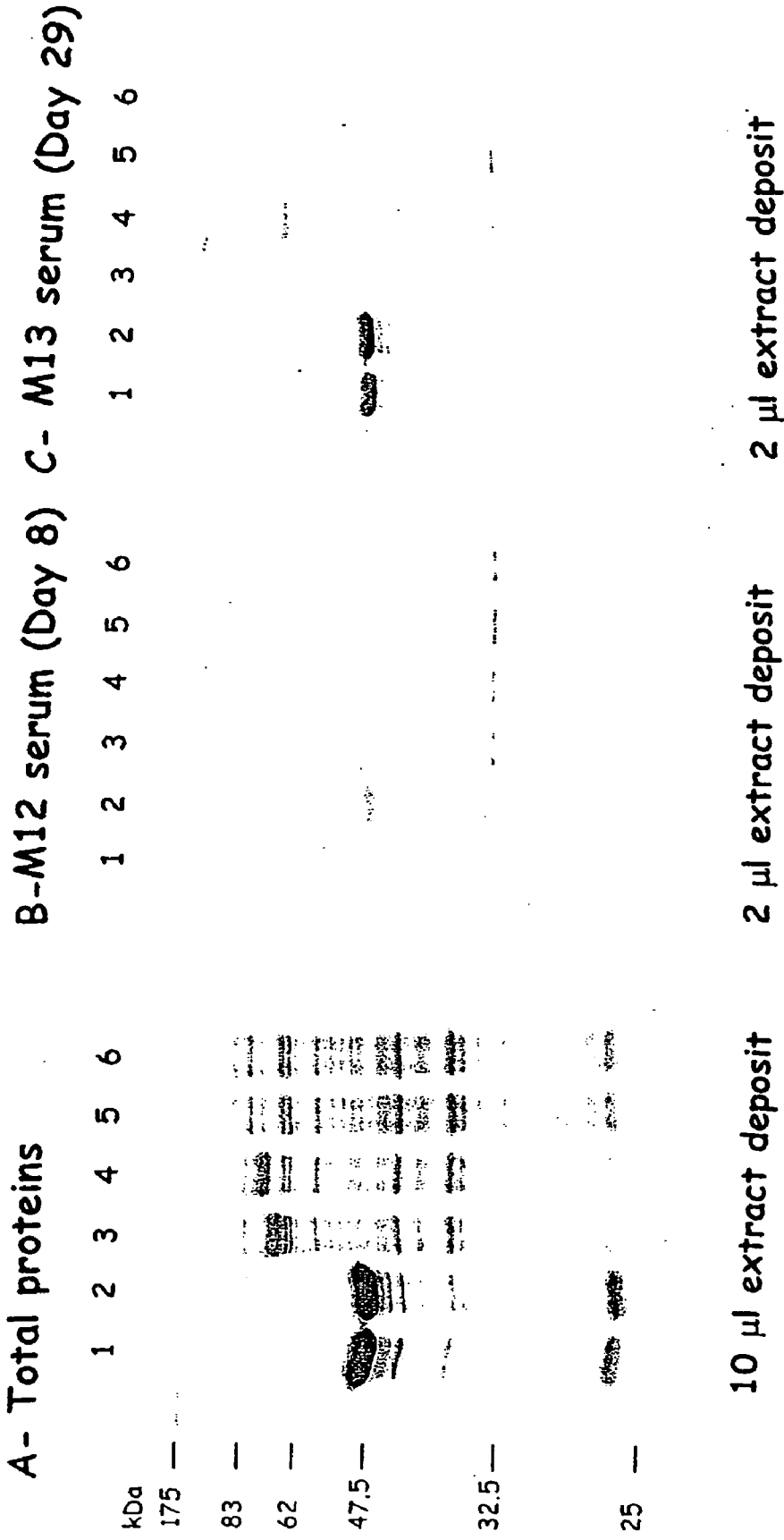


FIGURE 4

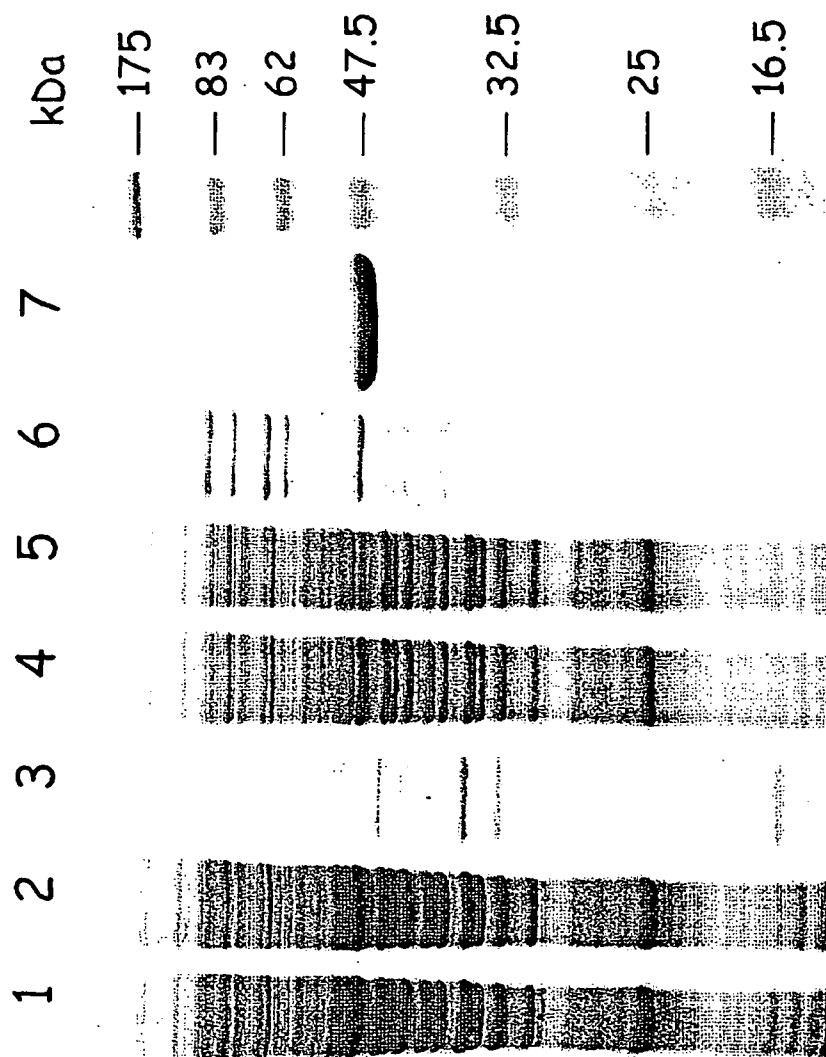


FIGURE 5

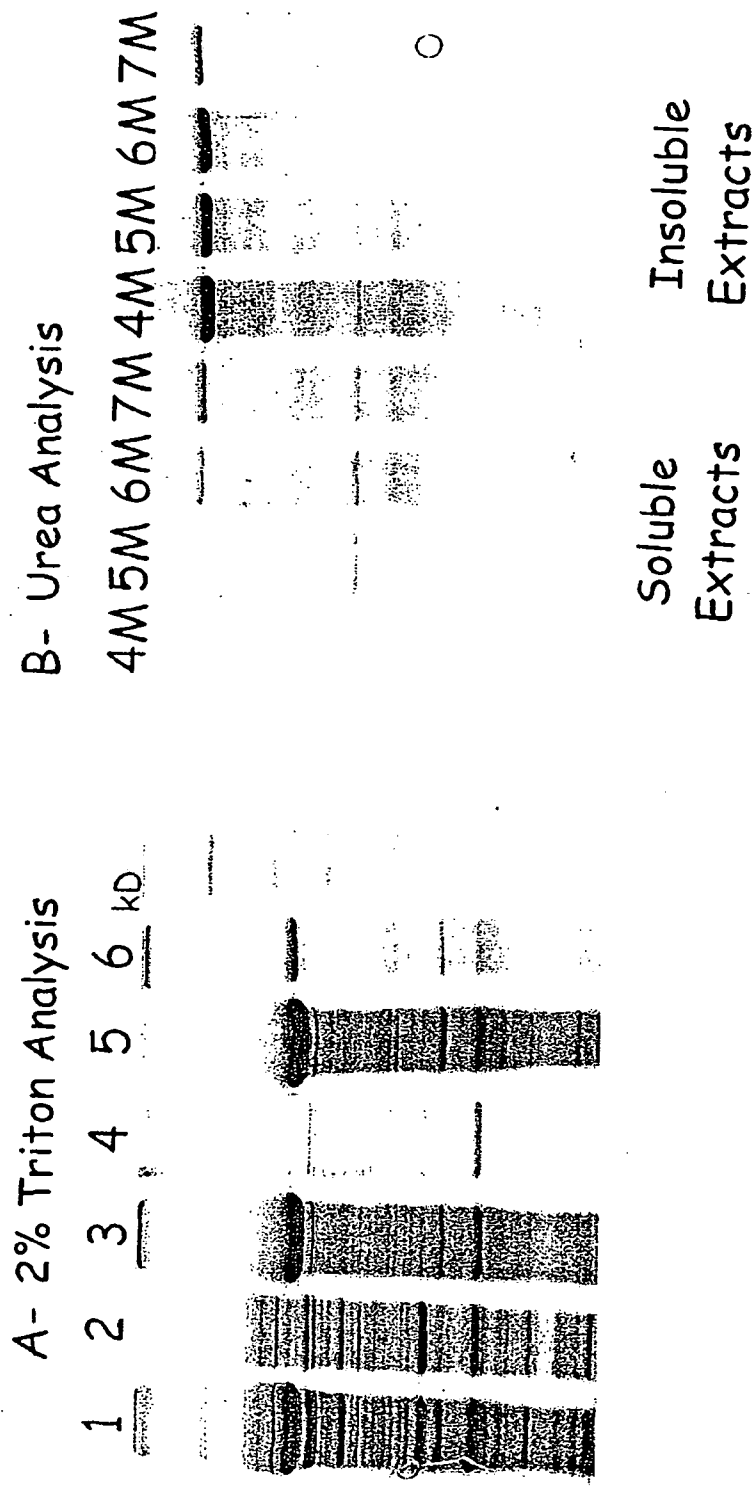


FIGURE 6

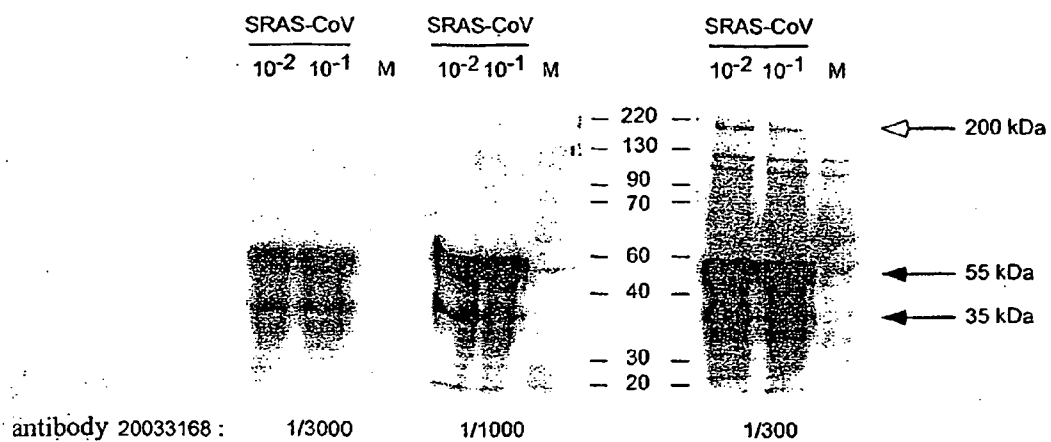


FIGURE 7

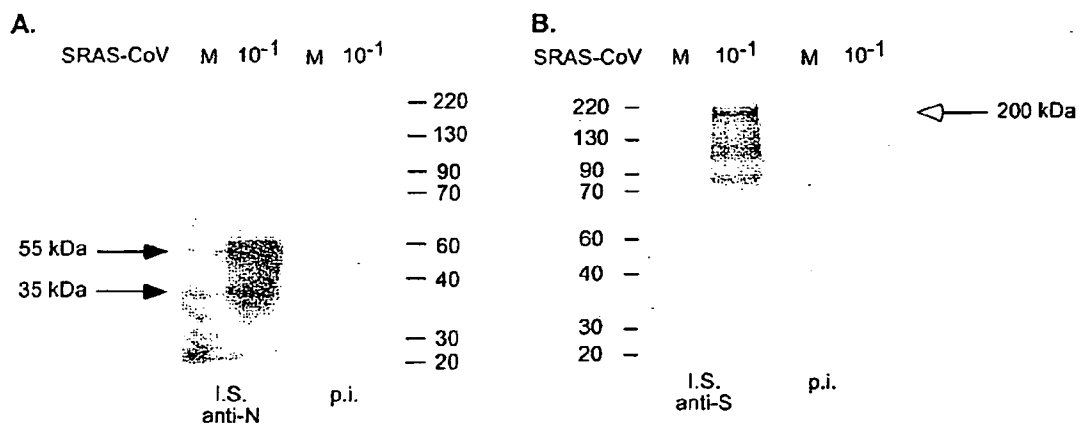
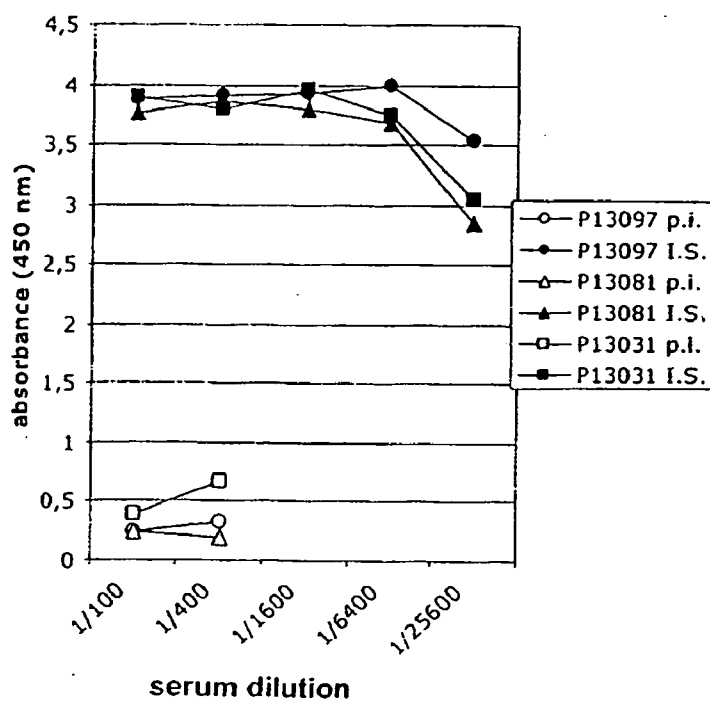


FIGURE 8

A



B

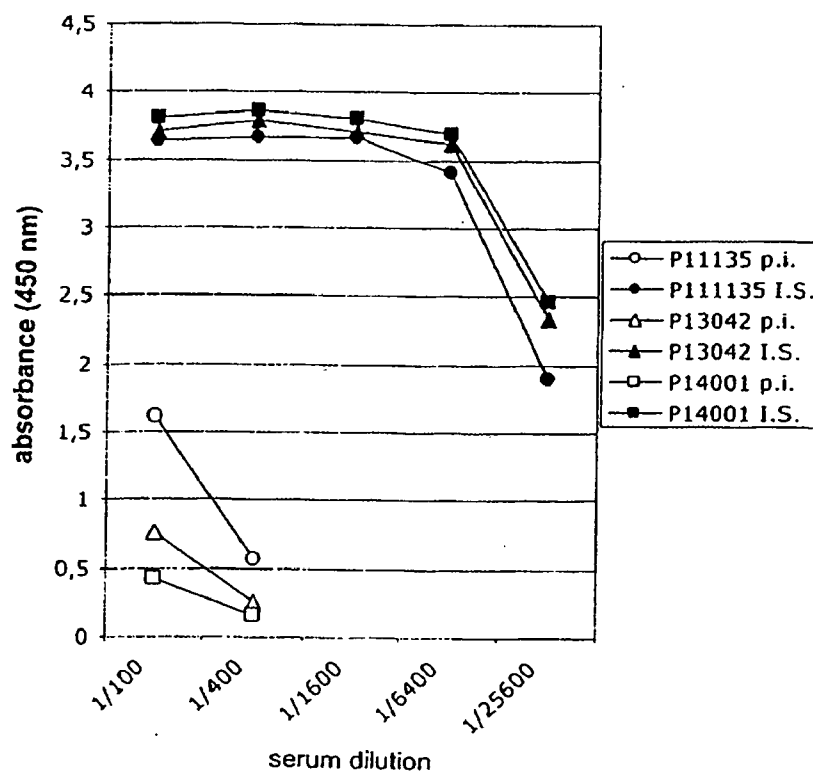


FIGURE 9

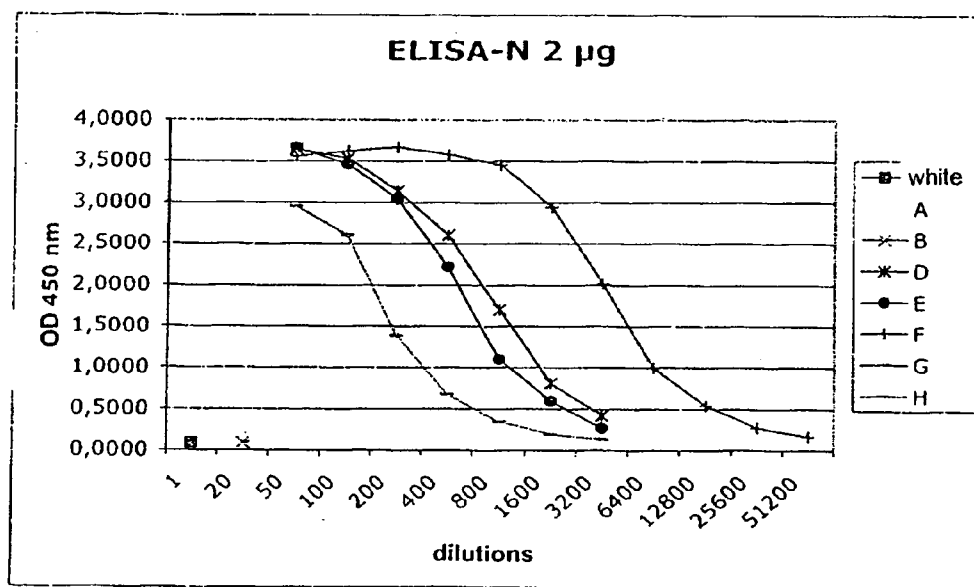
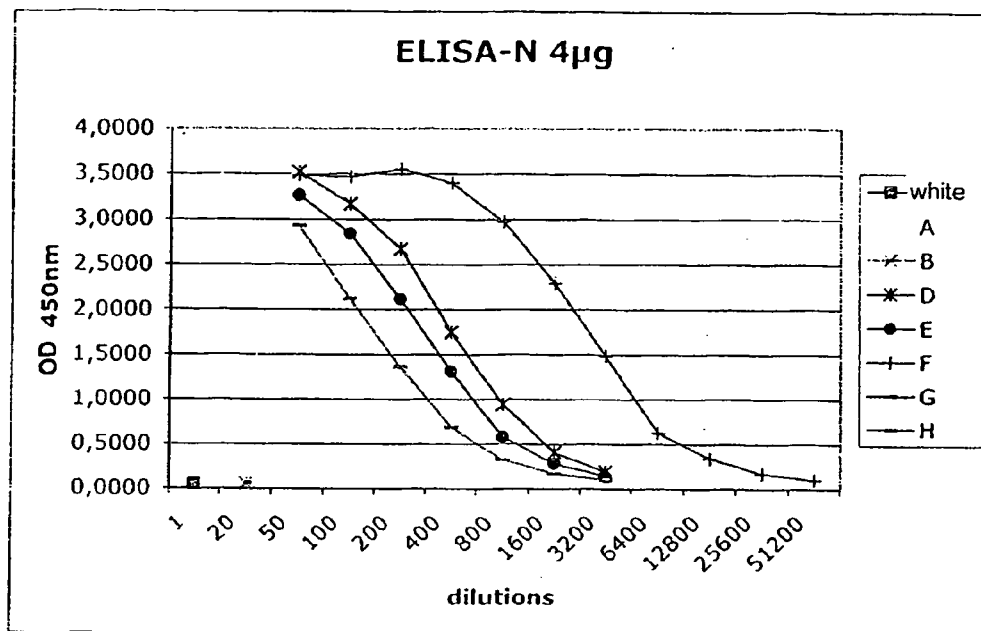


FIGURE 10a

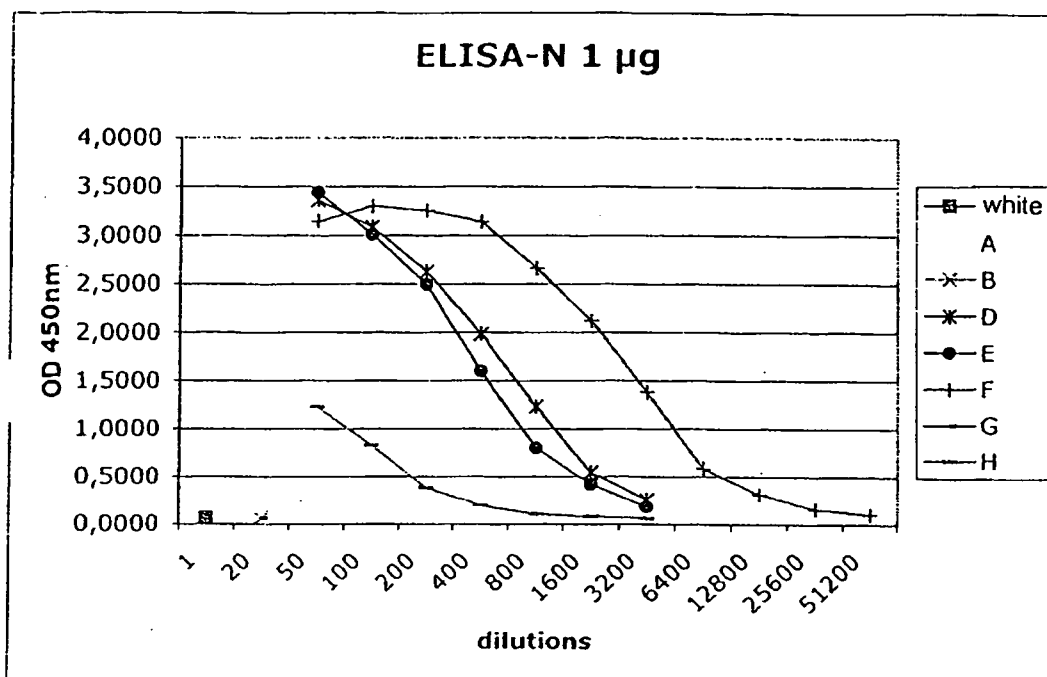


FIGURE 10b

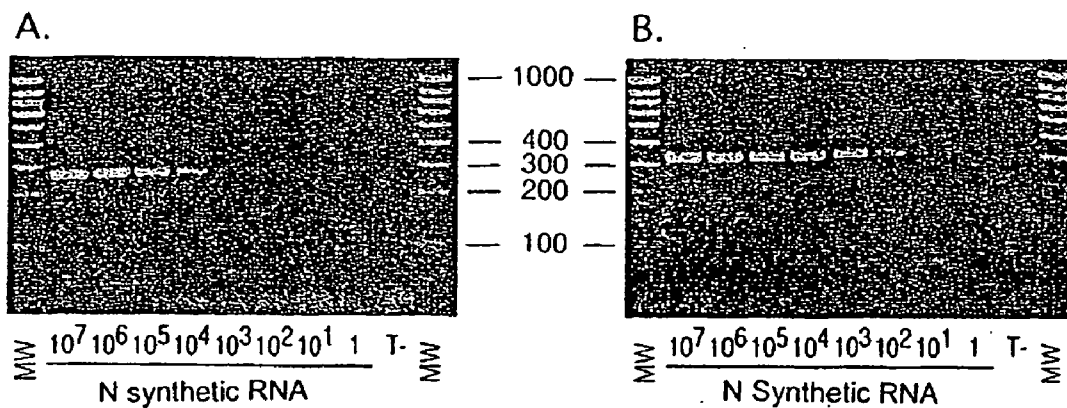


FIGURE 11

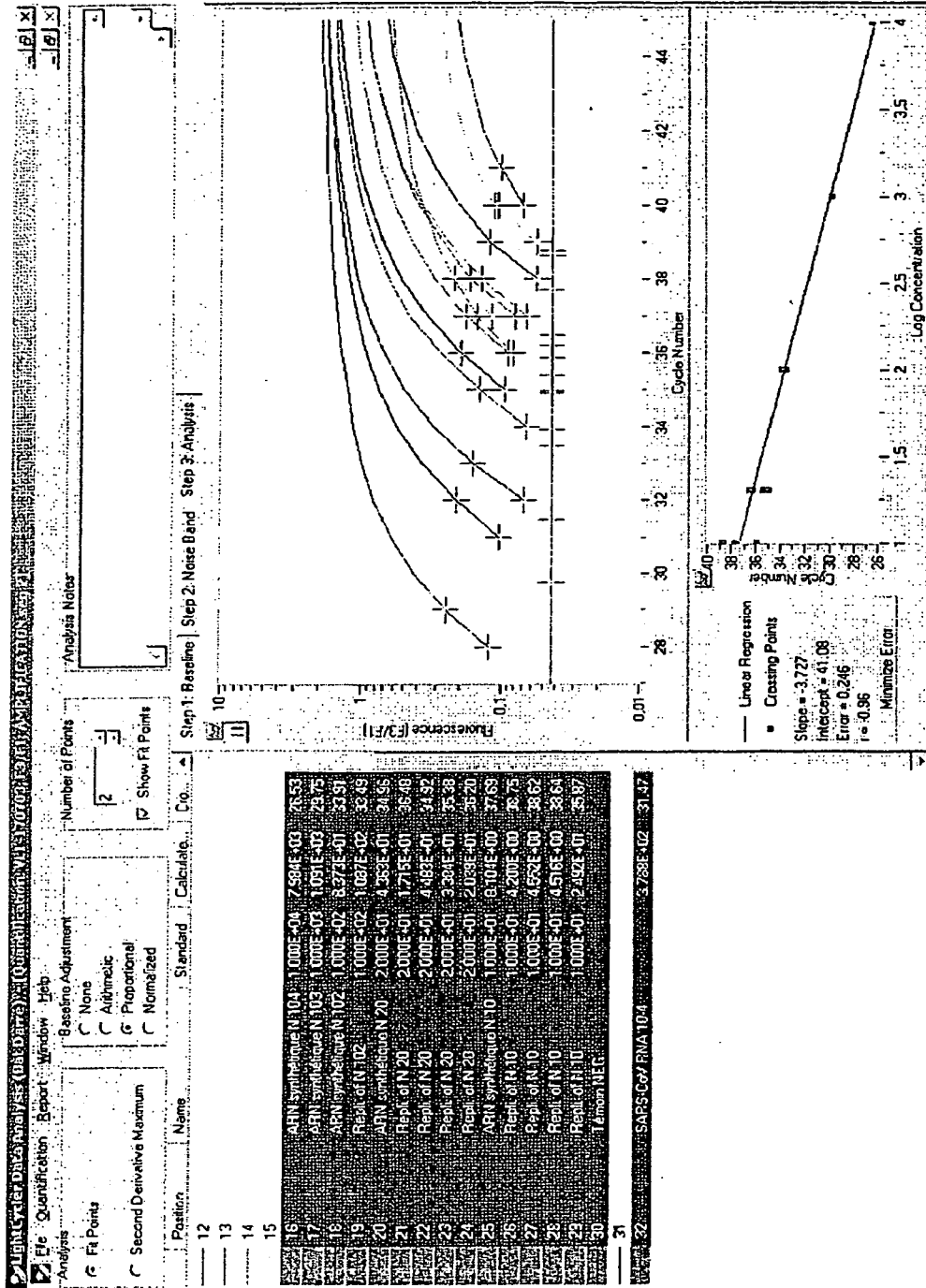


FIGURE 12

```

                >> XhoII
                >> Sau3AI
                >< TthHB8I >> NdeII
                >< TaqI >> MflI
                >< EcoRII >> Sau3AI >> MboI
                >< Ecl136I >> NdeII >> DpnII
                >< DsaV >> MboI>< MnlI>< DpnI
                >< BstOI >> DpnII >> BstYI
                >< BstNI >> DpnI >> BspAI
                >< BsiLI >> BspAI >> Bsp143I
                >< ApyI >> Bsp143I>< BglII
ATATTAGGTT TTTACCTACC CAGGAAAAGC CAACCAACCT CGATCTCTTG TAGATCTGTT CTCTAAACGA
    10          20          30          40          50          60          70

                >> VneI
                >< SphI
                >< SnoI
                >< RmaI
                >< PaeI >> SduI
                >< NspI >> NspII
                >< NspHI >> HgiAI
                >< NlaIII >> Bsp1286I
                >< MaeI >> BmyI
                >< ApaLI
                >< Alw44I
                >< Alw21I
>< Tru9I
>< MseI >> BbvI
>< DraI >> AluI >< Fnu4HI >> Alw21I
ACTTTAAAT CTGTGTAGCT GTCGCTCGGC TGCATGCCTA GTGCACCTAC GCAGTATAAA CAATAATAAA
    80          90          100          110          120          130          140

                >> SfcI
                >< PstI
                >< MnlI
                >< Ksp632I
                >< EarI
                >< Eam1104I
TTTTACTGTC GTTGACAAGA AACGAGTAAC TCGTCCCTCT TCTGCAGACT GCTTACGGTT TCGTCCGTGT
    150          160          170          180          190          200          210

                >< TthHB8I >> StyI
                >< TaqI >> RmaI >> ScrFI
                >< Sau3AI >> MaeI >> NciI
                >< NdeII >> EcoT14I >> MspI
                >< MboI >> Eco130I >> MaeIII
                >< DpnII >> BssT1I >> HpaII
                >< DpnI >> BsaJI >> HapII
                >< BspAI >> BlnI >> DsaV
                >< Bsp143I >> AvrII >> BcnI
TGCAGTCGAT CATCAGCATA CCTAGGTTTC GTCCGGGTGT GACCGAAAGG TAAGATGGAG AGCCTTGTTT
    220          230          240          250          260          270          280

                >> RmaI
                >< Esp3I >> MaeII
                >< HindII >> MaeII >< Eco57I >> BsmAI >> MaeI
                >< HincII >> AflIII >< DdeI >> Alw26I >> BsmBI
TTGGTGCAA CGAGAAAACA CACGTCCAAC TCAGTTTGCC TGTCTTCAG GTTAGAGACG TGCTAGTGCC
    290          300          310          320          330          340          350

```

FIGURE 13.1

```

                >> Sau96I
                >> PssI
                >> Pali
                >> NspIV
                >> MnlI
                >> HaeIII
                >> EcoO109I
                >> DraII>> MboII >> PmlI
                >> MnlI >> Cfr13I >> PmaCI
                >> Ksp632I >> BsuRI > < MaeII
                >> HinfI >> BsiZI>> EcoNI >> Eco72I
                >> EarI >> BshI >> BslI >> BsaAI
                >> PleI >> Eam1104I>> AsuI >> BsiYI>< BbrPI >> MnlI
TGGCTTCGGG GACTCTGTGG AAGAGGCCCT ATCGGAGGCA CGTGAACACC TCAAAAATGG CACTTGTGGT
    360          370          380          390          400          410          420

                >> Tru9I
                >> RsaI >> SfaNI
                >> RmaI >> Csp6I >> BspWI >> MseI
                >> MaeI >< AluI >> AfaI >< AluI >> MaeII
CTAGTAGAGC TGGAAAAGG CGTACTGCCC CAGCTTGAAC AGCCCTATGT GTTCATTAAA CGTTCTGATG
    430          440          450          460          470          480          490

                >> Pali
                >> HaeIII >> RsaI
                >> Tru9I >> GdiII McrI ><
                >> MseI >> EaeI >> Csp6I
                >> Esp4I >> BsuRI >> BsmI BsiEI ><
                >> AflII >> BshI >< AluI >> BscCI >< AfaI
CCTTAAGCAC CAATCACGGC CACAAGGTCG TTGAGCTGGT TGCAGAAATG GACGGCATTG AGTACGGTCC
    500          510          520          530          540          550          560

                >> NspI
                >> ScaI >> NspHI
                >> RsaI >> NlaIII
                > < Csp6I >> BslI >> MboII
                >< BsrI >> BsiYI >> MboII
                >< AciI >> AfaI >< AflIII >> MunI >< AciI
TAGCGGTATA ACACGGGAG TACTCGTCC ACATGTGGGC GAAACCCCAA TTGCATACCG CAATGTCTT
    570          580          590          600          610          620          630

                >> TthHB8I
                >> TaqI
                >> Sau3AI
                >> NdeII
                >> MboI
                >< DpnII
                > < DpnI
                >< ClaI
                >< Bsu15I
                >< BspDI
                >< BspAI
                >< Bsp143I
                >< Bsp106I
                >< BsiXI MaeIII >
                >< Cfr10I >< BscI>< SfaNI DdeI ><
                >< BscBI >< AluI >< BanIII BfrI ><
CTTCGTAAGA ACGGTAATAA GGGAGCCGGT GGTCATAGCT ATGGCATCGA TCTAAAGTCT TATGACTTAG
    640          650          660          670          680          690          700

```

FIGURE 13.2


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>< MvaI      >< Hin6I      >< SduI      >< Csp45I
>< Ecl136I   >< HhaI      >< NspII     >< BstBI
>< BstOI     >< MaeII     >< HgiAI     >< Bsp119I
>< BstNI     >< Eco47III  >< Bsp1286I >< BsiCI
>< BsiLI     >< CfoI      >< BmyI      >< Bpu14I
>< ApyI >< DdeI >< Bsp143II >< AluI   >< Alw21I   >< AsuII
CTGGTTCAC T GAGCGCTCTG ATAAGAGCTA CGAGCACCCAG ACACCCCTTCG AAATTAAGAG TGCCAAGAAA
990          1000          1010          1020          1030          1040          1050

                                >< Tru9I
                                >< BsmI
                                >< MseI
                                >< BscCI
                                >< MnlI
TTTGACACTT TCAAAGGGGA ATGCCCAAAG TTTGTGTTTC CTCTTAACTC AAAAGTCAAA GTCATTCAAC
1060          1070          1080          1090          1100          1110          1120

>< PmlI
>< PmaCI
>< MaeII
>< Eco72I
>< BsaAI
>< BbrPI
>< AflIII
CACGTGTTGA AAAGAAAAAG ACTGAGGGTT TCATGGGGCG TATACGCTCT GTGTACCCTG TTGCATCTCC
1130          1140          1150          1160          1170          1180          1190

>< SfaNI
>< MaeIII
ACAGGAGTGT AACAATATGC ACTTGTCTAC CTTGATGAAA TGTAATCATT GCGATGAAGT TTCATGGCAG
1200          1210          1220          1230          1240          1250          1260

                                >< SinI
                                >< Sau96I
                                PssI ><
                                >< Psp5II
                                >< PpuMI
                                >< NspIV
                                >< NspHII
                                >< Eco47I
                                >< DraII
                                >< Cfr13I
                                >< BsiZI
                                >< Bme18I
                                >< AvaII
                                >< AsuI

>< MaeII
ACGTGCGACT TTCTGAAAGC CACTTGTGAA CATTGTGGCA CTGAAAATTT AGTTATTGAA GGACCTACTA
1270          1280          1290          1300          1310          1320          1330

                                EcoO109I ><AflIII >
                                Van91I ><
                                SinI ><
                                Sau96I ><
                                PflMI ><
                                NspIV ><
                                NspHII >
                                Eco47I ><
                                Cfr13I ><
                                BslI ><
                                BsiZI ><
                                BsiYI ><
                                Bme18I ><
                                AvaII ><
                                AsuI ><

>< RsaI
>< NspI
>< NlaIV
>< NlaIII
>< NspHI>< KpnI
>< Eco64I
>< Csp6I
>< BscBI
>< BanI
>< Asp718
>< AfaI
>< AccBII

```

FIGURE 13. 4

```

    >< Acc65I          >< SfcI          >< NlaIII         AccB7I ><
CATGTGGGTA CCTACCTACT AATGCTGTAG TGAAATGCC ATGTCCTGCC TGTCAAGACC CAGAGATTGG
    1340          1350          1360          1370          1380          1390          1400

                                >< TthHB8I
                                >< TaqI>< MnlI
                                >< HinfI

    >< DdeI          >< PleI          >< AciI
ACCTGAGCAT AGTGTTCAG ATTATCACAA CCACTCAAAC ATTGAAACTC GACTCCGCAA GGGAGGTAGG
    1410          1420          1430          1440          1450          1460          1470

    >< RmaI          NlaIV ><
    >< MnlI          >< BsrI
    >< MaeI          >< BbvI          >< Fnu4HI         BscBI ><
ACTAGATGTT TTGGAGGCTG TGTGTTGGCC TATGTTGGCT GCTATAATAA GCGTGCCTAC TGGGTTCCTC
    1480          1490          1500          1510          1520          1530          1540

                                XhoII ><
                                Sau3AI ><
                                NdeII ><
                                MflI ><
                                MboI ><
                                >< MaeIII
                                >< Eco3II         DpnII ><
                                >< Pali          >< Eco3II
                                >< HaeIII         >< BsrI          >< MnlI DpnI ><
    >< RmaI          >< BsuRI         >< BsrI          >< BsmAI         BstYI ><
    >< MnlI          >< DdeI          >< BspWI         >< BsaI>< HphI     BspAI ><
    >< MaeI          >< BshI>< BglI         >< Alw26I        BspI43I >
GTGCTAGTGC TGATATTGGC TCAGGCCATA CTGGCATTAC TGGTGACAAT GTGGAGACCT TGAATGAGGA
    1550          1560          1570          1580          1590          1600          1610

                                >< Tru9I
                                >< MseI
                                >< MaeII         >< Tru9I
                                >< HpaI
                                >< HindII
                                >< HinfI >< PleI >< HincII
    >< Alwi >< DdeI         >< AflIII        >< MseI
TCTCCTTGAG ATRACTGAGTC GTGAACGTGT TAACATTAAC ATTGTTGGCG ATTTTCATT GAATGAAGAG
    1620          1630          1640          1650          1660          1670          1680

    >< MboII          PleI ><
    >< BstXI          >< SfaNI          >< HinfI
GTTGCCATCA TTTGGCATC TTTCTCTGCT TCTACAAGTG CCTTTATTGA CACTATAAAG AGTCTTGATT
    1690          1700          1710          1720          1730          1740          1750

                                >< StyI
                                >< MaeIII
                                >< EcoT14I
                                >< EcoI30I
                                >< PleI
                                >< MaeIII
                                >< BssTII         BslI ><
    >< HinfI>< AciI         >< BsaJI         BsiYI ><
ACAAGTCTTT CAAAACCATT GTTGAGTCCT GCGGTAAC TAAGTTACC AAGGGAAAGC CCGTAAAAGG
    1760          1770          1780          1790          1800          1810          1820

    >< Sau3AI          >< Van9II
    >< NdeII          >< PflMI
    >< MboI           >< DraIII
    >< DpnII          >< BslI
    >< DpnI >< Tru9I       >< EsiYI
    >< BspAI >< MseI         >< BbvI          >< MnlI
    >< BspI43I       >< AccB7I        Fnu4HI ><

```

FIGURE 135

```

TGCTTGAAC ATTGGACAAC AGAGATCACT TTTAACACCA CTGTGTGGTT TTCCTCACA GGCTGCTGGT
1830      1840      1850      1860      1870      1880      1890

      << ThaI
      << SfaNI
      << MvnI
      << HinPII
    << HinPII
    << Hin6I
    << Hin6I
      << HhaI
    << Sau3AI      << HhaI
    << NdeI        << CfoI
    << MboI        << CfoI
    << DpnII       << BstUI
      << DpnI      << BssHII
    << BspAI       << Bsp50I
      << Bsp143I   << AccII
GTTATCAGAT CAATTTTTCG GCGCACACTT GATGCAGCAA ACCACTCAAT TCCTGATTG CAAAGAGCAG
1900      1910      1920      1930      1940      1950      1960

      << TthHB8I
      << StyI
      << NcoI
      << HindII
      << HincII
      << HinfI
      << EcoT14I
      << Eco57I
      << TaqI>< Ecol30I
    << Sali << DsaI
    << RtrI << BssT1I
      << BsaHI
      << BbiII>< NlaIII
      << AclI << HgaI
    << MaeIII
      << BbvI
      << MaeII << AccI>< BsaJI HphI <<
CTGTCACCAT ACTTGATGGT ATTTCTGAAC AGTCATTACG TCTGTGCGAC GCCATGGTTT ATACTTCAGA
1970      1980      1990      2000      2010      2020      2030

      << RsaI
      << NdeI
      << BspMI
      << MaeIII << BsrI << AfaI << DdeI
CCTGCTCACC AACAGTGTCA TTATTATGGC ATATGTAACT GGTGGTCTTG TACAACAGAC TTCTCAGTGG
2040      2050      2060      2070      2080      2090      2100

      << StuI
      << Pali
      << HaeIII
      << Ecol47I
      << SduI << DdeI
      << NspII << BsuRI
      << Bsp1286I << BshI
      << BmyI << AatI << MnlI DdeI <<
TTGTCTAATC TTTTGGGCAC TACTGTTGAA AACTCAGGC CTATCTTTGA ATGGATTGAG GCGAAACTTA
2110      2120      2130      2140      2150      2160      2170

      << TfiI
      << HinfI
      << SfaNI << BsgI << FokI Tth111I <<
GTGCAGGAGT TGAATTTCTC AAGGATGCTT GGGAGATTCT CAAATTTCTC ATTACAGGTG TTTTGCAT
2180      2190      2200      2210      2220      2230      2240
    << AspI <<

```

FIGURE 13.6

```

Tru9I >>
MseI >>
HpaI >
HindII >
HincII >
>> Eco57I
CGTCAAGGGT CAAATACAGG TTGCTTCAGA TAACATCAAG GATTGTGTAA AATGCTTCAT TGATGTTGTT
2250      2260      2270      2280      2290      2300      2310

>> Sau3AI
>> NdeII
>> MboI
> < MaeIII
>> FbaI
>> DpnII
>> DpnI
>> BspAI
>> Bsp143I
>> TthHB8I
>> TaqI
AACAAGGCAC TCGAAATGTG CATTGATCAA GTCACTATCG CTGGCGCAA GTTGCATCA CTCAACTTAG
2320      2330      2340      2350      2360      2370      2380

>> HinPII
>> Hin6I
>> HhaI
>> CfoI
>> BspAI
>> BbsI
>> PvuII
>> MaeII
>> Bst1107I
>> BsaAI
>> BbvI
>> HphI
>> DrdI
>> AccI
GTGAAGTCTT CATCGCTCAA AGCAAGGGAC TTTACCGTCA GTGTATACGT GGCAAGGAGC AGCTGCAACT
2390      2400      2410      2420      2430      2440      2450

>> Tru9I
>> NlaIV
>> MseI
>> MnlI
>> Esp4I
>> Eco64I
>> BscBI
>> NlaIII >> BanI
>> AflII
>> BbvI
>> AccBII
>> MaeIII
ACTCATGCCT CTTAAGGCAC CAAAAGAAGT AACCTTTCTT GAAGGTGATT CACATGACAC AGTACTTACC
2460      2470      2480      2490      2500      2510      2520

> < XhoI
>> TthHB8I
>> TthHB8I >> TaqI
> < SlaI
> < PaeR7I
> < NspIII
>> HphI >> HinII
> < Eco88I
> < CcrI
>> Esp3I >> BsaHI
> < BcoI
>> BsmAI >> BbiII
> < AvaI >> HgaI
>> TaqI > < Ama87I >> BsmBI
>> DdeI >> MnlI
>> Alw26I >> AcyI
TCTGAGGAGG TTGTTCTCAA GAACGGTGAA CTCGAAGCAC TCGAGACGCC CGTTGATAGC TTCACAAATG
2530      2540      2550      2560      2570      2580      2590

```

FIGURE 13.7

```

                >< Pali >< NlaIII
                >< MaeIII >< MnlI
                >< BsuRI >< DdeI >< Tru9I
                >< BshI >< BfrI >< MseI
    >< AluI          >< BsrI
GAGCTATCGT TGGCACACCA GTCTGTGTAA ATGGCCTCAT GCTCTTAGAG ATTAAGGACA AAGAACAATA
    2600          2610          2620          2630          2640          2650          2660

                >< VneI
                Tru9I ><
                >< SnaI
                >< SduI
                >< NspII
                MseI ><
                >< HgiAI
                Bsp1286I >< BslI ><
                BsiYI ><
                >< BmyI
                >< ApaLI
                >< Tru9I >< Alw44I
                >< MseI >< Alw21I
CTGCGCATTG TCTCCTGGTT TACTGGCTAC AAACAATGTC TTTCGCTTAA AAGGGGGTGC ACCAATTAAA
    2670          2680          2690          2700          2710          2720          2730

                >< TfiI
    >< MaeIII          >< MboII > < MaeIII          >< HinfI AluI ><
GGTGTAACCT TTGGAGAAGA TACTGTTTGG GAAGTCAAG GTTACAAGAA TGTGAGAATC ACATTTGAGC
    2740          2750          2760          2770          2780          2790          2800

                >< RsaI
                >< NlaIV
                MaeIII ><
                >< MspI >< KpnI
                >< HpaII
                >< HapII
                > < Eco64I
                >< SduI >< Csp6I
                >< NspII >< TfiI >< BscBI
                >< HgiAI > < BanI
                >< Bsp1286I > < Asp718
                >< BmyI >< HinfI >< AfaI
                >< Alw21I > < AccB1I
                >< AflIII >< MseI >< AccI > < Acc65I
TTGATGAACG TGTGACAAA GTGCTTAATG AAAAGTGCTC TGTCTACACT GTTGAATCCG GTACCGAAGT
    2810          2820          2830          2840          2850          2860          2870

                >< Sau3AI
                >< NdeII
                >< MboI
                >< DpnII
                > < DpnI
                >< NspI
                >< NspHI >< MboII >< BspAI
                >< NlaIII > < BsrI > < Bsp143I
    >< DdeI          >< MnlI >< AlwNI          >< BbsI >< AlwNI
TACTGAGTTT GCATGTGTTG TAGCAGAGGC TGTTGTGAAG ACTTTACAAC CAGTTTCTGA TCTCCTTACC
    2880          2890          2900          2910          2920          2930          2940

                >< Sau3AI
                >< NdeII
                >< MboI
                >< DpnII
                >< DpnI
                >< BspAI

```

FIGURE 13.8

```

    >> NlaIII>> Bsp143I          >> AluI          >> SfaNI
AACATGGGTA TTGATCTTGA TGAGTGGAGT GTAGCTACAT TCTACTTATT TGATGATGCT GGTGAAGAAA
    2950          2960          2970          2980          2990          3000          3010

                                >> SfaNI
                                >> MnlI
    >> MboII          >> GsuI          >> Ksp632I          >> MnlI
                                >> BsaAI          >> EarI          >> MboII
    >> HphI          >> MaeII>> BpmI          >> MnlI          >> Eam1104I          >> MboII
ACTTTTCATC ACGTATGTAT TGTTCCCTTT ACCCTCCAGA TGAGGAAGAA GAGGACGATG CAGAGTGTGA
    3020          3030          3040          3050          3060          3070          3080

                                >> RsaI
                                >> RsaI
    >> NlaIII
                                >> MnlI          >> FokI
                                >> Csp6I          Eco31I >>
                                >> Csp6I          >> MamI BsmAI >>
    >> MboII          >> MboII          >> AfaI          >> BsiBI BsaI >>
    >> MboII          >> AfaI          >> BsaBIALw26I >>
GGAAGAAGAA ATTGATGAAA CCTGTGAACA TGAGTACGGT ACAGAGGATG ATTATCAAGG TCTCCCTCTG
    3090          3100          3110          3120          3130          3140          3150

    >> NlaIV>> PvuII>> XmnI
    >> Eco64I >> Psp5I          >> TthHB8I
    >> MnlI >> DdeI          >> TaqI          >> MnlI          >> MboII
    >> BscBI>> NspBII >> MnlI          >> Ksp632I          >> MboII >> MboII
    >> BanI          >> MnlI          >> EarI          >> BsrI
    >> AccBII >> AluI >> Asp700I          >> Eam1104I >> MboII>> BbsI
GAATTTGGTG CCTCAGCTGA AACAGTTCGA GTTGAGGAAG AAGAAGAGGA AGACTGGCTG GATGATACTA
    3160          3170          3180          3190          3200          3210          3220

                                >> Tru9I
    >> FokI          >> MseI          >> Eco57I
    >> DdeI          >> BsrI>> MboII BsrI >>
CTGAGCAATC AGAGATTGAG CCAGAACCAG AACCTACACC TGAAGAACCA GTTAATCAGT TTACTGGTTA
    3230          3240          3250          3260          3270          3280          3290

    >> Tru9I          >> MnlI
    >> MseI          >> Tru9I          >> HindII>> Tru9I          >> DraIII
    >> DraI          >> MseI          >> HincII>> MseI          >> BspWI
TTTAAAACTT ACTGACAATG TTGCCATTAA ATGTGTTGAC ATCGTTAAGG AGGCACAAAG TGCTAATCCT
    3300          3310          3320          3330          3340          3350          3360

                                >> VneI
                                >> SnoI
                                >> SduI
                                >> NspII
                                >> HgiAI
                                >> Bsp1286I
                                >> BmyI
                                >> ApaLI
    >> HphI          >> NlaIII          >> Alw44I
    >> BbvI          >> Fnu4HI          >> BspMI          >> Alw21I
ATGGTGATTG TAAATGCTGC TAACATACAC CTGAAACATG GTGGTGGTGT AGCAGGTGCA CTCACAAGG
    3370          3380          3390          3400          3410          3420          3430

                                >> Sau96I
                                >> Pali
                                >> NspIV
                                >> HaeIII
    >> NlaIV          >> Cfr13I

```

FIGURE 13.9

```

    >< Eco64I
    >< BscBI
    >< BanI
    >< AccB1I>< NlaIII
CAACCAATGG TGCCATGCAA AAGGAGAGTG ATGATTACAT TAAGCTAAAT GGCCCTCTTA CAGTAGGAGG
    3440          3450          3460          3470          3480          3490          3500

    >< BsuRI
    >< Tru9I
    >< MseI
    >< AluI
    >< AsuI
    >< MnlI
    >< SinI
    >< Sau96I
    >< NspIV
    >< NspHI>< NspHII
    >< Eco47I
    >< Cfr13I
    >< NlaIII
    >< BspMI
    >< Bsi2I
    >< Bme18I
    >< AvaII MnlI ><
    >< DdeI
    >< NspI>< AsuI FokI ><
GTCTTGTTTG CTTTCTGGAC ATAATCTTGC TAAGAAGTGT CTGCATGTTG TTGGACCTAA CCTAAATGCA
    3510          3520          3530          3540          3550          3560          3570

    >< Tru9I
    >< HphI> < MseI
    >< Esp4I
    >< AluI
    >< NdeI
    >< AflIII>< Fnu4HI
    >< BbvI
GGTGAGGACA TCCAGCTTCT TAAGGCAGCA TATGAAAATT TCAATTCACA GGACATCTTA CTGACCAT
    3580          3590          3600          3610          3620          3630          3640

    RsaI ><
    Csp6I ><
    AfaI ><
    >< Eco57I
    >< BcgI
TGTTGTCAGC AGGCATATTT GGTGCTAAAC CACTTCAGTC TTTACAAGTG TCGTGCAGA CGGTTCGTAC
    3650          3660          3670          3680          3690          3700          3710

    >< BsgI
    >< BcgI/a
    >< BspMI
    >< AluI
    >< NlaIII
ACAGGTTTAT ATTGCAGTCA ATGACAAAGC TCTTTATGAG CAGGTTGTCA TGGATTATCT TGATAACCTG
    3720          3730          3740          3750          3760          3770          3780

    >< RmaI
    >< MaeI
    >< MnlI
    >< Eco57I
    >< NlaIV
    >< BscBI
    >< TfiI
    >< MboII
    >< HinFI
    >< DdeI
AAGCCTAGAG TGGAAGCACC TAAACAAGAG GAGCCACCAA ACACAGAAGA TTCCAAAACCT GAGGAGAAT
    3790          3800          3810          3820          3830          3840          3850

    >< Tru9I
    >< StuI
    >< Pali
    >< MseI
    >< MnlI
    >< MaeIII
    >< HaeIII
    >< Eco065I
    >< Ecol47I
    >< Eco9I
    >< BsuRI
    >< BstXI ><
    >< BshI
    >< BstPI
    >< AatI
    >< BstEII
CTGTCGTACA GAAGCCTGTC GATGTGAAGC CAAAAATTAA GGCCTGCATT GATGAGGTTA CCACAACACT
    3860          3870          3880          3890          3900          3910          3920

    TfiI ><
    NlaIII ><
    HinFI ><
    >< DdeI
    >< EcoRV
    >< HindIII

```

FIGURE 13.10


```

>> BsrI      >> MboII      >> MaeIII      >> Eco32I      >> AluI
GGAAGAACT AAGTTTCTTA CCAATAAGTT ACTCTTGTTT GCTGATATCA ATGGTAAGCT TTACCATGAT
3930      3940      3950      3960      3970      3980      3990

      >> NspI
      >> NspHI
      >> NlaIII
>> MnlI      >> SfaNI
      >> DdeI      >> MboII >> BslI      >> NlaIII
      >> BfrI      >> HphI      >> BsiYI      >> FokI
TCTCAGAACA TGCTTAGAGG TGAAGATATG TCTTTCCTTG AGAAGGATGC ACCTTACATG GTAGGTGATG
4000      4010      4020      4030      4040      4050      4060

>> SpeI
>> RmaI
>> MaeI      >> EcoRV>> HphI      >> SfaNI
>> HphI      >> Eco32I      >> MnlI      >> DdeI
TTATCACTAG TGGTGATATC ACTTGTGTTG TAATACCCTC CAAAAAGGCT GGTGGCACTA CTGAGATGCT
4070      4080      4090      4100      4110      4120      4130

      >> ScrFI
      >> RsaI
      >> MvaI
      >> EcoRII
      >> Ecl136I
      >> DsaV
      >> Csp6I >> EcoNI
      >> BstOI
      >> BstNI
      >> BsiLI
      >> BsaJI
      >> BsaAI >> BslI
      >> MaeII>> ApyI
      >> AfaI >> BsiYI
>> AluI      >> BsrI      >> MboII      >> MaeII>> ApyI
CTCAAGAGCT TTGAAGAAAG TGCCAGTTGA TGAGTATATA ACCACGTACC CTGGACAAGG ATGTGCTGGT
4140      4150      4160      4170      4180      4190      4200

      >> Tru9I
      >> MseI
      >> DdeI      >> Esp4I      >> RsaI
>> MnlI      >> BspWI      >> Csp6I
>> FokI      >> AluI      >> AflIII      >> Eco57I >> AfaI
TATACACTTG AGGAAGCTAA GACTGCTCTT AAGAAATGCA AATCTGCATT TTATGTACTA CCTTCAGAAG
4210      4220      4230      4240      4250      4260      4270

      >> ScrFI
      >> MvaI
      >> EcoRII
      >> XmnI      >> Ecl136I      NlaIII >>
      >> Ksp632I >> RmaI      >> DsaV      Ksp632I >>
      >> EarI >> TfiI>> MboII      >> BstOI      >> EarI
      >> Eam1104I >> MaeI      >> BstNI      Eam1104I >>
      >> DdeI >> HinfI      >> BsiLI      BsmAI >>
      >> BspWI      >> Asp700I      >> ApyI      Alw26I >>
CACCTAATGC TAAGGAAGAG ATTCTAGGAA CTGTATCCTG GAATTTGAGA GAAATGCTTG CTCATGCTGA
4280      4290      4300      4310      4320      4330      4340

      >> VspI      >> Zsp2I
      >> Tru9I      >> Ppu10I
      >> MseI      >> NsiI
      >> MboII      >> NlaIII      >> FokI
      >> Eco57I >> Mph1103I >> FokI

```

FIGURE 13.11

```

                >< AsnI           >< EcoT22I          >< BspWI
                >< AseI           >< AvaIII           >< BglI           >< MaeII
AGAGACAAGA AAATTAATGC CTATATGCAT GGATGTTAGA GCCATAATGG CAACCATCCA ACGTAAGTAT
 4350         4360         4370         4380         4390         4400         4410

                >< SfaNI
                >< Tru9I           > < HindII           >< TfiI           >< SpeI
                >< MseI           > < HincII>< MboII           >< RmaI
                >< MnlI           >> < DrdI >< HinfI           >< MaeI
AAAGGAATTA AAATTCAAGA GGCATCGTT GACTATGGTG TCCGATTCTT CTTTATACT AGTAAAGAGC
 4420         4430         4440         4450         4460         4470         4480

                >< MaeIII
                >< SfcI           >> < Fnu4HI           >< MunI
                >> < AluI           >> < AluI           >> < AciI           >> < MaeIII >>
CTGTAGCTTC TATTATTACG AAGCTGAATC CTCTAAATGA GCCGCTTGTC ACAATGCCAA TTGGTTATGT
 4490         4500         4510         4520         4530         4540         4550

                >< ThaI
                >< MvnI
                >> < MboII
                >> < HinPII
                >> < HinPII
                >> < Hin6I
                >> < Hin6I
                >> < HhaI
                >> < HhaI
                >> < Tru9I           >> < Fnu4HI
                >> < NlaIII           >> < MseI           >> < CfoI
                >> < MnlI           >> < CfoI
                >> < Ksp632I           >> < BstUI
                >> < EarI           >> < BssHII>< BspWI           >> < Tru9I
                >> < Eam1104I           >> < Bsp50I           >> < MseI
                >> < BbvI           >> < AccII           >> < AluI           >> < HphI >>
GACACATGGT TTTAATCTTG AAGAGGCTGC GCGCTGTATG CGTTCTCTTA AAGCTCCTGC CGTAGTGTC
 4560         4570         4580         4590         4600         4610         4620

                >< MaeIII
                >< SfaNI           >> < AlwNI           >> < MnlI >> < MnlI>< DdeI
GTATCATCAC CAGATGCTGT TACTACATAT AATGGATACC TCACCTCGTC ATCAAAGACA TCTGAGGAGC
 4630         4640         4650         4660         4670         4680         4690

                >< SinI
                >< Sau96I
                >< NspIV
                >> < NspHII
                >> < Eco47I
                >> < NspII
                >> < Cfr13I
                >> < HgiAI
                >> < BsiZI
                >> < Bsp1286I           >> < Bme18I           >> < RsaI
                >> < BmyI           >> < AvaII           >> < Csp6I
                >> < Alw21I           >> < AsuI           >> < AfaI
ACTTTGTAGA AACAGTTTCT TTGGCTGGCT CTTACAGAGA TTGGTCCTAT TCAGGACAGC CTACAGAGTT
 4700         4710         4720         4730         4740         4750         4760

                > < TthHB8I
                > < TaqI
                >> < SduI
                >> < Van9I           >> < NspII
                >> < Tru9I           >> < RsaI           >> < PflMI           >> < Eco24I
                >> < MseI           >> < HphI           >> < BslI           >> < Bsp1286I
                >> < Esp4I           >> < Csp6I           >> < BsiYI           >> < BmyI GsuI >>

```

FIGURE 13.12

```

    >> AflIII >> MaeIII >> AfaI >> AccB7I >> BanIIBpmI >>
AGGTGTTGAA TTTCTTAAGC GTGGTGACAA AATTGTGTAC CACACTCTGG AGAGCCCCGT CGAGTTTCAT
    4770 4780 4790 4800 4810 4820 4830

    >> Tru9I
    >> P1eI >> EcoNI
    >> MnlI >> BslI
    >> BsmAI >> BsiYI
    >> MnlI >> HphI >> HinfI >> Alw26I >> AciI >> MseI
CTTGACGGTG AGGTTCTTTC ACTTGACAAA CTAAGAGATC TCTTATCCCT GCGGGAGGTT AAGACTATAA
    4840 4850 4860 4870 4880 4890 4900

    >> AluI >> NdeI
AAGTGTTTAC AACTGTGGAC AACACTAATC TCCACACACA GCTTGTGGAT ATGTCTATGA CATATGGACA
    4910 4920 4930 4940 4950 4960 4970

    >> SinI
    >> Sau96I
    >> NspIV
    >> NspHII
    >> Eco47I
    >> Cfr13I
    >> BsiZI
    >> Bme18I
    >> AvaII
    >> AsuI
    >> MaeIII >> Tru9I
    >> FokI >> MseI
    >> BspHI
GCAGTTTGGT CCAACATACT TGGATGGTGC TGATGTTACA AAAATTA AAC CTCATGTAAA TCATGAGGGT
    4980 4990 5000 5010 5020 5030 5040

    >> TthHB8I
    >> RsaI >> TaqI
    >> RmaI >> SnaBI >> ScaI
    >> MaeI >> MaeII >> HindIII >> RsaI
    >> Csp6I >> Eco105I >> Csp6I
    >> AfaI >> BsaAI >> AluI >> AfaI
AAGACTTTCT TTGTACTACC TAGTGATGAC ACACTACGTA GTGAAGCTTT CGAGTACTAC CATACTCTTG
    5050 5060 5070 5080 5090 5100 5110

    >> RsaI
    >> NspI
    >> NspHI
    >> NlaIII
    >> Csp6I >> Tru9I
    >> AflIII >> MseI
    >> AfaI >> DraI
    MnlI >
    BslI >>
    BsiYI >>
ATGAGAGTTT TCTTGTTAGG TACATGTCTG CTTTAAACCA CACAAAGAAA TGGAAATTC CTCAAGTTGG
    5120 5130 5140 5150 5160 5170 5180

    >> Tru9I >> Tru9I >> RmaI
    >> MseI >> MseI >> MunI >> MaeI
    AluI >
TGGTTTAACT TCAATTA AAT GGGCTGATAA CAATTGTTAT TTGTCTAGTG TTTTATTAGC ACTTCAACAG
    5190 5200 5210 5220 5230 5240 5250

    >> SfaNI
    >> SduI
    >> NspII
    >> Eco24I
    >> Bsp1286I
    >> BmyI >> HphI >
    >> BbvI >> Fnu4HI >>
    >> BanII >> BspWI
    >> MnlI

```

FIGURE 13.13

```

CTTGAAGTCA AATTCAATGC ACCAGCACTT CAAGAGGCTT ATTATAGAGC CCGTGCTGGT GATGCTGCTA
  5260      5270      5280      5290      5300      5310      5320
    >> VneI
    >> SnoI
      >> SduI
      >> NspII
      >> HgiAI
      >> Bsp1286I
      >> BmyI
    >> ApaLI
    >> Alw44I
      >> Alw21I
ACTTTTGTGC ACTCATACTC GCTTACAGTA ATAAACTGT TGGCGAGCTT GGTGATGTCA GAGAACTAT
  5330      5340      5350      5360      5370      5380      5390
      >> AluI
      >> HphI
      >< SphI
      >< PaeI
      >< NspI
      >< NspHI >< TfiI
      >< SfcI >< NlaIII >< HinfI
      >< Tru9I
      >< MseI
GACCCATCTT CTACAGCATG CTAATTGGA ATCTGCAAAG CGAGTTCCTTA ATGTGGTGTG TAAACATTGT
  5400      5410      5420      5430      5440      5450      5460
      >< Tru9I
      >< MseI
      >< AluI
      >< RsaI
      >< Csp6I
      >< AfaI
      >< Esp4I >
      >< AflIII >
GGTCAGAAAA CTACTACCTT AACGGGTGTA GAAGCTGTGA TGTATATGGG TACTCTATCT TATGATAATC
  5470      5480      5490      5500      5510      5520      5530
      >< RsaI
      >< MboII
      >< RmaIHinfI ><
      >< Csp6I
    >< Tru9I
    >< MseI
      >< SfaNI
      >< NlaIII
      >< MaeI >< BbsI
      >< AfaI
TTAAGACAGG TGTTCATT CCATGTGTGT GTGGTCGTGA TGCTACACAA TATCTAGTAC AACAGAGTC
  5540      5550      5560      5570      5580      5590      5600
      >< RsaI
      >< DdeI
      >< Csp6I
    >< BspI
      >< BspWI >< BspMI
      >< AfaI
TTCTTTTGTG ATGATGTCTG CACCACCTGC TGAGTATAAA TTACAGCAAG GTACATTCTT ATGTGCGAAT
  5610      5620      5630      5640      5650      5660      5670
      >< RsaI
      >< DdeI
      >< BsmAI
    >< Csp6I
      >< BsaI
      >< Alw26I
      >< MnlI ><
      >< AfaI >< BsrI
      >< HphI >
GAGTACACTG GTAACATCA GTGTGGTCAT TACACTCATA TAACTGCTAA GGAGACCCTC TATCGTATTG
  5680      5690      5700      5710      5720      5730      5740
      >< Eco31I
      >< DdeI
      >< BsmAI
      >< BsaI
      >< Alw26I
      >< MnlI ><
      >< HphI >
      >< SstI
      >< SduI
      >< SacI
      >< NspII
      >< HgiAI
      >< Eco24I
    >< Ecl136II
      >< Bsp1286I
      >< BmyI
      >< SinI
      >< Sau96I
      >< NspIV
      >< NspHII
      >< RsaI
      >< MaeIII
      >< Eco47I
      >< Cfr13I
      >< BsiZI
      >< Bme18I

```

FIGURE 13. 14

```

    >> BanII
    >> Alw21I
    >> AluI
    ACGGAGCTCA CCTTACAAAG ATGTCAGAGT ACAAAGGACC AGTGACTGAT GTTTTCTACA AGGAAACATC
    5750      5760      5770      5780      5790      5800      5810

    >> AvaII
    >> Csp6I>> AsuI
    > < AfaI >> BsrI>< AlwNI
    >> TthHB8I
    >> TaqI >> MaeIII
    TTACTACTACA ACCATCAAGC CTGTGTCGTA TAAACTCGAT GGAGTTACTT ACACAGAGAT TGAACCAAAA
    5820      5830      5840      5850      5860      5870      5880

    >> RsaI
    >> Csp6I
    >> SfcI >> BbvI
    >> FokI
    >> Fnu4HI
    >> AfaI
    TTGGATGGGT ATTATAAAAA GGATAATGCT TACTATACAG AGCAGCCTAT AGACCTTGTA CCAACTCAAC
    5890      5900      5910      5920      5930      5940      5950

    >> Tru9I >>
    >> SmaI >>
    >> MseI >>
    > < NspI
    >> MamI >>
    > < NspHI
    >> DraI >>
    > < NlaIII
    >> BsiBI >>
    >> AflIII
    >> BsaBI >>
    CATTACCAAA TGCGAGTTT GATAATTCA AACTCACATG TTCTAACACA AAATTTGCTG ATGATTTAAA
    5960      5970      5980      5990      6000      6010      6020

    >> MboII
    >> AluI
    >> AluI>> MaeIII
    TCAAATGACA GGCTTCACAA AGCCAGCTTC ACGAGAGCTA TCTGTCACAT TCTTCCCAGA CTTGAATGGC
    6030      6040      6050      6060      6070      6080      6090

    >> SfcI
    GATGTAGTGG CTATTGACTA TAGACTAT TCAGCGAGTT TCAAGAAAGG TGCTAAATTA CTGCATAAGC
    6100      6110      6120      6130      6140      6150      6160

    >> Tru9I
    >> ScrFI
    >> MvaI
    >> MseI
    >> EcoRII
    >> Ecl136I
    >> DsaV
    >> BstOI
    >> BstNI
    >> BsiLI
    >> MaeII >>
    >> DraIII
    >> MunI
    >> BstXI
    >> ApyI
    >> MaeII
    >> BstXI
    CAATTGTTG GCACATTAAC CAGGCTACAA CCAAGACAAC GTTCAAACCA AACACTTGGT GTTTACGTTG
    6170      6180      6190      6200      6210      6220      6230

    > < RsaI
    >> Csp6I
    > < AfaI>< BsrI
    >> MboII >>
    >> BbsI
    TCTTTGGAGT ACAAAGCCAG TAGATACTTC AAATTCATT GAAGTTCTGG CAGTAGAAGA CACACAAGGA
    6240      6250      6260      6270      6280      6290      6300

    >> HindII
    >> HincII
    >> MnlI
    >> MboII
    >> Eco57I
    ATGGACAATC TTGCTTGTA AAGTCAACAA CCCACCTCTG AAGAAGTAGT GGAAAATCCT ACCATACAGA
    6310      6320      6330      6340      6350      6360      6370

```

FIGURE 13.15

```

                << MaeIII
                << MaeII
AGGAAGTCAT AGAGTGTGAC GTGAAAAC TA CCGAAGTTGT AGGCAATGTC ATACTTAAAC CATCAGATGA
    6380          6390          6400          6410          6420          6430          6440

                << XhoII
                << Sau3AI
                << NlaIII
                << NdeII
                << MflI
                << MboI
                << DpnII
                << DpnI
                << BstYI
                << BspAI
    << Tru9I
    << MseI
    << BspHI << Bsp143I << Fnu4HI
    << MaeIII << MnlI << BbvI << AlwI
AGGTGTTAAA GTAACACAAG AGTTAGGTCA TGAGGATCTT ATGGCTGCTT ATGTGGAAAA CACAAGCATT
    6450          6460          6470          6480          6490          6500          6510

                << SauI
                << RmaI
                << MstII
    << MaeI
                << Eco81I
                << DdeI
                << CvnI
                << Bsu36I
                << Bse21I
                << BfrI << Tru9I
    << Tru9I << AxyI << MseI << MunI << NlaIII
    << MseI << AluJ << AocI << DraI << BbvI << Fnu4HI <<
ACCATTAAGA AACCTAATGA GCTTTCAC TA GCCTTAGGTT TAAAAACAAT TGCCACTCAT GGTATTGCTG
    6520          6530          6540          6550          6560          6570          6580

    << VspI << StyI
    << Tru9I << EcoT14I > < DdeI
    << MseI << Eco130I >> BslI
    << AsnI << BssTII >> BsiYI
    << AseI << BsaJI > < BfrI >> Fnu4HI
CAATTAATAG TGTTCCTGG AGTAAAATTT TGGCTTATGT CAAACCATTC TTAGGACAAG CAGCAATTAC
    6590          6600          6610          6620          6630          6640          6650

                << HinPII
                << Hin6I
                << HhaI << Tru9I
                << DdeI << MaeII << MseI
                << DraIII
    << BbvI << CfoI << AflIII
AACATCAAAT TGCCTAAGA GATTAGCACA ACGTGTGTTT AACAATTATA TGCCTTATGT GTTTACATTA
    6660          6670          6680          6690          6700          6710          6720

                << RsaI > < RsaI << XbaI
                << Csp6I << Csp6I << RmaI
    << MunI << AfaI > < AfaI << MaeI << AluI
TTGTTCCAAT TGTGTACTTT TACTAAAAGT ACCAATTCTA GAATTAGAGC TTCACTACCT ACAACTATTG
    6730          6740          6750          6760          6770          6780          6790

                << VspI
                << Tru9I
                << NaeI
                << MspI
                << MseI
    
```

FIGURE 13. 16

```

                >> HpaII
                >> HapII
                >> Cfr10I >> FokI
                >> Tru9I
                >> MseI
                >> SfaNI
                >> AsnI
                >> AseI>> HphI>> MaeIII
CTAAAAATAG TGTTAAGAGT GTTGCTAAAT TATGTTTGGG TGCCGGCATT AATTATGTGA AGTCACCCAA
6800          6810          6820          6830          6840          6850          6860

                >> Tru9I >> DdeI MaeIII >
                >> MseI >> BfrI >> BbvI
ATTTTCTAAA TTGTTACAAA TCGCTATGTG GCTATTGTG TTAAGTATTT GCTTAGGTTC TCTAATCTGT
6870          6880          6890          6900          6910          6920          6930

                >> SduI
                >> NspII
                >> HgiAI
                >> Bsp1286I
                >> BmyI
                >> Alw21I
                > < RsaI
                >> Csp6I
                >> Fnu4HI > < AfaI
GTAAGTCTG CTTTGGTGT ACTCTTATCT AATTTTGGTG CTCCTTCTTA TTGTAATGGC GTTAGAGAAT
6940          6950          6960          6970          6980          6990          7000

                Tru9I >>
                MseI >>
                >> Fnu4HI
                >> Tru9I > < MaeIII
                >> MseI >> MaeII
                >> Fnu4HI BbvI >
TGTATCTTAA TTCGCTAAC GTTACTACTA TGGATTCTG TGAAGTTCT TTTCTTGCA GCATTTGTTT
7010          7020          7030          7040          7050          7060          7070

                > < TfiI
                >> MamI
                >> Hinfi
                >> BsiBI
                >> BsaBI >> AluI
                >> XmnI>> MaeIII
                >> Asp700I
                >> PleI>> Hinfi
                >> AciI>> BsiYI
                >> RsaI >>
                >> HphI
                >> Csp6I >>
                >> AluI >
AAGTGGATTA GACTCCCTG ATTCTTATCC AGCTCTTGAA ACCATTCAGG TGACGATTTC ATCGTACAAG
7080          7090          7100          7110          7120          7130          7140

                >> Pali
                >> NspBII
                >> HaeIII
                >> GdiII
                >> Fnu4HI
                >> EaeI
                >> DdeI
                >> BsuRI
                >> RmaI
                >> BshI >> BslI
                >> MaeI
                >> AciI>> BsiYI
CTAGACTTGA CAATTTTAGG TCTGGCCGCT GAGTGGGTTT TGGCATATAT GTTGTTCACA AAATTCTTTT
7150          7160          7170          7180          7190          7200          7210

                >> BspMI
                >> AluI
                >> RmaI
                >> MaeI
ATTTATTAGG TCTTTCAGCT ATAATGCAGG TGTTCTTTGG CTATTTTGCT AGTCATTTCA TCAGCAATTC
7220          7230          7240          7250          7260          7270          7280

                RsaI >>
                >> MboII
                >> NlaIV
                >> Eco64I
                > < RsaI >> BscBI
                >> Csp6I >> BanI
                > < NlaIII
                > < AfaI>> AccBI
                >> RmaI >>
                >> BsiBI >>
                >> BsaBI >>
                >> AfaI >>

```

FIGURE 13.17


```

                >> FokI
                >> BsmAI
                >> MnlI                >> Alw26I                >> AciI
CCTCTACTTT GACAAGGCTG GTCAAAAGAC CTATGAGAGA CATCCGCTCT CCCATTTTGT CAATTTAGAC
    7710          7720          7730          7740          7750          7760          7770

                >> VspI
                >> Tru9I
                >> MseI
                >> AsnI
                >> AseI
                >< AluI                >> BcgI/a
AATTTGAGAG CTAACAACAC TAAAGGTTCA CTGCCTATTA ATGTCATAGT TTTTGATGGC AAGTCCAAAT
    7780          7790          7800          7810          7820          7830          7840

                >> SfcI                >> PvuII
                >> RsaI                >> Psp5I
                >> PleI                >> Csp6I                >> NspBII
                >> HinfI                >> DdeI                >> BcgI                >> AfaI                >> AluI
GCGACGAGTC TGCTTCTAAG TCTGCTTCTG TGTACTACAG TCAGCTGATG TGCCAACCTA TTCTGTTGCT
    7850          7860          7870          7880          7890          7900          7910

                TthHB8I ><
                TaqI ><
                SalI ><
                RtrI ><
                >< ScaI                HindII >
                >< RsaI                >< Tru9I                HincII >
                >< Csp6I                >< SfaNI >< Eco57I
                >< AluI                >< AfaI                >< MseI                AccI ><
TGACCAAGCT CTTGTATCAG ACGTTGGAGA TAGTACTGAA GTTTCGGTTA AGATGTTTGA TGCTTATGTC
    7920          7930          7940          7950          7960          7970          7980

                >< Tru9I
                >< MseI
                >< Esp4I                >> SfcI
                >< AflIII                >> BspWI >> AiuI
GACACCTTTT CAGCAACTTT TAGTGTTCTT ATGGAAAAAC TTAAGGCACT TGTGCTACA GCTCACAGCG
    7990          8000          8010          8020          8030          8040          8050

                >> PvuII
                >> Psp5I
                >> NspBII
                >> Fnu4HI
                >> AluI
                >> AluI                >> BbvI
AGTTAGCAAA GGGTGTAGCT TTAGATGGTG TCCTTTCTAC ATTCGTGTCA GCTGCCCGAC AAGGTGTTGT
    8060          8070          8080          8090          8100          8110          8120

                >> HindII
                >> HincII
                >> BsmAI                >> DdeI
                >> FokI >< Alw26I                >> BfrI
TGATACCGAT GTTGACACAA AGGATGTTAT TGAATGTCTC AAACTTTCAC ATCACTCTGA CTTAGAAGTG
    8130          8140          8150          8160          8170          8180          8190

                >> XhoII
                Sau3AI ><
                >> NdeII
                >> MflI
                >> MboI
                >> NlaIII >> HgaI
                >> HinfI >> DpnII
                DpnI ><
    
```

FIGURE 13.19

```

                                Bsp143I >>
                                >> BsaHI >> BstYI
                                >> BbiII >> BspAI
                                >> AcyI >> BglII
    >> MaeIII >> HphI >> NlaIII
ACAGGTGACA GTTGTAAACA TTTCATGCTC ACCTATAATA AGGTTGAAAA CATGACGCCC AGAGATCTTG
    8200      8210      8220      8230      8240      8250      8260

    >> NspI
    >> NspHI
    >> NlaIII
>> HinPII
>> Hin6I
    >> HhaI
    >> CfoI
                                >> BspWI >> MaeIII
GCGCATGTAT TGACTGTAAT GCAAGGCATA TCAATGCCCA AGTAGCAAAA AGTCACAATG TTCACTCAT
    8270      8280      8290      8300      8310      8320      8330

                                >> NspI
                                >> NspHI >> PvuII
                                >> NlaIII >> Psp5I
                                >> Eam1105I >> NspBII
                                >> BbvI >> Fnu4HI
                                >> AflIII >> AluI >> BbvI >> Fnu4HI
CTGGAATGTA AAAGACTACA TGTCTTTATC TGAACAGCTG CGTAAACAAA TTCGTAGTGC TGCCAAGAAG
    8340      8350      8360      8370      8380      8390      8400

                                >> RmaI
                                >> MaeI >> Eam1105I
AACAACATAC CTTTTAGACT AACTTGTGCT ACAACTAGAC AGGTTGTCAA TGTCATAACT ACTAAAATCT
    8410      8420      8430      8440      8450      8460      8470

                                >> Tru9I
                                >> Pali
                                >> MseI
                                >> HaeIII
                                >> ScaI >> Esp4I
                                >> RsaI >> Tru9I >> BsuRI
                                >> Csp6I >> MseI >> BshI
                                >> AfaI >> DraI >> AflII >> BbvI
CACTCAAGGG TGTAAGATT GTTAGTACTT GTTTAAACT TATGCTTAAG GCCACATTAT TGTGCGTTCT
    8480      8490      8500      8510      8520      8530      8540

                                >> RsaI
                                >> Csp6I
                                >> BsrI >> NlaIII
                                >> AfaI >> MaeIII
>> Fnu4HI
TGCTGCATTG GTTTGTTATA TCGTTATGCC AGTACATACA TTGTCAATCC ATGATGGTTA CACAAATGAA
    8550      8560      8570      8580      8590      8600      8610

                                >> MaeIII
                                >> MaeIII
                                >> FokI
ATCATTGGTT ACAAAGCCAT TCAGGATGGT GTCACCTCGT ACATCATTTT TACTGATGAT TGTTTTGCAA
    8620      8630      8640      8650      8660      8670      8680

                                SfcI >
                                >> NspI
                                >> NspHI >> Fnu4HI >>
                                >> NlaIII >> BbvI >>
                                >> HgaI >> BstXI >> BbvI >> AluI
ATAAACATGC TGGTTTTGAC GCATGGTTTA GCCAGCGTGG TGGTTCATAC AAAAAATGACA AAAGCTGCC
    8690      8700      8710      8720      8730      8740      8750

```

FIGURE 13. 20

```

                >< ScrFI
                >< ScrFI >< RsaI
                >< MvaI >< MspI
                >< EcoRII >< HpaII
                >< Ecl136I>< NciI
                >< DsaV >< HapII
                >< BstOI>< DsaV
                >< BstNI >< Csp6I
                >< BsiLI >< BcnIDdeI ><
                >< ApyI >< AfaI
                >< Fnu4HI
                >< AluI
TGTAGTAGCT GCTATCATT CAAGAGAGAT TGGTTTCATA GTGCCTGGCT TACCCGGTAC TGTGCTGAGA
    8760      8770      8780      8790      8800      8810      8820

                > < MaeIII >< HphI >< MnlI >< BspWI
GCAATCAATG GTGACTTCTT GCATTTTCTA CCTCGTGTTT TTAGTGCTGT TGGCAACATT TGCTACACAC
    8830      8840      8850      8860      8870      8880      8890

                Tru9I >
                SfaNI ><
                >< RsaI
                MseI >
                >< BspWI >< Fnu4HI >< Csp6I
                >< BbvI>< MnlI >< DdeI >< AfaI
CTTCCAAACT CATTGAGTAT AGTGATTTG CTACCTCTGC TTGCGTTCTT GCTGCTGAGT GTACAATTTT
    8900      8910      8920      8930      8940      8950      8960

                > < RmaI
                >< MnlI
                >< FokI
                > < MaeI
TAAGGATGCT ATGGGCAAAC CTGTGCCATA TTGTTATGAC ACTAATTGC TAGAGGGTTC TATTTCTTAT
    8970      8980      8990      9000      9010      9020      9030

                ScrFI >
                MvaI >
                MnlI ><
                EcoRII ><
                Ecl136I >
                DsaV ><
                BstOI >
                BstNI >
                >< NlaIV >< FokI >< BsiLI >
                >< BscBI >< ApyI >
                >< AluI
AGTGAGCTTC GTCCAGACAC TCGTTATGTG CTTATGGATG GTTCCATCAT ACAGTTTCCT AACACTTACC
    9040      9050      9060      9070      9080      9090      9100

                >< RsaI
                >< SfcI >< NspI
                >< ScaI >< NspHI
                >< RsaI >< NlaIII
                >< Csp6I >< NlaIII
                >< AfaI >< Csp6I
                >< BpmI >< DdeI >< AccI >< AfaI
TGGAGGGTTC TGTTAGAGTA GTAACAAC TTGATGCTGA G TACTGTAGA CATGGTACAT GCGAAAGGTC
    9110      9120      9130      9140      9150      9160      9170

                >< SstI
                >< SduI
                >< SacI
                NspII ><
                HgiAI ><
                Eco24I ><
                Bsp1286I ><

```

FIGURE 13.21

```

Ecl136II >>> BmyI
BanII ><
>< Tru9I
Alw21I ><
>< BsrI
>< MseI
>< AluI
AGAAGTAGGT ATTTGCCTAT CTACCAGTGG TAGATGGGTT CTTAATAATG AGCATTACAG AGCTCTATCA
9180 9190 9200 9210 9220 9230 9240

>< TfiI
>< SfaNI
>< HinfI >< AluI
>< MnlI
GGAGTTTTCT GTGGTGTGA TGGCATGAAT CTCATAGCTA ACATCTTTAC TCCTCTTG TG CAACCTGTGG
9250 9260 9270 9280 9290 9300 9310

>< MaeIII
HphI ><
>< Eco57I
>< BbvI Fnu4HI ><
GTGCTTTAGA TGTGTCTGCT TCACTAGTGG CTGGTGGTAT TATTGCCATA TTGGTGACTT GTGCTGCCTA
9320 9330 9340 9350 9360 9370 9380

>< RsaI
>< Csp6I >< NlaIII
>< MaeII
>< BbvI
>< Fnu4HI
>< AflIII
>< AfaI >< HphI
>< BspWI
CTACTTTATG AAATTCAGAC GTGTTTTTGG TGAGTACAAC CATGTTGTTG CTGCTAATGC ACTTTGTTT
9390 9400 9410 9420 9430 9440 9450

>< RsaI
>< NlaIV
>< KpnI
>< Eco64I
>< Csp6I
>< BscBI
>< Asp718
>< BanI >< AluI
>< AfaI
>< AccBI
>< Acc65I
>< AluI >< DsaV >< AccI
>< ScrFI
>< NciI
>< MspI
>< HpaII
>< HinfI
>< HapII >< PleI
>< BcnI >< DdeI
TTGATGTCTT TCACTATACT CTGTCTGTA CCAGCTTACA GCTTTCTGCC GGGAGTCTAC TCAGTCTTTT
9460 9470 9480 9490 9500 9510 9520

>< RsaI
>< Csp6I
>< AfaI >< HphI
>< HphI
NlaIII ><
ACTTGTA CT GACATTCTAT TTCACCAATG ATGTTTCATT CTTGGCTCAC CTTCAATGGT TTGCCATGTT
9530 9540 9550 9560 9570 9580 9590

TTCTCCTATT GTGCCTTTTT GGATAACAGC AATCTATGTA TTCTGTATTT CTCTGAAGCA CTGCCATTGG
9600 9610 9620 9630 9640 9650 9660

>< TthHB8I
>< RsaI
>< MnlI
>< MnlI
>< Csp6I
>< Tru9I
>< PleI
>< BcgI/a >< TaqI
>< MseI >< DdeI
>< NlaIII
>< BbvI
>< Eco57I >< BfrI >< HinfI >< MseI >< MaeIII
>< AfaI Fnu4HI ><
TTCTTTAACA ACTATCTTAG GAAAAGAGTC ATGTTTAATG GAGTTACATT TAGTACCTTC GAGGAGGCTG
9670 9680 9690 9700 9710 9720 9730

>< RsaI
>< Csp6I
>< BcgI
>< RsaI
>< Csp6I
>< BsmAI

```

FIGURE 13.22

```

    >> AfaI                >> AfaI                >> Alw26I
CTTTGTGTAC CTTTGTGCTC AACAAGGAAA TGTACCTAAA ATTGCGTAGC GAGACACTGT TGCCACTTAC
    9740          9750          9760          9770          9780          9790          9800

                                >> NlaIV
                                >> DdeI
                                >> RsaI
                                >> BscBI
                                >> Csp6I
                                >> BfrI   AluI >>
                                >> AfaI
ACAGTATAAC AGGTATCTTG CTCTATATAA CAAGTACAAG TATTCAGTG GAGCCTTAGA TACTACCAGC
    9810          9820          9830          9840          9850          9860          9870

    >> Fnu4HI
                                >> DdeI
                                >> BfrI
    >> Fnu4HI
    >> BbvI   >> AluI   >> BbvI                >> DdeI >> AlwNI
TATCGTGAAG CAGCTTGCTG CCACTTAGCA AAGGCTCTAA ATGACTTTAG CAACTCAGGT GCTGATGTTT
    9880          9890          9900          9910          9920          9930          9940

                                >> SfcI
                                >> PstI
                                >> BsmI
                                >> BscCI
TCTACCAACC ACCACAGACA TCAATCACTT CTGCTGTICT GCAGAGTGGT TTTAGGAAAA TGGCATTCCT
    9950          9960          9970          9980          9990          10000          10010

                                >> RsaI
                                >> NlaIII
                                >> MaeIII
                                >> Csp6I
                                >> AfaI
                                >> Tru9I
                                >> MseI
GTCAGGCAAA GTTGAAGGGT GCATGGTACA AGTAACCTGT GGAAGTACAA CTCTAATGG ATTGTGGTTG
    10020          10030          10040          10050          10060          10070          10080

                                XhoII >>
                                Sau3AI >>
                                >> Tru9I   NdeII >>
                                >> NspI     MflI >>
                                >> NspHI    MboI >>
                                >> NlaIII   DpnII >>
                                >> MseI     BstYI >>
                                >> MboII   BspAI >>
                                >> AccI     >> AflIII
                                >> Bst1107I
                                >> NspI
                                >> NspHI
                                >> NlaIII
                                >> BbsI
                                >> BglII >>
GATGACACAG TATACTGTCC AAGACATGTC ATTTGCACAG CAGAAGACAT GCTTAATCCT AACTATGAAG
    10090          10100          10110          10120          10130          10140          10150

                                Pali >
                                MscI >
                                HaeIII >
                                EaeI >>
                                BsuRI >
                                BshI >
                                Bali >
    >> DpnI >> MboII
    >> Bsp143I
    >> Bsp143I                >> AluI
ATCTGCTCAT TCGCAAATCC AACCATAGCT TTCTTGTTCA GGCTGGCAAT GTTCAACTTC GTGTTATTGG
    10160          10170          10180          10190          10200          10210          10220

                                >> DdeI > < Tru9I
                                >> BfrI > < MseI
                                >> DdeI
CCATTCTATG CAAAATTGTC TGCTTAGGCT TAAAGTTGAT ACTTCTAACC CTAAGACACC CAAGTATAAA
    10230          10240          10250          10260          10270          10280          10290

                                >> ScrFI
                                >> MvaI
                                >> EcoRII
                                >> Ecl136I
                                >> SphI

```

FIGURE 13.23

```

                >> DsaV
                >> BstOI
                >> BstNI
                >> BsiLI
                >> ApyI
                >> PaeI
                >> NspI
                >> NspHI
                >> RmaI >> NlaIII
                >> MaeI >> HphI
TTTGTCCGTA TCCAACCTGG TCAAACATTT TCAGTTCTAG CATGCTACAA TGGTTCACCA TCTGGTGTTT
10300      10310      10320      10330      10340      10350      10360

                >> Sau3AI
                >> NdeII
                >> MboI>> NlaIII
                >> DpnII
                >> Eco3II
                >> BsmAI
                >> BsaI>> NlaIII
                >> Alw26I
                >> Tru9I
                >> MseI
                >> Tru9I>> DpnI
                >> MseI >> Bsp143I
                >> BspAI>> AlwI
ATCAGTGTGC CATGAGACCT AATCATACCA TTAAGGTTTC TTTCCTTAAT GGATCATGTG GTAGTGTGG
10370      10380      10390      10400      10410      10420      10430

                >> Zsp2I
                >> Ppu10I
                >> NsiI>> SfaNI
                >> NdeI
                >> Mph1103I
                >> EcoT22I
                >> AvaIII
                >> AluI
                >> RsaI >>
                >> Csp6I >>
                >> AfaI >>
                >> Tru9I
                >> MseI
                >> MseI >>
                >> EcoT22I
                >> AvaIII
                >> AluI
                >> RsaI >>
                >> Csp6I >>
                >> AfaI >>
TTTTAACATT GATTATGATT GCGTGTCTTT CTGCTATATG CATCATATGG AGCTTCCAAC AGGAGTACAC
10440      10450      10460      10470      10480      10490      10500

                >> SinI
                >> Sau96I
                >> NspIV
                >> NspHII
                >> Eco47I
                >> Cfr13I
                >> BsiZI
                >> Bme18I
                >> AvaII
                >> AsuI>> BsgI
                >> HindII
                >> HincII
                >> BbvI
                >> BspMI
                >> SfcI
                >> RsaI >>
                >> PstI >>
                >> Fnu4HI
                >> Csp6I >>
                >> BspWI
                >> RsaI >>
                >> Csp6I >>
                >> AfaI >>
GCTGGTACTG ACTTAGAAGG TAAATTCTAT GGTCCATTTG TTGACAGACA AACTGCACAG GCTGCAGGTA
10510      10520      10530      10540      10550      10560      10570

                >> Tru9I
                >> MseI
                >> BbvI
                >> NlaIII
                >> Fnu4HI
                >> HphI >>
CAGACACAAC CATAACATTA AATGTTTGG CATGGCTGTA TGCTGCTGTT ATCAATGGTG ATAGGTGGTT
10580      10590      10600      10610      10620      10630      10640

                >> Tru9I
                >> TfiI
                >> MseI
                >> HphI
                >> HinfI
                >> Tru9I
                >> MseI
                >> RsaI
                >> Csp6I
                >> AfaI
TCTTAATAGA TTCACCACTA CTTTGAATGA CTTTAACCTT GTGGCAATGA AGTACAACCTA TGAACCTTTG
10650      10660      10670      10680      10690      10700      10710

                >> SinI
                >> Sau96I
                >> PssI
                >> Psp5II
                >> PpuMI
                >> NspIV
                >> NspHII
                >> NlaIV

```

FIGURE 13. 24


```

                >> Tru9I
                >> MseI
    > < RmaI          > < Esp4I
    > < MaeI          >> Eco57I
        >> AluI          > < AflIII          >> AluI
TGGCTGACAC TAGCTTGCT GGTATAGGC TTAAGGATTG TGTATGTAT GCTTCAGCTT TAGTTTTCCT
11210      11220      11230      11240      11250      11260      11270

                >> RmaI
                >> MaeII
                >> MaeI
    > < NlaIII      >> SfaNI      >> Enu4HI
    >> BspHI >> AluI >> BbvI      >> AflIII
TATTCTCATG ACAGCTCGCA CTGTTATGA TGATGCTGCT AGACGTGTT GGACACTGAT GAATGTCATT
11280      11290      11300      11310      11320      11330      11340

                                                >> Sau96I
                                                >> Pali
                                                >> NspIV
                                                >< NlaII
                                                >> HaeIII
                                                > < DdeI
    >> Sau3AI
    >> NdeII          >< Cfr13I
    >> MboI          >< BsuRI
    >> DpnII        >< BsiZI
    >< DpnI          >< BshI
    >> Bsp143I      > < BfrI
                >> AccI          >> BspAI >> AluI          >> AsuI
ACACTTGTTT ACAAAGTCTA CTATGGTAAT GCTTTAGATC AAGCTATTTC CATGTGGGCC TTAGTTATTT
11350      11360      11370      11380      11390      11400      11410

                >> RmaI
                >< NlaIII
                >> MaeI >> SfcI
    >> MaeIII      >> MnlI      >< MaeIII      >< AluI >< AluI
CTGTAACCTC TAACTATTCT GGTGTCGTTA CGACTATCAT GTTTTAGCT AGAGCTATAG TGTTTGTGTG
11420      11430      11440      11450      11460      11470      11480

                                                DdeI >
    TGTTGAGTAT TACCCATTGT TATTATTAC TGGCAACACC TTACAGTGTA TCATGCTTGT TTATTGTTTC
11490      11500      11510      11520      11530      11540      11550

                >> Pali
                >> HaeIII
                >> Fnu4HI      >< BsuRI
    >> BbvI      >> Fnu4HI      >< BspWI
    >> BbvI      >> BspWI      >< BshI      >> Eco57I      >< MaeIII
TTAGGCTATT GTTGCTGCTG CTACTTTGGC CTTTCTGTT TACTCAACCG TTAFTTCAGG CTTACTCTTG
11560      11570      11580      11590      11600      11610      11620

                                                >< ScrFI
                                                >< MvaI
    >< EcoRII
    >< Ecl136I
    >< DsaV
    >< BstOI
    >< BstNI
    >< BsiLI
    > < BsaJI
    >< BsaJI

    >> Eco31I
    >> BsmAI
    >> BsaI

```

FIGURE 13.26


```

    >> DrdI >> Alw26I >> ApyI DdeI >>
GTGTTTATGA CTA CTCTGGTC TCTACACAAG AATTTAGGTA TATGA ACTCC CAGGGGCTTT TGCCTCCTAA
11630 11640 11650 11660 11670 11680 11690

    >> Tru9I
    >> MseI
>> SfaNI >> HindIII >< Tru9I
>> MnlI >> AluI >< MseI >< MnlI >< NlaIII
GAGTAGTATT GATGCTTTCA AGCTTAACAT TAAGTTGTTG GGTATTGGAG GTAAACCATG TATCAAGGTT
11700 11710 11720 11730 11740 11750 11760

    >> VneI
    >> SnoI
    >> SduI
    >> NspII
    >> HqiAI
    >> Bsp1286I
    >> BmyI >> RsaI
    >> RsaI >> ApaLI >> MboII
    >> Csp6I >> Alw44I >> Csp6I DdeI >
    >> AfaI >> MaeII >> Alw21I >> AfaI BfrI >
GCTACTGTAC AGTCTAAAAT GTCTGACGTA AAGTGACAT CTGTGGTACT GCTCTCGGTT CTCAACAAC
11770 11780 11790 11800 11810 11820 11830

    >> NspII >< RsaI
    >> DraIII
    >> SduI >< Csp6I
    >> Bsp1286I
    >> MboII >> Hinfi >> PleI >> BmyI >< AfaI >> MboII
TTAGAGTAGA GTCATCTTCT AAATTGTGGG CACAATGTGT ACAACTCCAC AATGATATTC TTCTTGCAAA
11840 11850 11860 11870 11880 11890 11900

    >> TthHB8I
    >> TaqI SfcI >>
    >> HindIII >> MboII >> NlaIII
    >> AluI >< Eco57I >> BspWI AccI >>
AGACACA ACT GAAGCTTTCG AGAAGATGGT TTCTCTTTTG TCTGTTTTGC TATCCATGCA GGGTGCTGTA
11910 11920 11930 11940 11950 11960 11970

    >> VspI
    >> Tru9I >< Ksp632I
    >> MseI >< TthHB8I >< EarI
    >> AsnI >> TaqI >> MboII >< Eam1104I
    >> AseI >< MnlI >> BcgI/a >> Eco57I >> Eco57I >> BcgI
GACATTAATA GGTGTGCGA GGAAATGCTC GATAACCGTG CTACTCTTCA GGCTATGCTC TCAGAATTA
11980 11990 12000 12010 12020 12030 12040

    >> StuI
    >> ScrFI
    >> Pali
    >> MvaI >< HaeIII
    >> EcoRII >< Eco147I
    >> Ecl136I
    >> DsaV >> BsuRI
    >> BstOI
    >> BstNI
    >> BspWI
    >> BsiLI
    >> Fnu4HI >> BsaJI >< BshI TfiI >>
    >> NdeI >> BspWI >< MnlI >> BglI >> SfcI Hinfi >>
    >> AciI >> ApyI >< AatI >> >< AluI

```

FIGURE 13. 27

```

GTTCTTTACC ATCATATGCC GCTTATGCCA CTGCCCAGGA GGCCTATGAG CAGGCTGTAG CTAATGGTGA
12050      12060      12070      12080      12090      12100      12110

    >> XmnI          >> Tru9I                      >> SfaNI
    >> HphI          >> MseI                      >> DdeI
    >> Asp700I      >> Eco57I                      >> BbvI Fnu4HI >>
TTCTGAAGTC GTTCTCAAAA AGTTAAAGAA ATCTTTGAAT GTGGCTAAAAT CTGAGTTGA CCGTGATGCT
12120      12130      12140      12150      12160      12170      12180

                                                XhoII >>
                                                Sau3AI >>
                                                NdeII >>
                                                MnlI >
                                                >< MnlI
                                                >< MflI
                                                >< MboI
    >< Sau3AI
    >< NdeII
    >< MboI
    >< DpnII
    >< DpnI
    >< DpnI
    >< BspWI
    >< BspAI
    >< Bsp143I
    >< NlaIII
    >< NlaIII
    >< Bsp143I
    >< AfaIBglIII ><
    >< RsaIBspAI ><
    >< Csp6IBsp143I ><
    >< AfaIBglIII ><
GCCATGCAAC GCAAGTTGGA AAAGATGGCA GATCAGGCTA TGACCCAAAT GTACAAACAG GCAAGATCTG
12190      12200      12210      12220      12230      12240      12250

    >> SpeI
    >> RmaI
    >> MaeIII
    >> MaeI
    >> SpeI
    >> RmaI
    >> MaeIII
    >> MaeI
    >> Ksp632I > < HindIII
    >> Ksp632I > < DdeI >< SfaNI
    >> MboII
    >> Eam1104I >< BspWI
    >> BspWI
    >> EarI >< BfrI >< AluI
AGGACAAGAG GGCAAAAGTA ACTAGTGCTA TGCAAACAAT GCTCTTCACT ATGCTTAGGA AGCTTGATAA
12260      12270      12280      12290      12300      12310      12320

    >> ThaI
    >> MvnI
    >> HinPII
    >> Hin6I
    >> HhaI
    >> CfoI
    >> BstUI
    >> Tru9I
    >> MseI
    >> Bsp50I
    >> AccII
    >> ThaI
    >> MvnI
    >> HinPII
    >> Hin6I
    >> HhaI
    >> CfoI
    >> BstUI
    >> Tru9I
    >> MseI
    >> Bsp50I
    >> AccII
    >> SfcI >>
TGATGCACTT AACAAACATTA TCAACAATGC GCGTGATGGT TGTGTTCCAC TCAACATCAT ACCATTGACT
12330      12340      12350      12360      12370      12380      12390

    >> RsaI
    >> NlaIV
    >> Eco64I
    >> Csp6I
    >> BslI
    >> BsiYI >< KpnI
    >> BscBI
    >> BanI
    >> Asp718
    >> AfaI
    >> NlaIII
    >> BstXI
    >> AccB1I
    >> MaeIII
    >> Fnu4HI >> BbvI
    >> Acc65I
    >> BsgI >>
ACAGCAGCCA AACTCATGGT TGTGTGCCCT GATTATGGTA CCTACAAGAA CACTTGTGAT -GGTAACACCT
12400      12410      12420      12430      12440      12450      12460

    >> Zsp2I
    >> Ppu10I

```

FIGURE 13.28

```

    >> NsiI
    >> Mph1103I
    >> NdeI>> EcoT22I
    >> AvaIII >> SfaNI
    >> SfaNI
    >> AciI
    DdeI >>
    BfrI >>
    TTACATATGC ATCTGCACTC TGGGAAATCC AGCAAGTTGT TGATGCGGAT AGCAAGATTG TTCAACTTAG
    12470 12480 12490 12500 12510 12520 12530

    >> Pali
    >> HaeIII >> MnlI >> DdeIDdeI >>
    >> BsuRI >> MaeIII >> BspWI
    >> MseI>> HphI >> XcmI>> BshI >> AluI BspWI >>
    TGAAATTAAC ATGGACAATT CACCAAATTT GGCTTGGCCT CTTATTGTTA CAGCTCTAAG AGCCAACCTCA
    12540 12550 12560 12570 12580 12590 12600

    RsaI >>
    NlaIV >>
    KpnI >>
    >> Fnu4HI
    Eco64I >>
    Csp6I >>
    BscBI >>
    Asp718 >>
    AfaI >>
    >> AciI>> BanI
    AccB1I >>
    >> Tru9I
    >> PvuII
    >> Psp5I
    >> NspBII
    >> MseI
    >> AluI >> SfcI
    >> HinfI >> PleI
    >> DdeI>> BsrI
    >> PshAI
    Acc65I >>
    GCTGTAAAC TACAGAATAA TGAAGTGTAGT CCAGTAGCAC TACGACAGAT GTCCTGTGCG GCTGGTACCA
    12610 12620 12630 12640 12650 12660 12670

    >> TthHB8I
    >> TaqI
    >> SfuI
    >> NspV
    >> MnlI
    >> LspI
    >> Csp45I
    >> BstBI
    >> Bsp119I
    >> BsiCI
    >> Bpu14I
    >> AsuII
    >> RsaI
    >> Csp6I
    >> AluI
    >> AfaI
    CACAAACAGC TTGTAAGTAT GACAATGCAC TTGCCTACTA TAACAATTTCG AAGGGAGGTA GGTTTGTGCT
    12680 12690 12700 12710 12720 12730 12740

    >> XhoII
    >> Sau3AI
    >> NdeII
    >> MflI
    >> MboI
    >> DpnII
    >> DpnI
    >> BstYI
    >> BspAI
    >> Bsp143I
    >> BglII
    >> TfiI
    >> RmaI
    >> HinfI
    >> MaeI >> DdeI
    >> RsaI
    >> Csp6I
    >> Csp6I>> RsaI
    >> AfaI>> AfaI
    GGCATTACTA TCAGACCACC AAGATCTCAA ATGGGCTAGA TTCCCTAAGA GTGATGGTAC AGGTACAATT
    12750 12760 12770 12780 12790 12800 12810

    >> Sau96I
    >> PssI
    >> Pali
    >> NspIV

```

FIGURE 13.29

```

>< HaeIII
>< EcoO109I
>< DraII
>< CfrI3I
>< BsuRI
>< NlaIV
>< BsrI
>< BscBI
>< MaeIII
>< BsiZI
>< BshI
>< AsuI
>< RsaI
>< Csp6I
>< AfaI
TACACAGAAC TGGAACCACC TTGTAGGTTT GTTACAGACA CACCAAAAGG GCCTAAAGTG AAATACTTGT
12820 12830 12840 12850 12860 12870 12880

>< SfcI
>< MboII
MaeII ><
>< Fnu4HI >< RsaI
>< Eco57I >< Csp6I
>< BbsI
>< Tru9I
>< MseI >< MnlI
>< BbvI >< AluI >< AfaI
ACTTCATCAA AGGCTTAAAC AACCTAAATA GAGGTATGGT GCTGGGCAGT TTAGCTGCTA CAGTACGTCT
12890 12900 12910 12920 12930 12940 12950

>< RsaI
>< SfcI >< Csp6I
>< BspWI >< AfaI >< BspMI
TCAGGCTGGA AATGCTACAG AAGTACCTGC CAATTCAACT GTGCTTTCCT TCTGTGCTTT TGCAGTAGAC
12960 12970 12980 12990 13000 13010 13020

>< RmaI
>< MnlI
>< MaeI >< HphI
CCTGCTAAAG CATATAAGGA TTACCTAGCA AGTGGAGGAC AACCAATCAC CAACTGTGTG AAGATGTTGT
13030 13040 13050 13060 13070 13080 13090

>< SinI
>< Sau96I
>< NspIV
>< NspHII
>< NlaIII
>< Eco47I
>< Eam1105I
>< CfrI3I
>< BsiZI
>< Bme18I >< XcmI
>< AvaII >< PleI ><
>< RsaI >< RsaI
>< MboII >< Csp6I
>< Csp6I >< BsrI
>< AfaI >< AfaI
>< MaeIII
>< AluI >< AsuI >< HinfI
GTACACACAC TGGTACAGGA CAGGCAATTA CTGTAACACC AGAAGCTAAC ATGGACCAAG AGTCCTTTGG
13100 13110 13120 13130 13140 13150 13160

>< TfiI
>< SfaNI
>< MaelIII
>< NlaIII >< FokI
>< HinfI
TGGTGCTTCA TGTTGTCTGT ATTGTAGATG CCACATTGAC CATCCAAATC CTAAGGATT CTGTGACTTG
13170 13180 13190 13200 13210 13220 13230

>< RsaI
>< MaeII
>< Csp6I
>< AfaI
>< DdeI
>< BsrI >< BfrI
AAAGGTAAGT ACGTCCAAAT ACCTACCACT TGTGCTAATG ACCCAGTGGG TTTTACTT AGAAACACAG
13240 13250 13260 13270 13280 13290 13300

>< ThaI

```

FIGURE 13.30

```

>< SfaNI
>< MvnI
>< BstUI
>< Bsp50I
>< AciI
>< RsaI
>< Csp6I
>< AfaI >< AciI
>< SfcI >< MaeIII
>< AccIISfaNI ><
TCTGTACCGT CTGCGGAATG TGGAAAGGTT ATGGCTGTAG TTGTGACCAA CTCGCGGAAC CCTTGATGCA
13310 13320 13330 13340 13350 13360 13370

>< Zsp2I
>< SfaNI
>< Mph1103I>< Tru9I
>< Ppu10I>< MaeII
>< NsiI> < FokI
>< EcoT22I >< MseI
Fnu4HI ><
BsgI ><
>< BbvI
>< AciI>< AvaIII >< DraI >< AciI >< Fnu4HI AciI ><
GTCTGCGGAT GCATCAACGT TTTTAAACGG GTTTGCGGTG TAAGTGCAGC CCGTCTTACA CCGTGGCGCA
13380 13390 13400 13410 13420 13430 13440

>< SpeI
>< ScaI
>< RsaI
>< RmaI
>< MaeI
>< Csp6I >< SfcI
>< BspWI >< AfaI >< AccI >< BcgI/a >< BspWI
CAGGCACTAG TACTGATGTC GTCTACAGGG CTTTTGATAT TTACAACGAA AAAGTTGCTG GTTTTGCAAA
13450 13460 13470 13480 13490 13500 13510

>< ScrFI
>< MvaI
>< MnlI
>< EcoRII
>< Ecl136I
>< BstOI
>< BstNI
>< BslI
>< DsaV >< BsiYI
>< BsiLI
>< ApyI >< PleI
>< FokI >< HinfI
GTCCTAAAA ACTAATTGCT GTCGCTTCCA GGAGAAGGAT GAGGAAGGCA ATTTATTAGA CTCTTACTTT
13520 13530 13540 13550 13560 13570 13580

>< NlaIII
>< Ksp632I
>< EarI
>< Eam1104I
>< BsmAI >< Tru9I
>< MnlI >< Alw26I >< MboII >< MseI
GTAGTTAAGA GGCATACTAT GTCTAACTAC CAACATGAAG AGACTATTTA TAACTTGGTT AAAGATTGTC
13590 13600 13610 13620 13630 13640 13650

>< RsaI
>< NlaIV
>< NlaIII
>< KpnI
>< HphI
>< Eco64I
>< Csp6I
>< BscBI
>< BanI
>< Asp718

```

FIGURE 13.31

```

>< NspBII
>< AclI
CAGCGGTTGC TGTCATGAC TTTTCAAGT TTAGAGTAGA TGGTGACATG GTACCACATA TATCACGTCA
13660 13670 13680 13690 13700 13710 13720

>< MaeIII >< AfaI
> < AccBII MaeII ><
> < Acc65I > < HgaI

>< MnlI
>< MaeII
GCGTCTAACT AAATACACAA TGGCTGATTT AGTCTATGCT CTACGTCATT TTGATGAGGG TAATTGTGAT
13730 13740 13750 13760 13770 13780 13790

>< Tru9I
>< MseI >< MaeIII >< MunI
ACATTAAGG AAATACTCGT CACATACAAT TGCTGTGATG ATGATTATTT CAATAAGAAG GATTGGTATG
13800 13810 13820 13830 13840 13850 13860

>< ThaI
>< MvnI
>< MluI
>< BstUI
>< Bsp50I
>< TfiI >< AflIII >< DdeI >< RsaI
>< HinfI >< AccII >< BfrI >< Csp6I Tru9I >< HphI
ACTTCGTAGA GAATCCTGAC ATCTTACGCG TATATGCTAA CTTAGGTGAG CGTGACGCC AATCATTATT
13870 13880 13890 13900 13910 13920 13930

XhoII >
Sau3AI >
NdeII >
MflI >
MboI >
DpnII >
BstYI >
BspAI >

> < SfaNI
>< RsaI
>< Csp6I
>< AfaI >< SfaNI
AAAGACTGTA CAATTCTGCG ATGCTATGCG TGATGCAGGC ATTGTAGGCG TACTGACATT AGATAATCAG
13940 13950 13960 13970 13980 13990 14000

> < ScrFI
> < MvaI
>< Fnu4HI
>< EcoRII
> < Ecl136I
> < BstOI
> < BstNI

>< Tru9I
>< MseI >< RsaI >< BslI
>< DpnI >< Csp6I >< HphI >< BsiYI
>< Bsp143I >< BsrI >< Csp6I >< BsiLI
>< AlwI >< AfaI >< BbvI >< ApyI
GATCTTAATG GGAAGTGGTA CGATTTCGGT GATTTCGTAC AAGTAGCACC AGGCTGCGGA GTTCCTATTG
14010 14020 14030 14040 14050 14060 14070

>< SfaNI
>< HinfI
>< Fnu4HIpleI ><
>< DdeI
>< BspWI NdeI ><
>< RmaI
>< MnlI
>< MamI >< BsiBI
>< TfiI >< SfaNI >< BsaBI
>< HinfI >< FokI >< BbvI
TGGATTCATA TTAATCATTG CTGATGCCCA TCCTCACTTT GACTAGGGCA TTGGCTGCTG AGTCCCATAT
14080 14090 14100 14110 14120 14130 14140

>< Sau3AI
>< NdeII

```

FIGURE 13.32

```

>< MboI
>< MamI
>< DpnII
>< DpnI
>< BspWI
>< BspAI
>< Bsp143I
>< BsiBI
>< BsaBI >< FokI
GGATGCTGAT CTCGCAAAC CACTTATTAA GTGGGATTG CTGAAATATG ATTTTACGGA AGAGAGACTT
14150 14160 14170 14180 14190 14200 14210

>< XcmI
>< Tru9I
>< MseI
>< Ksp632I
>< Eam1104I
>< BsmAI
>< EarI AspI ><
>< Alw26I

>< SinI
>< Sau96I
>< NspIV
>< NspHII
>< NlaIV
>< TthHB8I
>< TaqI
>< McrI
>< Ksp632I
>< EarI
>< Eam1104I
>< BsmAI
>< MboII
>< Alw26I
TGTCTCTCG ACCGTTATTT TAAATATTGG GACCAGACAT ACCATCCCAA TTGTATTAAC TGTTTGGATG
14220 14230 14240 14250 14260 14270 14280

>< FokI
>< MseI
>< DraI
>< AsuI
>< Tru9I
>< MseI
>< MunI
>< MseI
>< SinI ><
>< Sau96I ><
>< NspIV ><
>< NspHII >
>< Eco47I ><
>< Cfr13I ><
>< Bsi2I ><
>< Bme18I ><
>< AvaI ><
>< AsuI ><
ATAGGTGTAT CCTTCATTGT GCAAACCTTA ATGTGTTATT TTCTACTGTG TTCCACCTA CAAGTTTTGG
14290 14300 14310 14320 14330 14340 14350

>< SpeI
>< RmaI
>< MaeI
>< SspI
>< BsrI
ACCCACTAGTA AGAAAAATAT TTGTAGATGG TGTTCCCTTT GTTGTTTCAA CTGGATACCA TTTTGGTGAG
14360 14370 14380 14390 14400 14410 14420

>< RsaI
>< HinfI >< PfiI
>< Csp6I
>< AfaI
>< HgaI >< AluI
>< FokI
>< AccII
>< BbvI
TTAGGAGTCG TACATAATCA GGATGTAAC TTACATAGCT CGCGTCTCAG TTTCAAGGAA CTTTGTAGTGT
14430 14440 14450 14460 14470 14480 14490

>< 2sp2I
>< SphI
>< Ppu10I
>< PaeI
>< NspI

```

FIGURE 13.33

```

    >< Sau3AI          >< NspHI
    >< NdeII           >< NsiI
    >< MboI            >< NlaIII
    >< DpnII           >< Mph1103I
    > < DpnI          >< Fnu4HI
    >< Fnu4HI>> BspWI >< EcoT22I
    >< BspAI           >< BspWI
    > < Bsp143I> < AvaIII > < AlwNI
    >< AlwI            >< AluI          >< AluI >< BbvI >< MaeI
    ATGCTGCTGA TCCAGCTATG CATGCAGCTT CTGGCAATTT ATTGCTAGAT AAACGCACTA CATGCTTTTC
    14500      14510      14520      14530      14540      14550      14560

    >< ScrFI
    >< NciI
    >< MspI
    >< HpaII
    >< HapII
    >< Fnu4HI
    >< AlwNI
    >< AluI
    >< DsaV           >< Tru9I
    >< BcnI          >< MseI
    AGTAGCTGCA CTAACAAACA ATGTTGCTTT TCAAACGTGC AAACCCGGTA ATTTTAATAA AGACTTTTAT
    14570      14580      14590      14600      14610      14620      14630

    >< Tru9I
    >< MseI
    >< MboII
    >< DdeI >>
    >< BbvI >>
    GACTTTGCTG TGTCTAAAGG TTTCTTTAAG GAAGGAAGTT CTGTTGAACT AAAACACTTC TTCTTTGCTC
    14640      14650      14660      14670      14680      14690      14700

    >< FokI
    >< Fnu4HI
    >< EcoRV >>
    >< Eco32I >>
    AGGATGGCAA CGCTGCTATC AGTGATTATG ACTATTATCG TTATAATCTG CCAACAATGT GTGATATCAG
    14710      14720      14730      14740      14750      14760      14770

    >< VspI
    >< Tru9I
    >< MseI
    >< AsnI
    >< AseI
    >< MaeIII
    ACAACTCCTA TTCGTAGTTG AAGTTGTTGA TAAATACTTT GATTGTTACG ATGGTGGCTG TATTAATGCC
    14780      14790      14800      14810      14820      14830      14840

    >< Tru9I
    >< MseI
    >< HpaI
    >< HindII
    >< HincII
    >< PvuII
    >< Psp5I
    > < XcmI
    >< NspBII
    >< Tru9I
    >< RmaI >>
    >< AluI
    >< MseI
    >< MaeI >>
    AACCAAGTAA TCGTTAACAA TCTGGATAAA TCAGCTGGTT TCCCATTTAA TAAATGGGGT AAGGCTAGAC
    14850      14860      14870      14880      14890      14900      14910

    >< SfaNI
    >< Sau3AI
    >< NdeII
    >< MboI
    >< DpnII
    >< DpnI
    >< PleI
    >< HinfI>> MnlI
    >< BspAI >> AlwI
    >< Bsp143I
    >< AlwI
    >< ThaI
    >< MvnI
    >< BstUI
    >< Bst1107I
    >< BspWI >> FokI
    >< Bsp50I
    >< AccII>> DdeI
    >< AccI
    TTTATTATGA CTCAATGAGT TATGAGGATC AAGATGCACT TTTCGCGTAT ACTAAGCGTA ATGTCATCCC
    14920      14930      14940      14950      14960      14970      14980

    >< SstI
    >< SduI
    >< SacI

```

FIGURE 13.34


```

                                << NspII
                                << HgiAI
                                << Eco24I
                                << Ecl136II
                                > < Bsp1286I
                                << BmyI
                                << BanII
                                << Alw21I
                                << AluI
                                << AluI
                                << Tru9I
                                << TfiI
                                << MseI
                                << HinfI
                                > < Esp4I
                                > < AflIII
                                << BspWI
                                > < AluI
                                << AluI
TACTATAACT CAAATGAATC TTAAGTATGC CATTAGTGCA AAGAATAGAG CTCGCACCGT AGCTGGTGTG
14990      15000      15010      15020      15030      15040      15050

                                RmaI ><
                                > < MnlI
                                MaeI ><
                                << Fnu4HI
                                << AciI
                                << ScaI
                                << SfcI>< RsaI
                                << BsmAI >< Csp6I
                                << Alw26I >< AfaI
TCTATCTGTA GTACTATGAC AAATAGACAG TTTCATCAGA AATTATTGAA GTCAATAGCC GCCACTAGAG
15060      15070      15080      15090      15100      15110      15120

                                << Tru9I
                                << MseI
                                << AluI
GAGTACTGT GGTAATTGGA ACAAGCAAGT TTTACGGTGG CTGGCATAAT ATGTTAAAAA CTGTTTACAG
15130      15140      15150      15160      15170      15180      15190

                                NspI ><
                                NspHI ><
                                NlaIII ><
                                << NlaIII
                                DdeI ><
                                BspWI ><
                                << MaeIII
                                BfrI ><
TGATGTAGAA ACTCCACACC TTATGGGTTG GGATTATCCA AAATGTGACA GAGCCATGCC TAACATGCTT
15200      15210      15220      15230      15240      15250      15260

                                > < Pali
                                > < HaeIII
                                > < BsuRI
                                > < BshI
                                << MnlI
                                << MaeIII
                                SfcI ><
AGGATAATGG CCTCTCTGT TCTTGCTCGC AAACATAACA CTTGCTGTAA CTTATCACAC CGTTTCTACA
15270      15280      15290      15300      15310      15320      15330

                                Tru9I ><
                                ScrFI >
                                MvaI >
                                << MseI
                                FokI ><
                                EcoRII ><
                                Ecl136I >
                                DsaV ><
                                BstOI >
                                BstNI >
                                << MstI
                                << HinPII
                                << Hin6I
                                > < HhaI
                                << FspI
                                << FdiII
                                << CfoI>< Tru9I
                                > < Fnu4HI
                                << AluI
                                << AviII >< MseI
                                << AciI
                                ApyI >
GGTTAGCTAA CGAGTGTGCG CAAGTATTAA GTGAGATGGT CATGTGTGGC GGCTCACTAT ATGTTAAACC
15340      15350      15360      15370      15380      15390      15400

                                > < SfaNI
                                << MspI
                                << HpaII
                                << HapII
                                << HphI
                                << BspWI
                                << Tru9I
                                MaeIII ><
                                << MseI
                                AluI ><

```

FIGURE 13.35


```

                << RsaI          << DpnII
                << MaeII        << DpnI          > < SspI
                << Tru9I        << Csp6I        << BstYI      HinPII >>
                << RmaI        << BsaAI          << BspMI      Hin6I >>
                << MaeI        << AflIII        << BspAI      HhaI >>
                << BspWI>< MseI    << AfaI        << AlwI>< Bsp143I  CfoI >>
AATGCTAGTT AAACAAGGAG ATGATTACGT GTACCTGCCT TACCCAGATC CATCAAGAAT ATTAGGCGCA
15830      15840      15850      15860      15870      15880      15890

                << RsaI          << SfaNI
                << TthHB8I      << Csp6I        << MaeIII
                << TaqI        << AfaI          BsrI >>
GGCTGTTTTG TCGATGATAT TGTCAAAACA GATGGTACAC TTATGATTGA AAGGTTCTGT TCACTGGCTA
15900      15910      15920      15930      15940      15950      15960

                > < FokI
                << BspWI
TTGATGCTTA CCCACTTACA AAACATCCTA ATCAGGAGTA TGCTGATGTC TTCACTTGT ATTTACAATA
15970      15980      15990      16000      16010      16020      16030

                << Van9I
                << PflMI
                << NspI
                > < Pali>< NspHI
                > < MscI>< NlaIII
                > < HaeIII
                > < BsuRI
                << BsrI
                << EaeI    << BslI    << NspI
                > < BshI>< BsiYI << NspHI
                << NlaIII    << AflIII << AflIII
                << MaeIII    << AluI > < BalI>< AccB7I >< NlaIII
CATTAGAAAG TTACATGATG AGCTTACTGG CCACATGTTG GACATGTATT CCGTAATGCT AACTAATGAT
16040      16050      16060      16070      16080      16090      16100

                << RsaI> < NlaIV
                << MnlI
                << Csp6I    << DdeI          << RsaI
                << BsrI >< MnlI    << Csp6I
                << AfaI> < BscBI    << AfaI          SfcI >>
AACACCTCAC GGTAAGGGA ACCTGAGTTT TATGAGGCTA TGTACACACC ACATACAGTC TTGCAGGCTG
16110      16120      16130      16140      16150      16160      16170

                << NlaIV
                << EcoNI
                << Eco3II
                << Eco64I>< BsmAI
                << BscBI >< BslI
                << BanI    << BsiYI
                << AciI    << BsaI
                << BspWI
                << AccB1I>< Alw26I BbvI >>
TAGGTGCTTG TGTATTGTGC AATTCACAGA CTTCACTTCG TTGCGGTGCC TGTATTAGGA GACCATTCTT
16180      16190      16200      16210      16220      16230      16240

                << Tth111I
                << Fnu4HI    << NlaIII
                << BspWI >< AspI          > < Tru9I
                << BspWI >< AspI          > < MseI
ATGTTGCAAG TGCTGCTATG ACCATGTCAT TTCAACATCA CACAAATTAG TGTTGTCTGT TAATCCCTAT
16250      16260      16270      16280      16290      16300      16310

                << ScrFI
                << MvaI

```

FIGURE 13.37

```

>< EcoRII
  >< Ecl136I
>< DsaV
  >< BstOI
  >< BstNI
  >< BsiLI
>< BsaJI
  >< ApyI
  >< MaeIII >< MaeIII
  >< MaeI
  >< RmaI
  >< MnlI
  >< BspWI ><
  >< AluI
GTTTGAATG CCCCAGGTTG TGATGTCACT GATGTGACAC AACTGTATCT AGGAGGTATG AGCTATTATT
16320      16330      16340      16350      16360      16370      16380

>< MaeIII
GCAAGTCACA TAAGCCTCCC ATTAGTTTTT CATTATGTGC TAATGGTCAG GTTTTTGGTT TATACAAAAA
16390      16400      16410      16420      16430      16440      16450

>< NspI
>< NspHI
>< NlaIII>< MaeIII>< MaeIII
>< AflIII
CACATGTGTA GGCAGTGACA ATGTCACTGA CTCAATGCG ATAGCAACAT GTGATTGGAC TAATGCTGGC
16460      16470      16480      16490      16500      16510      16520

>< RsaI
  >< P1eI
  >< DdeI
>< Csp6I
  >< BsmAI >< HinfI
  >< Alw26I
  >< AfaI
  >< HindIII
  >< AluI >< Fnu4HI
  >< BbvI
  >< MnlI
  >< DdeI ><
GATTACATAC TTGCCAACAC TTGTACTGAG AGACTCAAGC TTTTCGCAGC AGAAACGCTC AAAGCCACTG
16530      16540      16550      16560      16570      16580      16590

>< ThaI
  >< ScaI
>< RsaI >< RsaI
  >< MvnI
>< Csp6I >< Csp6I
  >< BstUI
  >< Bsp50I
  >< AfaI >< AfaI
  >< AccII
  >< MnlI >
AGGAAACATT TAAGCTGTCA TATGGTATTG CCACTGTACG CGAAGTACTC TCTGACAGAG AATGTCATCT
16600      16610      16620      16630      16640      16650      16660

MaeIII ><
  >< MaeII
  >< EcoO65I
  >< Eco9II
  >< BstPI
  >< BstEII
  >< BsrI
>< SfaNI
  >< NlaIII
  >< RmaI
  >< MaeI
TTCATGGGAG GTTGGAAAAC CTAGACCACC ATTGAACAGA AACTATGTCT TTACTGGTTA CCGTGTACT
16670      16680      16690      16700      16710      16720      16730

RsaI ><
  >< MnlI
  >< HphI
  >< Csp6I ><
  >< SfaNI
  >< MaeIII
  >< HphI AfaI ><
  >< RsaI
  >< RsaI
  >< Csp6I
  >< AfaI
  >< AfaI
  >< MaeIII
  >< HphI AfaI ><
AAAAATAGTA AAGTACAGAT TGGAGAGTAC ACCTTTGAAA AAGGTGACTA TGGTGATGCT GTTGTGTACA
16740      16750      16760      16770      16780      16790      16800

```

FIGURE 13. 38

```

    >> RsaI
    >> Csp6I
    >> AfaI
GAGGTACTAC GACATACAAG TTGAATGTTG GTGATTACTT TGTGTTGACA TCTCACACTG TAATGCCACT
    16810      16820      16830      16840      16850      16860      16870

    >> HphI
    >> HindII
    >> HincII
    DdeI >>
    BfrI >>

    >> VneI
    >> SnoI
    >> SduI
    >> NspII
    >> HgiAI
    >< SduI
    >< NspII
    >< HgiAI
    >< DraIII
    >< Bsp1286I
    >< BmyI
    >< BspWI >< DraIII
    >< RsaI
    >< Csp6I
    >< RmaI
    >< RmaI >< RmaI
    >< Bsp1286I
    >< Alw44I >< MaeI
    >< BmyI
    >< BsrI
    >< Alw21I
    >< Alw21I
    >< AfaI
    DdeI >
TAGTGCACCT ACTCTAGTGC CACAAGAGCA CTATGTGAGA ATTACTGGCT TGTACCCAAC ACTCAACATC
    16880      16890      16900      16910      16920      16930      16940

    StyI >>
    SinI >
    Sau96I >
    NspIV >
    EcoT14I ><
    Eco47I >
    Eco130I ><
    >< ScaI Cfr13I >
    BssT1I ><
    >< SphI >< RsaI Bsi2I >
    >< PaeI BsaJI ><
    >< NlaIII Bme18I >
    >< NspI >< Csp6I AvaII >
    >< NspHI >< AfaI AsuI >
TCAGATGAGT TTTCTAGCAA TGTTGCAAAT TATCAAAGG TCGGCATGCA AAAGTACTCT ACACTCCAAG
    16950      16960      16970      16980      16990      17000      17010

    >< ScrFI
    >< RsaI
    >< MvaI
    >< EcoRII
    >< Ecl136I
    >< Csp6I
    >< BstOI
    >< BstNI
    >< XcmI >< BslI
    >< NspHII >< BsiYI
    >< BsiLI
    >< ApyI >< BsrI
    >< DsaV >< AfaI >< HinfI >< PleI
GACCACCTGG TACTGGTAAG AGTCATTTTG CCATCGGACT TGCTCTCTAT TACCCATCTG CTCGCATAGT
    17020      17030      17040      17050      17060      17070      17080

    >< SfaNI
    >< SphI >< PvuII
    >< PaeI >< Psp5I
    >< NspI >< NspBII
    >< NspHI >< Fnu4HI
    >< Tru9I
    >< Bst1107I >< NlaIII >< BspWI
    >> SspI
    >< AccI >< NlaIII >< AluI >< BbvI
    >< MseI
GTATACGGCA TGCTCTCATG CAGCTGTTGA TGCCCTATGT GAAAAGGCAT TAAAATATTT GCCCATAGAT
    17090      17100      17110      17120      17130      17140      17150

```

FIGURE 13.39

```

> < ThaI
>< ThaI
> < MvnI
>< MvnI >< ThaI
> < HinPII
>< HinPII
>< HinPII >< MvnI
> < Hin6I
>< Hin6I
> < HhaI
>< HhaI >< HhaI
> < CfoI
>< CfoI >< CfoI
> < BstUI
>< BstUI >< BstUI
>< BssHII
>< BspMI
> < Bsp50I
>< Bsp50I>< Bsp50I
>< TfiI >< Hin6I> < AccII
>< HinfI >< AccII >< AccII
AAATGTAGTA GAATCATACC TGCGCGTGGC CGCGTAGAGT GTTTTGATAA ATTCAAAGTG AATCAACAC
17160 17170 17180 17190 17200 17210 17220
>< Zsp2I
>< Ppu10I
>< NsiI
>< Mph1103I
>< EcoT22I
>< BsqI >< AvaIII >< DrdI
TAGAACAGTA TGTTTTCTGC ACTGTAAATG CATTGCCAGA AACAACTGCT GACATTGTAG TCTTTGATGA
17230 17240 17250 17260 17270 17280 17290
>< RmaI
>< MaeI >< MaeII
AATCTCTATG GCTACTAATT ATGACTTGAG TGTTTGCAAT GCTAGACTTC GTGCAAAACA CTACGTCTAT
17300 17310 17320 17330 17340 17350 17360
>< Sau3AI
>< NdeII
>< MboI
>< DpnII
>< DpnI
>< BspAI
>< AlwI>< Bsp143I > < AciI >< RmaI
ATTGGCGATC CTGCTCAATT ACCAGCCCC CGCACATTGC TGACTAAAGG CACACTAGAA CCAGAATATT
17370 17380 17390 17400 17410 17420 17430
>< SinI
>< Sau96I
>< NspIV >< StyI
>< NspHII >< NspI
>< Eco47I >< NspHI
>< Cfr13I >< NlaIII
>< Bsi2I >< EcoT14I
>< BsgI >< Eco130I
>< Bme18I >< BssT1I
>< AvaII >< BsaJI
>< Tru9I
>< MseI
TTAATTCAGT GTGCAGACTT ATGAAAACAA TAGGTCCAGA CATGTTCCCTT GGAACCTGTC GCCGTTGTCC
17440 17450 17460 17470 17480 17490 17500

```

FIGURE 13. 40

```

    >> HindII
    >> HincII
    >> AluI
TGCTGAAATT GTTGACACTG TGAGTGCTTT AGTTTATGAC AATAAGCTAA AAGCACACAA GGATAAGTCA
17510      17520      17530      17540      17550      17560      17570

>> AluI
GCTCAATGCT TCAAAATGTT CTACAAAGGT GTTATTACAC >> NlaIII
17580      17590      17600      17610      17620      17630      17640

    >> MnlI
>> EcoNI
    >> BslI
    >> BsiYI
AAATAGGCGT TGTAAGAGAA TTTCTTACAC GCAATCCTGC TTGGAGAAAA >> HphI
17650      17660      17670      17680      17690      17700      17710
    >> SfcI
    >> DdeI
    >> TfiI
    >> AluI
    >> BfrI
    >> HinfI
TAATTCACAG AACGCTGTAG CTTCAAAAAT CTTAGGATG CCTACGCAGA CTGTTGATTC ATCACAGGGT
17720      17730      17740      17750      17760      17770      17780

    >> Tth111I
    >> AspI
    >> HindII
    >> HincII
    >> AclI
TCTGAATATG ACTATGTCAT ATTCACACAA ACTACTGAAA CAGCACACTC TTGTAATGTC AACCGCTTCA
17790      17800      17810      17820      17830      17840      17850

    >> XhoII
    >> Sau3AI
    >> NdeII
    >> MflI
    >> MboI
    >> MamI
    >> DpnII
    >> DpnI
    >> BstYI
    >> BspAI
    >> Bsp143I
    >> BsiBI
    >> BsaBI
    >> BspWI
    >> BglII
ATGTGGCTAT CACAAGGGCA AAAATTGGCA TTTTGTGCAT AATGTCTGAT AGAGATCTTT ATGACAACT
17860      17870      17880      17890      17900      17910      17920

    >> XbaI
    >> RmaI
    >> MaeI
    >> MaeII
    >> MaeIII
    >> BsrI
GCAATTTACA AGTCTAGAAA TACCACGTCG CAATGTGGCT ACATTACAAG CAGAAAATGT AACTGGACTT
17930      17940      17950      17960      17970      17980      17990

    >> Sau3AI
    >> NdeII
    >> MboII
    >> MboI
    >> FokI
    >> DpnII
    >> DpnI
    >> BspAI
    >> Bsp143I
    >> Tru9I
    >> MseI
    >> SfcI
    >> BbsI
    >> BsrI
    >> NlaIV
    >> Eco64I
    >> BscBI
    >> BanI
    >> Acc81I
    >> MnlI
    >> DdeI

```

FIGURE 13. 41


```

TGTGACACT GAAAATAACA CAGAATTCAC CAGAGTTAAT GCAAAACCTC CACCAGGTGA CCAGTTTAAA
18350      18360      18370      18380      18390      18400      18410

                << ScrFI
                << MvaI
                << EcoRII
                << Ecl136I
                << DsaV
                << BstOI
                << BstNI
                << BsiLI
                << BsaJI
                << NlaIII
                << ApyI
                << Tru9I>< Csp6I
                << MseI >> AfaI
                >> RsaI
                >> DdeI >>

CATCTTATAC CACTCATGTA TAAAGGCTTG CCCTGGAATG TAGTGCCTAT TAAGATAGTA CAAATGCTCA
18420      18430      18440      18450      18460      18470      18480

                << NlaIII
                << HinPII
                << Tth111I
                << Hin6I
                << HinfI
                << HhaI
                << AspI
                << PleI
                << CfoI
                << AluI

GTGATACACT GAAAGGATTG TCAGACAGAG TCGTGTTCGT CCTTTGGGCG CATGGCTTTG AGCTTACATC
18490      18500      18510      18520      18530      18540      18550

                << SinI
                << Sau96I
                << NspIV
                << NspHII
                << Eco47I
                << Cfr13I
                << ScaI
                << BsiZI
                << RsaI
                << Bme18I
                << Csp6I
                << AvaII
                << MaeII
                << AfaI
                << AsuI
                << AflIII
                << MaeIII>< MaeII

AATGAAGTAC TTTGTCAAGA TTGGACCTGA AAGAACGTGT TGTCTGTGTG ACAAACGTGC AACTTGCTTT
18560      18570      18580      18590      18600      18610      18620

                >< TfiI
                >< Tth111I
                >< HinfI
                >< AspI

TCTACTTCAT CAGATACTTA TGCCTGCTGG AATCATTCTG TGGGTTTGA CTATGTCTAT AACCCATTTA
18630      18640      18650      18660      18670      18680      18690

                >< ScrFI
                >< RsaI >>
                >< MvaI
                >< EcoRII
                >< Ecl136I >>
                >< DsaV
                >< Csp6I >>
                >< BstXI >>
                >< MaeIII
                >< BstOI
                >< EcoO65I
                >< BstNI
                >< Eco91I
                >< BsiLI
                >< BstPI
                >< ApyI
                >< Eco57I> << BstEII
                >< MaeIII >< NlaIII
                >< AfaI >>

TGATTGATGT TCAGCAGTGG GGCTTTACGG GTAACCTTCA GAGTAACCAT GACCAACATT GCCAGGTACA
18700      18710      18720      18730      18740      18750      18760

                << SfaNI
                << RmaI
                << NspI
                << NspHI
    
```

FIGURE 13.43

```

                >> NlaIII                >> RmaI
                >> MaeI                >> NlaIII                Tru9I >>
<> NlaIII    >> BspWI                >> MaeI                >> NlaIII
    > < AflIII                >> BspHI                MseI >>
TGGAAATGCA CATGTGGCTA GTTGTGATGC TATCATGACT AGATGTTTAG CAGTCCATGA GTGCTTTGTT
    18770      18780      18790      18800      18810      18820      18830

    >> Thai
    >> MvnI
    >> HinPII
    >> Hin6I
    >> Hhai
    >> CfoI
    >> BstUI
    >> Bsp50I
    >> AccII
                >> EcoNI > < MnlI
                >> BslI
                >> BsiYI
                >> DdeI
                >> Tru9I
                >> MseI
AAGCGCGTTG ATTGGTCTGT TGAATACCCCT ATTATAGGAG ATGAAGTGGAG GGTAAATTCT GCTTGCAGAA
    18840      18850      18860      18870      18880      18890      18900

    >> RsaI
    >> Csp6I
    >> AfaI
                >> NlaIII
                >> BspWI
                >> MboII
                >> BsrI
                >> BspHI
                > < NlaIII
AAGTACAACA CATGGTTGTG AAGTCTGCAT TGCTTGCTGA TAAGTTTCCA GTTCTTCATG ACATTGGAAA
    18910      18920      18930      18940      18950      18960      18970

                >> SauI
                >> MstII
                >> Eco8II
                >> DdeI
                >> CvnI
                >> Bsu36I
                >> Bse2II
                >> AxyI
                >> AocI
                >> MnlI
                >> SfaNI
                >> Bpu1102I
                NlaIII >>
                >> EspI
                >> Eco57I MaeIII >>
                >> DdeI
                >> CelII
TCCAAAGGCT ATCAAGTGTG TGCCTCAGGC TGAAGTAGAA TGGAAGTTCT ACGATGCTCA GCCATGTAGT
    18980      18990      19000      19010      19020      19030      19040

                >> MnlI
                >> Ksp632I
    >> HindIII
    >> EarI
    >> AluI
    >> MboII
    >> Eam1104I
GACAAAGCTT ACAAATAGA GGAAGTCTTC TATCTTATG CTACACATCA CGATAAATTC ACTGATGGTG
    19050      19060      19070      19080      19090      19100      19110

                >> Sau3AI
                >> NdeII
                >> MboI
                >> MaeII > < MaeIII
                >> DpnII
                >> DpnI
                >> BspAI
                >> MaeIII
                >> Bsp143I
                >> MunI
                >> HinFI >
                >> DrdI >>
TTTGTGTTGTT TTGGAATTGT AACGTTGATC GTTACCCAGC CAATGCAATT GTGTGTAGGT TTGACACAAG
    19120      19130      19140      19150      19160      19170      19180

                >> Zsp2I >>
                >> SphI
                > < Ppu10I
                >> PaeI
                >> NspI
                >> NspHI
                >> NlaIII
                >> Mph1103I >>
                >> ScrFI
                >> MvaI
                >> EcoRII
    
```

FIGURE 1344

```

                >> Ecl136I
                >> DsaV
                >> BstOI
                >> BstNI
                >> BsiLI
                >> ApyI
                >> PleI
    AGTCTTGTCA AACTTGAAC TACCAGGCTG TGATGGTGGT AGTTTGTATG TGAATAAGCA TGCATTCCAC
    19190      19200      19210      19220      19230      19240      19250

                >> Tru9I
                > < MunI
                >> TthHB8I
                >> MseI
    >> BcgI/a >> TaqI
                >> DraI
                >> AluI
                >> BcgI
    ACTCCAGCTT TCGATAAAAG TGCATTTACT AATTAAAGC AATTGCCTTT CTTTACTAT TCTGATAGTC
    19260      19270      19280      19290      19300      19310      19320

                >> PleI
                >> NlaIII
                >> BsmAI
                >> HinFI >> Alw26I
                >> SfaNI >>
                >> MaeII
                >> BsaAI >>
                >> AflIII >>
    CTTGTGAGTC TCATGGCAA CAAGTAGTGT CGGATATTGA TTATGTTCCA CTCAAATCTG CTACGTGTAT
    19330      19340      19350      19360      19370      19380      19390

                Zsp2I >
                >> ScaI
                PpulOI ><
                >> RsaINsiI >
                Mph1103I >
                >< SfaNIEcoT22I >
                > < RsaI >> Csp6I
                >< Csp6I
                >< AvaIII >>
                >> NlaIII > < AfaI >> AfaI
    TACACGATGC AATTAGGTG GTGCTGTTTG CAGACACCAT GCAAATGAGT ACCGACAGTA CTTGGATGCA
    19400      19410      19420      19430      19440      19450      19460

                >> FokI
    TATAATATGA TGATTTCTGC TGGATTAGC CTATGGATT ACAAACAATT TGATACTTAT AACCTGTGGA
    19470      19480      19490      19500      19510      19520      19530

                >> ScrFI
                >> MvaI
                >> MaeIII
                >> EcoRII
                >> Ecl136I
                >> DsaV
                >> BstOI
                >> BstNI
                >> BsiLI
                >> ApyI
                >> Tru9I
                >> MseI
    ATACATTTAC CAGGTTACAG AGTTAGAAA ATGTGGCTTA TAATGTTGTT AATAAAGGAC ACTTTGATGG
    19540      19550      19560      19570      19580      19590      19600

                >> SgrAI
                >> NaeI
                >> MspI
                >> HpaII
                >> HapII
                >> Cfr10I
                >> BspWI
                > < VspI
                > < Tru9I
                > < MseI
                > < AsnI
                > < AseI
    ACACGCCGGC GAAGCACCTG TTCCATCAT TAATAATGCT GTTACACAA AGGTAGATGG TATTGATGTG
    19610      19620      19630      19640      19650      19660      19670
    
```

FIGURE 13. 45


```

    << AccI
AAAGTAGACG GCATTATTCA ACAGTTGCCT GAAACCTACT TTA CTCAGAG CAGAGACTTA GAGGATTTTA
    20100      20110      20120      20130      20140      20150      20160

    << TthHB8I
    << TaqI
        << SstI
        << SduI
        << SacI
    > < PaeR7I
    > < NspIII
        << NspII
        << HgiAI
    > < Eco88I
    > < XhoI>< Eco24I
        << Ecl136II
    << XcmI
    << Sau3AI
    << NdeII
    << MboI
    << DpnII
    << DpnI
    << BspAI
    << Bsp143I
AGCCCAGATC ACAAATGGAA ACTGACTTTC TCGAGCTCGC TATGGATGAA TTCATACAGC GATATAAGCT
    20170      20180      20190      20200      20210      20220      20230

    << TthHB8I
    << TaqI
    << SfuI
    << NspV
    << LspI
    << Csp45I
    << BstBI
    << Bsp119I
    << BsiCI
    << Bpu14I
    << AsuII << BcgI
    << MboII
    << BbsI Tru9I <<
    << NlaIII << AciIMseI <<
CGAGGGCTAT GCCTTCGAAC ACATCGTTTA TGGAGATTTT AGTCATGGAC AACTTGGCGG TCTTCATTTA
    20240      20250      20260      20270      20280      20290      20300

    << HphI
    << HinPII
    << Hin6I
    << EspI > < HhaI << TfiI
    << DdeI << HaeII
    << CelII << Eco47III << Tru9I
    << Bpu1102I > < CfoI << HinfI << MseI
    << BfrI << Bsp143II << MnlI
ATGATAGGCT TAGCCAAGCG CTCACAAGAT TCACCACTTA AATTAGAGGA TTTTATCCCT ATGGACAGCA
    20310      20320      20330      20340      20350      20360      20370

    << MstI
    << HinPII
    << Hin6I
    << HhaI
    << FspI
    << FdiII
    << CfoI
    << SfaNI << AviII
    Sau3AI <<
    NdeII <<
    MboI <<
    DpnII <<
    DpnI <<
    BspAI <<
    Bsp143I <<
CAGTGAAAAA TTA CTTCATA ACAGATGCGC AAACAGGTTT ATCAAATGT GTGTGTCTCG TGATTGAYCT
    20380      20390      20400      20410      20420      20430      20440

    << TthHB8I

```

FIGURE 13.47

```

>< Tth111I
  >< TaqI
    >< AspI          > < MaeIII          MaeIII ><
TTFACTTGAT GACTTTGTCG AGATAATAAA GTCACAAGAT TTGTCAGTGA TTCAAAAAGT GGTC AAGGTT
  20450      20460      20470      20480      20490      20500      20510

                                >< NspI
                                >< NspHI
                                >< NlaIII
                                >< FokI

>< MunI          > < NlaIII          >< AflIII
ACAATTGACT ATGCTGAAAT TTCATTCATG CTTTGGTGTA AGGATGGACA TGTTGAAACC TTCTACCCAA
  20520      20530      20540      20550      20560      20570      20580

                                >< SfaNI
                                >< ScrFI
                                >< MvaI
                                >< EcoRII
                                >< Ecl136I
                                >< DsaV
                                >< BstOI          >< SfaNI
                                >< BstNI          >< RsaI   BspWI ><
                                >< BsiLI          > < Csp6I   BsmI >
                                >< ApyI          >< AfaI   BscCI ><
AACTACAAGC AAGTCAAGCG TGGCAACCAG GTGTTCGCAT GCCTAACTTG TACAAGATGC AAAGAATGCT
  20590      20600      20610      20620      20630      20640      20650

>< Eco57I >< MaeIII          >< HphI
TCTTGAAAAG TGTGACCTTC AGAATTATGG TGAAAATGCT GTTATACCAA AAGGAATAAT GATGAATGTC
  20660      20670      20680      20690      20700      20710      20720

                                > < RsaI
                                >< Csp6I

>< Bst1107I          >< Tru9I          >< AluI
>< AccI          >< MseI          > < AfaINlaIII ><
GCAAAGTATA CTCAACTGTG TCAATACTTA AATACACTTA CTTTAGCTGT ACCCTACAAC ATGAGAGTTA
  20730      20740      20750      20760      20770      20780      20790

                                >< ScrFI
                                >< RsaI
                                >< MvaI
                                >< EcoRII >< NspBII
                                >< Ecl136I >< SduI
                                > < Csp6I >< NspII
                                >< BstOI >< PvuII >< HgiAI
                                >< BstNI >< DdeI
                                >< BsiLI >< Psp5I >< Bsp1286I
                                >< ApyI >< AluI >< BmyI
                                >< DsaV >< AfaI >< Alw21I
TTCAC TTTGG TGCTGGCTCT GATAAAGGAG TTGCACCAGG TACAGCTGTG CTCAGACAAT GGTTGCCAAC
  20800      20810      20820      20830      20840      20850      20860

                                >< XhoII
                                >< Tru9I
                                >< Sau3AI
                                >< NdeII
>< TthHB8I >< MseI
>< MflI
>< MboI
>< MamI
>< DpnII
>< TfiI >< DpnI

```

FIGURE 13. 48


```

                >< Zsp2I
                >< SphI
                >< Ppu10I
                >< PaeI
                >< NspI
                >< NspHI
                >< NsiI
                >< NlaIII
                > < NlaIII
                >< Mph1103I
                >< EcoT22I
                > < AvaIII >< MnlI
GGGCTAACTA TCTTGGCAAG CCGAAGGAAC AAATTGATGG CTATACCATG CATGCTAACT ACATTTTCTG
 21220      21230      21240      21250      21260      21270      21280

                Tru9I ><
                >< MboII >< Tru9I
                >< GsuI >< MseI ><
                >< BsrI >< MseI ><
                >< BpmI >< MnlI ><
                >< BbsI >< NlaIII >< MnlI
GAGGAACACA AATCCTATCC AGTTGTCTTC CTATTCACCTC TTTGACATGA GCAAATTTCC TCTTAAATTA
 21290      21300      21310      21320      21330      21340      21350

                >< Tru9I
                >< MseI
                >< Esp4I> < TfiI
                >< BsmAI >< Ksp632I ><
                >< Alw26I >< MboII >< EarI
                >< AflII> < HinfI >< Eam1104I ><
AGAGGAACTG CTGTAATGTC TCTTAAGGAG AATCAAATCA ATGATATGAT TTATTCTCTT CTGGAAAAAG
 21360      21370      21380      21390      21400      21410      21420

                >< Tru9I
                >< MseI
                >< HindII
                >< HincII
                >< HpaI AflIII >
GTAGGCTTAT CATTAGAGAA AACAACAGAG TTGTGGTTTC AAGTGATATT CTTGTTAACA ACTAAACGAA
 21430      21440      21450      21460      21470      21480      21490

                >< VneI
                >< SnoI
                >< SduI
                >< NspII
                >< HpaII
                >< HgiAI
                >< HapII
                >< Cfr10I
                >< Bsp1286I
                >< MspI>< BmyI
                >< ApaLI
                >< Alw44I
                >< NspI >< SpeI >< ApaLI
                >< NspHI >< RmaI >< Alw44I
                >< NlaIII >< MaeI >< MaeIII >< AgeI >< Alw21I
CATGTTTATT TTCTTATTAT TTCTTACTCT CACTAGTGGT AGTGACCTTG ACCGGTGCAC CACTTTTGAT
 21500      21510      21520      21530      21540      21550      21560

                > < AluI >< MnlI
GATGTTCAAG CTCTAATTA CACTCAACAT ACTTCATCTA TGAGGGGGGT TTAATATCCT GATGAAATTT
 21570      21580      21590      21600      21610      21620      21630

                >< Sau3AI
    
```

FIGURE 13. 50


```

>> NdeII
>> MboI
>> DpnII
  >> DpnI           >> Tru9I
>> BspAI           >> MseI > < MboII
  >> Bsp143I       >> DdeI
TTAGATCAGA CACTCTTTAT TTAAGTCAGG ATTTATTTCT TCCATTTTAT TCTAATGTTA CAGGGTTTCA
21640      21650      21660      21670      21680      21690      21700

  >> VspI
  >> Tru9I
  >> MseI
  >> AsnI           >> Tru9I           >> FokI
  >> AseI >> MaeII >> MseI >> BbvI   > < Fnu4HI
TACTATTAAT CATACGTTTG GCAACCTGTG CATACCTTTT AAGGATGGTA TTTATTTTGC TGCCACAGAG
21710      21720      21730      21740      21750      21760      21770

                >> BslI
                >> DsaI >> BsiYI           >> NlaIII
                >> BsaJI           > < MaeIII
AAATCAAATG TTGTCCGTGG TTGGGTTTTT GGTCTACCA TGAACAACAA GTCACAGTGG GTGATTATTA
21780      21790      21800      21810      21820      21830      21840

                >> NspI
>> Tru9I           >> NspHI
>> MseI           >> NlaIII
>> HphI           >> MaeII           >> MaeIII
TTAACAATTC TACTAATGTT GTTATACGAG CATGTAACCT TGAATTGTGT GACAACCCTT TCTTTGCTGT
21850      21860      21870      21880      21890      21900      21910

  >> StyI           >> Zsp2I
    >> NlaIII       >> Tru9I
  >> NcoI >> RsaI   >> Ppu10I TthHB8I >>
  >> EcoT14I       >> NsiI           >> TaqI
  >> Eco130I       >> MseI           SfaNI >>
  >> DsaI >> Csp6I >> Mph1103I   RsaI >>
  >> BssT1I       >> TthHB8I >> EcoT22I   Csp6I >>
  >> BsaJI >> AfaI >> TaqI >> AfaIII   AfaI >>
TTCTAAACCC ATGGGTACAC AGACACATAC TATGATATTC GATAATGCAT TTAATTGCAC TTTTCGAGTAC
21920      21930      21940      21950      21960      21970      21980

                >> Tru9I
                >> MseI
                >> DraI
ATATCTGATG CCTTTTCGCT TGATGTTTCA GAAAAGTCAG GTAATTTTAA ACACCTTACGA GAGTTTGTGT
21990      22000      22010      22020      22030      22040      22050

                >> Sau3AI
                >> NdeII
                >> MboI
                >> DpnII
                >> DpnI
>> Tru9I           >> BspAI
>> MseI           >> SfcI           Bsp143I ><
>> DraI           >> SfcI           Bsp143I ><
TTAAAAATAA AGATGGGTTT CTCTATGTTT ATAAGGGCTA TCAACCTATA GATGTAGTTC GTGATCTACC
22060      22070      22080      22090      22100      22110      22120

                >> Tru9I
                > < Tru9I
                >> MseI
                >> MnlI
TTCTGGTTTT AACACTTTGA AACCTATTTT TAAGTTGCCT CTTGGTATTA ACATTACAAA TTTAGAGCC
22130      22140      22150      22160      22170      22180      22190

```

FIGURE 13.51

```

> < SduI>< SfcI
    >< PvuII
    >< Esp5I
> < NspII
    >< NspBII
> < MaeII > < Fnu4HI
> < Bspl286I >< PstI           Tru9I >
    >< BmyI>< Fnu4HI           MseI >
    >< BspMI
    >< HphI
    >< BbvI
    >< AluI
    >< BbvI
ATTCTTACAG CCTTTTCACC TGCTCAAGAC ATTTGGGGCA CGTCAGCTGC AGCCTATTTT GTTGGCTATT
22200      22210      22220      22230      22240      22250      22260

    >< SfaNI
    >< RsaI
> < Csp6I
    >< AfaI
    >< AlwNI
    >< DraI
TAAAGCCAAC TACATTTATG CTCAAGTATG ATGAAAATGG TACAATCACA GATGCTGTTG ATTGTTCTCA
22270      22280      22290      22300      22310      22320      22330

    > < Tru9I
    > < MseI
    >< AluI
AAATCCACTT GCTGAACTCA AATGCTCTGT TAAGAGCTTT GAGATTGACA AAGGAATTTA CCAGACCTCT
22340      22350      22360      22370      22380      22390      22400

    >< SauI
    >< MstII
    >< Eco81I
    >< DdeI
    >< CvnI
    >< Bsu36I
    >< Bse21I
    >< AxyI
    >< TfiI
    >< MnlI
    >< AocI
    >< MnlI
    >< HinfI
    >< SspI
    >< MnlI
AATTTCAGGG TTGTTCCCTC AGGAGATGTT GTGAGATTCC CTAATATTAC AAACCTGTGT CCTTTTGGAG
22410      22420      22430      22440      22450      22460      22470

    >< Zsp2I
    >< Ppu10I
    >< NsiI
    > < NlaIII
    >< Mph1103I
    >< EcoT22I
    >< Tru9I
    >< MseI
    >< AvaIII
AGGTTTTTAA TGCTACTAAA TTCCCTTCTG TCTATGCATG GGAGAGAAAA AAAATTTCTA ATTGTGTTGC
22480      22490      22500      22510      22520      22530      22540

    >< SduI
    >< NspII
    >< HgiAI
    >< Bspl286I
    >< BmyI
    >< Alw21I
    >< Tru9I
    >< MseI
    >< DdeI ><
TGATTACTCT GTGCTCTACA ACTCAACATT TTTTCAACC TTAAAGTGCT ATGGCGTTTC TGCCACTAAG
22550      22560      22570      22580      22590      22600      22610

    >< Sau3AI
    >< NdeII
    >< MboI
    >< DpnII
    >< DpnI

```

FIGURE 13.52

```

    >< BspAI
      >< Bsp143I
TTGAATGATC TTTGCTTCTC CAATGTCTAT GCAGATTCTT TTGTAGTCAA GGGAGATGAT GTAAGACAAA
22620      22630      22640      22650      22660      22670      22680

    >< ScrFI
    >< MvaI
>< HinPII
>< Hin6I
  >< HhaI
  >< HaeII
  >< EcoRII
  >< Ecl136I
  >< DsaV
  >< CfoI
  >< BstOI
  >< BstNI
  >< Bsp143II
  >< BsiLI
  >< ApyI      > < BsrI
TAGCGCCAGG ACAAACCTGGT GTTATTGCTG ATTATAATTA TAAATTGCCA GATGATTTC A TGGGTGTGT
22690      22700      22710      22720      22730      22740      22750

    >< SfaNI
    >< RmaI
    >< MaeI
CCTTGCTTGG AATACTAGGA ACATTGATGC TACTTCAACT GGTAATTATA ATTATAAATA TAGGTATCTT
22760      22770      22780      22790      22800      22810      22820

    >< Sau96I
    >< Pali
    >< NspIV
  > < HindIII
    >< HaeIII
    >< Eco0109I
    >< DraII
  >< DdeI
    >< Cfr13I
    >< BsuRI
    >< BsiZI
    >< BshI
  >< BfrI >< PssI
  >< NlaIII >< AsuI>< BsmAI
    >< AluI      >< Alw26I
AGACATGGCA AGCTTAGGCC CTTTGAGAGA GACATATCTA ATGTGCCTTT CTCCCCTGAT GGCAAACCTT
22830      22840      22850      22860      22870      22880      22890

    >< Tru9I
    >< Pali
    >< MscI
    >< HaeIII
  >< EaeI>< MseI
  >< Tru9I      >< BsuRI
  >< MseI      >< BshI
  >< BspMI     >< Ball
GCACCCACCC TGCTCTTAAT TGTTATTGGC CATTAAATGA TTATGGTTTT TACACCACTA CTGGCATTGG
22900      22910      22920      22930      22940      22950      22960

    >< Sau96I ><
    >< PalINspIV ><
  > < MspI NspHII ><
    >< HaeIII

```

FIGURE 13.53

```

                > < HpaII Eco47I ><
                >< DsaI
                > < HapII Cfr13I ><
                >< BsuRISinI ><
                >< GdiII BsiZI ><
                >< BsaJI
                >< ScaI
                >< RsaI
                >< Csp6I
                >< AfaI
                >< Tru9I >< EaeI Bme18I ><
                >< MseI >< Cfr10I AvaII ><
                >< DraI >< BshI AsuI ><
CTACCAACCT TACAGAGTTG TAGTACTTTC TTTTGAACCT TTAATGCAC CGGCCACGGT TTGTGGACCA
  22970      22980      22990      23000      23010      23020      23030

                >< Tru9I
                >< Tru9I >< RsaI
                >< PleI >< Csp6I
                >< MseI >< BsrI ><
                > < Tru9I
                > < MseI >< BsrI
                >< MseI >< HinfI >< AfaI
AAATTATCCA CTGACCTTAT TAAGAACCAG TGTGTCAATT TTAATTTTAA TGGACTCACT GGTACTGGTG
  23040      23050      23060      23070      23080      23090      23100

>< Tru9I
>< MseI
>< MboII
>< HpaI
>< HindII
>< HincII
                >< Pali
                >< HaeIII
                >< GdiII
                >< EaeI
                >< BsuRI
                >< BshI
                Tffi ><
                HinfI ><
TGTTAACTCC TTCTTCAAAG AGATTTCAAC CATTTCACA AATTGGCCGT GATGTTTCTG ATTTCACTGA
  23110      23120      23130      23140      23150      23160      23170

                > < XhoII
                >< TthHB8I
                >< TaqI
                > < Sau3AI
                > < NdeII
                > < MflI
                > < MboI
                > < DpnII
                >< DpnI
                > < BstYI
                > < BspAI
                > < SspI
                >< AlwI >< Bsp143I
                >< HphI
TCCGTTCTGA GATCCTAAAA CATCTGAAAT ATTAGACATT TCACCTTGCT CTTTTGGGGG TGTAAGTGTA
  23180      23190      23200      23210      23220      23230      23240

                >< ScrFI
                >< MvaI
                >< EcoRII
                >< Ecl136I
                >< DsaV
                >< BstOI
                >< BstNI
                >< BsiLI
                >< ApyI
                >< Tru9I
                >< MseI
                >< HpaI
                >< HindII
                >< Eco57I
                >< BsgI >< HincII
ATTACACCTG GAACAAATGC TTCATCTGAA GTTGCTGTTT TATATCAAGA TGTTAACTGC ACTGATGTTT
  23250      23260      23270      23280      23290      23300      23310

                >< Sau3AI
                >< NlaIII
                >< NdeII
                >< MboI
                >< DpnII
                >< DpnI
                >< HinpII

```

FIGURE 13. 54

```

                >> BspWI                >> Hin6I
                >> BspAI                > < HhaI                PleI >>
>< SfcI                >< BspI43I        >< AluI> < CfoI                >< BsrI
CTACAGCAAT TCATGCAGAT CAACTCACAC CAGCTTGGCG CATATATTCT ACTGGAAACA ATGTATTCCA
    23320         23330         23340         23350         23360         23370         23380

                >> TthHB8I
                >< TaqI
                >< SalI
                >< RtrI
                >< NspI
                >< EspI >< NspHI
                >< DdeI >< NlaIII
                >< CelII >< HindII
                >< Bpu102I>> HincII
>< HinfI                >< AluI >< AccI
GACTCAAGCA GGCTGTCTTA TAGGAGCTGA GCATGTCGAC ACTTCTTATG AGTGCGACAT TCCTATTGGA
    23390         23400         23410         23420         23430         23440         23450

                > < SnaBI
                >< ScaI
                >< RsaI
                >< RmaI
                >< MaeII >< MaeI
                > < Eco105I
                >< Csp6I
                > < BsaAI
                >< AfaI
>< AluI                >< MaeI
GCTGGCATTG GTGCTAGTTA CCATACAGTT TCTTTATTAC GTAGTACTAG CCAAAAATCT ATTGTGGCTT
    23460         23470         23480         23490         23500         23510         23520

                >> MunI
ATACTATGTC TTTAGGTGCT GATAGTCAA TTGCTTACTC TAATAACACC ATTGCTATAC CTACTAACTT
    23530         23540         23550         23560         23570         23580         23590

                RsaI >>
                >< MnlI
                Csp6I >>
                AfaI >>
>< SfcI
TTCAATTAGC ATTACTACAG AAGTAATGCC TGTTCCTATG GCTAAAACCT CCGTAGATTG TAATATGTAC
    23600         23610         23620         23630         23640         23650         23660

                > < TfiI
                > < HinfI
                >< AciI                > < AluI
ATCTGCGGAG ATTCTACTGA ATGTGCTAAT TTGCTTCFCC AATATGGTAG CTTTGCACA CAACTAAATC
    23670         23680         23690         23700         23710         23720         23730

>< VneI
    >< SduI
    >< NspII
    >< HqiAI
>< SnaI>> DdeI                >< Sau3AI                >< PmlI
    >< Bsp1286I                >< NdeII                >< PmaCI
    >< BmyI                    >< MboI                    >< MaeII
    >< BbvI                    >< DpnI                    >< Eco72I
    >< ApaI                    >> Bsp143I                >< BsaAI
    >< Alw44I                >< DpnII >> AlwI
    >< Alw21I                >> Fnu4HI                >< BspAI                >< AflIII
GTGCACTCTC AGGTATTGCT GCTGAACAGG ATCGCAACAC ACGTGAAGTG TTCGCTCAAG TCAAACAAAT
    23740         23750         23760         23770         23780         23790         23800

```

FIGURE 13.55

```

>< RsaI
>< Csp6I
>< AfaI
GTACAAAACC CCAACTTTGA AATATTTTGG TGGTTTAAAT TTTTCACAAA TATTACCTGA CCCTCTAAAG
23810 23820 23830 23840 23850 23860 23870

>< MnlI
>< MnlI
>< DdeI >< MnlI
>< Tru9I >< SfaNI >< HphI NlaIII ><
>< MseI >< MaeIII BspHI ><
CCAAC TAAGA GGTCTTTTAT TGAGGACTTG CTCITTAATA AGGTGACACT CGCTGATGCT GGCTTCATGA
23880 23890 23900 23910 23920 23930 23940

>< XhoII
>< Sau3AI
>< StyI >< RmaI
>< RmaI >< NdeII
>< MaeI >< MflI
>< EcoT14I >< MboI >< MstI
>< Eco130I >< MaeI >< HinPII
>< BssT1I >< VspI >< DpnII >< Hin6I
>< BsmI >< HphI >< DpnI >< HhaI
>< BscCI >< Tru9I >< BstYI >< FspI
>< BsaJI >< MseI >< BspAI >< FdiII
>< BlnI >< AsnI >< Bsp143I >< CfoI
>< AvrII >< AseI >< BglII >< AviII
AGCAATATGG CGAATGCCTA GGTGATATTA ATGCTAGAGA TCTCATTTGT GCGCAGAAGT TCAATGGACT
23950 23960 23970 23980 23990 24000 24010

>< RmaIRsaI ><
>< MnlI >< Fnu4HI >< Fnu4HI Csp6I ><
>< BspWI >< BbvI >< BbvI >< BspWI >< MaeIAfaI ><
TACAGTGTG CCACCTCTGC TCACTGATGA TATGATTGCT GCCTACACTG CTGCTCTAGT TAGTGGTACT
24020 24030 24040 24050 24060 24070 24080

>< MboII
>< HinPII
>< Hin6I
>< HhaI
>< HaeII
>< Fnu4HI >< Ksp632I
>< CfoI >< EarI
>< FokI >< BspWI >< Eam1104I
>< BbvI >< Bsp143II
GCCACTGCTG GATGGACATT TGGTGCTGGC GCTGCTCTTC AAATACCTTT TGCTATGCAA ATGGCATATA
24090 24100 24110 24120 24130 24140 24150

>< MaeIII
>< Tru9I ><
>< MseI ><
GGTTCAATGG CATTGGAGTT ACCCAAATG TTCTCTATGA GAACCAAAAA CAAATCGCCA ACCAATTTAA
24160 24170 24180 24190 24200 24210 24220

>< MaeII ><
>< TfiI >< Fnu4HI
>< HinfI >< BbvI >< AluI
CAAGGCGATT AGTCAAATC AAGAATCACT TACAACAACA TCAACTGCAT TGGGCAAGCT GCAAGACGTT
24230 24240 24250 24260 24270 24280 24290

>< Tru9I
>< MseI
>< HpaI
>< HindII >< BsmI >< Tru9I >< DdeI
>< HincII >< BscCI >< MseI >< Tru9I >< BfrI
>< MseI >< AluI

```

FIGURE 13. 56

```

GTTAACCAGA ATGCTCAAGC ATTAACACA CTTGTTAAAC AACTTAGCTC TAATTTGGT GCAATTTCAA
24300      24310      24320      24330      24340      24350      24360

      << Thal
      << SpoI
      << NruI
      << MvnI
      << BstUI          << TthHB8I
      << Bsp68I        << TaqI          << RsaI
      << EcoRV << Bsp50I << MnlI          << Csp6I          << Tru9I
      << Eco32I << AccII << MnlI        << AciI>< AfaI          << MseI
GTGTGCTAAA TGATATCCTT TCGCGACTTG ATAAAGTCGA GCGCGAGGTA CAAATTGACA GGTTAATTAC
24370      24380      24390      24400      24410      24420      24430

      << MaeIII << BbvI          << Fnu4HI BbvI >>
AGGCAGACTT CAAAGCCTTC AACCTATGT AACACAACAA CTAATCAGGG CTGCTGAAAT CAGGCCTTCT
24440      24450      24460      24470      24480      24490      24500

      << Fnu4HI
      << BspWI          << DdeI          << HindII
      << HincII
GCTAATCTTG CTGCTACTAA AATGTCTGAG TGTGTTCTTG GACAATCAAA AAGAGTTGAC TTTGTGGAA
24510      24520      24530      24540      24550      24560      24570

      > < NspI
      > < NspHI
      > < NlaIII
      << MaeIII
      << MaeII
      << FokI
      << BsaAI >>
      << Fnu4HI << BbsI
      << AciI>< BbvI          << AflIII
AGGGCTACCA CCTTATGTCC TTCCACAAG CAGCCCGCA TGGTGTGTC TTCCTACATG TCACGTATGT
24580      24590      24600      24610      24620      24630      24640

      << ScrFI
      << MvaI
      << EcoRII
      << Ecl136I
      << BstOI
      << BstNI          << HinPII
      << MnlI << BslI          << Hin6I
      << DsaV>< BsiYI          << HhaI
      << BsiLI          << HaeII
      << BsaJI>< HphI          << CfoI          << NlaIII
      << ApyI          << Bsp143II << BspHI          EcoNI >>
GCCATCCCAG GAGAGGAACT TCACCACAGC GCCAGCAATT TGTCATGAAG GCAAAGCATA CTTCCCTCGT
24650      24660      24670      24680      24690      24700      24710

      << MnlI
      << BslI          << Tru9I
      << BsiYI          << MseI          << MnlI
GAAGGTGTTT TTGTGTTTAA TGGCACTTCT TGGTTTATTA CACAGAGGAA CTTCTTTTCT CCACAAATAA
24720      24730      24740      24750      24760      24770      24780

      << DdeI          << Tru9I
      << BsmAI          << SfaNI
      << SfcI          << Alw26I          << MseIAlwI <<
TTACTACAGA CAATACATTT GTCTCAGGAA ATTGTGATGT CGTTATTGGC ATCATTAAACA ACACAGTTTA
24790      24800      24810      24820      24830      24840      24850

<< Sau3AI
<< NdeII

```

FIGURE 13.57

```

>< MboI           >< P1eI           > < ScaI
>< DpnII          >< MnlI           > < Ksp632I       > < RsaI
>< DpnI           >< DdeI   >< HinfI         >< MboII
>< BspAI          >< BspWI          > < Eam1104I     >< Csp6I
>< Bsp143I        >< AluI           > < EarI   > < AluI   > < AfaI   > < HphI
TGATCCTCTG CAACCTGAGC TTGACTCATT CAAAGAAGAG CTGGACAAGT ACTTCAAAAA TCATACATCA
24860      24870      24880      24890      24900      24910      24920

    >< Sau3AI
    >< NdeII
    >< MboI
>< MãmI
>< DpnII
    >< DpnI
    >< BspAI
    >< Bsp143I
    >< BsiBI           >< Tru9I           >< HindII
    >< BsaBI           >< MseI            >< HincII           AciI ><
CCAGATGTTG ATCTTGGCGA CATTTCAGGC ATTAACGCTT CTGTCGTCAA CATTCAAAAA GAAATTGACC
24930      24940      24950      24960      24970      24980      24990

                >< Tru9I
                > < TfiI
    >< MnlI           >< SwaI
>< EcoNI           >< MseI
    >< BslI           > < HinfI
>< MnlI>< BsiYI     >< DraI
GCCTCAATGA GGTTCGCTAAA AATTTAAATG AATCACTCAT TGACCTTCAA GAATTGGGAA AATATGAGCA
25000      25010      25020      25030      25040      25050      25060

    >< StyI
    >< Pali
    >< HaeIII
    >< EcoT14I
    >< Eco130I
    >< BsuRI
    >< BssTII
    >< Tru9I>< BshI           NlaIII ><
    >< MseI >< BsaJI           MaeIII ><
    ATATATTAATA TGGCCTTGGT ATGTTTGGCT CGGCTTCATT GCTGGACTAA TTGCCATCGT CATGGTTACA
25070      25080      25090      25100      25110      25120      25130

                > < SphI
                > < PaeI
                > < NspI
    >< SpeI           > < RmaI           > < NspHI
    >< NlaIII        > < NlaIII
    > < MaeI           >< MnlI>< BbvI Fnu4HI ><
ATCTTGCTTT GTTGCATGAC TAGTTGTTGC AGTTGCCTCA AGGGTGCATG CTCTTGTTGGT TCTTGCTGCA
25140      25150      25160      25170      25180      25190      25200

                >< FokI
                >< DdeI
>< MnlI >< P1eI>< HinfI >< BsrI
AGTTTGATGA GGATGACTCT GAGCCAGTTC TCAAGGGTGT CAAATTACAT TACACATAAA CGAACTTATG
25210      25220      25230      25240      25250      25260      25270

    >< Sau3AI
    >< NdeII
    >< MboI
    >< DpnII
    > < DpnI

```

FIGURE 13.58


```

                >< BspAI
                > < Bsp143I
                >< BsgI      >< AlwI      >< BsrI      BspWI >
GATTTGTTTA TGAGATTTT TACTCTTGA TCAATTACTG CACAGCCAGT AAAAATTGAC AATGCTTCTC
25280      25290      25300      25310      25320      25330      25340

                >< ScaI
                >< RsaI
                >< Csp6I      >< SfcI
                >< AfaI      >< NlaIII      >< AciI      >< MnlI      FokI >
CTGCAAGTAC TGTTTCATGCT ACAGCAACGA TACCGCTACA AGCCTCACTC CCTTTCGGAT GGCTTGTTAT
25350      25360      25370      25380      25390      25400      25410

                > < HinPII
                > < Hin6I
                >< HhaI      >> RmaI >>
                >> HaeII      >< HinPII      NheI ><
                >< Eco47III      >< Hin6I      MaeI ><
                >< CfoI      >< HhaI      Fnu4HI ><
                >> BspWI      >> Bsp143II      >< CfoI      AluI ><
TGCGTTCGCA TTTCTTGCTG TTTTTCAGAG CGTACCAAAA ATAATTGCGC TCAATAAAAG ATGGCAGCTA
25420      25430      25440      25450      25460      25470      25480

                >< EcoNI
                >< BslI
                >< BsiYI      >> MaeIII
                >< BbvI      >< BsrI      >< BbvI      > < Fnu4HI      BbvI ><
GCCCTTATA AGGGCTTCCA GTTCATTTGC AATTTACTGC TGCTATTTGT TACCATCTAT TCACATCTTT
25490      25500      25510      25520      25530      25540      25550

                >> Zsp2I >>
                >> Ppu10I >>
                > < SfcI      >< HinPII      NsiI ><
                >> PstI      >< Hin6I      >< RsaI      Mph1103I ><
                > < Fnu4HI      >< HhaI      >< Csp6I      EcoT22I ><
                >< BspMI      >< MnlI      >< CfoI      >< AfaI      >< MnlI      AvaIII ><
TGCTTGTTCG TGCAGGTATG GAGGCGCAAT TTTTGTACCT CTATGCCTTG ATATATTTTC TACAATGCAT
25560      25570      25580      25590      25600      25610      25620

                >< SfaNI
                >< NspI
                >< NspHI
                >< NlaIII      >> SfaNI
CAACGCATGT AGAATTATTA TGAGATGTTG GCTTTGTTGG AAGTGCAAAT CCAAGAACCC ATTACTTTAT
25630      25640      25650      25660      25670      25680      25690

                >> Bst1107I
                >> AccI      MaeIII >>
GATGCCAACT ACTTTGTTG CTGGCACACA CATAACTATG ACTACTGTAT ACCATATAAC AGTGTCCACAG
25700      25710      25720      25730      25740      25750      25760

                >> MboII
                >> BstXI >>
                >> HphI      >> Eco57I      >> BbsI MnlI >
                >< MunI >< MaeIII >< MaeIII      >> Eco57I      >> BbsI MnlI >
ATACAATTGT CGTTACTGAA GGTGACGGCA TTTCAACACC AAAACTCAAA GAAGACTACC AAATTGGTGG
25770      25780      25790      25800      25810      25820      25830

                >> RsaI
                >> < NlaIII
                >> HphI
                >> Tru9I >< Tth111I >< Csp6I
                >> DdeI      >> DdeI      >< MseI >< AspI      >< AfaI

```

FIGURE 13.59

```

TATTCTGAG GATAGGCACT CAGGTGTAA AGACTATGTC GTTGACATG GCTATTTAC CGAAGTTTAC
25840      25850      25860      25870      25880      25890      25900

      > < HinfI>< P1eI          >> BsrI          Tru9I ><
      >> AluI >< AccI          >> SfcI >< AlwNI          MseI ><
TACCAGCTTG AGTCTACACA AATTACTACA GACACTGGTA TTGAAAATGC TACATTCTTC ATCTTTAACA
25910      25920      25930      25940      25950      25960      25970

      >> Tru9I          >< TthHB8I
      >> MseI          >< TaqI          >< Ksp632I
>< AluI          >> Eco57I          >< EarI BspWI ><
AGCTTGTTAA AGACCCACCG AATGTGCAAA TACACACAAT CGACGGCTCT TCAGGAGTTG CTAATCCAGC
25980      25990      26000      26010      26020      26030      26040

>< XhoII
>< Sau3AI
>< NlaIV
>< NdeII
>< MflI
>< MboI
>< DpnII
>< DpnI
>< BstYI
>< BstI
>< BspAI
>< Bsp143I
>< BscBI
>< BamHI >< AlwI
AATGGATCCA ATTTATGATG AGCCGACGAC GACTACTAGC GTGCCTTTGT AAGCACAAGA AAGTGAGTAC
26050      26060      26070      26080      26090      26100      26110

      >< Tru9I
      >> RsaI
      >< MseI
      >< MboII
      >> MaeII
      >< RsaI
      >> Csp6I          >< Tru9I >< Csp6I
      >< AfaI          >> AfaI          >< MseI >< AfaI
GAACTTATGT ACTCATTCTG TTCGGAAGAA ACAGGTACGT TAATAGTTAA TAGCGTACTT CTTTTTCTTG
26120      26130      26140      26150      26160      26170      26180

      >< TthHB8I
      >< TaqI
      >> RmaI          >> HinPII          >< RsaI
      >< MaeIII          >> Hin6I          Fnu4HI ><
      >< MaeI >> RmaI          >> HhaI          >< Csp6I
      >< FokI >> MaeI          >> CfoI >> BbvI >< AfaI
CTTTCGTGGT ATTCTTGCTA GTCACACTAG CCATCCTTAC TGCCTTCGA TTGTGTGCGT ACTGCTGCAA
26190      26200      26210      26220      26230      26240      26250

      >> Tru9I
      >> MseI
>< SspI >< MaeII
      >> HpaI          >> BstUI          Ksp632I >
      >> HindII          >> MaeII >< Bsp50I >< MboII EarI >
      >> HincII          >> AccI >> AccII          Eam1104I >
TATTGTTAAC GTGAGTTTAG TAAAACCAAC GGTTCACGTC TACTCGCGTG TTAAAAATCT GAATCTTCT
26260      26270      26280      26290      26300      26310      26320

```

FIGURE 13.60

```

>> Sau3AI
>> NdeII
>> MboI
>> DpnII
>> MboII>> DpnI
>> XmnI >> BspAI> < Eco57I >> Tru9I
>> Asp700I>< Bsp143I >> MseI
GAAGGAGTTC CTGATCTTCT GGTCTAAACG AACTAACTAT TATTATTATT CTGTTTGAA CTTTAACATT
26330 26340 26350 26360 26370 26380 26390

>> ScrFI
>> MvaI
>> EcoRII
>> Ecl136I
>> DsaV NlaIV >>
>> RsaI >> BstOI
>> MnlI >> Tru9I >> BstNI RmaI >>
>> Csp6I >> MseI >> BsiLI MaeI >>
> < NlaIII >> AfaI > < AluI >> ApyIBscBI >>
GCTTATCATG GCAGACAACG GTACTATTAC CGTTGAGGAG CTAAACAAC TCCTGGAACA ATGGAACCTA
26400 26410 26420 26430 26440 26450 26460

>> ScrFI
>> RmaI
>> MvaI
>> MaeI
>> EcoRII
>> Ecl136I
>> DsaV
>> BstOI
>> BstNI
>> BsiLI
>> ApyI >> MaeIII
GTAATAGGTT TCCTATTCCT AGCCTGGATT ATGTTACTAC AATTTGCCTA TTCTAATCGG AACAGGTTTT
26470 26480 26490 26500 26510 26520 26530

>> Pali
>> MscI
>> MnlI >> MaeIII
>> HaeIII
>> EaeI
>> BsuRI
>> BsrI
>> BspWI
>> BshI
>> BspWI
>> BshI
>> BsiLI
>> BsiLI
>> BbvI Fnu4HI >>
TGTACATAAT AAAGCTTGTT TTCCTCTGGC TCTTGCGCC AGTAACACTT GCTTGTTTTG TGCTTGCTGC
26540 26550 26560 26570 26580 26590 26600

>> VspI
>> Tru9I
>> MseI >> HphI
>> SfcI >> AsnI >> BsrI
>> AccI >> AseI>> MaeIII>> AciI
TGTCTACAGA ATTAATTGGG TGACTGGCGG GATTGCGATT GCAATGGCTT GTATTGTAGG CTTGATGTGG
26610 26620 26630 26640 26650 26660 26670

>> EspI
>> Eco57I
>> DdeI
>> CelII >> RsaI
>> Bpu1102I >> Csp6I

```

FIGURE 13.61

```

>< BfrI
  >< AluI
CTTAGCTACT TCGTTGCTTC CTCAGGCTG TTTGCTCGTA CCCGCTCAAT GTGGTCATTC AACCCAGAAA
  26680      26690      26700      26710      26720      26730      26740

  >< AfaI
    >< AciI
      MboII >
    >< ScrFI
    >< NciI
    >< MspI
    >< HpaII
    >< HapII
    >< DsaV>< MnlI
    >< BslI
    >< BsiYI
    >< BsaJI >< MunI      > < XcmI
    >< BcnI      >< MaeIII >< AciI >< NlaIII
CAAACATTCT TCTCAATGTG CCTCTCCGGG GGACAATTGT GACCAGACCG CTCATGGAAA GTGAACTTGT
  26750      26760      26770      26780      26790      26800      26810

  Tru9I ><
    SinI >
    Sau96I >
    PpuMI >
    NspIV >
    MseI ><
    >< MaeIII
  >< Sau3AI
  >< NdeII
  >< MboI
  >< FbaI
  >< DpnII
  >< DpnI
  >< BspAI
  >< Bsp143I
  >< BsiQI
  >< BclI
  >< BspMI
  >< Psp5II
  >< NspHII
  GACCTGCCAA AAGAGATCAC TGTGGCTACA TCACGAACGC TTTCTTATTA CAAATTAGGA GCGTCGCAGC
  26820      26830      26840      26850      26860      26870      26880

  >< BclI
  >< MaeIII
  >< BshI
  >< AvrII >< CfoI AsuI >
CATTGGTGCT GTGATCATTG GTGGTCACTT GCGAATGGCC GGACACTCCC TAGGGCGCTG TGACATTAAG
  26820      26830      26840      26850      26860      26870      26880

  >< Sau3AI
  >< NdeII
  >< MboI
  >< DpnII
  >< DpnI
  >< PssI >< BspMI
  >< Psp5II
  >< NspHII
  >< BspAI
  >< Bsp143I
  >< XmnI
  >< Asp700I > < HgaI Fnu4HI ><
GACCTGCCAA AAGAGATCAC TGTGGCTACA TCACGAACGC TTTCTTATTA CAAATTAGGA GCGTCGCAGC
  26890      26900      26910      26920      26930      26940      26950

  >< TfiI
  >< HinfI
  >< BbvI
  >< BbvI
  >< Fnu4HI >< AciI
  >< Tru9I
  >< MseI
GTGTAGGCAC TGATTCAGGT TTTGCTGCAT ACAACCGCTA CCGTATTGGA AACTATAAAT TAAATACAGA
  26960      26970      26980      26990      27000      27010      27020

  >< MspI
  >< HpaII
  >< HapII
  >< Cfr10I
  >< BcgI/a
  >< RsaI
  >< RmaI
  >< Csp6I
  >< MaeI>< BcgI
  >< AfaI >< MaeIII
  HindII ><
  HincII ><
  >< SspI

```

FIGURE 13.62

```

CCACGCCGGT AGCAACGACA ATATTGCTTT GCTAGTACAG TAAGTGACAA CAGATGTTTC ATCTTGTGTA
 27030      27040      27050      27060      27070      27080      27090

  << ScrFI
  << MvaI
    << MaeIII
  << EcoRII
    << Ecl136I
  << DsaV
    << BstOI
    << BstNI
    << BsiLI
    << ApyI
                                << MnlI
                                HinFI ><
CTTCCAGGTT ACAATAGCAG AGATATTGAT TATCATTATG AGGACTTTCA GGATTGCTAT TTGGAACTTT
 27100      27110      27120      27130      27140      27150      27160

    << MaeII
    << BsmAI
    << Alw26I
    << Tru9I
    > < MnlI
    << MseI
    << DdeI
    << MboII
GACGTTATAA TAAGTTCAAT AGTGAGACAA TTATTTAAGC CTCTAACTAA GAAGAATTAT TCGGAGTTAG
 27170      27180      27190      27200      27210      27220      27230

                                << Ksp632I
                                << MboII
                                << EarI
    << MboII
    << NlaIII Eam1104I ><
ATGATGAAGA ACCTATGGAG TTAGATTATC CATAAACGA ACATGAAAAT TATTCTCTTC CTGACATTGA
 27240      27250      27260      27270      27280      27290      27300

                                > < RsaI >< RsaI
                                << Csp6I >< Csp6I
    > < AluI
    << MnlI
    > < AfaI >< AfaI
TTGTATTTAC ATCTTGCGAG CTATATCACT ATCAGGAGTG TGTTAGAGGT ACGACTGTAC TACTAAAAGA
 27310      27320      27330      27340      27350      27360      27370

    << MnlI
    << HphI
    << HphI
    << MnlI
ACCTTGCCCA TCAGGAACAT ACGAGGGCAA TTCACCATTT CAGCCTCTTG CTGACAATAA ATTTGCACTA
 27380      27390      27400      27410      27420      27430      27440

                                Sau3AI >
                                > < PvuII
                                > < Psp5I
                                > < NspBII
                                << TthHB8I
                                << TaqI
                                << RsaI
                                << Csp6I
                                << BbvI
    << RmaI
    << MaeI
    << AfaI
    > < AluI
ACTTGCCTA GCACACACTT TGCTTTTGCT TGTGCTGACG GTACTCGACA TACCTATCAG CTGCGTGCAA
 27450      27460      27470      27480      27490      27500      27510

                                << SstI
                                << SduI
                                << SacI
                                << NspII
                                << HgiAI
                                << Eco24I
    > < Ecl136II
                                << BspWI
                                << Bsp1286I
                                << BmyI
                                << BanII
                                << Alw21I

<< HphI
<< DpnI
                                << MnlI

```

FIGURE 13. 63

```

>< Bsp143I          >< MnlI          > < AluI    BbvI ><
GATCAGTTTC ACCAAAACCTT TTCATCAGAC AAGAGGAGGT TCAACAAGAG CTCTACTCGC CACTTTTTTCT
  27520      27530      27540      27550      27560      27570      27580

                                     SstI ><
                                     SduI ><
                                     SacI ><
                                     NspII ><
                                     HgiAI ><
                                     Eco24I ><
                                     Ecl136II ><
                                     Bsp1286I ><
                                     BmyI ><
                                     BanII ><
                                     Alw21I ><
                                     AluI ><
>< RmaI    >< Tru9I
>< MaeI    >< MseI          >< Tru9I
>< Fnu4HI  >< HphI          >< MseI
CATTGTTGCT GCTCTAGTAT TTTTAATACT TTGCTTCACC ATTAAGAGAA AGACAGAATG AATGAGCTCA
  27590      27600      27610      27620      27630      27640      27650

>< Tru9I          >< Tru9I
>< MseI          >< MseI
CTTTAATTGA CTTCTATTGG TGCTTTTTAG CCTTCTGCT ATTCCTTGTT TTAATAATGC TTATTATATT
  27660      27670      27680      27690      27700      27710      27720

>< XhoII
>< XbaI
> < ScrFI
>< Sau3AI
>< RmaI
>< NdeII
> < MvaI
>< MflI
>< MboI
>< EcoRII>< MaeI
> < Ecl136I
>< DpnII
>< DpnI
>< BstYI
> < BstOI
> < BstNI
>< TthHB8I >< BspAI          > < RsaI
>< DsaV>< Bsp143I          >< MboII
> < BsiLI          >< Csp6I
>< TaqI > < ApyI > < AlwI > < AfaI          >< NlaIII
TTGGTTTTCA CTCGAAATCC AGGATCTAGA AGAACCTGT ACCAAAGTCT AACGAACAT GAAACTTCTC
  27730      27740      27750      27760      27770      27780      27790

                                     >< HinP1I
                                     >< Hin6I
                                     >< HhaI
                                     >< RsaI >< HaeII
>< SfcI          >< Eco47III
>< Csp6I>< CfoI SfaNI ><
>< AfaI >< Bsp143II
ATTGTTTTGA CTTGTATTTT TCTATGCAGT TGCATATGCA CTGTAGTACA GCGCTGTGCA TCTAATAAAC
  27800      27810      27820      27830      27840      27850      27860

>< XhoII
>< Sau3AI
>< NdeII
> < MnlI
>< MflI

```

FIGURE 13.64

```

    << MboI
    << DpnII
      << DpnI      << RsaI
    << BstYI    << MboII
  << NlaIII>< BspAI    << Csp6I >< RmaI
    << AlwI >< Bsp143I  << AfaI >< MaeI
CTCATGTGCT TGAAGATCCT TGTAAGGTAC AACACTAGGG GTAATACTTA TAGCACTGCT TGGCTTTGTG
  27870      27880      27890      27900      27910      27920      27930

<< SduI
  << RmaI
  << NspII
  << MaeI
  << HgiAI
  << Bsp1286I
  << BmyI
  << Alw21I
                                << NspI
                                << NspHI
                                << NlaIII >< MaeIII
CTCTAGGAAA GGTTTTACCT TTTTCATAGAT GGCACACTAT GGTTCAAACA TGCACACCTA ATGTTACTAT
  27940      27950      27960      27970      27980      27990      28000

  > < XhoII
  > < Sau3AI > < Van9II
    << PvuII
    << Psp5I
  > < NdeII > < PflMI
  > < MflI>< NspBII
  > < DpnII      << HinPII
    << Bsp143I    << Hin6I
  > < BstYI > < BslI >< HhaI >< RmaI >< Asp718 >< Eco9II
  > < BspAI > < BsiYI>< CfoI >< MaeI >< AfaI >< BstPI
  > < MboI>< AluI>< BspWI >< BspWI >< AccBII >< BstEII
<< AlwI >< DpnI > < AccB7I >< AluI >< Acc65I >< BbvI
CAACTGTCAA GATCCAGCTG GTGGTGCCT TATAGCTAGG TGTTGGTACC TTCATGAAGG TCACCAAAC
  28010      28020      28030      28040      28050      28060      28070

                                << SinI
                                << Sau96I
                                << NspIV
                                NspHII <<
                                NlaIV ><
                                << Eco47I
                                << Cfr13I
                                << BsiZI
                                BscBI ><
                                << Bme18I
                                << AvaII
                                << AsuI
  << Fnu4HI >< MaeII
    << Esp3I >< Csp6I >< Tru9I
    << BsmAI >< BsmBI >< MseI >< Tru9I
    << Alw26I >< AfaI >< DraI >< MseI
GCTGCATTTA GAGACGTACT TGTTGTTTAA AATAAACGAA CAAATTAATA TGTCTGATAA TGGACCCCAA
  28080      28090      28100      28110      28120      28130      28140

                                << SinI
                                << Sau96I
                                << NspIV
                                << NspHII
                                << NlaIV
                                << Eco47I
                                << Cfr13I
                                << BsiZI
                                << BscBI
    << SduI
    << NspII
    << Bsp1286I
    << BmyI
  << MaeII >< AciI
                                << AvaII >< TfiI
                                << AsuI >< HinfI
                                >> MnlI

```

FIGURE 13. 65

```

TCAAACCAAC GTAGTGCCCC CCGCATTACA TTTGGTGGAC CCACAGATTC AACTGACAAT AACCAGAATG
28150      28160      28170      28180      28190      28200      28210

      << HinFII >> StyI
      << HaeII
      << Pali >> Hin6I >> EcoT14I
      << HaeIII >> HhaI >> Eco130I
      << BspWI >> BssTII
      << BsuRI >> Bsp143II
      << HgaI >> BshI >> CfoI >> BsaJI >> HgaI
GAGGACGCAA TGGGCAAGG CCAAAACAGC GCCGACCCCA AGGTTTACCC AATAATACTG CGTCTTGGTT
28220      28230      28240      28250      28260      28270      28280

      << TthHB8I
      << ScrFI
      << Pali
      << PaeR7I
      << NspIII
      << MvaI
      << HaeIII
      << EcoRII
      << Eco88I
      << XhoI >> Eco1136I
      << DsaV
      << BsuRI
      << SlaI >> BstOI
      << MnlI >> TaqI >> BstNI
      << CcrI >> BsiLI
      << HinfI >> BshI
      << TfiI >> BcoI >> BsaJI
      << MnlI >> DdeI >> Aval >> ApyI
      << AluI >> DdeI >> NlaIII >> BfrI >> Ama87I >> MnlI
CACAGCTCTC ACTCAGCATG GCAAGGAGGA ACTTAGATTC CCTCGAGGCC AGGGCGTTCC AATCAACACC
28290      28300      28310      28320      28330      28340      28350

      << SinI
      << Sau96I
      << NspIV
      << NspHII
      << Eco47I
      << Cfr13I
      << BsiZI
      << Bme18I >> Ksp632I
      << AvaII >> Eam1104I
      << AsuI >> EarI >> AluI >> MboII >> MaeIII
AATAGTGGTC CAGATGACCA AATGGCTAC TACCGAAGAG CTACCGACG AGTTCGTGGT GGTGACGGCA
28360      28370      28380      28390      28400      28410      28420

      << SstI
      << SduI
      << SacI
      << NspII
      << HgiAI
      << EspI
      << Eco24I >> Sau96I
      << Eco1136II >> StyI >> Pali
      << DdeI >> RmaI >> NspIV
      << CclII >> MaeI >> HaeIII
      << Bsp1286I >> EcoT14I >> Cfr13I
      << Bpu1102I >> Eco130I >> BsuRI
      << BmyI >> BssTII >> BsrI
      << BanII >> RsaI >> BsaJI >> Bsi2I

```

FIGURE 13.66


```

    >> Alw21I    >> Csp6I    >> BlnI    >> BshI>> HindIII
>> HphI >> AluI    >> AfaI    >> AvrII    >> AsuI    >> AluI
AAATGAAAGA GCTCAGCCCC AGATGGTACT TCTATTACCT AGGAACTGGC CCAGAAGCTT CACTTCCCTA
28430      28440      28450      28460      28470      28480      28490

>> HinPII
>> Hin6I
>> HhaI
>> HaeII
>> CfoI
>> Bsp143II
CGGCCTAAC AAAGAAGGCA TCGTATGGGT TGCAACTGAG GGAGCCTTGA ATACACCCAA AGACCACATT
28500      28510      28520      28530      28540      28550      28560

>> NlaIV
>> Eco64I
>> BscBI
>> BanI
>> AciI
>> AccBII >> BbvI    >> Fnu4HI    >> MnlI
GGCACCCGCA ATCCTAATAA CAATGCTGCC ACCGTGCTAC AACTTCCTCA AGGAACAACA TTGCCAAAAG
28570      28580      28590      28600      28610      28620      28630

>> MnlI >> MnlI >> Fnu4HI >> Ksp632I >> EarI >> Eam1104I
GCTTCTACGC AGAGGGGAGC AGAGGCGGCA GTC AAGCCTC TTCTCGCTCC TCATCACGTA GTCGGGTAA
28640      28650      28660      28670      28680      28690      28700

>> ScrFI
>> MvaI
>> EcoRII
>> Ecl136I
>> DsaV>> Fnu4HI
>> BstOI
>> BstNI
>> BsiLI
>> ApyI
TCAAGAAAT TCAACTCCTG GCAGCAGTAG GGGAAATTCT CCTGCTCGAA TGGCTAGCGG AGGTGGTGAA
28710      28720      28730      28740      28750      28760      28770

>> ThaI
>> MvnI
>> HphI >> MnlI
>> HinPII
>> Hin6I
>> HhaI
>> BstUI >> RmaI
>> Bsp50I >> MaeI
>> BbvI >> CfoI>> Fnu4HI
>> AccII>> BspWI >> AluI
ACTGCCCTCG CGCTATTGCT GCTAGACAGA TTGAACCAGC TTGAGAGCAA AGTTTCTGGT AAAGGCCAAC
28780      28790      28800      28810      28820      28830      28840

>> PalI>> MaeIII
>> HaeIII
>> BsuRI >> DdeI
>> Fnu4HI
>> DdeI
RsaI >>
>> MnlI
MaeII >>
Csp6I >>

```

FIGURE 13.67

```

> < BshI > < BbvI >> MnlI >> BspWI >> SfaNI AfaI ><
AACAAACAAGG CCAAAGTGTG ACTAAGAAAT CTGCTGCTGA GGCATCTAAA AAGCCTCGCC AAAAACGTAC
28850 28860 28870 28880 28890 28900 28910

>> Tth111I
>> SinI
>> Sau96I
>> NspIV
>> NspHII
> < MaeII
>> Eco47I
>> Cfr13I
>> BsmBI
>> RsaI >> BsiZI >> StyI
>> MaeIII >> Bme18I >> EcoT14I
>> MaeII >> Esp3I >> AvaII >> Eco130I
>> Csp6I >> BsmAI >> AsuI >> BssT1I
>> AfaI >> Alw26I> < AspI >> BsaJI
TGCCACAAAA CAGTACAACG TCACTCAAGC ATTTGGGAGA CGTGGTCCAG AACAAACCA AGGAAATTC
28920 28930 28940 28950 28960 28970 28980

>> SinI
>> Sau96I
>> NspIV
>> NspHII
>> NlaIV >> Pali
>> Eco47I >> HaeIII
>> Cfr13I >> GdiII
>> BsiZI >> Fnu4HI
>> BscBI >> EaeI
>> Bme18I >> BsuRI
>> AvaII >> BshI >> BspWI >
>> AsuI >> AciI >> BspWI
GGGGACCAAG ACCTAATCAG ACAAGGAAGT GATTACAAAC ATTGGCCGCA AATTGCACAA TTTGCTCCAA
28990 29000 29010 29020 29030 29040 29050

>> BsmI >> NlaIII
>> BscCI >> MnlI >> MaeIII >> MaeIII >> NlaIII
GTGCCTCTGC ATTCTTTGGA ATGTCACGCA TTGGCATGGA AGTCACACCT TCGGGAACAT GECTGACTTA
29060 29070 29080 29090 29100 29110 29120

>> XhoII
>> Sau3AI
>> NdeII
>> MflI
>> MboI
>> FokI
>> Tru9I >> DpnII
>> NlaIV >> DpnI
>> NlaIII >> BstYI >> Tth111I
>> MseI >> BspAI >> MaeII
>> BscBI >> BstXI>> AlwI> < Bsp143I >> AspI BspWI ><
TCATGGAGCC ATTAATTGG ATGACAAAGA TCCACAATTC AAAGACAACG TCATACTGCT GAACAAGCAC
29130 29140 29150 29160 29170 29180 29190

EspI ><
DdeI ><
CclII ><
Bpu1102I ><
AluI ><
>> HgaI
ATTGACGCAT ACAAACATT CCCACCAACA GAGCCTAAAA AGGACAAAAA GAAAAAGACT GATGAAGCTC
29200 29210 29220 29230 29240 29250 29260

```

FIGURE 13.68

```

                >> PleI
    >> Fnu4HI          >> MboII
    >> BspWI           >> MboII >> Ksp632I >> GsuI
    >> BsmAI           >> MaeIII >> EarI >> Fnu4HI
    >> Alw26I         >> HinfI >> Eam1104I >> BpmI
    >> AciI           >> Fnu4HI >> BbvI >> AciI >> NlaIII
AGCCTTTGCC GCAGAGACAA AAGAAGCAGC CCACTGTGAC TCTTCTCCT GCGGCTGACA TGGATGATTT
29270      29280      29290      29300      29310      29320      29330

                >> NlaIII >> HinfI NlaIII >>
    >> FokI           >> AluI >> TfiI >> DdeI >> BspHI
CTCCAGACAA CTTCAA AATT CCATGAGTGG AGCTTCTGCT GATTCAACTC AGGCATAAAC ACTCATGATG
29340      29350      29360      29370      29380      29390      29400

                >> MaeII >> AccI
ACCACACAAG GCAGATGGGC TATGTAAACG TTTTCGCAAT TCCGTTTACG ATACATAGTC TACTCTGTG
29410      29420      29430      29440      29450      29460      29470

                >> Tru9I
                >> Tru9I
                >> MseI
    >> XmnI           >> HpaI
    >> EcoRI >> MaeIII >> HindII Tru9I >>
    >> Asp700I >> BsgI >> HincII MseI >>
CAGAATGAAT TCTCGTAACT AACAGCACA AGTAGGTTTA GTTAACTTTA ATCTCACATA GCAATCTTTA
29480      29490      29500      29510      29520      29530      29540

                XorII >
                TthHB8I >
                TaqI >
                Sau3AI >>
                RsaI >>
    >> ThaIPvuI >
                NdeII >>
                >> MnlI
    >> MvnIMcrI >
                MboI >>
                DpnII >>
                DpnI >>
                Csp6I >>
                >> BstUI
                >> HaeIII BspCI >
                BspAI >>
    >> TthHB8I >> Bsp50I
                >> Pali Bspl43I >>
                >> BsuRI BsiEI >
                >> BshIAfaI >>
                >> TaqI >> AciI
    >> MnlI           >> MnlI >> AccII
    >> MaeIII
ATCAATGTGT AACATTAGGG AGGACTTGAA AGAGCCACCA CATTTCATC GAGGCCACGC GGAGTACGAT
29550      29560      29570      29580      29590      29600      29610

                >> SduI
                >> NspII
                >> MboII >> VspI
    >> RsaI           >> RmaI >> Fnu4HI >> Eco24I >> Tru9I
    >> Csp6I         >> MaeI >> EarI >> Bsp1286I >> MseI
    >> AfaI         >> BbvI > < AluI >> Eam1104I >> BanII >> AseI

```

FIGURE 13.69

```
CGAGGGTACA GTGAATAATG CTAGGGAGAG CTGCCTATAT GGAAGAGCCC TAATGTGTAA AATTAATTTT
 29620      29630      29640      29650      29660      29670      29680

                >< Tru9I   >< DdeI
                >< MseI   >< BfrI
                >< NlaIII  > < AluI
AGTAGTGCTA TCCCATGTG ATTTTAATAG CTTCTTAGGA GAATGACAAA AAAAAAAAAA AAAAAA
 29690      29700      29710      29720      29730      29740
```

FIGURE 13. 70

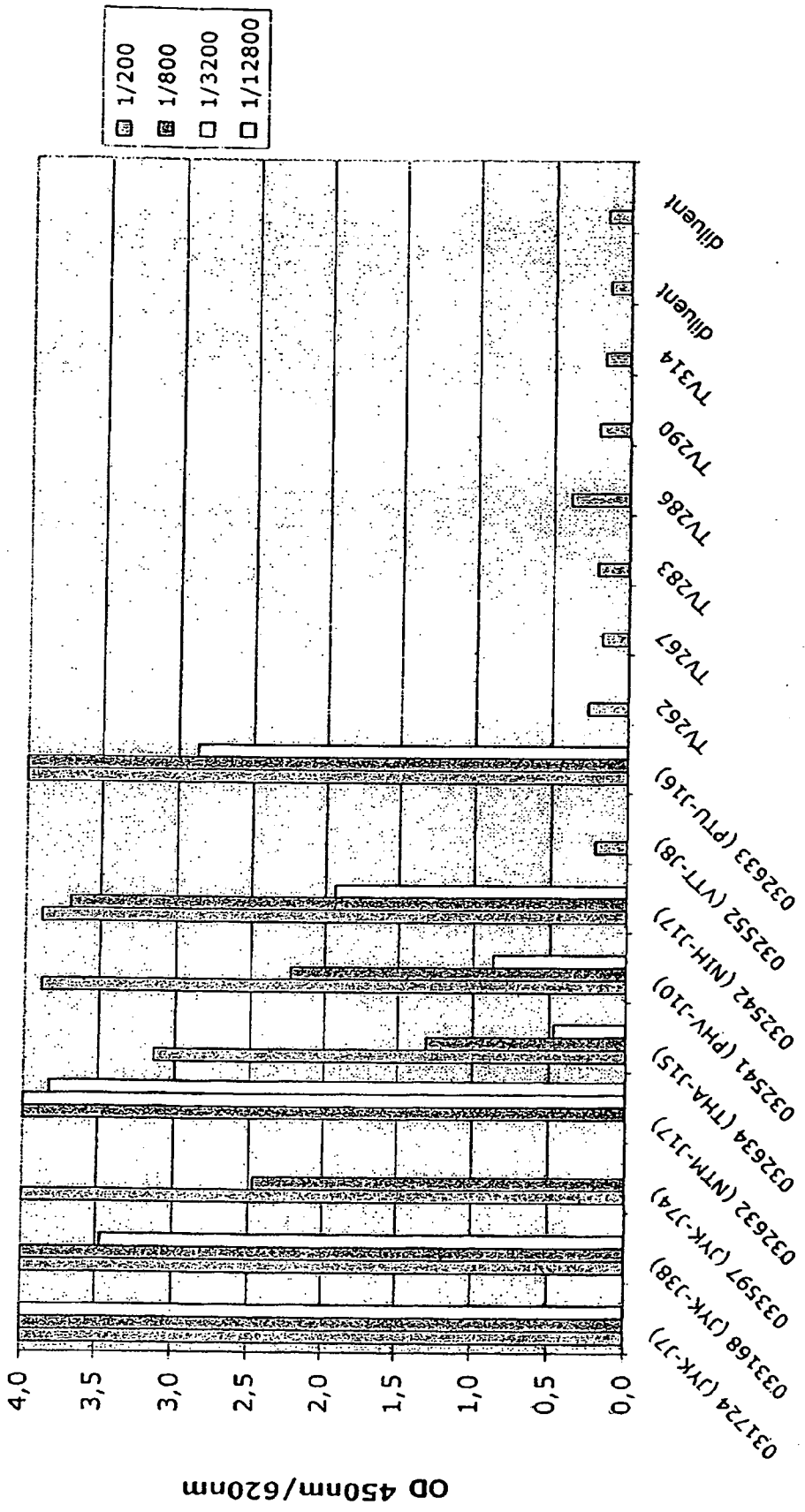


FIGURE 14

SRAS serology: Indirect N Technique (Second set)

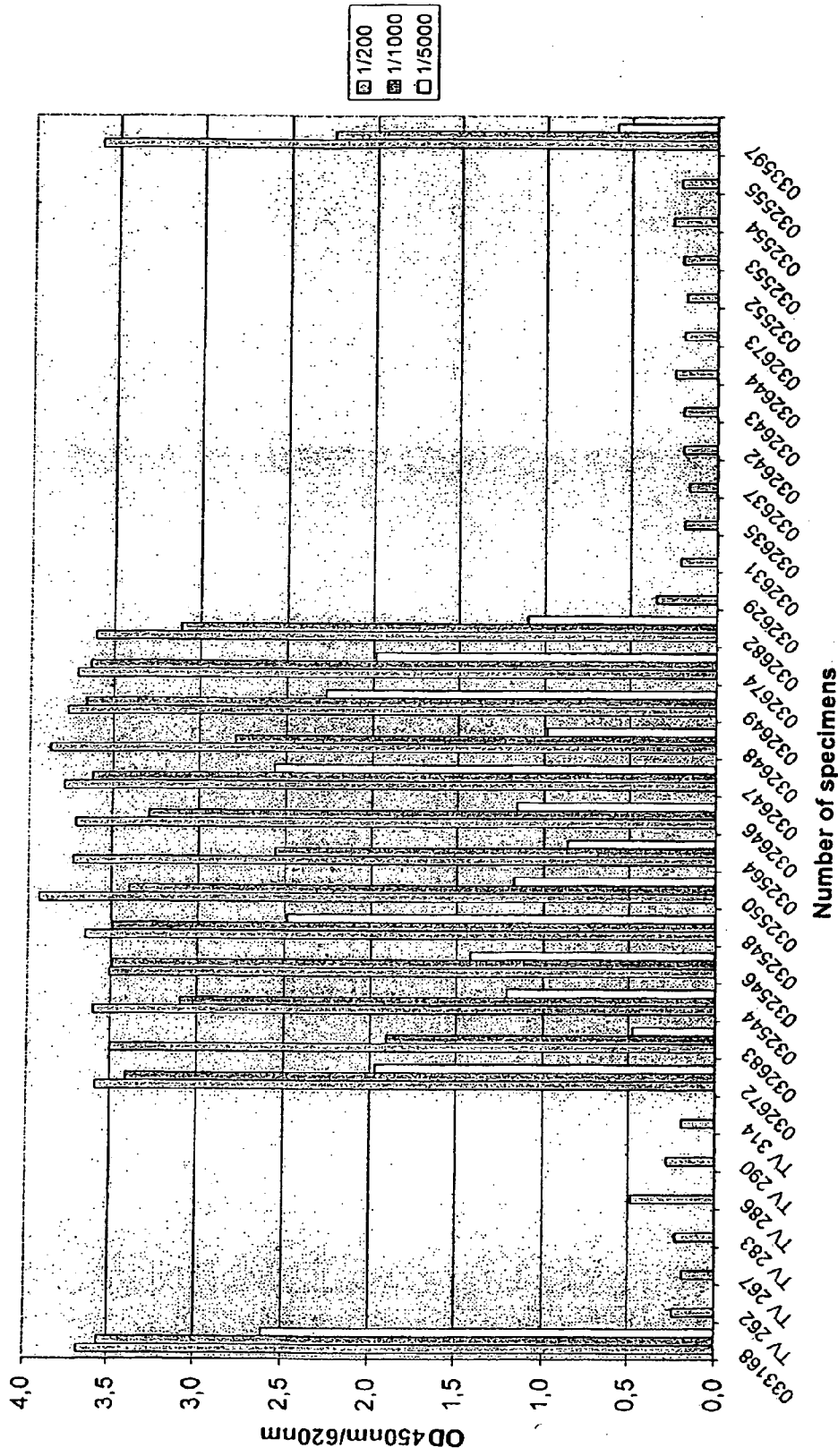


FIGURE 15

SRAS serology: Double Epitope Technique (First set)

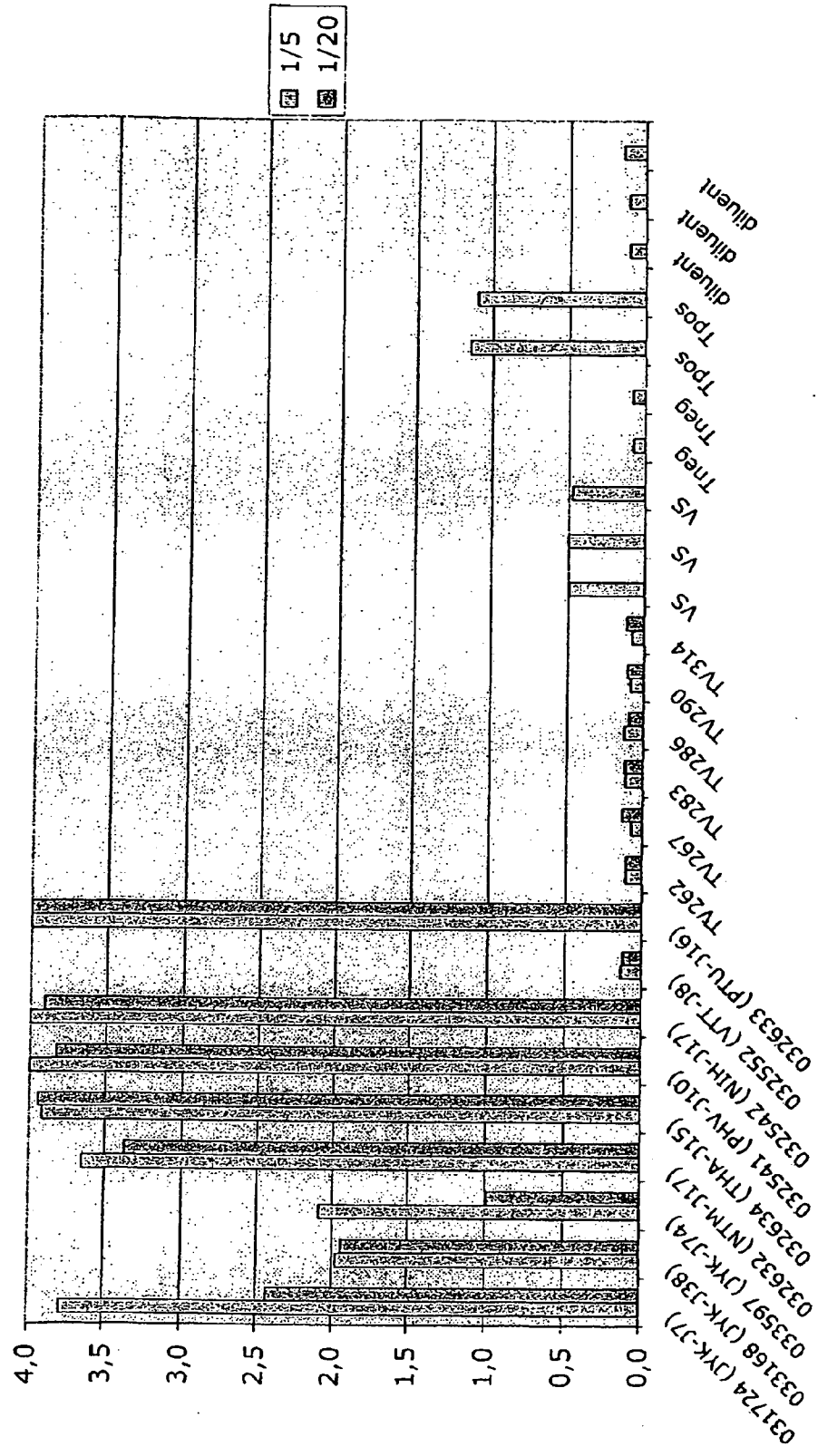
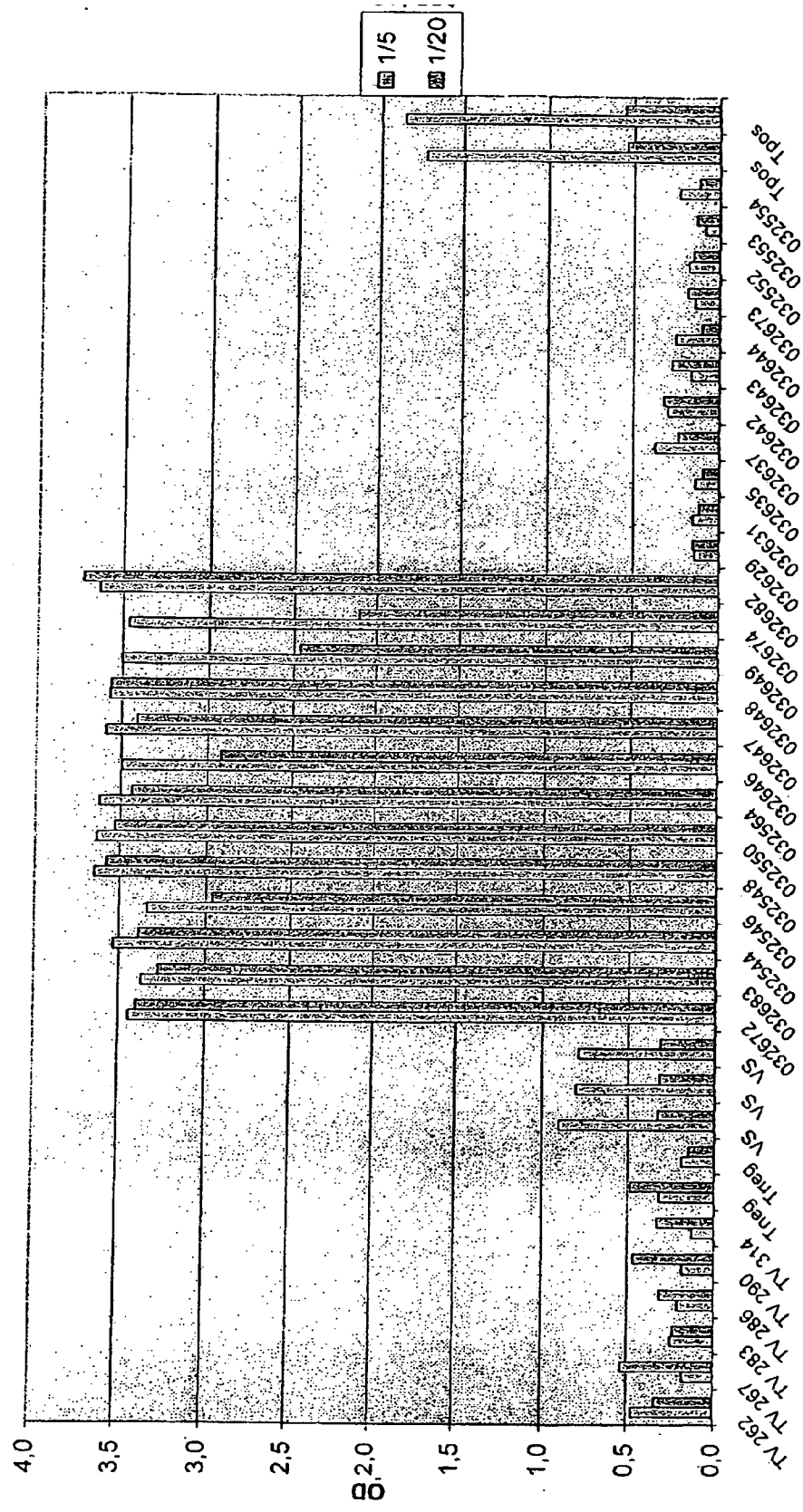


FIGURE 16

SRAS serology: Double Epitope Technique (Second set)



Number of specimens

FIGURE 17

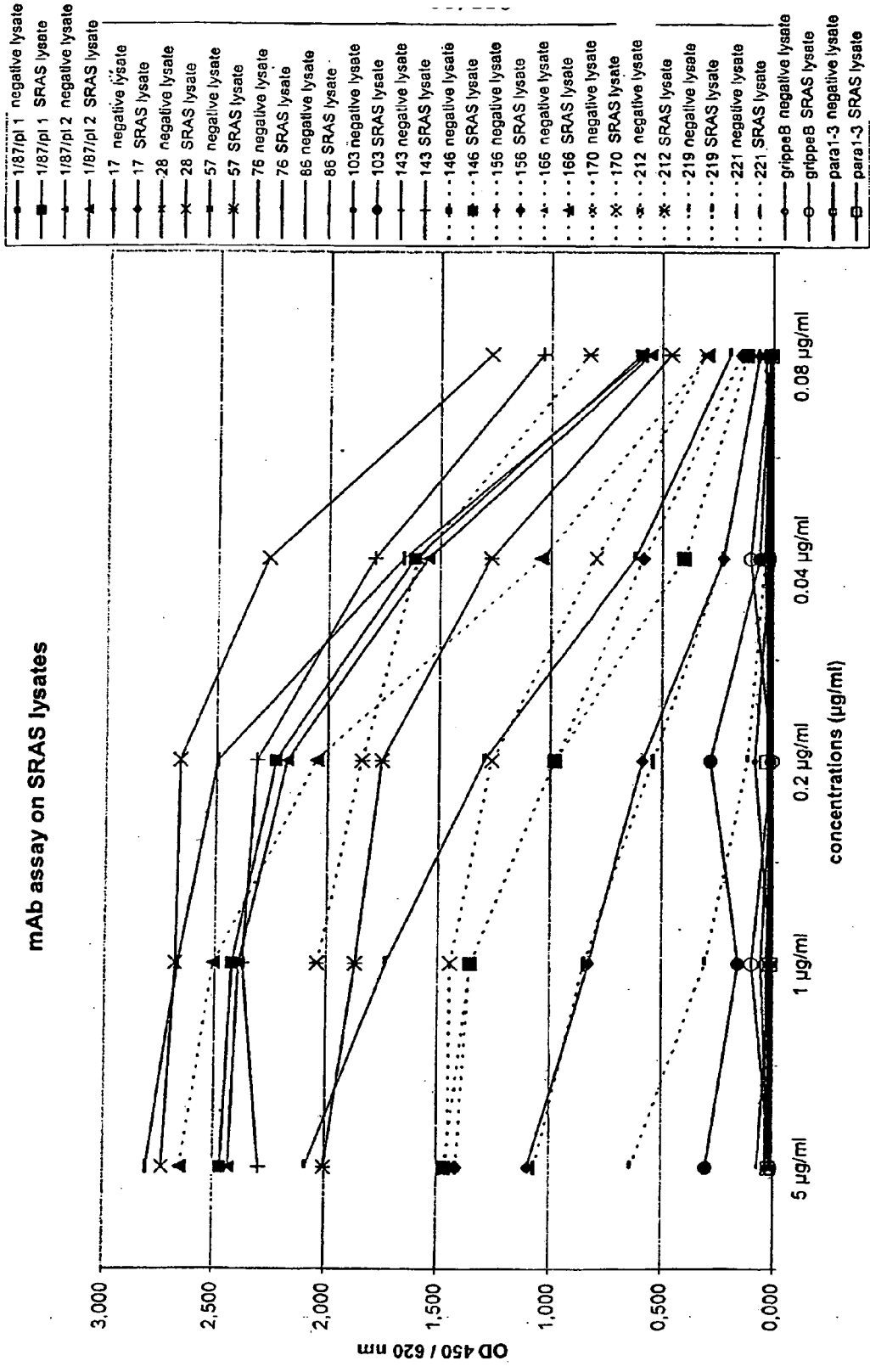


FIGURE 18

- 187 negative lysate
- 187 lysat 229E
- 5-11H,6 negative lysate
- 5-11H,6 lysat 229E
- 17 lysat negative lysate
- 17 lysat 229E
- 28 negative lysate
- 28 lysat 229E
- 57 negative lysate
- 57 lysat 229E
- 76 negative lysate
- 76 lysat 229E
- 86 negative lysate
- 86 lysat 229E
- 103 negative lysate
- 103 lysat 229E
- 143 negative lysate
- 143 lysat 229E
- 146 negative lysate
- 146 lysat 229E
- 156 negative lysate
- 156 lysat 229E
- 166 negative lysate
- 166 lysat 229E
- 170 negative lysate
- 170 lysat 229E
- 212 negative lysate
- 212 lysat 229E
- 219 negative lysate
- 219 lysat 229E
- 221 negative lysate
- 221 lysat 229E
- grippeB negative lysate
- grippeB lysat 229E
- para1-3 negative lysate
- para1-3 lysat 229E

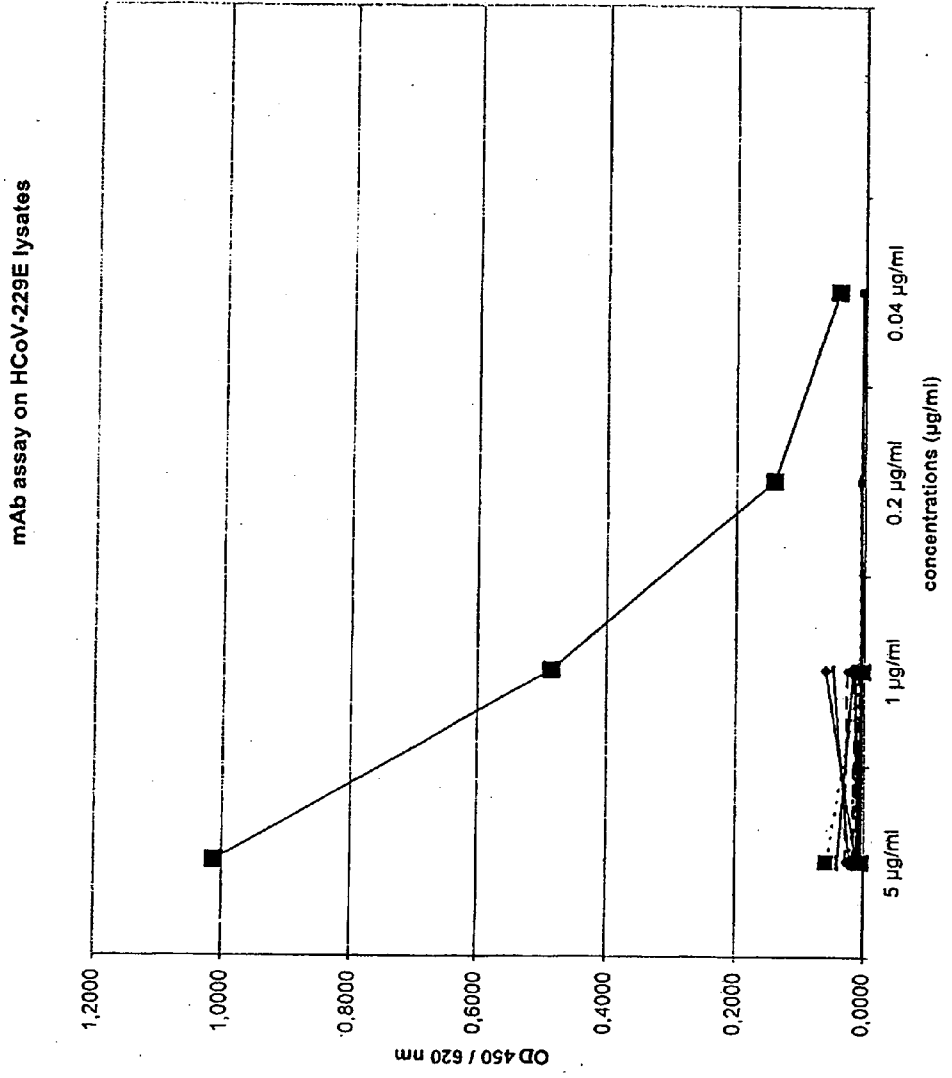


FIGURE 19

#para1-3

#grippeB

#221

#219

#212

#170

#166

#156

#146

#143

#103

#86

#76

#57

#28

#17

1/87

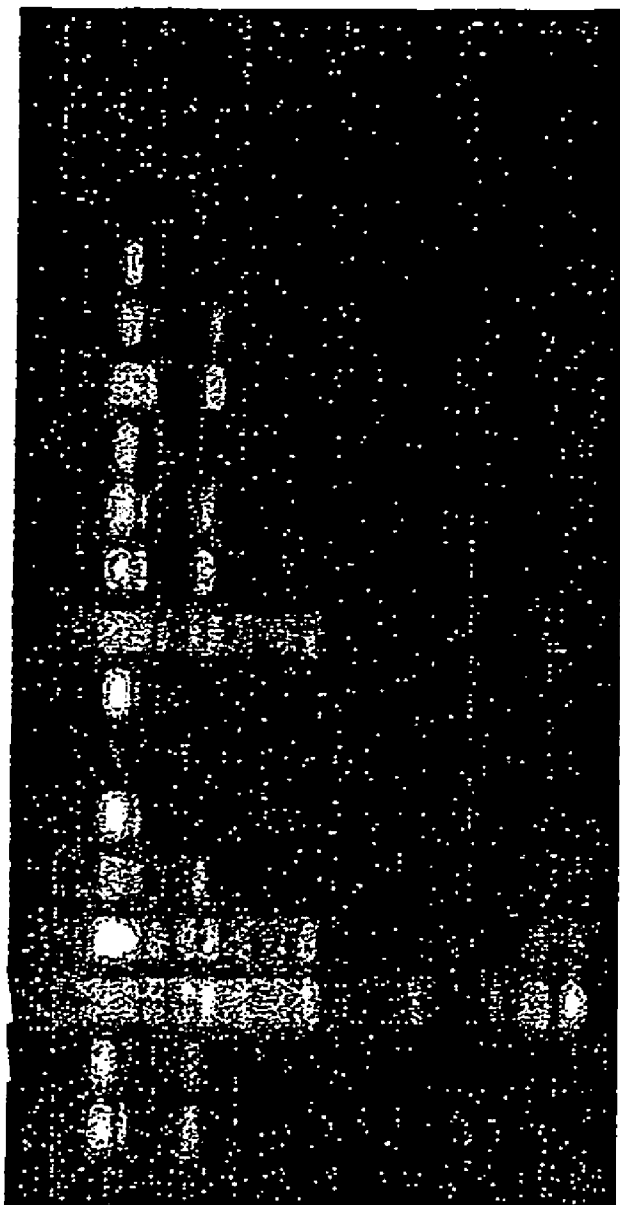


FIGURE 20

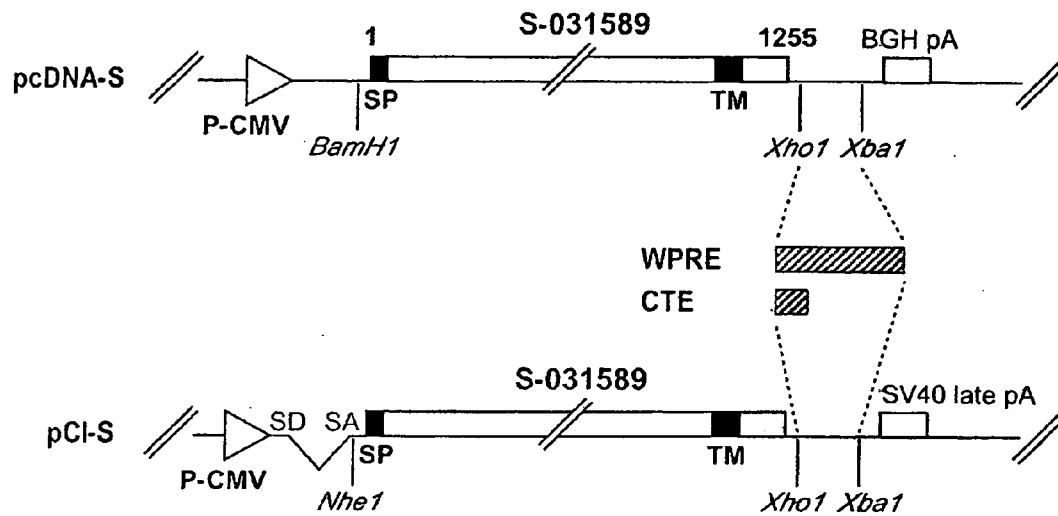


FIGURE 21

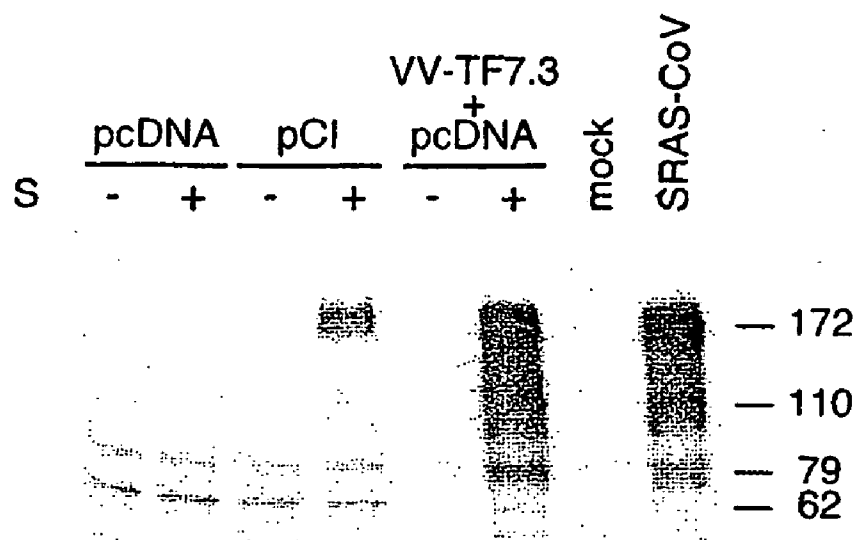
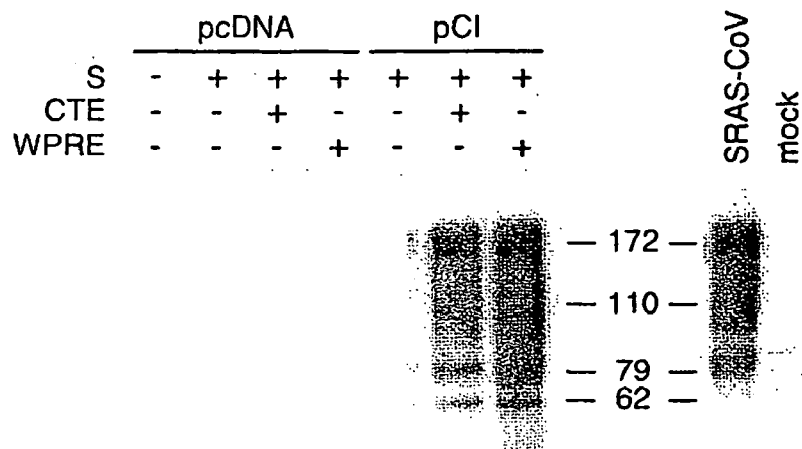


FIGURE 22

A.



B.

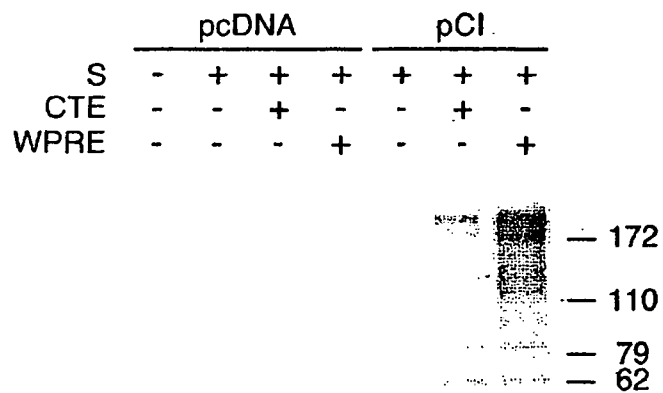


FIGURE 23

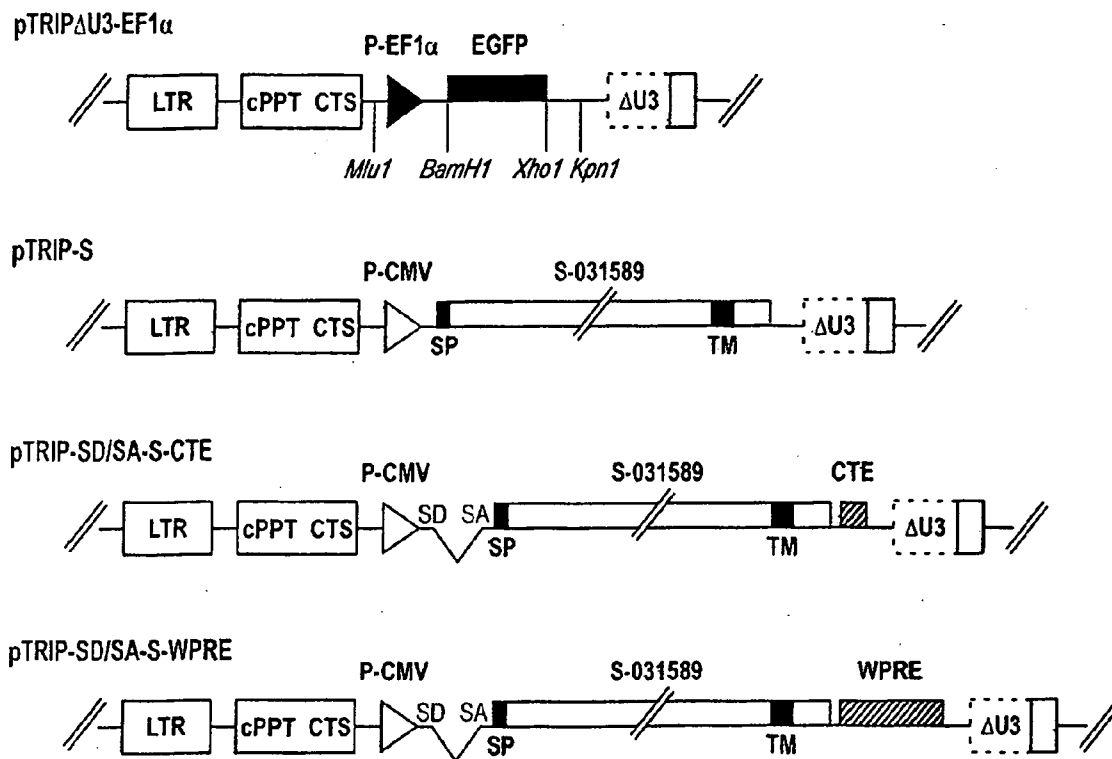


FIGURE 24

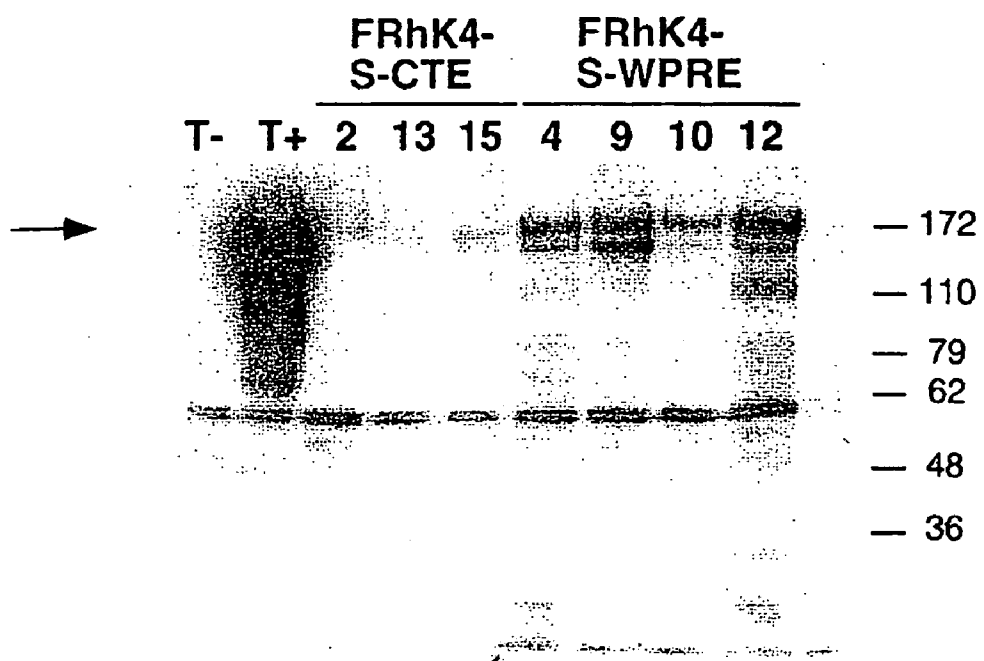


FIGURE 25

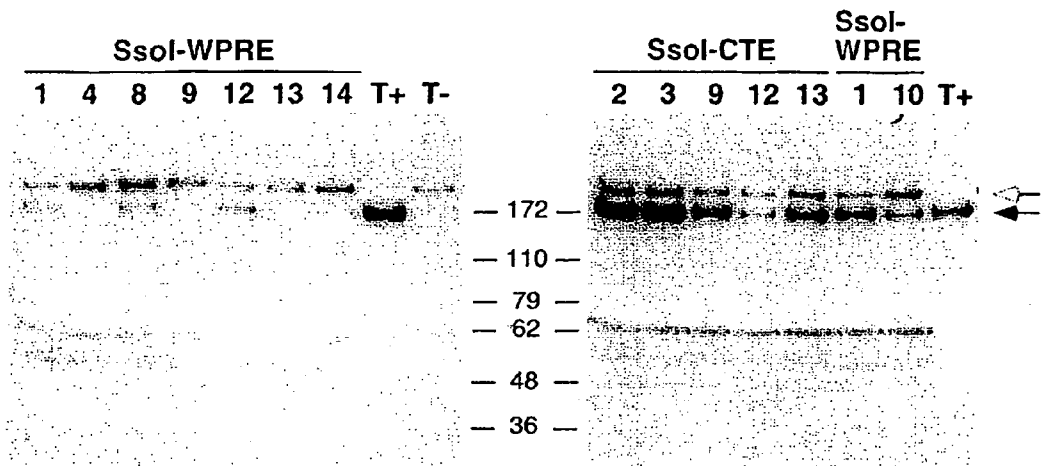
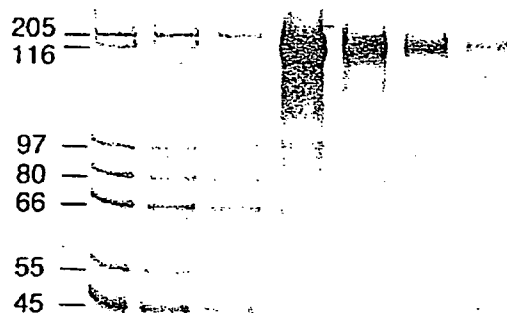


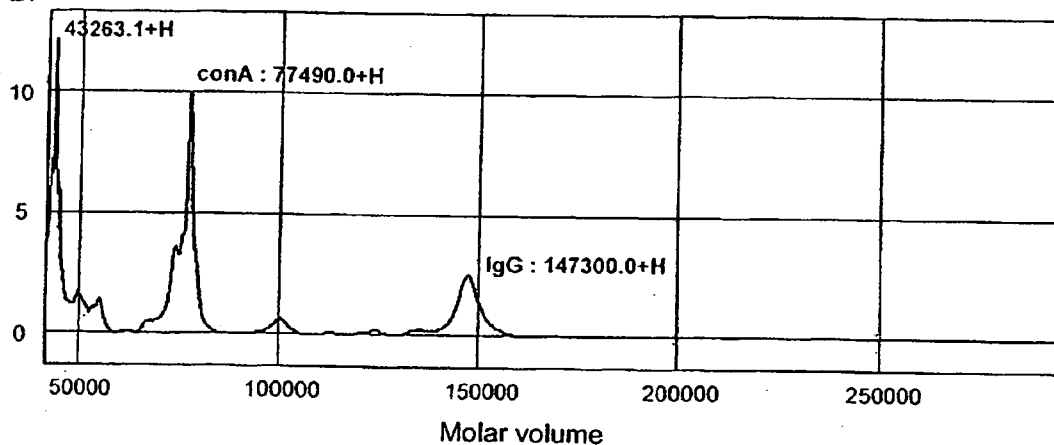
FIGURE 26

A.

MM (ng)			Ssol (μg)		
120	60	30	2	.5	.13 .03



B.



C.

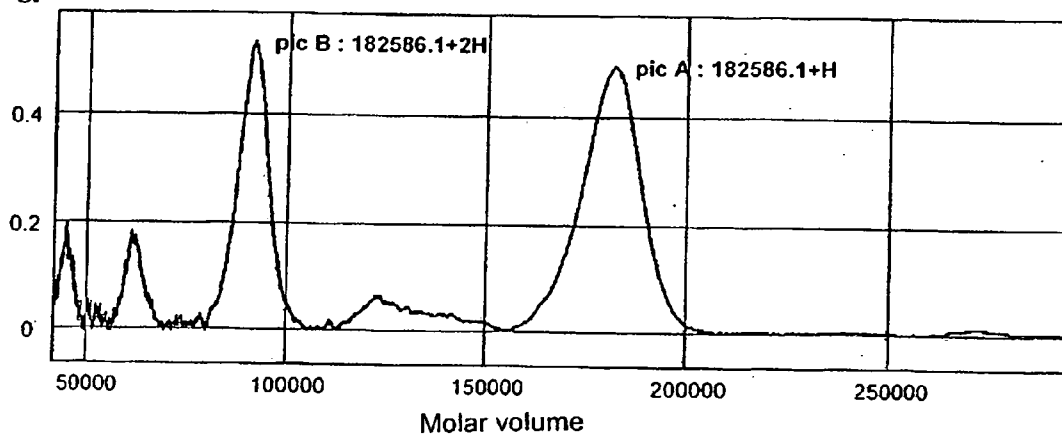


FIGURE 27 A-C

D.

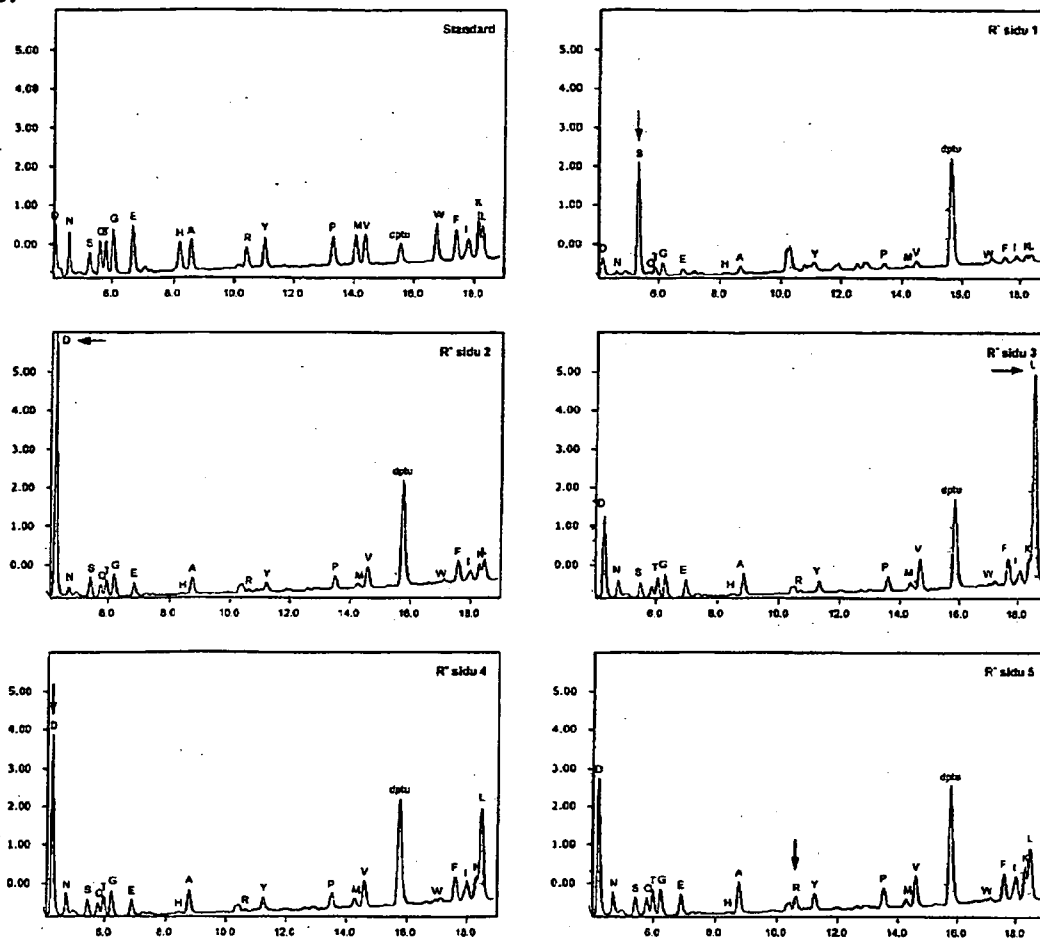


FIGURE 27 D

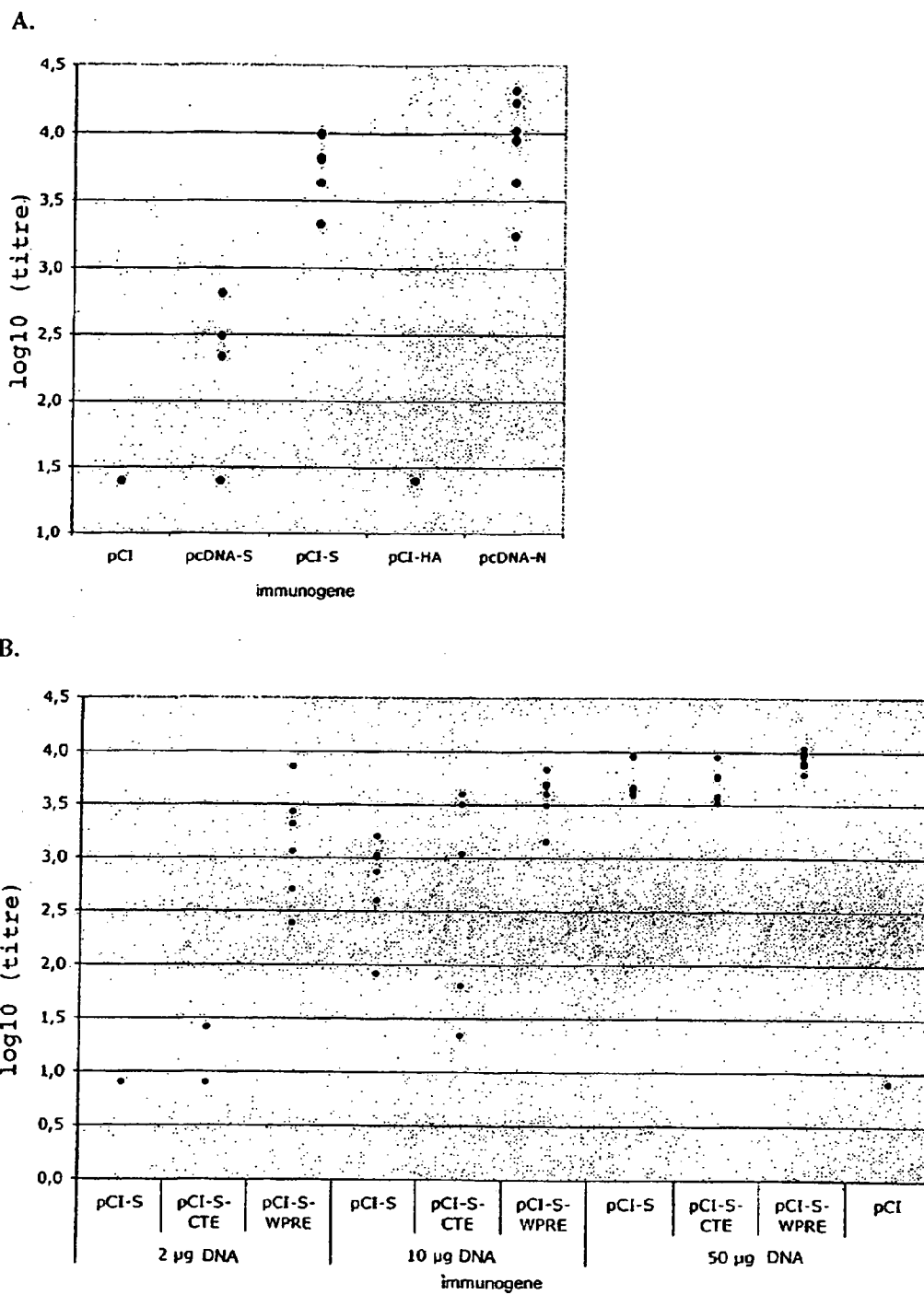


FIGURE 28

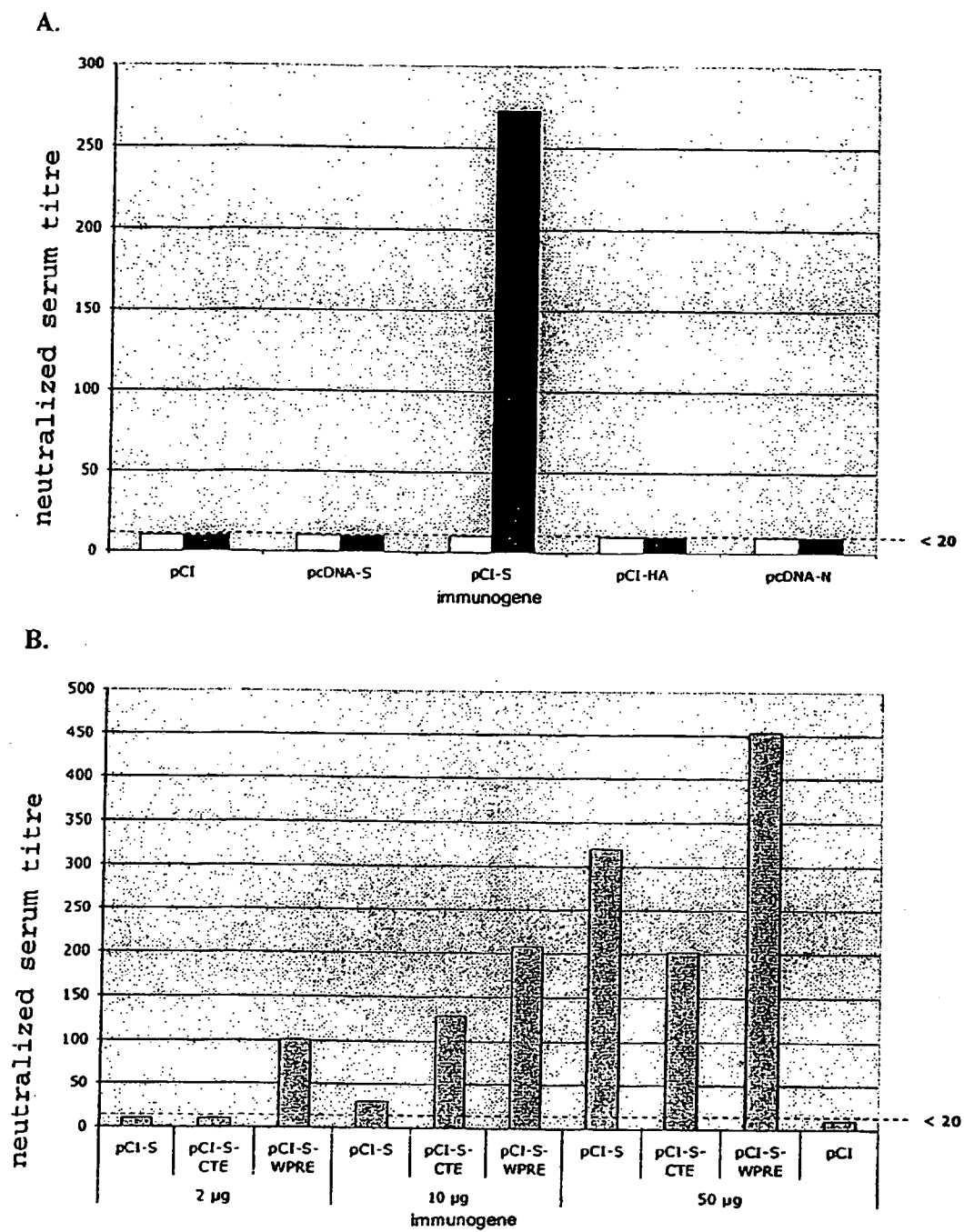
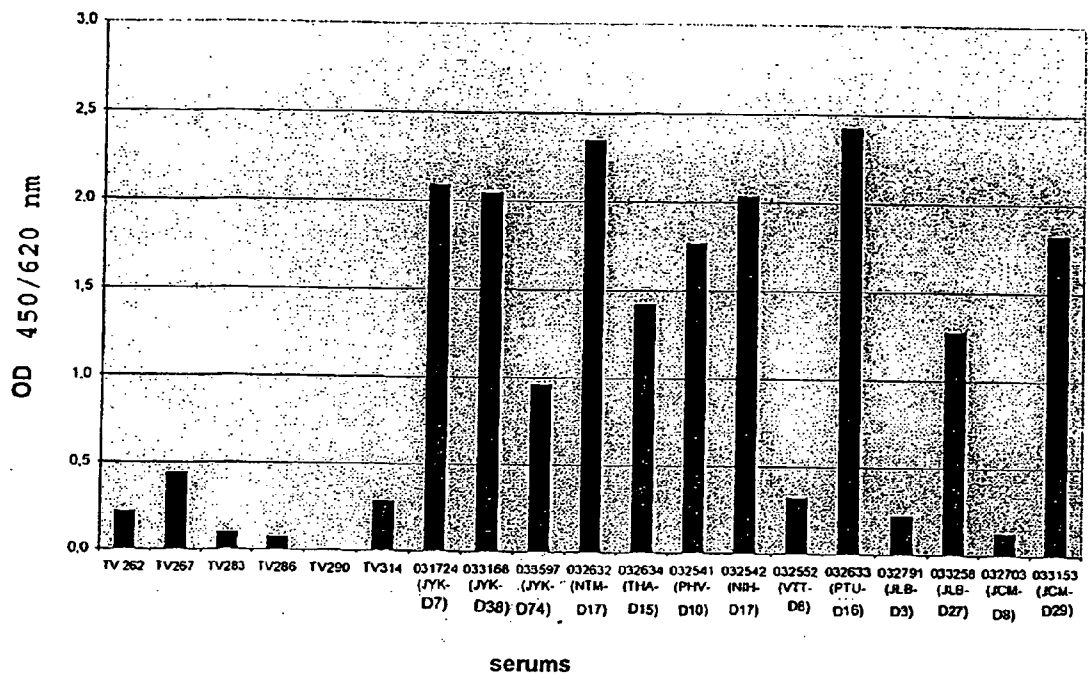


FIGURE 29



serums
FIGURE 30

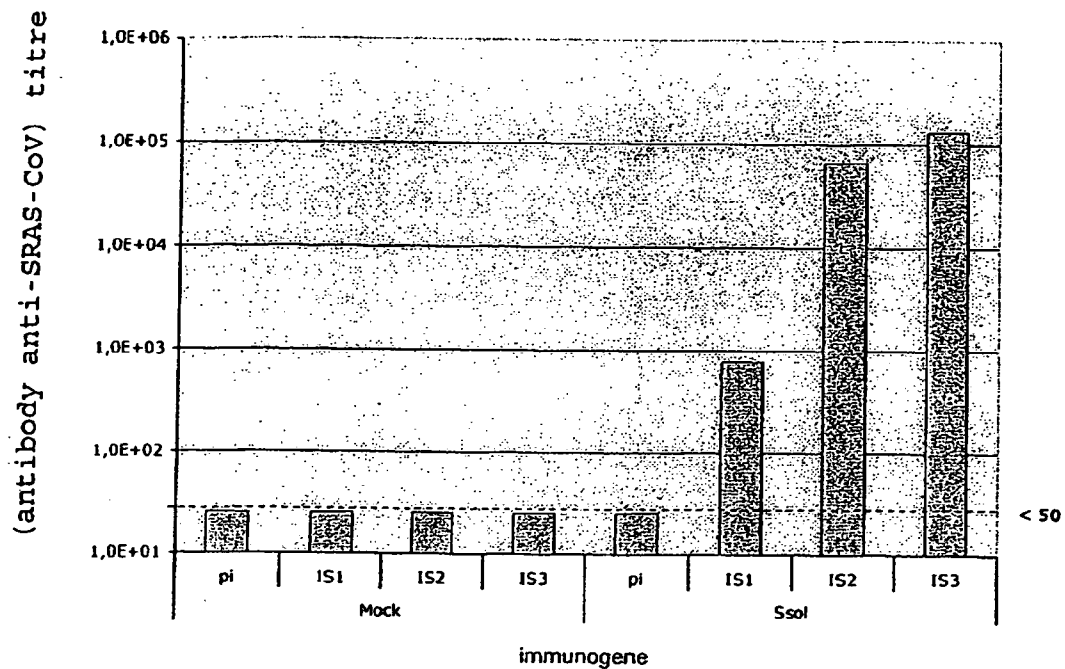


FIGURE 31

I-3059 2697 GGTGCTGGCGCTGCTCTTCAAATACCTTTTGCTATGCAAATGGCATATAGGTTCAATGGC
S-040530 2620 "A"C"A"C"C"G"G"C"C"C"C"C"C"C"C"C"C"C"C"C"C"C"

I-3059 2757 ATTGGAGTTACCCAAAATGTTCTCTATGAGAACCAAAAACAAATCGCCAACCAATTTAAC
S-040530 2680 "C"C"G"G"G"C"G"G"C"G"G"G"G"G"G"G"G"G"G"G"G"G"G"G"

I-3059 2817 AAGGCGATTAGTCAAATCAAGAATCACTTACAACAACATCAACTGCATTGGGCAAGCTG
S-040530 2740 "C"C"C"C"G"C"G"GAGC"G"C"C"C"C"C"C"C"C"C"

I-3059 2877 CAAGACGTTGTTAACCAGAATGCTCAAGCATTAAACACACTTGTAAACAACCTTAGCTCT
S-040530 2800 "G"G"G"G"G"G"G"G"G"G"C"C"G"CC"G"C"G"G"G"G"G"G"AGC

I-3059 2937 AATTTTGGTGCAATTTCAAGTGTGCTAAATGATATCCTTTGCGGACTTGATAAAGTCGAG
S-040530 2860 "C"C"C"C"C"CAGCTC"G"C"C"C"GAGCA"G"C"C"C"C"C"

I-3059 2997 GCGGAGGTACAAATGACAGGCTAATTACAGGCAGACTTCAAAGCCTTCAAACCTATGTA
S-040530 2920 "C"A"G"G"C"C"C"G"C"C"AC"C"G"GTG"G"G"G"C"G"

I-3059 3057 ACACAACAATAATCAGGGCTGCTGAAATCAGGGCTTCTGCTAATCTGCTGCTACTAAA
S-040530 2980 "C"G"G"G"G"A"C"C"G"C"C"C"CAGC"C"G"C"C"C"C"

I-3059 3117 ATGCTGAGTGTGTTCTTGGACAATCAAAAAGAGTTGACTTTTGTGGAAAGGGCTACCAC
S-040530 3040 "AGC"C"G"G"C"GAGC"G"G"G"C"C"C"C"C"C"C"C"

I-3059 3177 CTTATGTCCTTCCACAAGCAGCCCGCATGGTGTGCTTCCTACATGTCAGGTATGTG
S-040530 3100 "G"AG"C"G"C"C"C"C"C"G"G"G"G"G"C"G"C"C"

I-3059 3237 CCATCCAGGAGAGGAACCTCACCACAGCGCCAGCAATTTGTCATGAAGGCAAAGCATAAC
S-040530 3160 "TAG"C"C"C"C"C"C"C"C"C"C"C"C"C"C"G"G"C"

I-3059 3297 TTCCCTCGTGAAGGTGTTTTGTGTTAATGGCACTTCTTGGTTTATTACACAGAGGAAC
S-040530 3220 "C"G"G"C"G"C"C"C"C"C"CAGC"C"C"C"C"

I-3059 3357 TTCTTTCTCCACAAATAATFACTACAGACAATACATTTGTCTCAGGAAATGTGATGTC
S-040530 3280 "CAGC"C"G"C"C"C"C"C"C"C"C"C"C"

I-3059 3417 GTTATTGGCATCATTAAACAACACAGTTTATGATCCTCTGCAACCTGAGCTTGACTCATT
S-040530 3340 "G"C"C"C"C"C"C"C"C"C"C"C"C"C"C"G"C"C"C"AGC"

I-3059 3477 AAAGAAGAGCTGGACAAGTACTTCAAAAATCATAACATCACCAGATGTTGATCTTGCCGAC
S-040530 3400 "G"G"G"G"G"A"G"C"C"C"C"C"C"C"G"C"G"

I-3059 3537 ATTCAGGCATTAACGCTTCTGTCGTCACATTCAAAAGAAATGACCGCCTCAATGAG
S-040530 3460 "CAGC"C"C"G"G"G"C"G"G"G"C"A"A"G"C"

I-3059 3597 GTCGCTAAAAATTTAAATGAATCACTCATTGACCTTCAAGAATGGGAAAATATGAGCAA
S-040530 3520 "G"C"G"CC"G"C"GAGC"G"C"G"G"G"GC"C"G"C"

I-3059 3657 TATATTAAATGGCCTTGGTATGTTGGCTCGGCTTCAATGCTGGACTAATGGCCATCGTC
S-040530 3580 "C"C"G"C"C"C"C"G"G"C"C"C"C"C"G"C"

I-3059 3717 ATGGTTACAATCTTGCTTTGTTGCATGACTAGTTGTTGCAGTTGCCTCAAGGGTGCATGC
S-040530 3640 "G"C"C"C"G"C"C"C"C"C"C"C"C"C"TC"C"G"GA"C"

I-3059 3777 TCTTGTGGTTCTTGCTGCAAGTTGATGAGGATGACTCTGAGCCAGTTCTCAAGGGTGTG
S-040530 3700 AGC"CAGC"C"C"C"AGC"C"G"G"C"

I-3059 3837 AAATTACATTACACATAAACGAACTTATGGATTTGTTTATGAGATTTTTACTCTTGGAT
S-040530 3760 "GC"G"C"C"GT "CGA"

I-3059 3897 CAATTACTGCACAGCCAGTAAAATTGACAATGCTTCTCCTGCAAGT
S-040530

FIGURE 32.3

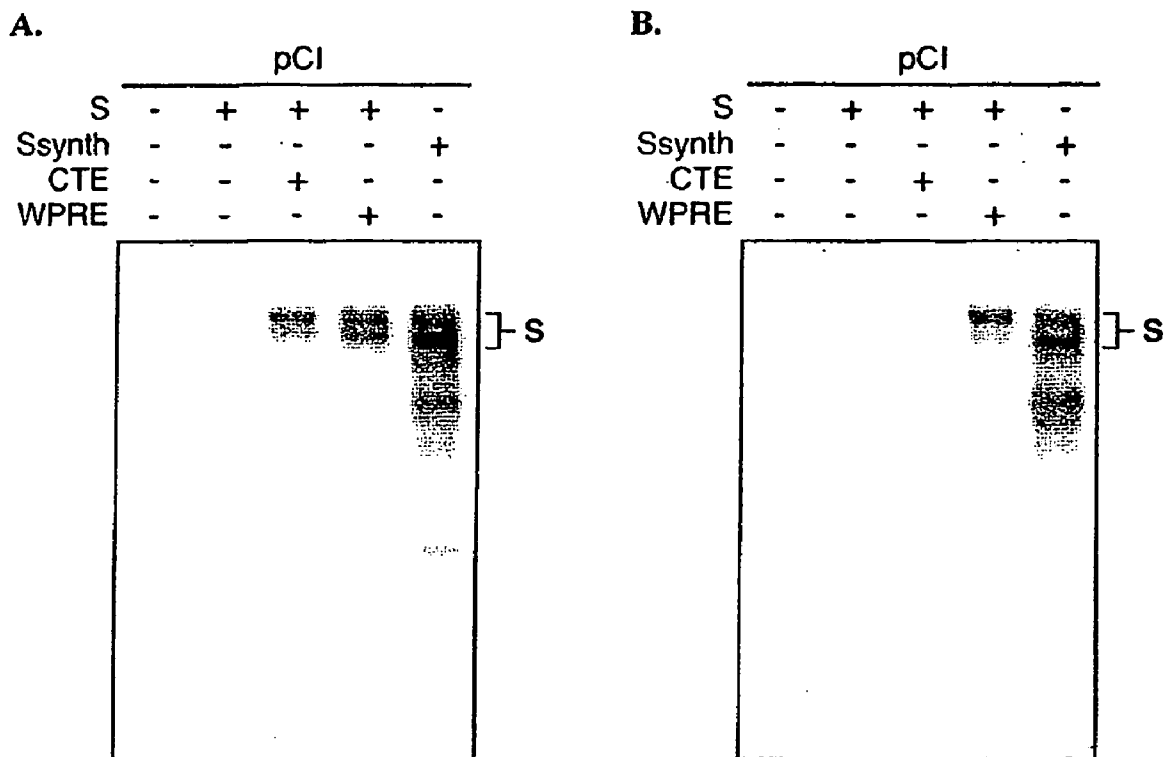
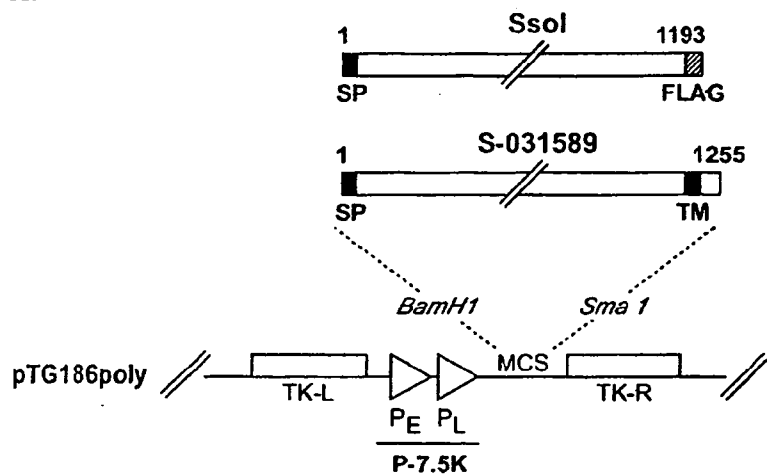
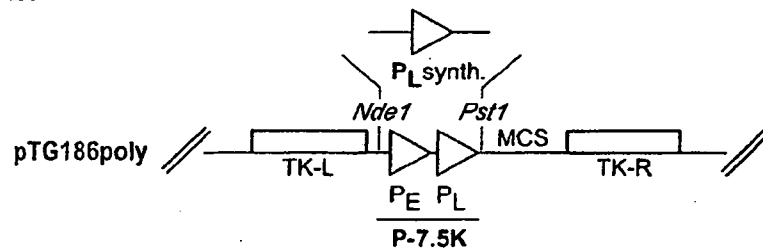


FIGURE 33

A.



B.



C.

CATATG AGC [T]₂₀GGCATATAAATA GACTC GGCGCGCC AT CTGCAG
NdeI promoteur 480 *AscI* *PstI*

FIGURE 34 A-C

D.

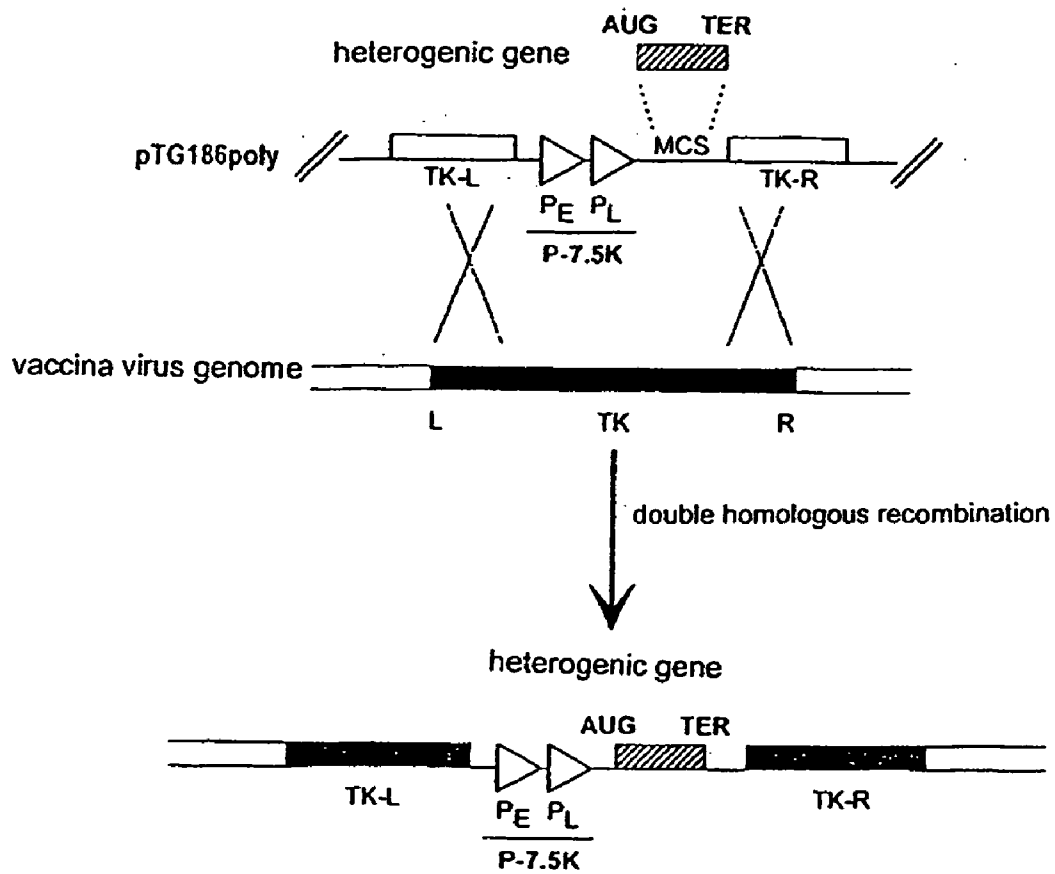
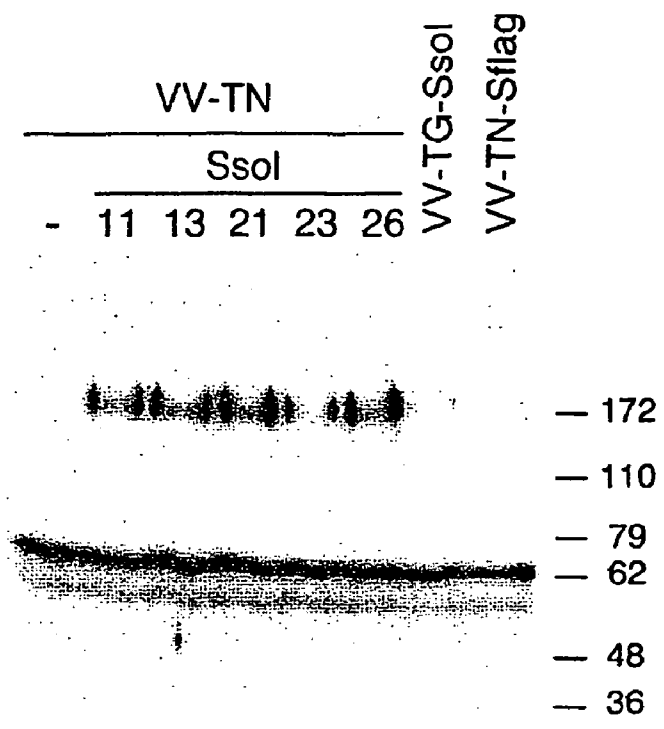


FIGURE 34 D

A.



B.

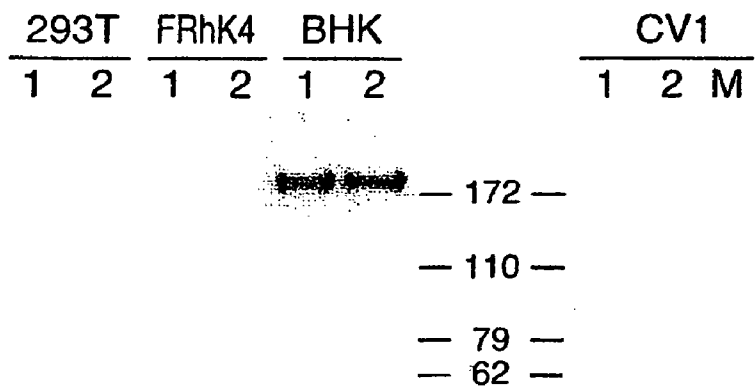


FIGURE 36

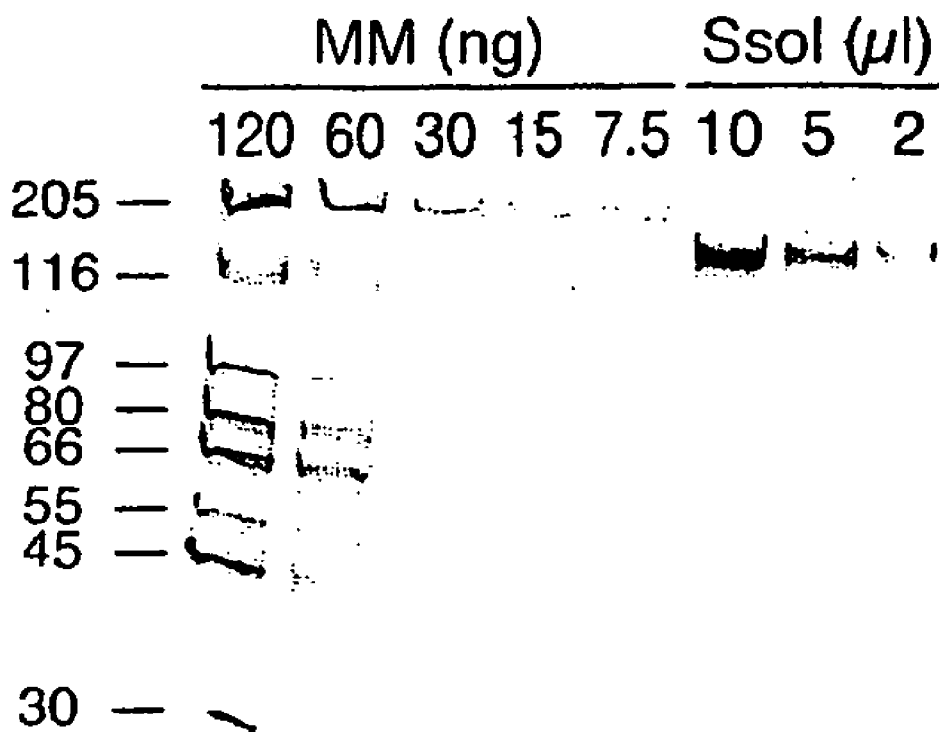
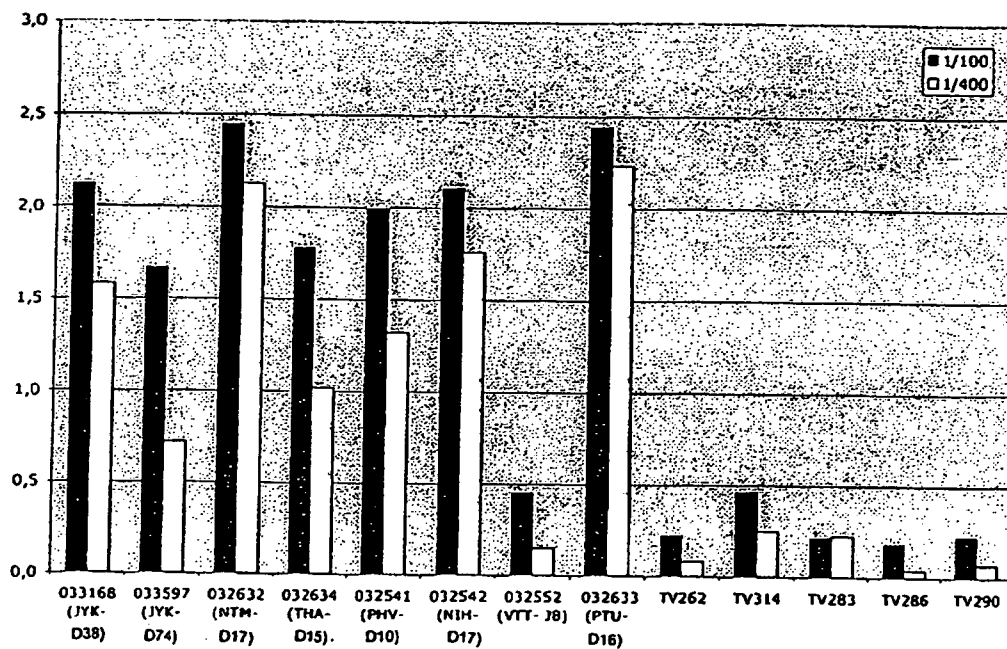


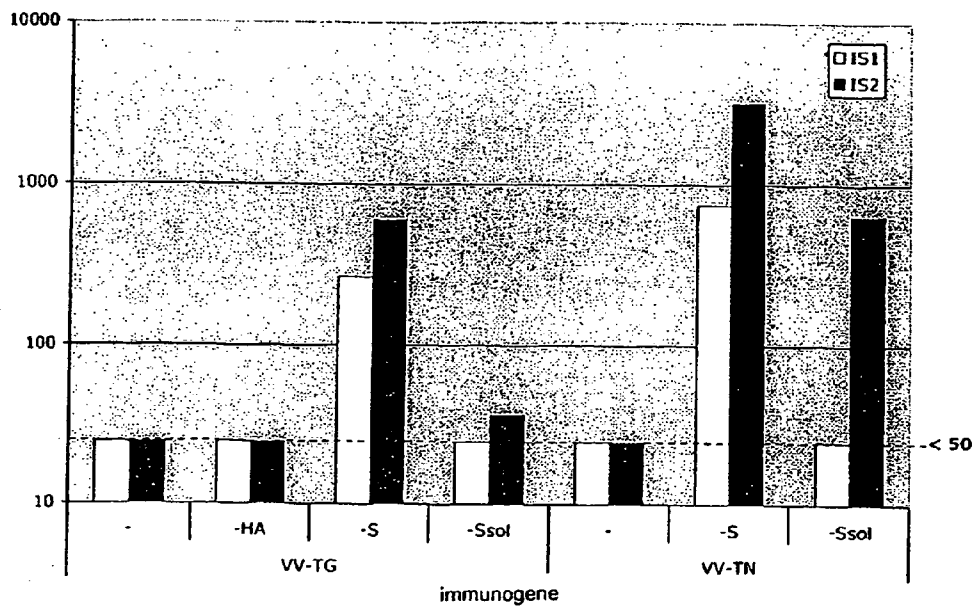
FIGURE 37



serums

FIGURE 38

A.



B.

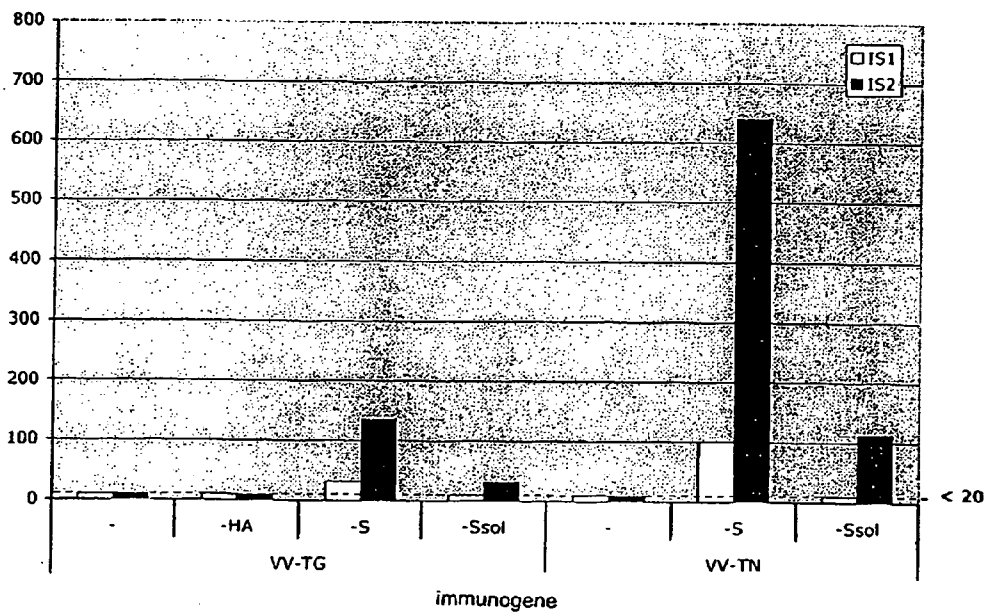


FIGURE 39

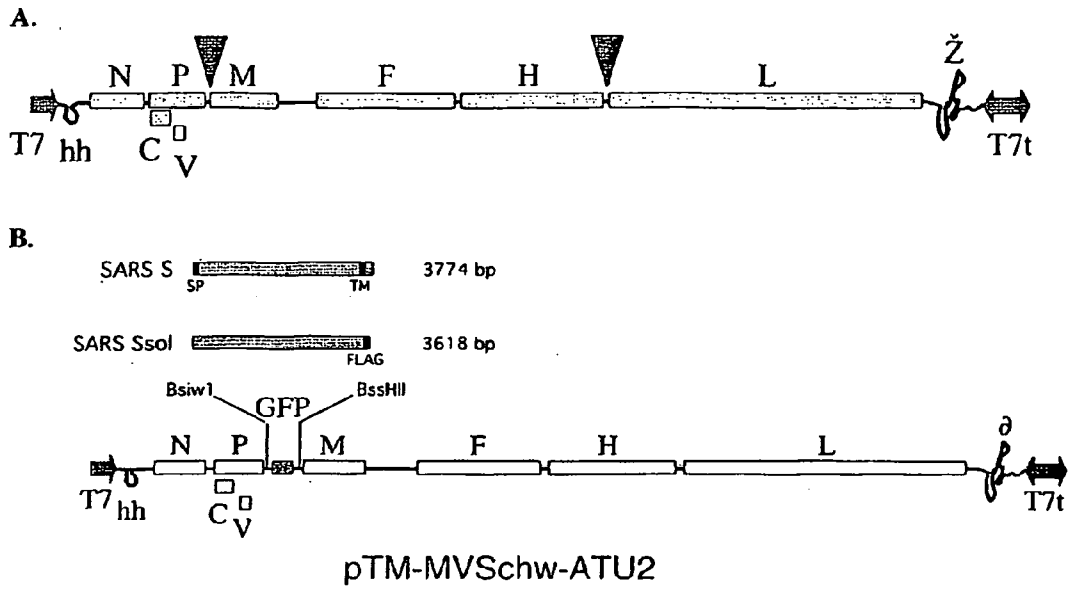


FIGURE 40

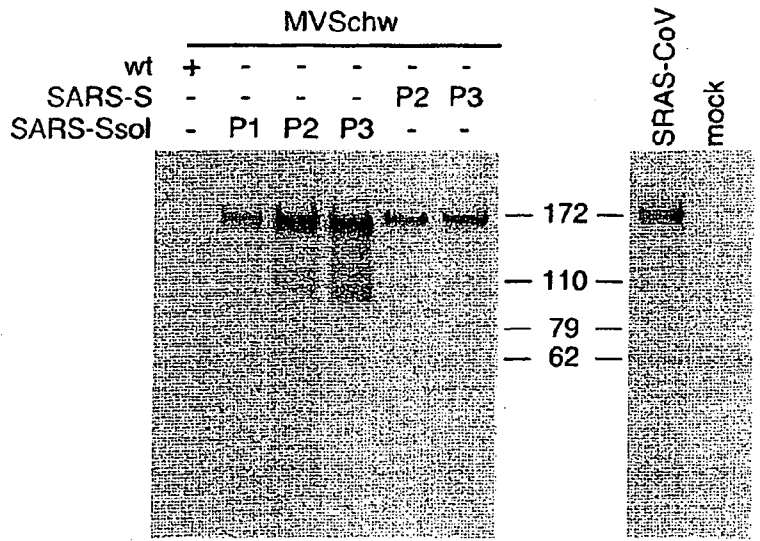


FIGURE 41

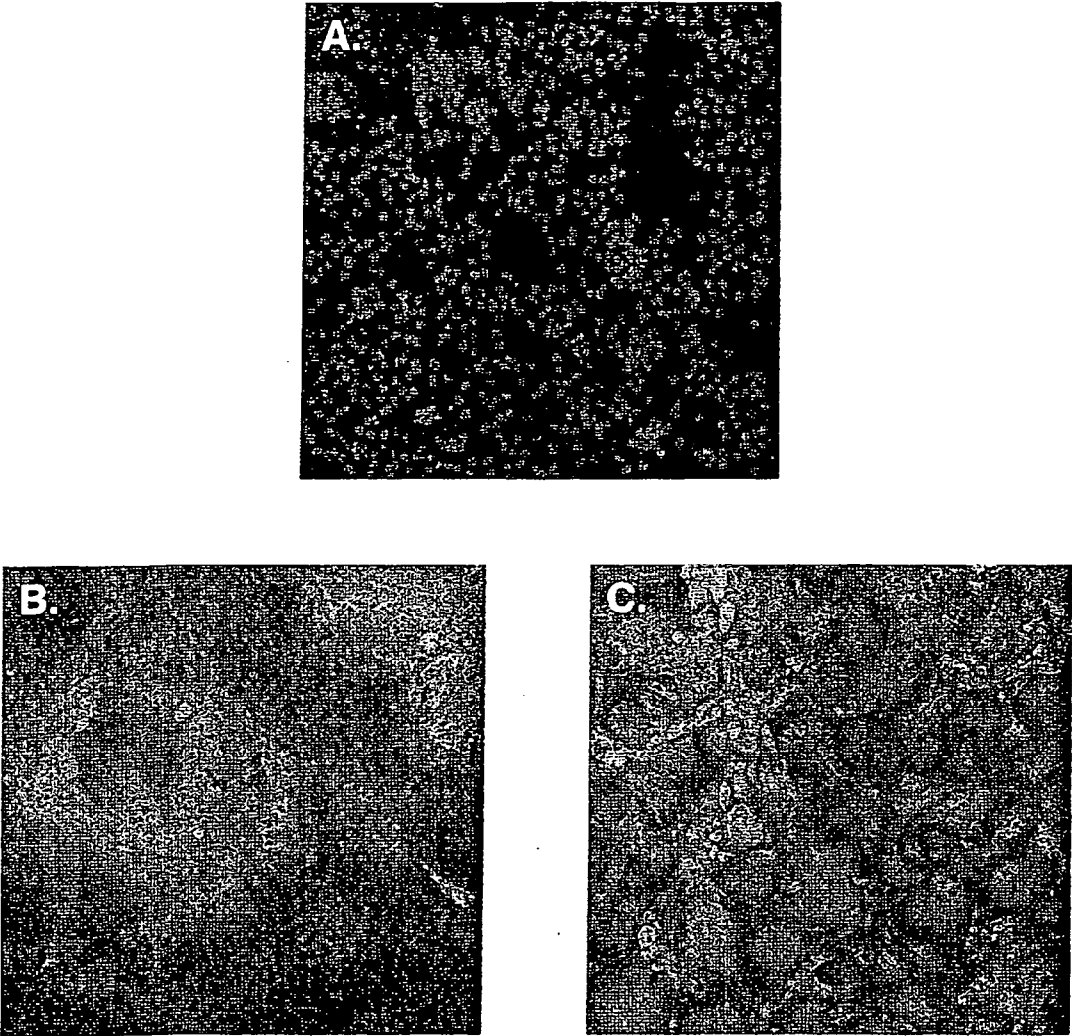


FIGURE 42

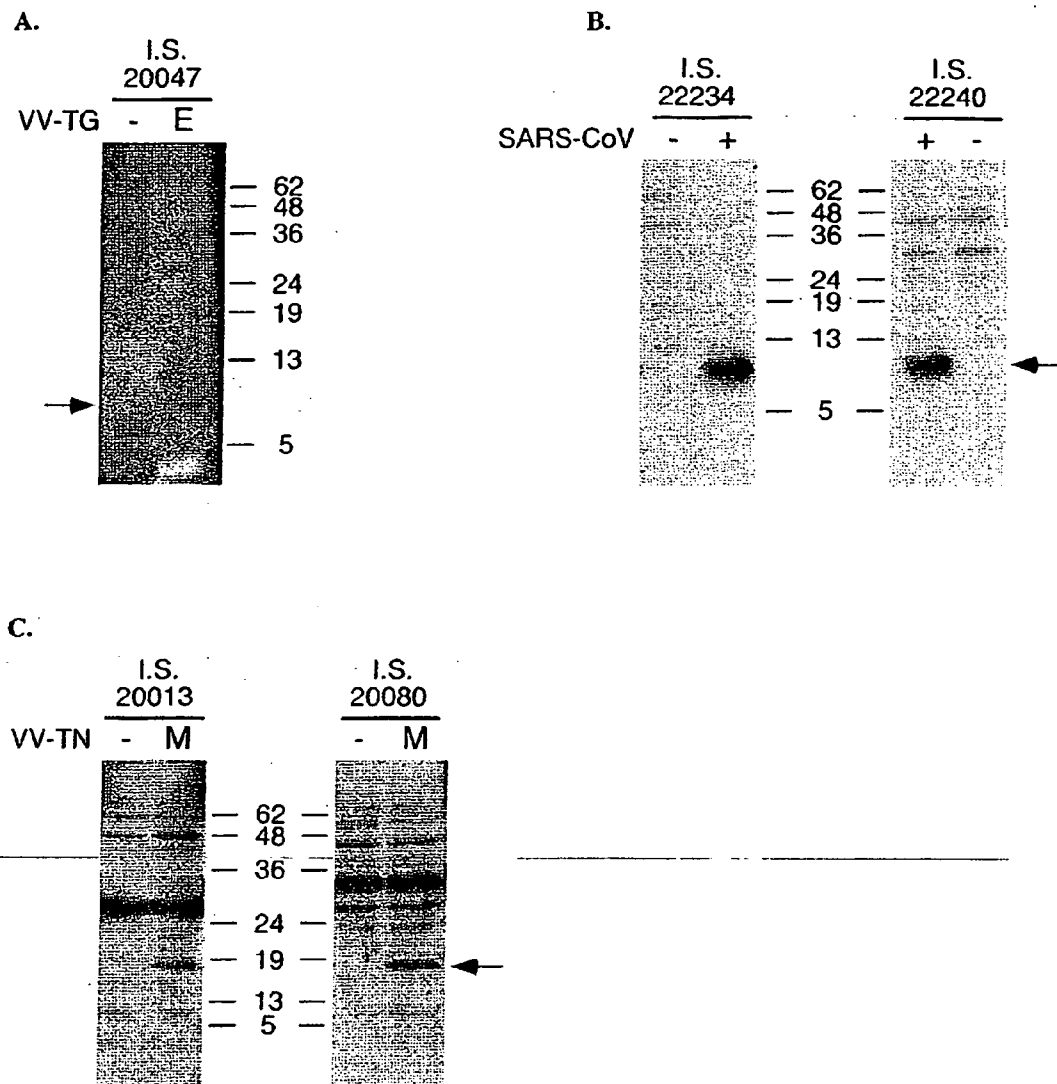


FIGURE 43

NOVEL STRAIN OF SARS-ASSOCIATED CORONAVIRUS AND APPLICATIONS THEREOF

[0001] The present invention relates to a novel strain of severe acute respiratory syndrome (SARS)-associated coronavirus derived from a sample recorded under No. 031589 and collected in Hanoi (Vietnam), to nucleic acid molecules derived from its genome, to the proteins and peptides encoded by said nucleic acid molecules and to their applications, in particular as diagnostic reagents and/or as vaccine.

[0002] Coronavirus is a virus containing single-stranded RNA, of positive polarity, of approximately 30 kilobases which replicates in the cytoplasm of the host cells; the 5' end of the genome has a capped structure and the 3' end contains a polyA tail. This virus is enveloped and comprises, at its surface, peplomeric structures called spicules.

[0003] The genome comprises the following open reading frames or ORFs, from its 5' end to its 3' end: ORF1a and ORF1b corresponding to the proteins of the transcription-replication complex, and ORF-S, ORF-E, ORF-M and ORF-N corresponding to the structural proteins S, E, M and N. It also comprises ORFs corresponding to proteins of unknown function encoded by: the region situated between ORF-S and ORF-E and overlapping the latter, the region situated between ORF-M and ORF-N, and the region included in ORF-N.

[0004] The S protein is a membrane glycoprotein (200-220 kDa) which exists in the form of spicules or spikes emerging from the surface of the viral envelope. It is responsible for the attachment of the virus to the receptors of the host cell and for inducing the fusion of the viral envelope with the cell membrane.

[0005] The small envelope protein (E), also called sM (small membrane), which is a nonglycosylated transmembrane protein of about 10 kDa, is the protein present in the smallest quantity in the virion. It plays a powerful role in the coronavirus budding process which occurs at the level of the intermediate compartment in the endoplasmic reticulum and the Golgi apparatus.

[0006] The M protein or matrix protein (25-30 kDa) is a more abundant membrane glycoprotein which is integrated into the viral particle by an M/E interaction, whereas the incorporation of S into the particles is directed by an S/M interaction. It appears to be important for the viral maturation of coronaviruses and for the determination of the site where the viral particles are assembled.

[0007] The N protein or nucleocapsid protein (45-50 kDa) which is the most conserved among the coronavirus structural proteins is necessary for encapsidating the genomic RNA and then for directing its incorporation into the virion. This protein is probably also involved in the replication of the RNA.

[0008] When the host cell is infected, the reading frame (ORF) situated in 5' of the viral genome is translated into a polyprotein which is cleaved by the viral proteases and then releases several nonstructural proteins such as the RNA-dependent RNA polymerase (Rep) and the ATPase helicase (Hel). These two proteins are involved in the replication of the viral genome and in the generation of transcripts which are used in the synthesis of the viral proteins. The mecha-

nisms by which these subgenomic mRNAs are produced are not completely understood; however, recent facts indicate that the sequences for regulation of transcription at the 5' end of each gene represent signals which regulate the discontinuous transcription of the subgenomic mRNAs.

[0009] The proteins of the viral membrane (S, E and M proteins) are inserted into the intermediate compartment, whereas the replicated RNA (+ strand) is assembled with the N (nucleocapsid) protein. This protein-RNA complex then combines with the M protein contained in the membranes of the endoplasmic reticulum and the viral particles form when the nucleocapsid complex buds into the endoplasmic reticulum. The virus then migrates across the Golgi complex and eventually leaves the cell, for example by exocytosis. The site of attachment of the virus to the host cell is at the level of the S protein.

[0010] Coronaviruses are responsible for 15 to 30% of colds in humans and for respiratory and digestive infections in animals, especially cats (FIPV: Feline infectious peritonitis virus), poultry (IBV: Avian infectious bronchitis virus), mice (MHV: Mouse hepatitis virus), pigs (TGEV: Transmissible gastroenteritis virus, PEDV: Porcine Epidemic diarrhea virus, PRCoV: Porcine Respiratory Coronavirus, HEV: Hemagglutinating encephalomyelitis Virus) and bovines (BCoV: Bovine coronavirus).

[0011] In general, each coronavirus affects only one species; in immunocompetent individuals, the infection induces optionally neutralizing antibodies and cell immunity, capable of destroying the infected cells.

[0012] An epidemic of atypical pneumonia, called severe acute respiratory syndrome (SARS) has spread in various countries (Vietnam, Hong Kong, Singapore, Thailand and Canada) during the first quarter of 2003, from an initial focus which appeared in China in the last quarter of 2002. The severity of this disease is such that its mortality rate is about 3 to 6%. The determination of the causative agent of this disease is underway by numerous laboratories worldwide.

[0013] In March 2003, a new coronavirus (SARS-CoV or SARS virus) was isolated, in association with cases of severe acute respiratory syndrome (T. G. KSLAZEK et al., *The New England Journal of Medicine*, 2003, 348, 1319-1330; C. DROSTEN et al., *The New England Journal of Medicine*, 2003, 348, 1967-1976; Peiris et al., *Lancet*, 2003, 361, 1319).

[0014] Genomic sequences of this new coronavirus have thus been obtained, in particular those of the Urbani isolate (Genbank accession No. AY274119.3 and A. MARRA et al., *Science*, May 1, 2003, 300, 1399-1404) and the Toronto isolate (Tor2, Genbank accession No. AY278741 and A. ROTA et al., *Science*, 2003, 300, 1394-1399).

[0015] The organization of the genome is comparable with that of other known coronaviruses, thus making it possible to confirm that SARS-CoV belongs to the Coronaviridae family; open reading frames ORF1a and 1b and open reading frames corresponding to the S, E, M and N proteins, and to proteins encoded by: the region situated between ORF-S and ORF-E (ORF3), the region situated between ORF-S and ORF-E and overlapping ORF-E (ORF4), the region situated between ORF-M and ORF-N (ORF7 to

ORF11) and the region corresponding to ORF-N (ORF13 and ORF14), have in particular been identified.

[0016] Seven differences have been identified between the sequences of the Tor2 and Urbani isolates; 3 correspond to silent mutations (c/t at position 16622 and a/g at position 19064 of ORF1b, t/c at position 24872 of ORF-S) and 4 modify the amino acid sequence of respectively: the proteins encoded by ORF1a (c/t at position 7919 corresponding to the A/V mutation), the S protein (g/t at position 23220 corresponding to the A/S mutation), the protein encoded by ORF3 (a/g at position 25298 corresponding to the R/G mutation) and the M protein (t/c at position 26857 corresponding to the S/P mutation).

[0017] In addition, phylogenetic analysis shows that SARS-CoV is distant from other coronaviruses and that it did not appear by mutation of human respiratory coronaviruses nor by recombination between known coronaviruses (for a review, see Holmes, J. C. I., 2003, 111, 1605-1609).

[0018] The determination and the taking into account of new variants are important for the development of reagents for the detection and diagnosis of SARS which are sufficiently sensitive and specific, and immunogenic compositions capable of protecting populations against epidemics of SARS.

[0019] The inventors have now identified another strain of SARS-associated coronavirus which is distinguishable from the Tor2 and Urbani isolates.

[0020] The subject of the present invention is therefore an isolated or purified strain of severe acute respiratory syndrome-associated human coronavirus, characterized in that its genome has, in the form of complementary DNA, a serine codon at position 23220-23222 of the gene for the S protein or a glycine codon at position 25298-25300 of the gene for ORF3, and an alanine codon at position 7918-7920 of ORF1a or a serine codon at position 26857-26859 of the gene for the M protein, said positions being indicated in terms of reference to the Genbank sequence AY274119.3.

[0021] According to an advantageous embodiment of said strain, the DNA equivalent of its genome has a sequence corresponding to the sequence SEQ ID No: 1; this coronavirus strain is derived from the sample collected from the bronchoalveolar washings from a patient suffering from SARS, recorded under the No. 031589 and collected at the Hanoi (Vietnam) French hospital.

[0022] In accordance with the invention, said sequence SEQ ID No: 1 is that of the deoxyribonucleic acid corresponding to the ribonucleic acid molecule of the genome of the isolated coronavirus strain as defined above.

[0023] The sequence SEQ ID No: 1 is distinguishable from the Genbank sequence AY274119.3 (Tor2 isolate) in that it possesses the following mutations:

[0024] g/t at position 23220; the alanine codon (gct) at position 577 of the amino acid sequence of the Tor2 S protein is replaced by a serine codon (tct),

[0025] a/g at position 25298; the arginine codon (aga) at position 11 of the amino acid sequence of the protein encoded by the Tor2 ORF3 is replaced by a glycine codon (gga).

[0026] In addition, the sequence SEQ ID No: 1 is distinguishable from the Genbank sequence AY278741 (Urbani isolate) in that it possesses the following mutations:

[0027] t/c at position 7919; the valine codon (ggt) in position 2552 of the amino acid sequence of the protein encoded by ORF1a is replaced by an alanine codon (gct),

[0028] t/c at position 16622: this mutation does not modify the amino acid sequence of the proteins encoded by ORF1b (silent mutation),

[0029] g/a at position 19064: this mutation does not modify the amino acid sequence of the proteins encoded by ORF1b (silent mutation),

[0030] c/t at position 24872: this mutation does not modify the amino acid sequence of the S protein, and

[0031] c/t at position 26857: the proline codon (ccc) at position 154 of the amino acid sequence of the M protein is replaced by a serine codon (tcc).

[0032] Unless otherwise stated, the positions of the nucleotide and peptide sequences are indicated with reference to the Genbank sequence AY274119.3.

[0033] The subject of the present invention is also an isolated or purified polynucleotide, characterized in that its sequence is that of the genome of the isolated coronavirus strain as defined above.

[0034] According to an advantageous embodiment of said polynucleotide, it has the sequence SEQ ID No: 1.

[0035] The subject of the present invention is also an isolated or purified polynucleotide, characterized in that its sequence hybridizes under high stringency conditions with the sequence of the polynucleotide as defined above.

[0036] The terms "isolated or purified" mean modified "by the hand of humans" from the natural state; in other words if an object exists in nature, it is said to be isolated or purified if it is modified or extracted from its natural environment or both. For example, a polynucleotide or a protein/peptide naturally present in a living organism is neither isolated nor purified; on the other hand, the same polynucleotide or protein/peptide separated from coexisting molecules in its natural environment, obtained by cloning, amplification and/or chemical synthesis is isolated for the purposes of the present invention. Furthermore, a polynucleotide or a protein/peptide which is introduced into an organism by transformation, genetic manipulation or by any other method, is "isolated" even if it is present in said organism. The term purified as used in the present invention means that the proteins/peptides according to the invention are essentially free of association with the other proteins or polypeptides, as is for example the product purified from the culture of recombinant host cells or the product purified from a nonrecombinant source.

[0037] For the purposes of the present invention, high stringency hybridization conditions are understood to mean temperature and ionic strength conditions chosen such that they make it possible to maintain the specific and selective hybridization between complementary polynucleotides.

[0038] By way of illustration, high stringency conditions for the purposes of defining the above polynucleotides are

advantageously the following: the DNA-DNA or DNA-RNA hybridization is performed in two steps: (1) prehybridization at 42° C. for 3 hours in phosphate buffer (20 mM, pH 7.5) containing 5×SSC (1×SSC corresponds to a 0.15 M NaCl+0.015 M sodium citrate solution), 50% formamide, 7% sodium dodecyl sulfate (SDS), 10× Denhardt's, 5% dextran sulfate and 1% salmon sperm DNA; (2) hybridization for 20 hours at 42° C. followed by 2 washings of 20 minutes at 20° C. in 2×SSC+2% SDS, 1 washing of 20 minutes at 20° C. in 0.1×SSC+0.1% SDS. The final washing is performed in 0.1×SSC+0.1% SDS for 30 minutes at 60° C.

[0039] The subject of the present invention is also a representative fragment of the polynucleotide as defined above, characterized in that it is capable of being obtained either by the use of restriction enzymes whose recognition and cleavage sites are present in said polynucleotide as defined above, or by amplification with the aid of oligonucleotide primers specific for said polynucleotide as defined above, or by transcription in vitro, or by chemical synthesis.

[0040] According to an advantageous embodiment of said fragment, it is selected from the group consisting of: the cDNA corresponding to at least one open reading frame (ORF) chosen from: ORF1a, ORF1b, ORF-S, ORF-E, ORF-M, ORF-N, ORF3, ORF4, ORF7 to ORF11, ORF13 and ORF14 and the cDNA corresponding to the noncoding 5' or 3' ends of said polynucleotide.

[0041] According to an advantageous feature of this embodiment, said fragment has a sequence selected from the group consisting of:

[0042] the sequences SEQ ID NO: 2 and 4 representing the cDNA corresponding to the ORF-S which encodes the S protein,

[0043] the sequences SEQ ID NO: 13 and 15 representing the cDNA corresponding to the ORF-E which encodes the E protein,

[0044] the sequences SEQ ID NO: 16 and 18 representing the cDNA corresponding to the ORF-M which encodes the M protein,

[0045] the sequences SEQ ID NO: 36 and 38 representing the cDNA corresponding to the ORF-N which encodes the N protein,

[0046] the sequences representing the cDNA corresponding respectively: to ORF1a and ORF1b (ORF1ab, SEQ ID NO: 31), to ORF3 and ORF4 (SEQ ID NO: 7, 8), to ORF7 to 11 (SEQ ID NO: 19, 20) to ORF13 (SEQ ID NO: 32) and to ORF14 (SEQ ID NO: 34), and

[0047] the sequences representing the cDNAs corresponding respectively to the noncoding 5' (SEQ ID NO: 39 and 72) and 3' (SEQ ID NO: 40, 73) ends of said polynucleotide.

[0048] The subject of the present invention is also a cDNA fragment encoding the S protein, as defined above, characterized in that it has a sequence selected from the group consisting of the sequences SEQ ID NO: 5 and 6 (Sa and Sb fragments).

[0049] The subject of the present invention is also a cDNA fragment corresponding to ORF1a and ORF1b as defined

above, characterized in that it has a sequence selected from the group consisting of the sequences SEQ ID NO: 41 to 54 (L0 to L12 fragments).

[0050] The subject of the present invention is also a polynucleotide fragment as defined above, characterized in that it has at least 15 consecutive bases or base pairs of the genome of said strain including at least one of those situated in position 7979, 16622, 19064, 23220, 24872, 25298 and 26857. Preferably this is a fragment of 20 to 2500 bases or base pairs, preferably from 20 to 400.

[0051] According to an advantageous embodiment of said fragment, it includes at least one pair of bases or base pairs corresponding to the following positions: 7919 and 23220, 7919 and 25298, 16622 and 23220, 19064 and 23220, 16622 and 25298, 19064 and 25298, 23220 and 24872, 23220 and 26857, 24872 and 25298, 25298 and 26857.

[0052] The subject of the present invention is also primers of at least 18 bases capable of amplifying a fragment of the genome of a SARS-associated coronavirus or of the DNA equivalent thereof.

[0053] According to an embodiment of said primers, they are selected from the group consisting of:

[0054] the pair of primers No. 1 corresponding respectively to positions 28507 to 28522 (sense primer, SEQ ID NO: 60) and 28774 to 28759 (antisense primer, SEQ ID NO: 61) of the sequence of the polynucleotide as defined above,

[0055] the pair of primers No. 2 corresponding respectively to positions 28375 to 28390 (sense primer, SEQ ID NO: 62) and 28702 to 28687 (antisense primer, SEQ ID NO: 63) of the sequence of the polynucleotide as defined above, and

[0056] the pair of primers consisting of the primers SEQ ID Nos: 55 and 56.

[0057] The subject of the present invention is also a probe capable of detecting the presence of the genome of a SARS-associated coronavirus or of a fragment thereof, characterized in that it is selected from the group consisting of: the fragments as defined above and the fragments corresponding to the following positions of the polynucleotide sequence as defined above: 28561 to 28586, 28588 to 28608, 28541 to 28563 and 28565 to 28589 (SEQ ID NO: 64 to 67).

[0058] The probes and primers according to the invention may be labeled directly or indirectly with a radioactive or nonradioactive compound by methods well known to persons skilled in the art so as to obtain a detectable and/or quantifiable signal. Among the radioactive isotopes used, there may be mentioned ³²P, ³³P, ³⁵S, ³H or ¹²⁵I. The nonradioactive entities are selected from ligands such as biotin, avidin, streptavidin, digoxigenin, haptens, dyes, luminescent agents such as radioluminescent, chemoluminescent, bioluminescent, fluorescent and phosphorescent agents.

[0059] The invention encompasses the labeled probes and primers derived from the preceding sequences.

[0060] Such probes and primers are useful for the diagnosis of infection by a SARS-associated coronavirus.

[0061] The subject of the present invention is also a method for the detection of a SARS-associated coronavirus, from a biological sample, which method is characterized in that it comprises at least:

[0062] (a) the extraction of nucleic acids present in said biological sample,

[0063] (b) the amplification of a fragment of ORF-N by RT-PCR with the aid of a pair of primers as defined above, and

[0064] (c) the detection, by any appropriate means, of the amplification products obtained in (b).

[0065] The amplification products (amplicons) in (b) are 268 bp for the pair of primers No. 1 and 328 bp for the pair of primers No. 2.

[0066] According to an advantageous embodiment of said method, the step (b) of detection is carried out with the aid of at least one probe corresponding to positions 28561 to 28586, 28588 to 28608, 28541 to 28563 and 28565 to 28589 of the sequence of the polynucleotide as defined above.

[0067] Preferably, the SARS-associated coronavirus genome is detected and optionally quantified by PCR in real time with the aid of the pair of primers No. 2 and probes corresponding to positions 28541 to 28563 and 28565 to 28589 labeled with different compounds, in particular different fluorescent agents.

[0068] The real time RT-PCR which uses this pair of primers and this probe is very sensitive since it makes it possible to detect 10^2 copies of RNA and up to 10 copies of RNA; it is in addition reliable and reproducible.

[0069] The invention encompasses the single-stranded, double-stranded and triple-stranded polydeoxyribonucleotides and polyribonucleotides corresponding to the sequence of the genome of the isolated strain of coronavirus and its fragments as defined above, and to their sense or antisense complementary sequences, in particular the RNAs and cDNAs corresponding to the sequence of the genome and of its fragments as defined above.

[0070] The present invention also encompasses the amplification fragments obtained with the aid of primers specific for the genome of the purified or isolated strain as defined above, in particular with the aid of primers or pairs of primers as defined above, the restriction fragments formed by or comprising the sequence of fragments as defined above, the fragments obtained by transcription in vitro from a vector containing the sequence SEQ ID NO: 1 or a fragment as defined above, and fragments obtained by chemical synthesis. Examples of restriction fragments are deduced from the restriction map of the sequence SEQ ID NO: 1 illustrated by FIG. 13. In accordance with the invention, said fragments are either in the form of isolated fragments, or in the form of mixtures of fragments. The invention also encompasses fragments modified, in relation to the preceding ones, by removal or addition of nucleotides in a proportion of about 15%, relative to the length of the above fragments and/or modified in terms of the nature of the nucleotides, as long as the modified nucleotide fragments retain a capacity for hybridization with the genomic or antigenomic RNA sequences of the isolate as defined above.

[0071] The nucleic acid molecules according to the invention are obtained by conventional methods, known per se, following standard protocols such as those described in *Current Protocols in Molecular Biology* (Frederick M. AUSUBEL, 2000, Wiley and son Inc., Library of Congress, USA). For example, they may be obtained by amplification of a nucleic sequence by PCR or RT-PCR or alternatively by total or partial chemical synthesis.

[0072] The subject of the present invention is also a DNA or RNA chip or filter, characterized in that it comprises at least one polynucleotide or one of its fragments as defined above.

[0073] The DNA or RNA chips or filters according to the invention are prepared by conventional methods, known per se, such as for example chemical or electrochemical grafting of oligonucleotides on a glass or nylon support.

[0074] The subject of the present invention is also a recombinant cloning and/or expression vector, in particular a plasmid, a virus, a viral vector or a phage comprising a nucleic acid fragment as defined above. Preferably, said recombinant vector is an expression vector in which said nucleic acid fragment is placed under the control of appropriate elements for regulating transcription and translation. In addition, said vector may comprise sequences (tags) fused in phase with the 5' and/or 3' end of said insert, which are useful for the immobilization and/or detection and/or purification of the protein expressed from said vector.

[0075] These vectors are constructed and introduced into host cells by conventional recombinant DNA and genetic engineering methods which are known per se. Numerous vectors into which a nucleic acid molecule of interest may be inserted in order to introduce it and to maintain it in a host cell are known per se; the choice of an appropriate vector depends on the use envisaged for this vector (for example replication of the sequence of interest, expression of this sequence, maintenance of the sequence in extrachromosomal form or alternatively integration into the chromosomal material of the host), and on the nature of the host cell.

[0076] In accordance with the invention, said plasmid is selected in particular from the following plasmids:

[0077] the plasmid, called SARS-S, contained in the bacterial strain deposited under the No. I-3659, on Jun. 20, 2003, at the Collection Nationale de Cultures de Microorganismes, 25 rue du Docteur Roux, 75724 Paris Cedex 15; it contains the cDNA sequence encoding the S protein of the SARS-CoV strain derived from the sample recorded under the No. 031589, said sequence corresponding to the nucleotides at positions 21406 to 25348 (SEQ ID NO: 4), with reference to the Genbank sequence AY274119.3,

[0078] the plasmid, called SARS-S1, contained in the bacterial strain deposited under the No. I-3020, on May 12, 2003, at the Collection Nationale de Cultures de Microorganismes, 25 rue du Docteur Roux, 75724 Paris Cedex 15; it contains a 5' fragment of the cDNA sequence encoding the S protein of the SARS-CoV strain derived from the sample recorded under the No. 031589, as defined above, said fragment corresponding to the nucleotides at positions 21406 to 23454 (SEQ ID NO: 5), with reference to the Genbank sequence AY274119.3 Tor2,

- [0079] the plasmid, called SARS-S2, contained in the bacterial strain deposited under the No. I-3019, on May 12, 2003, at the Collection Nationale de Cultures de Microorganismes, 25 rue du Docteur Roux, 75724 Paris Cedex 15; it contains a 3' fragment of the cDNA sequence encoding the S protein of the SARS-CoV strain derived from the sample recorded under the number No. 031589, as defined above, said fragment corresponding to the nucleotides at positions 23322 to 25348 (SEQ ID NO: 6), with reference to the Genbank sequence accession No. AY274119.3,
- [0080] the plasmid, called SARS-SE, contained in the bacterial strain deposited under the No. I-3126, on Nov. 13, 2003, at the Collection Nationale de Cultures de Microorganismes, 25 rue du Docteur Roux, 75724 Paris Cedex 15; it contains the cDNA corresponding to the region situated between ORF-S and ORF-E and overlapping ORF-E of the SARS-CoV strain derived from the sample recorded under the No. 031589, as defined above, said region corresponding to the nucleotides at positions 25110 to 26244 (SEQ ID NO: 8), with reference to the Genbank sequence accession No. AY274119.3,
- [0081] the plasmid, called SARS-E, contained in the bacterial strain deposited under the No. I-3046, on May 28, 2003, at the Collection Nationale de Cultures de Microorganismes, 25 rue du Docteur Roux, 75724 Paris Cedex 15; it contains the cDNA sequence encoding the E protein of the SARSCoV strain derived from the sample recorded under the No. 031589, as defined above, said sequence corresponding to the nucleotides at positions 26082 to 26413 (SEQ ID NO: 15), with reference to the Genbank sequence accession No. AY274119.3,
- [0082] the plasmid, called SARS-M, contained in the bacterial strain deposited under the No. I-3047, on May 28, 2003, at the Collection Nationale de Cultures de Microorganismes, 25 rue du Docteur Roux, 75724 Paris Cedex 15; it contains the cDNA sequence encoding the M protein of the SARS-CoV strain derived from the sample recorded under the No. 031589, as defined above; said sequence corresponding to the nucleotides at positions 26330 to 27098 (SEQ ID NO: 18), with reference to the Genbank sequence accession No. AY274119.3,
- [0083] the plasmid, called SARS-MN, contained in the bacterial sequence deposited under the No. I-3125, on Nov. 13, 2003, at the Collection Nationale de Cultures de Microorganismes, 25 rue du Docteur Roux, 75724 Paris Cedex 15; it contains the cDNA sequence corresponding to the region situated between ORF-M and ORF-N of the SARS-CoV strain derived from the sample recorded under the No. 031589 and collected in Hanoi, as defined above, said sequence corresponding to the nucleotides at positions 26977 to 28218 (SEQ ID NO: 20), with reference to the Genbank accession No. AY274119.3,
- [0084] the plasmid, called SARS-N, contained in the bacterial strain deposited under the No. I-3048, on Jun. 5, 2003, at the Collection Nationale de Cultures de Microorganismes, 25 rue du Docteur Roux, 75724 Paris Cedex 15; it contains the cDNA encoding the N protein of the SARS-CoV strain derived from the sample recorded under the No. 031589, as defined above, said sequence corresponding to the nucleotides at positions 28054 to 29430 (SEQ ID NO: 38), with reference to the Genbank sequence accession No. AY274119.3; thus, this plasmid comprises an insert of sequence SEQ ID NO: 38 and is contained in a bacterial strain which was deposited under the No. I-3048, on Jun. 5, 2003, at the Collection Nationale de Cultures de Microorganismes, 25 rue du Docteur Roux, 75724 Paris Cedex 15,
- [0085] the plasmid, called SARS-5'NC, contained in the bacterial strain deposited under the No. I-3124, on Nov. 7, 2003, at the Collection Nationale de Cultures de Microorganismes, 25 rue du Docteur Roux, 75724 Paris Cedex 15; it contains the cDNA corresponding to the noncoding 5' end of the genome of the SARS-CoV strain derived from the sample recorded under the No. 031589, as defined above, said sequence corresponding to the nucleotides at positions 1 to 204 (SEQ ID NO: 39), with reference to the Genbank sequence accession No. AY274119.3,
- [0086] the plasmid called SARS-3'NC, contained in the bacterial strain deposited under the No. I-3123 on Nov. 7, 2003, at the Collection Nationale de Cultures de Microorganismes, 25 rue du Docteur Roux, 75724 Paris Cedex 15; it contains the cDNA sequence corresponding to the noncoding 3' end of the genome of the SARS-CoV strain derived from the sample recorded under the No. 031589, as defined above, said sequence corresponding to that situated between the nucleotide and position 28933 to 29727 (SEQ ID NO: 40), with reference to the Genbank sequence accession No. AY274119.3, ends with a series of nucleotides a.,
- [0087] the expression plasmid, called pIV2.3N, containing a cDNA fragment encoding a C-terminal fusion of the N protein (SEQ ID NO: 37) with a polyhistidine tag,
- [0088] the expression plasmid, called pIV2.3S_C, containing a cDNA fragment encoding a C-terminal fusion of the fragment corresponding to positions 475 to 1193 of the amino acid sequence of the S protein (SEQ ID NO: 3) with a polyhistidine tag,
- [0089] the expression plasmid, pIV2.3S_L, containing a cDNA fragment encoding a C-terminal fusion of the fragment corresponding to positions 14 to 1193 of the amino acid sequence of the S protein (SEQ ID NO: 3) with a polyhistidine tag,
- [0090] the expression plasmid, called pIV2.4N, containing a cDNA fragment encoding a N-terminal fusion of the N protein (SEQ ID NO: 3) with a polyhistidine tag,
- [0091] the expression plasmid, called pIV2.4S_C or pIV2.4S₁, containing an insert encoding a N-terminal fusion of the fragment corresponding to positions 475 to 1193 of the amino acid sequence of the S protein (SEQ ID NO: 3) with a polyhistidine tag, and
- [0092] the expression plasmid, called pIV2.4S_L, containing a cDNA fragment encoding an N-terminal fusion of the fragment corresponding to positions 14 to

1193 of the amino acid sequence of the S protein (SEQ ID NO: 3) with a polyhistidine tag.

[0093] According to an advantageous feature of the expression plasmid as defined above, it is contained in a bacterial strain which was deposited under the No. I-3117, on Oct. 23, 2003, at the Collection Nationale de Cultures de Microorganismes, 25 rue du Docteur Roux, 75724 Paris Cedex 15.

[0094] According to another advantageous feature of the expression plasmid as defined above, it is contained in a bacterial strain which was deposited under the No. I-3118, on Oct. 23, 2003, at the Collection Nationale de Cultures de Microorganismes, 25 rue du Docteur Roux, 75724 Paris Cedex 15.

[0095] According to another feature of the expression plasmid as defined above, it is contained in a bacterial strain which was deposited at the CNCM, 25 rue du Docteur Roux, 75724 Paris Cedex 15 under the following numbers:

- [0096] a) strain No. I-3118, deposited on Oct. 23, 2003,
- [0097] b) strain No. I-3019, deposited on May 12, 2003,
- [0098] c) strain No. I-3020, deposited on May 12, 2003,
- [0099] d) strain No. I-3059, deposited on Jun. 20, 2003,
- [0100] e) strain No. I-3323, deposited on Nov. 22, 2004,
- [0101] f) strain No. I-3324, deposited on Nov. 22, 2004,
- [0102] g) strain No. I-3326, deposited on Dec. 1, 2004,
- [0103] h) strain No. I-3327, deposited on Dec. 1, 2004,
- [0104] i) strain No. I-3332, deposited on Dec. 1, 2004,
- [0105] j) strain No. I-3333, deposited on Dec. 1, 2004,
- [0106] k) strain No. I-3334, deposited on Dec. 1, 2004,
- [0107] l) strain No. I-3335, deposited on Dec. 1, 2004,
- [0108] m) strain No. I-3336, deposited on Dec. 1, 2004,
- [0109] n) strain No. I-3337, deposited on Dec. 1, 2004,
- [0110] o) strain No. I-3338, deposited on Dec. 2, 2004,
- [0111] p) strain No. I-3339, deposited on Dec. 2, 2004,
- [0112] q) strain No. I-3340, deposited on Dec. 2, 2004,
- [0113] r) strain No. I-3341, deposited on Dec. 2, 2004.

[0114] The subject of the present invention is also a nucleic acid insert of viral origin, characterized in that it is contained in any of the strains as defined above in a)-r).

[0115] The subject of the present invention is also a nucleic acid containing a synthetic gene allowing optimized expression of the S protein in eukaryotic cells, characterized in that it possesses the sequence SEQ ID NO: 140.

[0116] The subject of the present invention is also an expression vector containing a nucleic acid containing a synthetic gene allowing optimized expression of the S protein, which vector is contained in the bacterial strain deposited at the CNCM, on Dec. 1, 2004, under the No. I-3333.

[0117] According to one embodiment of said expression vector, it is a viral vector, in the form of a viral particle or in the form of a recombinant genome.

[0118] According to an advantageous feature of this embodiment, this is a recombinant viral particle or a recombinant viral genome capable of being obtained by transfection of a plasmid according to paragraphs g), h) and k) to r) as defined above, in an appropriate cellular system, that is to say, for example, cells transfected with one or more other plasmids intended to transcomplement certain functions of the virus that are deleted in the vector and that are necessary for the formation of the viral particles.

[0119] The expression "S protein family" is understood here to mean the complete S protein, its ectodomain and fragments of this ectodomain which are preferably produced in a eukaryotic system.

[0120] The subject of the present invention is also a lentiviral vector encoding a polypeptide of the S protein family, as defined above.

[0121] The subject of the present invention is also a recombinant measles virus encoding a polypeptide of the S protein family, as defined above.

[0122] The subject of the present invention is also a recombinant vaccinia virus encoding a polypeptide of the S protein family, as defined above.

[0123] The subject of the present invention is also the use of a vector according to paragraphs e) to r) as defined above, or of a vector containing a synthetic gene for the S protein, as defined above, for the production, in a eukaryotic system, of the SARS-associated coronavirus S protein or of a fragment of this protein.

[0124] The subject of the present invention is also a method for producing the S protein in a eukaryotic system, comprising a step of transfecting eukaryotic cells in culture with a vector chosen from the vectors contained in the bacterial strains mentioned in paragraphs e) to r) above or a vector containing a synthetic gene allowing optimized expression of the S protein.

[0125] The subject of the present invention is also a cDNA library characterized in that it comprises fragments as defined above, in particular amplification fragments or restriction fragments, cloned into a recombinant vector, in particular an expression vector (expression library).

[0126] The subject of the present invention is also cells, in particular prokaryotic cells, modified by a recombinant vector as defined above.

[0127] The subject of the present invention is also a genetically modified eukaryotic cell expressing a protein or a polypeptide as defined above. Quite obviously, the terms "genetically modified eukaryotic cell" do not denote a cell modified with a wild-type virus.

[0128] According to an advantageous embodiment of said cell, it is capable of being obtained by transfection with any of the vectors mentioned in paragraphs i) to l) above.

[0129] According to an advantageous feature of this embodiment, this is the cell FRhK4-Sso1-30, deposited at the CNCM on Nov. 22, 2004, under the No. I-3325.

[0130] The recombinant vectors as defined above and the cells transformed with said expression vectors are advantageously used for the production of the corresponding proteins and peptides. The expression libraries derived from

said vectors, and the cells transformed with said expression libraries are advantageously used to identify the immunogenic epitopes (B and T epitopes) of the SARS-associated coronavirus proteins.

[0131] The subject of the present invention is also the purified or isolated proteins and peptides, characterized in that they are encoded by the polynucleotide or one of its fragments as defined above.

[0132] According to an advantageous embodiment of the invention, said protein is selected from the group consisting of:

[0133] the S protein having the sequence SEQ ID NO: 3 or its ectodomaine

[0134] the E protein having the sequence SEQ ID NO: 14

[0135] the M protein having the sequence SEQ ID NO: 17

[0136] the N protein having the sequence SEQ ID NO: 37

[0137] the proteins encoded by the ORFs: ORF1a, ORF1b, ORF3, ORF4 and ORF7 to ORF11, ORF13 and ORF14 and having the respective sequence, SEQ ID NO: 74, 75, 10, 12, 22, 24, 26, 28, 30, 33 and 35.

[0138] The terms "ectodomaine of the S protein" and "soluble form of the S protein" will be used interchangeably below.

[0139] According to an advantageous embodiment of the invention, said polypeptide consists of the amino acids corresponding to positions 1 to 1193 of the amino acid sequence of the S protein.

[0140] According to another advantageous embodiment of the invention, said peptide is selected from the group consisting of:

[0141] a) the peptides corresponding to positions 14 to 1193 and 475 to 1193 of the amino acid sequence of the S protein,

[0142] b) the peptides corresponding to positions 2 to 14 (SEQ ID NO: 69) and 100 to 221 of the amino acid sequence of the M protein; these peptides correspond respectively to the ectodomaine and to the endodomaine of the M protein, and

[0143] c) the peptides corresponding to positions 1 to 12 (SEQ ID NO: 70) and 53 to 76 (SEQ ID NO: 71) of the amino acid sequence of the E protein; these peptides correspond respectively to the ectodomaine and to the C-terminal end of the E protein, and

[0144] d) the peptides of 5 to 50 consecutive amino acids, preferably of 10 to 30 amino acids, inclusive or partially or completely overlapping the sequence of the peptides as defined in a), b) or c).

[0145] The subject of the present invention is also a peptide, characterized in that it has a sequence of 7 to 50 amino acids including an amino acid residue selected from the group consisting of:

[0146] the alanine situated at position 2552 of the amino acid sequence of the protein encoded by ORF1a,

[0147] the serine situated at position 577 of the amino acid sequence of the S protein of the SARS-CoV strain as defined above,

[0148] the glycine at position 11 of the amino acid sequence of the protein encoded by ORF3 of the SARS-CoV strain as defined above,

[0149] the serine at position 154 of the amino acid sequence of the M protein of the SARS-CoV strain as defined above.

[0150] The subject of the present invention is also an antibody or a polyclonal or monoclonal antibody fragment which can be obtained by immunization of an animal with a recombinant vector as defined above, a cDNA library as defined above or alternatively a protein or a peptide as defined above, characterized in that it binds to at least one of the proteins encoded by SARS-CoV as defined above.

[0151] The invention encompasses the polyclonal antibodies, the monoclonal antibodies, the chimeric antibodies such as the humanized antibodies, and fragments thereof (Fab, Fv, scFv).

[0152] A subject of the present invention is also a hybridoma producing a monoclonal antibody against the N protein, characterized in that it is chosen from the following hybridomas:

[0153] the hybridoma producing the monoclonal antibody 87, deposited at the CNCM on Dec. 1, 2004 under the number I-3328,

[0154] the hybridoma producing the monoclonal antibody 86, deposited at the CNCM on Dec. 1, 2004 under the number I-3329,

[0155] the hybridoma producing the monoclonal antibody 57, deposited at the CNCM on Dec. 1, 2004 under the number I-3330, and

[0156] the hybridoma producing the monoclonal antibody 156, deposited at the CNCM on Dec. 1, 2004 under the number I-3331.

[0157] The subject of the present invention is also a polyclonal or monoclonal antibody or antibody fragment directed against the N protein, characterized in that it is produced by a hybridoma as defined above.

[0158] For the purposes of the present invention, the expression chimeric antibody is understood to mean, in relation to an antibody of a particular animal species or of a particular class of antibody, an antibody comprising all or part of a heavy chain and/or of a light chain of an antibody of another animal species or of another class of antibody.

[0159] For the purposes of the present invention, the expression humanized antibody is understood to mean a human immunoglobulin in which the residues of the CDRs (Complementary Determining Regions) which form the antigen-binding site are replaced by those of a nonhuman monoclonal antibody possessing the desired specificity, affinity or activity. Compared with the nonhuman antibodies, the humanized antibodies are less immunogenic and possess a prolonged half-life in humans because they possess only a small proportion of nonhuman sequences given that practically all the residues of the FR (Framework) regions and of

the constant (Fc) region of these antibodies are those of a consensus sequence of human immunoglobulins.

[0160] A subject of the present invention is also a protein chip or filter, characterized in that it comprises a protein, a peptide or alternatively an antibody as defined above.

[0161] The protein chips according to the invention are prepared by conventional methods known per se. Among the appropriate supports on which proteins may be immobilized, there may be mentioned those made of plastic or glass, in particular in the form of microplates.

[0162] The subject of the present invention is also reagents derived from the isolated strain of SARS-associated coronavirus, derived from the sample recorded under the No. 031589, which are useful for the study and diagnosis of the infection caused by a SARS-associated coronavirus, said reagents are selected from the group consisting of:

[0163] (a) a pair of primers, a probe or a DNA chip as defined above,

[0164] (b) a recombinant vector or a modified cell as defined above,

[0165] (c) an isolated coronavirus strain or a polynucleotide as defined above,

[0166] (d) a protein or a peptide as defined above,

[0167] (e) an antibody or an antibody fragment as defined above, and

[0168] (f) a protein chip as defined above.

[0169] These various reagents are prepared and used according to conventional molecular biology and immunology techniques following standard protocols such as those described in *Current Protocols in Molecular Biology* (Frederick M. AUSUBEL, 2000, Wiley and Son Inc., Library of Congress, USA), in *Current Protocols in Immunology* (John E. Cologan, 2000, Wiley and Son Inc., Library of Congress, USA) and in *Antibodies: A Laboratory Manual* (E. Howell and D. Lane, Cold Spring Harbor Laboratory, 1988).

[0170] The nucleic acid fragments according to the invention are prepared and used according to conventional techniques as defined above. The peptides and proteins according to the invention are prepared by recombinant DNA techniques, known to persons skilled in the art, in particular with the aid of the recombinant vectors as defined above. Alternatively, the peptides according to the invention may be prepared by conventional techniques of solid or liquid phase synthesis, known to persons skilled in the art.

[0171] The polyclonal antibodies are prepared by immunizing an appropriate animal with a protein or a peptide as defined above, optionally coupled to KLH or to albumin and/or combined with an appropriate adjuvant such as (complete or incomplete) Freund's adjuvant or aluminum hydroxide; after obtaining a satisfactory antibody titer, the antibodies are harvested by collecting serum from the immunized animals and enriched with IgG by precipitation, according to conventional techniques, and then the IgGs specific for the SARS-CoV proteins are optionally purified by affinity chromatography on an appropriate column to which said peptide or said protein is attached, as defined above, so as to obtain a monospecific IgG preparation.

[0172] The monoclonal antibodies are produced from hybridomas obtained by fusion of B lymphocytes from an animal immunized with a protein or a peptide as defined above with myelomas, according to the Köhler and Milstein technique (Nature, 1975, 256, 495-497); the hybridomas are cultured in vitro, in particular in fermenters or produced in vivo, in the form of ascites; alternatively, said monoclonal antibodies are produced by genetic engineering as described in American U.S. Pat. No. 4,816,567.

[0173] The humanized antibodies are produced by general methods such as those described in International application WO 98/45332.

[0174] The antibody fragments are produced from the cloned V_H and V_L regions, from the mRNAs of hybridomas or splenic lymphocytes of an immunized mouse; for example, the Fv, scFv or Fab fragments are expressed at the surface of filamentous phages according to the Winter and Milstein technique (Nature, 1991, 349, 293-299); after several selection steps, the antibody fragments specific for the antigen are isolated and expressed in an appropriate expression system, by conventional techniques for cloning and expression of recombinant DNA.

[0175] The antibodies or fragments thereof as defined above are purified by conventional techniques known to persons skilled in the art, such as affinity chromatography.

[0176] The subject of the present invention is additionally the use of a product selected from the group consisting of: a pair of primers, a probe, a DNA chip, a recombinant vector, a modified cell, an isolated coronavirus strain, a polynucleotide, a protein or a peptide, an antibody or an antibody fragment and a protein chip as defined above, for the preparation of a reagent for the detection and optionally genotyping/serotyping of a SARS-associated coronavirus.

[0177] The proteins and peptides according to the invention, which are capable of being recognized and/or of inducing the production of antibodies specific for the SARS-associated coronavirus, are useful for the diagnosis of infection with such a coronavirus; the infection is detected, by an appropriate technique—in particular EIA, ELISA, RIA, immunofluorescence—in a biological sample collected from an individual capable of being infected.

[0178] According to an advantageous feature of said use, said proteins are selected from the group consisting of the S, E, M and/or N proteins and the peptides as defined above.

[0179] The S, E, M and/or N proteins and the peptides derived from these proteins as defined above, for example the N protein, are used for the indirect diagnosis of a SARS-associated coronavirus infection (serological diagnosis; detection of an antibody specific for SARS-CoV), in particular by an immunoenzymatic method (ELISA).

[0180] The antibodies and antibody fragments according to the invention, in particular those directed against the S, E, M and/or N proteins and the derived peptides as defined above, are useful for the direct diagnosis of a SARS-associated coronavirus infection; the detection of the protein(s) of SARS-CoV is carried out by an appropriate technique, in particular EIA, ELISA, RIA, immunofluorescence, in a biological sample collected from an individual capable of being infected.

[0181] The subject of the present invention is also a method for the detection of a SARS-associated coronavirus, from a biological sample, which method is characterized in that it comprises at least:

[0182] (a) bringing said biological sample into contact with at least one antibody or one antibody fragment, one protein, one peptide or alternatively one protein or peptide chip or filter as defined above, and

[0183] (b) visualizing by any appropriate means antigen-antibody complexes formed in (a), for example by EIA, ELISA, RIA, or by immunofluorescence.

[0184] According to one advantageous embodiment of said process, step (a) comprises:

[0185] (a₁) bringing said biological sample into contact with at least a first antibody or an antibody fragment which is attached to an appropriate support, in particular a microplate,

[0186] (a₂) washing the solid phase, and

[0187] (a₃) adding at least a second antibody or an antibody fragment, different from the first, said antibody or antibody fragment being optionally appropriately labeled.

[0188] This method, which makes it possible to capture the viral particles present in the biological sample, is also called immunocapture method.

[0189] For example:

[0190] step (a₁) is carried out with at least a first monoclonal or polyclonal antibody or a fragment thereof, directed against the S, M and/or E protein, and/or a peptide corresponding to the ectodomain of one of these proteins (M2-14 or E1-12 peptides)

[0191] step (a₃) is carried out with at least one antibody or an antibody fragment directed against another epitope of the same protein or preferably against another protein, preferably against an inner protein such as the N nucleoprotein or the endodomain of the E or M protein, more preferably still these are antibodies or antibody fragments directed against the N protein which is very abundant in the viral particle; when an antibody or an antibody fragment directed against an inner protein (N) or against the endodomain of the E or M proteins is used, said antibody is incubated in the presence of detergent, such as Tween 20 for example, at concentrations of the order of 0.1%.

[0192] step (b) for visualizing the antigen-antibody complexes formed is carried out, either directly with the aid of a second antibody labeled for example with biotin or an appropriate enzyme such as peroxidase or alkaline phosphatase, or indirectly with the aid of an anti-immunoglobulin serum labeled as above. The complexes thus formed are visualized with the aid of an appropriate substrate.

[0193] According to a preferred embodiment of this aspect of the invention, the biological sample is mixed with the visualizing monoclonal antibody prior to its being brought into contact with the capture monoclonal antibodies. Where appropriate, the serum-visualizing antibody mixture is incubated for at least 10 minutes at room temperature before being applied to the plate.

[0194] The subject of the present invention is also an immunocapture test intended to detect an infection by the SARS-associated coronavirus by detecting the native nucleoprotein (N protein), in particular characterized in that the antibody used for the capture of the native viral nucleoprotein is a monoclonal antibody specific for the central region and/or for a conformational epitope.

[0195] According to one embodiment of said test, the antibody used for the capture of the N protein is the monoclonal antibody mAb87, produced by the hybridoma deposited at the CNCM on Dec. 1, 2004 under the number I-3328.

[0196] According to another embodiment of said immunocapture test, the antibody used for the capture of the N protein is the monoclonal antibody mAb86, produced by the hybridoma deposited at the CNCM on Dec. 1, 2004 under the number I-3329.

[0197] According to another embodiment of said immunocapture test, the monoclonal antibodies mAb86 and mAb87 are used for the capture of the N protein.

[0198] In the immunocapture tests according to the invention, it is possible to use, for visualizing the N protein, the monoclonal antibody mAb57, produced by the hybridoma deposited at the CNCM on Dec. 1, 2004 under the number I-3330, said antibody being conjugated with a visualizing molecule or particle.

[0199] In accordance with said immunocapture test, a combination of the antibodies mAb57 and mAb87, conjugated with a visualizing molecule or particle, is used for the visualization of the N protein.

[0200] A visualizing molecule may be a radioactive atom, a dye, a fluorescent molecule, a fluorophore, an enzyme; a visualizing particle may be for example: colloidal gold, a magnetic particle or a latex bead.

[0201] The subject of the present invention is also a reagent for detecting a SARS-associated coronavirus, characterized in that it is selected from the group consisting of:

[0202] (a) a pair of primers or a probe as defined above,

[0203] (b) a recombinant vector as defined above or a modified cell as defined above,

[0204] (c) an isolated coronavirus strain as defined above or a polynucleotide as defined above,

[0205] (d) an antibody or an antibody fragment as defined above,

[0206] (e) a combination of antibodies comprising the monoclonal antibodies mAb86 and/or mAb87, and the monoclonal antibody mAb57, as defined above,

[0207] (f) a chip or a filter as defined above.

[0208] The subject of the present invention is also a method for the detection of a SARS-associated coronavirus infection, from a biological sample, by indirect IgG ELISA using the N protein, which method is characterized in that the plates are sensitized with an N protein solution at a concentration of between 0.5 and 4 µg/ml, preferably to 2 µg/ml, in a 10 mM PBS buffer pH 7.2, phenol red at 0.25 ml/l.

[0209] The subject of the present invention is additionally a method for the detection of a SARS-associated coronavirus infection, from a biological sample, by double epitope ELSA, characterized in that the serum to be tested is mixed with the visualizing antigen, said mixture then being brought into contact with the antigen attached to a solid support.

[0210] According to one variant of the tests for detecting SARS-associated coronaviruses, these tests combine an ELSA using the N protein, and another ELSA using the S protein, as described below.

[0211] The subject of the present invention is also an immune complex formed of a polyclonal or monoclonal antibody or antibody fragment as defined above, and of a SARS-associated coronavirus protein or peptide.

[0212] The subject of the present invention is additionally a SARS-associated coronavirus detection kit, characterized in that it comprises at least one reagent selected from the group consisting of: a pair of primers, a probe, a DNA or RNA chip, a recombinant vector, a modified cell, an isolated coronavirus strain, a polynucleotide, a protein or a peptide, an antibody, and a protein chip as defined above.

[0213] The subject of the present invention is additionally an immunogenic composition, characterized in that it comprises at least one product selected from the group consisting of:

- [0214] a) a protein or a peptide as defined above,
- [0215] b) a polynucleotide of the DNA or RNA type or one of its representative fragments as defined above, having a sequence chosen from:
 - [0216] (i) the sequence SEQ ID NO: 1 or its RNA equivalent
 - [0217] (ii) the sequence hybridizing under high stringency conditions with the sequence SEQ ID NO: 1,
 - [0218] (iii) the sequence complementary to the sequence SEQ ID NO: 1 or to the sequence hybridizing under high stringency conditions with the sequence SEQ ID NO: 1,
 - [0219] (iv) the nucleotide sequence of a representative fragment of the polynucleotide as defined in (i), (ii) or (iii),
 - [0220] (v) the sequence as defined in (i), (ii), (iii) or (iv), modified, and
- [0221] c) a recombinant expression vector comprising a polynucleotide as defined in b), and
- [0222] d) a cDNA library as defined above,

said immunogenic composition being capable of inducing protective humoral or cellular immunity specific for the SARS-associated coronavirus, in particular the production of an antibody directed against a specific epitope of the SARS-associated coronavirus.

[0223] The proteins and peptides as defined above, in particular the S, M, E and/or N proteins and the derived peptides, and the nucleic acid (DNA or RNA) molecules encoding said proteins or said peptides are good candidate vaccines and may be used in immunogenic compositions for the production of a vaccine against the SARS-associated coronavirus.

[0224] According to an advantageous embodiment of the compositions according to the invention, they additionally contain at least one pharmaceutically acceptable vehicle and optionally carrier substances and/or adjuvants.

[0225] The pharmaceutically acceptable vehicles, the carrier substances and the adjuvants are those conventionally used.

[0226] The adjuvants are advantageously chosen from the group consisting of oily emulsions, saponin, mineral substances, bacterial extracts, aluminum hydroxide and squalene.

[0227] The carrier substances are advantageously selected from the group consisting of unilamellar liposomes, multilamellar liposomes, micelles of saponin or solid microspheres of a saccharide or auriferous nature.

[0228] The compositions according to the invention are administered by the general route, in particular by the intramuscular or subcutaneous route or alternatively by the local, in particular nasal (aerosol) route.

[0229] The subject of the present invention is also the use of an isolated or purified protein or peptide having a sequence selected from the group consisting of the sequences SEQ ID NO: 3, 10, 12, 14, 17, 22, 24, 26, 28, 30, 33, 35, 37, 69, 70, 71, 74 and 75 to form an immune complex with an antibody specifically directed against an epitope of the SARS-associated coronavirus.

[0230] The subject of the present invention is also an immune complex consisting of an isolated or purified protein or peptide having a sequence selected from the group consisting of the sequences SEQ ID NO: 3, 10, 12, 14, 17, 22, 24, 26, 28, 30, 33, 35, 37, 69, 70, 71, 74 and 75, and of an antibody specifically directed against an epitope of the SARS-associated coronavirus.

[0231] The subject of the present invention is also the use of an isolated or purified protein or peptide having a sequence selected from the group consisting of the sequences SEQ ID NO: 3, 10, 12, 14, 17, 22, 24, 26, 28, 30, 33, 35, 37, 69, 70, 71, 74 and 75 to induce the production of an antibody capable of specifically recognizing an epitope of the SARS-associated coronavirus.

[0232] The subject of the present invention is also the use of an isolated or purified polynucleotide having a sequence selected from the group consisting of the sequences SEQ ID NO: 1, 2, 4, 7, 8, 13, 15, 16, 18, 19, 20, 31, 36 and 38 to induce the production of an antibody directed against the protein encoded by said polynucleotide and capable of specifically recognizing an epitope of the SARS-associated coronavirus.

[0233] The subject of the present invention is also monoclonal antibodies recognizing the native S protein of a SARS-associated coronavirus.

[0234] The subject of the present invention is also the use of a protein or a polypeptide of the S protein family, as defined above, or of an antibody recognizing the native S protein, as defined above, to detect an infection by a SARS-associated coronavirus, in a biological sample.

[0235] The subject of the present invention is also a method for detecting an infection by a SARS-associated coronavirus, in a biological sample, characterized in that the

detection is carried out by ELISA using the recombinant S protein, expressed in a eukaryotic system.

[0236] According to an advantageous embodiment of said method, it is a double epitope ELISA method, and the serum to be tested is mixed with the visualizing antigen, said mixture then being brought into contact with the antigen attached to a solid support.

[0237] The subject of the present invention is also an immune complex consisting of a monoclonal antibody or antibody fragment recognizing the native S protein, and of a protein or a peptide of the SARS-associated coronavirus.

[0238] The subject of the present invention is also an immune complex consisting of a protein or a polypeptide of the S protein family, as defined above, and of an antibody specifically directed against an epitope of the SARS-associated coronavirus.

[0239] The subject of the present invention is additionally a SARS-associated coronavirus detection kit or box, characterized in that it comprises at least one reagent selected from the group consisting of: a protein or polypeptide of the S protein family, as defined above, a nucleic acid encoding a protein or peptide of the S protein family, as defined above,

a cell expressing a protein or polypeptide of the S protein family, as defined above, or an antibody recognizing the native S protein of a SARS-associated coronavirus.

[0240] The subject of the present invention is an immunogenic and/or vaccine composition, characterized in that it comprises a polypeptide or a recombinant protein of the S protein family, as defined above, obtained in a eukaryotic expression system.

[0241] The subject of the present invention is also an immunogenic and/or vaccine composition, characterized in that it comprises a vector or recombinant virus, expressing a protein or a polypeptide of the S protein family, as defined above.

[0242] In addition to the preceding features, the invention further comprises other features, which will emerge from the description which follows, which refers to examples of use of the polynucleotide representing the genome of the SARS-CoV strain derived from the sample recorded under the number 031589, and derived cDNA fragments which are the subject of the present invention, and to Table I presenting the sequence listing:

TABLE I

Identification number	Sequence	Sequence listing	
		Position of the cDNA with reference to Genbank AY274119.3	Deposit number at the of the CNCM corresponding plasmid
SEQ ID NO: 1	genome of the strain derived from the sample 031589	—	—
SEQ ID NO: 2	ORF-S*	21406-25348	—
SEQ ID NO: 3	S protein	—	—
SEQ ID NO: 4	ORF-S**	21406-25348	I-3059
SEQ ID NO: 5	Sa fragment	21406-23454	I-3020
SEQ ID NO: 6	Sb fragment	23322-25348	I-3019
SEQ ID NO: 7	ORF-3 + ORF-4*	25110-26244	—
SEQ ID NO: 8	ORF-3 + ORF-4**	25110-26244	I-3126
SEQ ID NO: 9	ORF3	—	—
SEQ ID NO: 10	ORF-3 protein	—	—
SEQ ID NO: 11	ORF4	—	—
SEQ ID NO: 12	ORF-4 protein	—	—
SEQ ID NO: 13	ORF-E*	26082-26413	—
SEQ ID NO: 14	E protein	—	—
SEQ ID NO: 15	ORF-E**	26082-26413	I-3046
SEQ ID NO: 16	ORF-M*	26330-27098	—
SEQ ID NO: 17	M protein	—	—
SEQ ID NO: 18	ORF-M**	26330-27098	I-3047
SEQ ID NO: 19	ORF7 to 11*	26977-28218	—
SEQ ID NO: 20	ORF7 to 11**	26977-28218	I-3125
SEQ ID NO: 21	ORF7	—	—
SEQ ID NO: 22	ORF7 protein	—	—
SEQ ID NO: 23	ORF8	—	—
SEQ ID NO: 24	ORF8 protein	—	—
SEQ ID NO: 25	ORF9	—	—
SEQ ID NO: 26	ORF9 protein	—	—
SEQ ID NO: 27	ORF10	—	—
SEQ ID NO: 28	ORF10 protein	—	—
SEQ ID NO: 29	ORF11	—	—
SEQ ID NO: 30	ORF11 protein	—	—
SEQ ID NO: 31	OrF1ab	265-21485	—
SEQ ID NO: 32	ORF13	28130-28426	—
SEQ ID NO: 33	ORF13 protein	—	—
SEQ ID NO: 34	ORF14	—	—
SEQ ID NO: 35	ORF14 protein	28583-28795	—

TABLE I-continued

Identification number	Sequence	Sequence listing	
		Position of the cDNA with reference to Genbank AY274119.3	Deposit number at the of the CNCM corresponding plasmid
SEQ ID NO: 36	ORF-N*	28054-29430	—
SEQ ID NO: 37	N protein	—	—
SEQ ID NO: 38	ORF-N**	28054-29430	I-3048
SEQ ID NO: 39	noncoding 5**	1-204	I-3124
SEQ ID NO: 40	noncoding 3**	28933-29727	I-3123
SEQ ID NO: 41	ORF1ab	30-500	—
	Fragment L0		
SEQ ID NO: 42	Fragment L1	211-2260	—
SEQ ID NO: 43	Fragment L2	2136-4187	—
SEQ ID NO: 44	Fragment L3	3892-5344	—
SEQ ID NO: 45	Fragment L4b	4932-6043	—
SEQ ID NO: 46	Fragment L4	5305-7318	—
SEQ ID NO: 47	Fragment L5	7275-9176	—
SEQ ID NO: 48	Fragment L6	9032-11086	—
SEQ ID NO: 49	Fragment L7	10298-12982	—
SEQ ID NO: 50	Fragment L8	12815-14854	—
SEQ ID NO: 51	Fragment L9	14745-16646	—
SEQ ID NO: 52	Fragment L10	16514-18590	—
SEQ ID NO: 53	Fragment L11	18500-20602	—
SEQ ID NO: 54	Fragment L12	20319-22224	—
SEQ ID NO: 55	Sense N primer	—	—
SEQ ID NO: 56	Antisense N primer	—	—
	Sense S _C primer	—	—
SEQ ID NO: 57	Sence S _L primer	—	—
SEQ ID NO: 58	Antisense S _C and S _L primer	—	—
SEQ ID NO: 59	Sense primer series 1	28507-28522	—
SEQ ID NO: 60	Antisense primer series 1	28774-28759	—
SEQ ID NO: 61	Sense primer series 2	28375-28390	—
SEQ ID NO: 62	Antisense primer series 2	28702-28687	—
SEQ ID NO: 63	Probe 1/series 1	28561-28586	—
SEQ ID NO: 64	Probe 2/series 1	28588-28608	—
SEQ ID NO: 65	Probe 1/series 2	28541-28563	—
SEQ ID NO: 66	Probe 2/series 2	28565-28589	—
SEQ ID NO: 67	Anchor primer 14T	—	—
SEQ ID NO: 68	Peptide M2-14	—	—
SEQ ID NO: 69	Peptide E1-12	—	—
SEQ ID NO: 70	Peptide E53-76	—	—
SEQ ID NO: 71	Noncoding 5*	1-204	—
SEQ ID NO: 72	Noncoding 3*	28933-29727	—
SEQ ID NO: 73	ORF1a protein	—	—
SEQ ID NO: 74	ORF1b protein	—	—
SEQ ID NO: 75	Primers	—	—
SEQ ID NO: 76-139	Pseudogene of S	—	—
SEQ ID NO: 140	Primers	—	—
SEQ ID NO: 141-148	Aa1-13 of S	—	—
SEQ ID NO: 149	Polypeptide	—	—
SEQ ID NO: 150	Primers	—	—
SEQ ID NO: 151-158			

* PCR amplification product (amplicon)

** Insert cloned into the plasmid deposited at the CNCM and to the appended drawings in which:

[0243] FIG. 1 illustrates Western-blot analysis of the expression in vitro of the recombinant proteins N, S_C and S_L from the expression vectors pIVEX. Lane 1: pIV2.3N. Lane 2: pIV2.3S_C. Lane 3: pIV2.3S_L. Lane 4: pIV2.4N. Lane 5: pIV2.4S₁ or pIV2.4S_C. Lane 6: pIV2.4S_L. The expression of the GFP protein expressed from the same vector is used as a control.

[0244] FIG. 2 illustrates the analysis, by polyacrylamide gel electrophoresis under denaturing conditions (SDS-PAGE) and staining with Coomassie blue, of the expression in vivo of the N protein from the expression vectors pIVEX. The *E. coli* BL21(DE3)pDIA17 strain transformed with the recombinant vectors pIVEX is cultured at 30° C. in LB medium, in the presence or in the absence of inducer (IPTG 1 mM). Lane 1: pIV2.3N. Lane 2: pIV2.4N.

[0245] FIG. 3 illustrates the analysis, by polyacrylamide gel electrophoresis under denaturing conditions (SDS-

PAGE) and staining with Coomassie blue, of the expression in vivo of the S_L and S_C polypeptides from the expression vectors pIVEX. The *E. coli* BL21(DE3)pDIA17 strain transformed with the recombinant vectors pIVEX is cultured at 30° C. in LB medium, in the presence or in the absence of inducer (IPTG 1 mM). Lane 1: pIV2.3 S_C . Lane 2: pIV2.3 S_L . Lane 3: pIV2.4 S_L . Lane 4: pIV2.4 S_L .

[0246] FIG. 4 illustrates the antigenic activity of the recombinant N, S_L and S_C proteins produced in the *E. coli* BL21(DE3)pDIA17 strain transformed with the recombinant vectors pIVEX. A: electrophoresis (SDS-PAGE) of the bacterial lysates. B and C: Western-blot with the sera, obtained from the same patient infected with SARS-CoV, collected 6 days (B: serum M12) and 29 days (C: serum M13) respectively after the onset of the SARS symptoms. Lane 1: pIV2.3N. Lane 2: pIV2.4N. Lane 3: pIV2.3 S_C . Lane 4: pIV2.4 S_L . Lane 5: pIV2.3 S_L . Lane 6: pIV2.4 S_L .

[0247] FIG. 5 illustrates the purification on an Ni-NTA agarose column of the recombinant N protein produced in the *E. coli* BL21(DE3)pDIA17 strain from the vector pIV2.3N. Lane 1: total bacterial extract. Lane 2: soluble extract. Lane 3: insoluble extract. Lane 4: extract deposited on the Ni-NTA column. Lane 5: unbound proteins. Lane 6: fractions of peak 1. Lane 7: fractions of peak 2.

[0248] FIG. 6 illustrates the purification of the recombinant S_C protein from the inclusion bodies produced in the *E. coli* BL21(DE3)pDIA17 strain transformed with pIV2.4 S_L . A: Treatment with Triton X-100 (2%): Lane 1: total bacterial extract. Lane 2: soluble extract. Lane 3: insoluble extract. Lane 4: supernatant after treatment with Triton X-100 (2%). Lanes 5 and 6: pellet after treatment with Triton X-100 (2%). B: Treatment with 4 M, 5 M, 6 M and 7 M urea of the soluble and insoluble extracts.

[0249] FIG. 7 represents the immunoblot produced with the aid of a lysate of cells infected with SARS-CoV and a serum from a patient suffering from atypical pneumopathy.

[0250] FIG. 8 represents immunoblots produced with the aid of a lysate of cells infected with SARS-CoV and rabbit immunosera specific for the nucleoprotein N (A) and for the spicule protein S (B). I.S.: immune serum. p.i.: preimmune serum. The anti-N immune serum was used at 1/50 000 and the anti-S immune serum at 1/10 000.

[0251] FIG. 9 illustrates the ELISA reactivity of the rabbit monospecific polyclonal sera directed against the N protein or the short fragment of the S protein (S_C), toward the corresponding recombinant proteins used for immunization. A: rabbits P13097, P13081 and P13031 immunized with the purified recombinant N protein. B: rabbits P11135, P13042 and P14001 immunized with a preparation of inclusion bodies corresponding to the short fragment of the S protein (S_C). I.S.: immune serum. p.i.: preimmune serum.

[0252] FIG. 10 illustrates the ELISA reactivity of the purified recombinant N protein, toward sera from patients suffering from atypical pneumonia caused by SARS-CoV. FIG. 10a: ELISA plates prepared with the N protein at the concentration of 4 µg/ml and 2 µg/ml. FIG. 10B: ELISA plate prepared with the N protein at the concentration of 1 µg/ml. The sera designated A, B, D, E, F, G, H correspond to those of Table IV.

[0253] FIG. 11 illustrates the amplification by RT-PCR of decreasing quantities of synthetic RNA of the SARS-CoV N

gene (10^7 to 1 copy), with the aid of pairs of primers No. 1 (N/+28507, N/-28774) (A) and No. 2 (N/+28375, N/-28702) (B). T: amplification performed in the absence of RNA. MW: DNA marker.

[0254] FIG. 12 illustrates the amplification by RT-PCR in real time of synthetic RNA for the SARS-CoV N gene: decreasing quantities of synthetic RNA as replica (repli.; lanes 16 to 29) and of viral RNA diluted $1/20 \times 10^4$ (lane 32) were amplified by RT-PCR in real time with the aid of the kit "Light Cycler RNA Amplification Kit Hybridization Probes" and pairs of primers and probes of the No. 2 series, under the conditions described in Example 8.

[0255] FIG. 13 (FIGS. 13.1 to 13.7) represents the restriction map of the sequence SEQ ID NO: 1 corresponding to the DNA equivalent of the genome of the SARS-CoV strain derived from the sample recorded under the number 031589.

[0256] FIG. 14 shows the result of the SARS serology test by indirect N ELISA (1st series of sera tested).

[0257] FIG. 15 shows the result of the SARS serology test by indirect N ELISA (2nd series of sera-tested).

[0258] FIG. 16 presents the result of the SARS serology test by double epitope N ELISA (1st series of sera tested).

[0259] FIG. 17 shows the result of the SARS serology test by double epitope N ELISA (2nd series of sera tested).

[0260] FIG. 18 illustrates the test of reactivity of the anti-N monoclonal antibodies by ELISA on the native nucleoprotein N of SARS-CoV. The antibodies were tested in the form of hybridoma culture supernatants by indirect ELISA using an irradiated lysate of VeroE6 cells infected with SARS-CoV as antigen (SARS lysate curves). A negative control for reactivity is performed for each antibody on a lysate of uninfected VeroE6 cells (negative lysate curves). Several monoclonal antibodies of known specificity were used as negative control antibodies: para1-3 directed against the antigens of the parainfluenza viruses type 1-3 (Bio-Rad) and influenza B directed against the antigens of the influenza virus type B (Bio-Rad).

[0261] FIG. 19 illustrates the test of reactivity of the anti-N of SARS-CoV monoclonal antibodies by ELISA on the native antigens of the human coronavirus 229E (HCoV-229E). The antibodies were tested in the form of hybridoma culture supernatants by an indirect ELISA test using a lysate of MRC-5 cells infected with the human coronavirus 229E as antigen (229E lysate curves). A negative control for immunoreactivity was performed for each antibody on a lysate of noninfected MRC-5 cells (negative lysate curves). The monoclonal antibody 5-11H.6 directed against the S protein of the human coronavirus 229E (Sizun et al. 1998, J. Virol. Met. 72: 145-152) is used as positive control antibody. The antibodies para1-3 directed against the antigens of the parainfluenza virus type 1-3 (Bio-Rad) and influenza B directed against the antigens of the influenza virus type B (Bio-Rad) were added to the panel of monoclonal antibodies tested.

[0262] FIG. 20 shows a test of reactivity of the anti-N of SARS-CoV monoclonal antibodies by Western blotting on the denatured native nucleoprotein N of SARS-CoV. A lysate of VeroE6 cells infected with SARS-CoV was prepared in the loading buffer according to Laemmli and caused to migrate in a 12% SDS polyacrylamide gel and then the

proteins were transferred onto PVDF membrane. The anti-N monoclonal antibodies tested were used for the immunoassay at the concentration of 0.05 µg/ml. The visualization is carried out with anti-mouse IgG(H+L) antibodies coupled to peroxidase (NA934V, Amersham) and the ECL+ system. Two monoclonal antibodies were used as negative controls for reactivity: influenza B directed against the antigens of the influenza virus type B (Bio-Rad) and para1-3 directed against the antigens of the parainfluenza virus type 1-3 (Bio-Rad).

[0263] FIG. 21 presents the plasmids for expression in mammalian cells of the SARS-CoV S protein. The cDNA for the SARS-CoV S was inserted between the BamHI and XhoI sites of the expression plasmid pcDNA3.1(+) (Clontech) in order to obtain the plasmid pcDNA-S and between the NheI and XhoI sites of the expression plasmid pCI (Promega) in order to obtain the plasmid PCI-S. The WPRE and CTE sequences were inserted between each of the two plasmids pcDNA-S and pCI-S between the XhoI and XbaI sites in order to obtain the plasmids pcDNA-S-CTE, pcDNA-S-WPRE, pCI-S-CTE and pCI-S-WPRE, respectively.

[0264] SP: signal peptide predicted (aa 1-13) with the software signalP v2.0 (Nielsen et al., 1997, Protein Engineering, 10: 1-6)

[0265] TM: transmembrane region predicted (aa 1196-1218) with the software TMHMM v2.0 (Sonnhammer et al., 1998, Proc. of Sixth Int. Conf. on Intelligent Systems for Molecular Biology, pp. 175-182, AAAI Press). It should be noted that the amino acids W-1194 and P1195 are possibly part of the transmembrane region with the respective probabilities of 0.13 and 0.42

[0266] P-CMV: cytomegalovirus immediate/early promoter. BGH pA: polyadenylation signal of the bovine growth hormone gene

[0267] SV40 late pA: SV40 virus late polyadenylation signal

[0268] SD/SA: splice donor and acceptor sites

[0269] WPRE: sequences of the "Woodchuck Hepatitis Virus posttranscriptional regulatory element" of the woodchuck hepatitis virus

[0270] CTE: sequences of the "constitutive transport element" of the Mason-Pfizer simian retrovirus

[0271] FIG. 22 illustrates the expression of the S protein after transfection of VeroE6 cells. Cellular extracts were prepared 48 hours after transfection of VeroE6 cells with the plasmids pcDNA, pcDNA-S, pCI and pCI-S. Cellular extracts were also prepared 18 hours after infection with the recombinant vaccinia virus VV-TF7.3 and transfection with the plasmids pcDNA or pcDNA-S. As a control, extracts of VeroE6 cells were prepared 8 hours after infection with SARS-CoV at a multiplicity of infection of 3. They were separated on an 8% SDS acrylamide gel and analyzed by Western blotting with the aid of an anti-S rabbit polyclonal antibody and an anti-rabbit IgG(H+L) polyclonal antibody coupled to peroxidase (NA934V, Amersham). A molecular mass ladder (kDa) is presented in the figure.

[0272] SARS-CoV: extract of VeroE6 cells infected with SARS-CoV

[0273] Mock: control extract of noninfected cells

[0274] FIG. 23 illustrates the effect of the CTE and WPRE sequences on the expression of the S protein after transfection of VeroE6 and 293T cells. Cellular extracts were prepared 48 hours after transfection of VeroE6 cells (A) or 293T cells (B) with the plasmids pcDNA, pcDNA-S, pcDNA-S-CTE, pcDNA-S-WPRE, pCI-S, pCI-S-CTE and pCI-S-WPRE separated on 8% SDS polyacrylamide gel and analyzed by Western blotting with the aid of an anti-S rabbit polyclonal antibody and an anti-rabbit IgG(H+L) polyclonal antibody coupled to peroxidase (NA934V, Amersham). A molecular mass ladder (kDa) is presented in the figure.

[0275] SARS-CoV: extract of VeroE6 cells prepared 8 hours after infection with SARS-CoV at a multiplicity of infection of 3.

[0276] Mock: control extract of noninfected VeroE6 cells

[0277] FIG. 24 presents defective lentiviral vectors with central DNA flap for the expression of SARS-CoV S. The cDNA for the SARS-CoV S protein was cloned in the form of a BamHI-XhoI fragment into the plasmid pTRIPΔU3-CMV containing a defective lentiviral vector TRIP with central DNA flap (Sirven et al., 2001, Mol. Ther., 3: 438-448) in order to obtain the plasmid pTRIP-S. The optimum expression cassettes consisting of the CMV virus immediate/early promoter, a splice signal, cDNA for S and either of the posttranscriptional signals CTE or WPRE were substituted for the cassette EF1α-EGFP of the defective lentiviral expression vector with central DNA flap TRIPΔU3-EF1α (Sirven et al., 2001, Mol. Ther., 3: 438-448) in order to obtain the plasmids pTRIP-SD/SA-S-CTE and pTRIP-SD/SA-S-WPRE.

[0278] SP: signal peptide

[0279] TM: transmembrane region

[0280] P-CMV: cytomegalovirus immediate/early promoter

[0281] P-EF1α: EF1α gene promoter

[0282] SD/SA: splice donor and acceptor sites

[0283] WPRE: sequences of the "Woodchuck Hepatitis Virus posttranscriptional regulatory element" of the woodchuck hepatitis virus

[0284] CTE: sequences of the "constitutive transport element" of the Mason-Pfizer simian retrovirus

[0285] LTR: long terminal repeat

[0286] ΔU3: LTR deleted for the "promoter/enhancer" sequences

[0287] cPPT: "polypurine tract cis-active sequence"

[0288] CTS: "central termination sequence"

[0289] FIG. 25 shows the Western-blot analysis of the expression of the SARS-CoV S by cell lines transduced with the lentiviral vectors TRIP-SD/SA-S-WPRE and TRIP-SD/SA-S-CTE. Cellular extracts were prepared from established lines FrhK4-S-CTE and FrhK4-S-WPRE after transduction with the lentiviral vectors TRIP-SD/SA-S-CTE and TRIP-SD/SA-S-WPRE respectively. They were separated on an 8% SDS acrylamide gel and analyzed by Western blotting

with the aid of an anti-S rabbit polyclonal antibody and an anti-rabbit IgG(H+L) conjugate coupled to peroxidase. A molecular mass ladder (kDa) is presented in the figure.

[0290] T-: control extract of FrhK-4 cells

[0291] T+: extract of FrhK-4 cells prepared 24 hours after infection with SARS-CoV at a multiplicity of infection of 3.

[0292] FIG. 26 relates to the analysis of the expression of Ssol polypeptide by cell lines transduced with the lentiviral vectors TRIP-SD/SA-Ssol-WPRE and TRIP-SD/SA-Ssol-CTE. The secretion of the Ssol polypeptide was determined in the supernatant of a series of cell clones isolated after transduction of FrhK-4 cells with the lentiviral vectors TRIP-SD/SA-Ssol-WPRE and TRIP-SD/SA-Ssol-CTE. 5 μ l of supernatant, diluted 1/2 in loading buffer according to Laemmli, were analyzed by Western blotting, visualized with an anti-FLAG monoclonal antibody (M2, Sigma) and an anti-mouse IgG(H+L) conjugate coupled to peroxidase. T-: supernatant of the parental FRhK-4 line. T+: supernatant of BHK cells infected with a recombinant vaccinia virus expressing the Ssol polypeptide. The solid arrow indicates the Ssol polypeptide, while the empty arrow indicates a cross reaction with a protein of cellular origin.

[0293] FIG. 27 shows the results relating to the analysis of the purified Ssol polypeptide

[0294] A. 8, 2, 0.5 and 0.125 μ g of recombinant Ssol polypeptide purified by anti-FLAG affinity chromatography and gel filtration (G75) were separated on 8% SDS polyacrylamide gel. The Ssol polypeptide and variable quantities of molecular mass markers (MM) were visualized by staining with silver nitrate (Gelcode SilverSNAP stain kit II, Pierce).

[0295] B. Standard markers for analysis by SELDI-TOF mass spectrometry

[0296] IgG: bovine IgG of MM 147300

[0297] ConA: conalbumin of MM 77490

[0298] HRP: horseradish peroxidase analyzed as a control and of MM 43240

[0299] C. Analysis by mass spectrometry (SELDI-TOF) of the recombinant Ssol polypeptide.

[0300] The peaks A and B correspond to the single and double charged Ssol polypeptide.

[0301] D. Sequencing of the N-terminal end of the recombinant Ssol polypeptide. 5 Edman degradation cycles in liquid phase were carried out on an ABI494 sequencer (Applied Biosystems).

[0302] FIG. 28 illustrates the influence of a splicing signal and of the CTE and WPRE sequences on the efficacy of the gene immunization with the aid of plasmid DNA encoding the SARS-CoV S

[0303] A. Groups of 7 BALB/c mice were immunized twice at 4 weeks' interval with the aid of 50 μ g of plasmid DNA of pCI, pcDNA-S, pCI-S, pcDNA-N and pCI-HA.

[0304] B. Groups of 6 BALB/c mice were immunized twice at 4 weeks' interval with the aid of 2 μ g, 10 μ g or 50 μ g of plasmid DNA of pCI, pCI-S, pCI-S-CTE and pCI-S-WPRE.

[0305] The immune sera collected 3 weeks after the second immunization were analyzed by indirect ELISA using a lysate of VeroE6 cells infected with SARS-CoV as antigen. The anti-SARS-CoV antibody titers are calculated as the reciprocal of the dilution producing a specific OD of 0.5 after visualization with an anti-mouse IgG polyclonal antibody coupled to peroxidase (NA931V, Amersham) and TMB (KPL).

[0306] FIG. 29 shows the seroneutralization of the infectivity of SARS-CoV with the antibodies induced in mice after gene immunization with the aid of plasmid DNA encoding SARS-CoV S. Pools of immune sera collected 3 weeks after the second immunization were prepared for each of the groups of experiments described in FIG. 28 and evaluated for their capacity to seroneutralize the infectivity of 100 TCID₅₀ of SARS-CoV on FRhK-4 cells. 4 points are produced for each of the 2-fold dilutions tested from 1/20. The seroneutralizing titer is calculated according to the Reed and Munsch method as the reciprocal of the dilution neutralizing the infectivity of 2 wells out of 4.

[0307] A. Groups by BALB/c mice immunized twice at 4 weeks' interval with the aid of 50 μ g of plasmid DNA of pCI, pcDNA-S, pCI-S, pcDNA-N and pCI-HA. \square : pre-immune serum. \blacksquare : immune serum.

[0308] B. Groups of BALB/c mice immunized twice at 4 weeks' interval with the aid of 2 μ g, 10 μ g or 50 μ g of plasmid DNA of pCI, pCI-S, pCI-S-CTE and pCI-S-WPRE.

[0309] FIG. 30 illustrates the immunoreactivity of the recombinant Ssol polypeptide toward sera from patients suffering from SARS. The reactivity of sera from patients was analyzed by indirect ELISA test against solid phases prepared with the aid of the purified recombinant Ssol polypeptide. The antibodies from patients reacting with the solid phase at a dilution of 1/400 are visualized with a human anti-IgG(H+L) polyclonal antibody coupled to peroxidase (Amersham NA933V) and TMB plus, H202 (KPL). The sera of probable SARS cases are identified by a National Reference Center for Influenza Viruses serial number and by the initials of the patient and the number of days elapsed since the onset of symptoms, where appropriate. The TV sera are control sera from subjects which were collected in France before the SARS epidemic which occurred in 2003.

[0310] FIG. 31 shows the induction of antibodies directed against SARS-CoV after immunization with the recombinant Ssol polypeptide. Two groups of 6 mice were immunized at 3 weeks' interval with 10 μ g of recombinant Ssol polypeptide (Ssol group) adjuvanted with aluminum hydroxide or, as a control, of adjuvant alone (mock group). Three successive immunizations were performed and the immune sera were collected 3 weeks after each of the three immunizations (IS1, IS2, IS3). The immune sera were analyzed per pool for each of the 2 groups by indirect ELISA using a lysate of VeroE6 cells infected with SARS CoV as antigen. The anti-SARS-CoV antibody titers are calculated as the reciprocal of the dilution producing a specific OD of 0.5 after visualization with an anti-mouse IgG polyclonal antibody coupled to peroxidase (Amersham) and TMB (KPL).

[0311] FIG. 32 presents the nucleotide alignment of the sequences of the synthetic gene 040530 with the sequence of the wild-type gene of the SARS-CoV isolate 031589. I-3059

corresponds to nucleotides 21406-25348 of the SARS-CoV isolate 031589 deposited at the C.N.C.M. under the number I-3059 (SEQ ID NO: 4, plasmid pSARS-S) S-040530 is the sequence of the synthetic gene 040530.

[0312] FIG. 33 illustrates the use of a synthetic gene for the expression of the SARS-CoV S. Cellular extracts prepared 48 hours after transfection of VeroE6 cells (A) or 293T cells (B) with the plasmids pCI, PCI-S, pCI-S-CTE, pCI-S-WPRE and pCI-Ssynth were separated on 8% SDS acrylamide gel and analyzed by Western blotting with the aid of an anti-S rabbit polyclonal antibody and an anti-rabbit IgG(H+L) polyclonal antibody coupled to peroxidase (NA934V, Amersham). The Western blot is visualized by luminescence (ECL+, Amersham) and acquisition on a digital imaging device (FluorS, BioRad). The levels of expression of the S protein were measured by quantifying the 2 predominant bands identified on the image.

[0313] FIG. 34 presents a diagram for the construction of recombinant vaccinia viruses VV-TG-S, VV-TG-Ssol, VV-TN-S and VV-TN-Ssol

[0314] A. The cDNAs for the S protein and the Ssol polypeptide of SARS-CoV were inserted between the BamHI and SmaI sites of the transfer plasmid pTG186 in order to obtain the plasmids pTG-S and pTG-Ssol.

[0315] B. The sequences of the synthetic promoter 480 were then substituted for those of the 7.5 promoter by exchange of the NdeI-PstI fragments of the plasmids pTG186poly, pTG-S and pTG-Ssol in order to obtain the transfer plasmids pTN480, pTN-S and pTN-Ssol.

[0316] C. Sequence of the synthetic promoter 480 as contained between the NdeI and PstI sites of the transfer plasmids of the pTN series. An AscI site was inserted in order to facilitate subsequent handling. The restriction sites and the promoter sequence are underlined.

[0317] D. The recombinant vaccinia viruses are obtained by double homologous recombination *in vivo* between the TK cassette of the transfer plasmids of the pTG and pTN series and the TK gene of the Copenhagen strain of the vaccinia virus.

[0318] SP: signal peptide predicted (aa 1-13) with the software signalP v2.0 (Nielsen et al., 1997, Protein Engineering, 10: 1-6)

[0319] TM: transmembrane region predicted (aa 1196-1218) with the software TMHMM v2.0 (Sonnhammer et al., 1998, Proc. of Sixth Int. Conf. on Intelligent Systems for Molecular Biology, pp. 175-182, AAAI Press). It should be noted that the amino acids W1194 and P1195 possibly form part of the transmembrane region with respective probabilities of 0.13 and 0.42.

[0320] TK-L, TK-R: left- and right-hand parts of the vaccinia virus thymidine kinase gene

[0321] MCS: multiple cloning site

[0322] PE: early promoter

[0323] PL: late promoter

[0324] PL synth: synthetic late promoter 480

[0325] FIG. 35 illustrates the expression of the S protein by recombinant vaccinia viruses, analyzed by Western blot-

ting. Cellular extracts were prepared 18 hours after infection of CV1 cells with the recombinant vaccinia viruses VV-TG, VV-TG-S and VV-TN-S at an M.O.I. of 2 (A). As a control, extracts of VeroE6 cells were prepared 8 hours after infection with SARS-CoV at a multiplicity of infection of 2. Cellular extracts were also prepared 18 hours after infection of CV1 cells with the recombinant vaccinia viruses VV-TG-S, VV-TG-Ssol, VV-TN, VV-TN-S and VV-TN-Ssol (B). They were separated on 8% SDS acrylamide gels and analyzed by Western blotting with the aid of an anti-S rabbit polyclonal antibody and an anti-rabbit IgG(H+L) polyclonal antibody coupled to peroxidase (NA934V, Amersham). "1 μ l" and "10 μ l" indicates the quantities of cellular extracts deposited on the gel. A molecular mass ladder (kDa) is presented in the figure.

[0326] SARS-CoV: extract of VeroE6 cells infected with SARS-CoV

[0327] Mock: control extract of noninfected cells

[0328] FIG. 36 shows the result of a Western-blot analysis of the secretion of the Ssol polypeptide by the recombinant vaccinia viruses.

[0329] A. Supernatants of CV1 cells infected with the recombinant vaccinia virus VV-TN, various clones of the VV-TN-Ssol virus and with the viruses VV-TG-Ssol or VV-TN-Sflag were harvested 18 hours after infection of CV1 cells at an M.O.I. of 2.

[0330] B. Supernatants of 293T, FRhK-4, BHK-21 and CV1 cells infected in duplicate (1.2) with the recombinant vaccinia virus VV-TN-Ssol at an M.O.I. of 2 were harvested 18 hours after infection. The supernatant of CV1 cells infected with the virus VV-TN was also harvested as a control (M).

[0331] All the supernatants were separated on 8% SDS acrylamide gel according to Laemmli and analyzed by Western blotting with the aid of an anti-FLAG mouse monoclonal antibody and an anti-mouse IgG(H+L) polyclonal antibody coupled to peroxidase (NA931V, Amersham) (A) or with the aid of an anti-S rabbit polyclonal antibody and an anti-rabbit IgG(H+L) polyclonal antibody coupled to peroxidase (NA934V, Amersham) (B).

[0332] A molecular mass ladder (kDa) is presented in the figure.

[0333] FIG. 37 shows the analysis of the Ssol polypeptide, purified on SDS polyacrylamide gel

[0334] 10, 5 and 2 μ l of recombinant Ssol polypeptide purified by anti-FLAG affinity chromatography were separated on 4 to 15% gradient SDS polyacrylamide gel. The Ssol polypeptide and variable quantities of molecular mass markers (MM) were visualized by staining with silver nitrate (Gelcode SilverSNAP stain kit II, Pierce).

[0335] FIG. 38 illustrates the immunoreactivity of the recombinant Ssol polypeptide produced by the recombinant vaccinia virus VV-TN-Ssol toward sera of patients suffering from SARS. The reactivity of sera from patients was analyzed by indirect ELISA test against solid phases prepared with the aid of the purified recombinant Ssol polypeptide. The antibodies from patients reacting with the solid phase at a dilution of 1/100 and 1/400 are visualized with a human anti-IgG(H+L) polyclonal antibody coupled to peroxidase

(Amersham NA933V) and TMB plus H202 (KPL). The sera of probable SARS cases are identified by a National Reference Center for Influenza Virus serial number and by the initials of the patient and the number of days elapsed since the onset of symptoms, where appropriate. The TV sera are control sera from subjects which were collected in France before the SARS epidemic which occurred in 2003.

[0336] FIG. 39 shows the anti-SARS-CoV antibody response in mice after immunization with the recombinant vaccinia viruses. Groups of 7 BALB/c mice were immunized by the i.v. route twice at 4 weeks' interval with 106 pfu of recombinant vaccinia viruses VV-TG, VV-TG-HA, VV-TG-S, W-TG-Ssol, VV-TN, VV-TN-S, VV-TN-Ssol.

[0337] A. Pools of immune sera collected 3 weeks after each of the two immunizations were prepared for each of the groups and were analyzed by indirect ELISA using a lysate of VeroE6 cells infected with SARS-CoV as antigen. The anti-SARS-CoV antibody titers are calculated as the reciprocal of the dilution producing a specific OD of 0.5 after visualization with an anti-mouse IgG polyclonal antibody coupled to peroxidase (NA931V, Amersham) and TMB (KPL).

[0338] B. The pools of immune sera were evaluated for their capacity to seroneutralize the infectivity of 100 TCID50 of SARS-CoV on FRhK-4 cells. 4 points are produced for each of the 2-fold dilutions tested from 1/20. The seroneutralizing titer is calculated according to the Reed and Munsch method as the reciprocal of the dilution neutralizing the infectivity of 2 wells out of 4.

[0339] FIG. 40 describes the construction of the recombinant viruses MVSchw2-SARS-S and MVSchw2-SARS-Ssol.

[0340] A. The measles vector is a complete genome of the Schwarz vaccine strain of the measles virus (MV) into which an additional transcription unit has been introduced (Combredet, 2003, Journal of Virology, 77: 11546-11554). The expression of the additional open reading frames (ORF) is controlled by cis-acting elements necessary for the transcription, for the formation of the cap and for the polyadenylation of the transgene which were copied from the elements present at the N/P junction. 2 different vectors allow the insertion between the P (phosphoprotein) and M (matrix) genes on the one hand and the H (hemagglutinin) and L (polymerase) genes on the other hand.

[0341] B. The recombinant genomes MVSchw2-SARS-S and MVSchw2-SARS-Ssol of the measles virus were constructed by inserting the ORFs of the S protein and of the Ssol polypeptide into an additional transcription unit located between the P and M genes of the vector.

[0342] The various genes of the measles virus (MV) are indicated: N (nucleoprotein), PVC (V/C phosphoprotein and protein), M (matrix), F (fusion), H (hemagglutinin), L (polymerase). T7=RNA polymerase promoter, hh=hammerhead ribozyme, T7t=T7 phage RNA polymerase terminator sequence, δ=ribozyme of the hepatitis δ virus, (2), (3)=additional transcription units (ATU).

[0343] Size of the MV genome: 15 894 nt.

[0344] SP: signal peptide

[0345] TM: transmembrane region

[0346] FLAG: FLAG tag

[0347] FIG. 41 illustrates the expression of the S protein by the recombinant measles viruses, analyzed by Western blotting.

[0348] Cytoplasmic extracts were prepared after infection of Vero cells by different passages of the viruses MVSchw2-SARS-S and MVSchw2-SARS-Ssol and the wild-type virus MWSchw as control. Cellular extracts in loading buffer according to Laemmli were also prepared 8 hours after infection of VeroE6 cells with SARS-CoV at a multiplicity of infection of 3. They were separated on 8% SDS acrylamide gel and analyzed by Western blotting with the aid of an anti-S rabbit polyclonal antibody and an anti-rabbit IgG(H+L) polyclonal antibody coupled to peroxidase (NA934V, Amersham).

[0349] A molecular mass ladder (kDa) is presented in the figure.

[0350] Pn: nth passage of the virus after coculture of 293-3-46 and Vero cells

[0351] SARS-CoV: extract of VeroE6 cells infected with SARS-CoV

[0352] Mock: control extract of noninfected VeroE6 cells

[0353] FIG. 42 shows the expression of the S protein by the recombinant measles viruses, analyzed by immunofluorescence

[0354] Vero cells in monolayers on glass slides were infected with the wild-type virus MWSchw (A) or the viruses MVSchw2-SARS-S (B) and MVSchw2-SARS-Ssol (C). When the syncytia have reached 30 to 40% confluence (A., B.) or 90-100% (C), the cells were fixed, permeabilized and labeled with anti-SARS-CoV rabbit polyclonal antibodies and an anti-rabbit IgG(H+L) conjugate coupled to FITC (Jackson).

[0355] FIG. 43 illustrates the Western-blot analysis of the immunoreactivity of rabbit sera directed against the peptides E1-12, E53-76 and M2-14. The rabbit 20047 was immunized with the peptide E1-12 coupled to KLH. The rabbits 22234 and 22240 were immunized with the peptide E53-76 coupled to KLH. The rabbits 20013 and 20080 were immunized with the peptide M2-14 coupled to KLH. The immune sera were analyzed by Western blotting with the aid of extracts of cells infected with SARS-CoV (B) or with the aid of extracts of cells infected with a recombinant vaccinia virus expressing the protein E (A) or M (C) of the SARS-CoV 031589 isolate. The immunoblots were visualized with the aid of an anti-rabbit IgG(H+L) conjugate coupled to peroxidase (NA934V, Amersham).

[0356] The position of the E and M proteins is indicated by an arrow.

[0357] A molecular mass ladder (kDa) is presented in the figure.

[0358] It should be understood, however, that these examples are given solely by way of illustration of the subject of the invention, and do not constitute in any manner a limitation thereto.

EXAMPLE 1

Cloning and Sequencing of the Genome of the SARS-CoV Strain Derived from the Sample Recorded Under the Number 031589

[0359] The RNA of the SARS-CoV strain was extracted from the sample of bronchoalveolar washing recorded under the number 031589, performed on a patient at the Hanoi (Vietnam) French hospital suffering from SARS.

[0360] The isolated RNA was used as template to amplify the cDNAs corresponding to the various open reading frames of the genome (ORF1a, ORF1b, ORF-S, ORF-E, ORF-M, ORF-N (including ORF-13 and ORF-14), ORF3, ORF4, ORF7 to ORF11), and at the noncoding 5' and 3' ends. The sequences of the primers and of the probes used for the amplification/detection were defined based on the available SARS-CoV nucleotide sequence.

[0361] In the text which follows, the primers and the probes are identified by: the letter S, followed by a letter which indicates the corresponding region of the genome (L for the 5' end including ORF1a and ORF1b; S, M and N for ORF-S, ORF-M, ORF-N, SE and MN for the corresponding intergene regions), and then optionally by Fn, Rn, with n between 1 and 6 corresponding to the primers used for the nested PCR (F1+R1 pair for the first amplification, F2+R2 pair for the second amplification, and the like), and then by +/- or -/- corresponding to a sense or antisense primer and finally by the positions of the primers with reference to the Genbank sequence AY27411.3; for the sense and antisense S and N primers and the other sense primers only, when a single position is indicated, it corresponds to that of the 5' end of a probe or of a primer of about 20 bases; for the antisense primers other than the S and N primers, when a single position is indicated, it corresponds to that of the 3' end of a probe or of a primer of about 20 bases.

[0362] The amplification products thus generated were sequenced with the aid of specific primers in order to determine the complete sequence of the genome of the SARS-CoV strain derived from the sample recorded under the number 031589. These amplification products, with the exception of those corresponding to ORF1a and ORF1b, were then cloned into expression vectors in order to produce the corresponding viral proteins and the antibodies directed against these proteins, in particular by DNA-based immunization.

1. Extraction of the RNAs

[0363] The RNAs were extracted with the aid of the QIamp viral RNA extraction mini kit (QIAGEN) according to the manufacturer's recommendations. More specifically: 140 μ l of the sample and 560 μ l of AVL buffer were vigorously mixed for 15 seconds, incubated for 10 minutes at room temperature and then briefly centrifuged at maximum speed. 560 μ l of 100% ethanol were added to the supernatant and the mixture thus obtained was very vigorously stirred for 15 sec. 630 μ l of the mixture were then deposited on the column.

[0364] The column was placed on a 2 ml tube, centrifuged for 1 min at 8000 rpm, and then the remainder of the preceding mixture was deposited on the same column, centrifuged again, for 1 min at 8000 rpm, and the column was transferred over a clean 2 ml tube. Next, 500 μ l of AW1

buffer were added to the column, and then the column was centrifuged for 1 min at 8000 rpm and the eluate was discarded. 500 μ l of AW2 buffer were added to the column which was then centrifuged for 3 min at 14 000 rpm and transferred onto a 1.5 ml tube. Finally, 60 μ l of AVE buffer were added to the column which was incubated for 1 to 2 min at room temperature and then centrifuged for 1 min at 8000 rpm. The eluate corresponding to the purified RNA was recovered and frozen at -20° C.

2. Amplification, Sequencing and Cloning of the cDNAs

2.1) cDNA Encoding the S Protein

[0365] The RNAs extracted from the sample were subjected to reverse transcription with the aid of random sequence hexameric oligonucleotides (pdN6), so as to produce cDNA fragments.

[0366] The sequence encoding the SARS-CoV S glycoprotein was amplified in the form of two overlapping DNA fragments: 5' fragment (SARS-Sa, SEQ ID NO: 5) and 3' fragment (SARS-Sb, SEQ ID NO: 6), by carrying out two successive amplifications with the aid of nested primers. The amplicons thus obtained were sequenced, cloned into the PCR plasmid vector 2.1-TOPO™ (INVITROGEN), and then the sequence of the cloned cDNAs was determined.

a) Cloning and Sequencing of the Sa and Sb Fragments

a.1) Synthesis of the cDNA

[0367] The reaction mixture containing: RNA (5 μ l), H₂O for injection (3.5 μ l), 5 \times reverse transcriptase buffer (4 μ l), 5 mM dNTP (2 μ l), pdN6 100 μ g/ml (4 μ l), RNasin 40 IU/ μ l (0.5 μ l) and reverse transcriptase AMV-RT, 10 IU/ μ l, PROMEGA (1 μ l) was incubated in a thermocycler under the following conditions: 45 min at 42° C., 15 min at 55° C., 5 min at 95° C., and then the cDNA obtained was kept at $+4^{\circ}$ C.

a.2) First PCR Amplification

[0368] The 5' and 3' ends of the S gene were respectively amplified with the pairs of primers S/F1+/21350-21372 and S/R1-/23518-23498, S/F3+/23258-23277 and S/R3-/25382-25363. The 50 μ l reaction mixture containing: cDNA (2 μ l), 50 μ M primers (0.5 μ l), 10 \times buffer (5 μ l), 5 mM dNTP (2 μ l), Taq Expand High Fidelity, Roche (0.75 μ l) and H₂O (39, 75 μ l) was amplified in a thermocycler, under the following conditions: an initial step of denaturation at 94° C. for 2 min was followed by 40 cycles comprising: a step of denaturation at 94° C. for 30 sec, a step of annealing at 55° C. for 30 sec and then a step of extension at 72° C. for 2 min 30 sec, with 10 sec of additional extension at each cycle, and then a final step of extension at 72° C. for 5 min.

a.3) Second PCR Amplification

[0369] The products of the first PCR amplification (5' and 3' amplicons) were subjected to a second PCR amplification step (nested PCR) under conditions identical to those of the first amplification, with the pairs of primers S/F2+/21406-21426 and S/R2-/23454-23435 and S/F4+/23322-23341 and S/R4-/25348-25329, respectively for the 5' amplicon and the 3' amplicon.

a.4) Cloning and Sequencing of the Sa and Sb Fragments

[0370] The Sa (5' end) and Sb (3' end) amplicons thus obtained were purified with the aid of the QIAquick PCR

purification kit (QIAGEN), following the manufacturer's instructions, and then they were cloned into the vector PCR2.1-TOPO (Invitrogen kit), to give the plasmids called SARS-S1 and SARS-S2.

[0371] The DNA of the Sa and Sb clones was isolated and then the corresponding insert was sequenced with the aid of the Big Dye kit, Applied Biosystem® and universal primers M13 forward and M13 reverse, and primers: S/S/+21867, S/S/+22353, S/S/+22811, S/S/+23754, S/S/+24207, S/S/+24699, S/S/+24348, S/S/-24209, S/S/-23630, S/S/-23038, S/S/-22454, S/S/-21815, S/S/-24784, S/S/+21556, S/S/+23130 and S/S/+24465 following the manufacturer's instructions; the sequences of the Sa and Sb fragments thus obtained correspond to the sequences SEQ ID NO: 5 and SEQ ID NO: 6 in the sequence listing appended as an annex.

[0372] The plasmid, called SARS-S1, was deposited under the No. I-3020, on May 12, 2003, at the Collection Nationale de Cultures de Microorganismes, 25 rue du Docteur Roux, 75724 Paris Cedex 15; it contains a 5' fragment of the sequence of the S gene of the SARS-CoV strain derived from the sample recorded under the No. 031589, as defined above, said fragment called Sa corresponding to the nucleotides at positions 21406 to 23454 (SEQ ID NO: 5), with reference to the Genbank sequence AY274119.3 Tor2.

[0373] The plasmid, called TOP10F⁺-SARS-S2, was deposited under the No. I-3019, on May 12, 2003, at the Collection Nationale de Cultures de Microorganismes, 25 rue du Docteur Roux, 75724 Paris Cedex 15; it contains a 3' fragment of the sequence of the S gene of the SARS-CoV strain derived from the sample recorded under the No. 031589, as defined above, said fragment called Sb corresponding to the nucleotides at positions 23322 to 25348 (SEQ ID NO: 6), with reference to the Genbank sequence accession No. AY274119.3.

b) Cloning and Sequencing of the Complete cDNA (SARS-S Clone of 4 kb)

[0374] The complete S cDNA was obtained from the abovementioned clones SARS-S1 and SARS-S2, in the following manner:

[0375] 1) A PCR amplification reaction was carried out on a SARS-S2 clone in the presence of the abovementioned primer S/R4/-/25348-25329 and of the primer S/S/+24696-24715: an amplicon of 633 bp was obtained,

[0376] 2) Another PCR amplification reaction was carried out on another SARS-S2 clone, in the presence of the primers S/F4/+23322-23341 mentioned above and S/S/-/24803-24784: an amplicon of 1481 bp was obtained.

[0377] The amplification reaction was carried out under the conditions as defined above for the amplification of the Sa and Sb fragments, with the exception that 30 amplification cycles comprising a step of denaturation at 94° C. for 20 sec and a step of extension at 72° C. for 2 min 30 sec were carried out.

[0378] 3) The 2 amplicons (633 bp and 1481 bp) were purified under the conditions as defined above for the Sa and Sb fragments.

[0379] 4) Another PCR amplification reaction with the aid of the abovementioned primers S/F4/+23322-23341 and

S/R4/-/25348-25329 was carried out on the purified amplicons obtained in 3). The amplification reaction was carried out under the conditions as defined above for the amplification of the Sa and Sb fragments, except that 30 amplification cycles were performed.

[0380] The 2026 bp amplicon thus obtained was purified, cloned into the vector PCR2.1-TOPO and then sequenced as above, with the aid of the primers as-defined above for the Sa and Sb fragments. The clone thus obtained was called clone 3'.

[0381] 5) The clone SARS-S1 obtained above and the clone 3"were digested with EcoR I, the bands of about 2 kb thus obtained were gel purified and then amplified by PCR with the abovementioned primers S/F2/+21406-21426 and S/R4/-/25348-25329. The amplification reaction was carried out under the conditions as defined above for the amplification of the Sa and Sb fragments, except that 30 amplification cycles were performed. The amplicon of about 4 kb was purified and sequenced. It was then cloned into the vector PCR2.1-TOPO in order to give the plasmid, called SARS-S, and the insert obtained in this plasmid was sequenced as above, with the aid of the primers as defined above for the Sa and Sb fragments. The cDNA sequences of the insert and of the amplicon encoding the S protein correspond respectively to the sequences SEQ ID NO: 4 and SEQ ID NO: 2 in the sequence listing appended as an annex, they encode the S protein (SEQ ID NO: 3).

[0382] The sequence of the amplicon corresponding to the cDNA encoding the S protein of the SARS-CoV strain derived from the sample No. 031589 has the following two mutations compared with the corresponding sequences of respectively the Tor2 and Urbani isolates, the positions of the mutations being indicated with reference to the complete sequence of the genome of the Tor2 isolate (Genbank AY274119.3):

[0383] g/t in position 23220; the alanine codon (gct) in position 577 of the amino acid sequence of the S protein of Tor2 is replaced with a serine codon (tct),

[0384] c/t in position 24872: this mutation does not modify the amino acid sequence of the S protein, and

the plasmid, called SARS-S, was deposited under the No. I-3059, on Jun. 20, 2003, at the Collection Nationale de Cultures de Microorganismes, 25 rue du Docteur Roux, 75724 Paris Cedex 15; it contains the cDNA sequence encoding the S protein of the SARS-CoV strain derived from the sample recorded under the No. 031589, said sequence corresponding to the nucleotides at positions 21406 to 25348 (SEQ ID NO: 4), with reference to the Genbank sequence AY274119.3.

2.2) cDNA Encoding the M and E Proteins

[0385] The RNAs derived from the sample 031589, extracted as above, were subjected to a reverse transcription, combined, during the same step (Titan One Step RT-PCR® kit, Roche), with a PCR amplification reaction, with the aid of the pairs of primers:

[0386] S/E/F1/+26051-26070 and S/E/R1/-/26455-26436 in order to amplify ORF-E, and

[0387] S/M/F1/+26225-26244 and S/M/R1/-/27148-27129 in order to amplify ORF-M.

[0388] A first reaction mixture containing: 8.6 μ l of H₂O for injection, 1 μ l of dNTP (5 mM), 0.2 μ l of each of the primers (50 μ M), 1.25 μ l of DTT (100 mM) and 0.25 μ l of RNAsin (40 IU/ μ l) was combined with a second reaction mixture containing: 1 μ l of RNA, 7 μ l of H₂O for injection, 5 μ l of 5 \times RT-PCR buffer and 0.5 μ l of enzyme mixture and the combined mixtures were incubated in a thermocycler under the following conditions: 30 min at 42° C., 10 min at 55° C., 2 min at 94° C. followed by 40 cycles comprising a step of denaturation at 94° C. for 10 sec, a step of annealing at 55° C. for 30 sec and a step of extension at 68° C. for 45 sec, with 3 sec increment per cycle and finally a step of terminal extension at 68° C. for 7 min.

[0389] The amplification products thus obtained (M and E amplicons) were subjected to a second PCR amplification (nested PCR) using the Expand High-Fi® kit, Roche), with the aid of the pairs of primers:

[0390] S/E/F2/+26082-26101 and S/E/R2/-/26413-26394 for the amplicon E, and

[0391] S/M/F2/+26330-26350 and S/M/R2/-/27098-27078 for the amplicon M.

[0392] The reaction mixture containing: 2 μ l of the product of the first PCR, 39.25 μ l of H₂O for injection, 5 μ l of 10 \times buffer containing MgCl₂, 2 μ l of dNTP (5 mM), 0.5 μ l of each of the primers (50 μ M) and 0.75 μ l of enzyme mixture was incubated in a thermocycler under the following conditions: a step of denaturation at 94° C. for 2 min was followed by 30 cycles comprising a step of denaturation at 94° C. for 15 sec, a step of annealing at 60° C. for 30 sec and a step of extension at 72° C. for 45 sec, with 3 sec increment per cycle, and finally a step of terminal extension at 72° C. for 7 min. The amplification products obtained corresponding to the cDNAs encoding the E and M proteins were sequenced as above, with the aid of the primers: S/E/F2/+26082 and S/E/R2/-/26394, S/M/F2/+26330, S/M/R2/-/27078 cited above and the primers S/M/+26636-26655 and S/M/-/26567-26548. They were then cloned, as above, in order to give the plasmids called SARS-E and SARS-M. The DNA of these clones was then isolated and sequenced with the aid of the universal primers M13 forward and M13 reverse and the primers S/M/+26636 and S/M/-/26548 mentioned above.

[0393] The sequence of the amplicon representing the cDNA encoding the E protein (SEQ ID NO: 13) of the SARS-CoV strain derived from the sample No. 031589 does not contain differences in relation to the corresponding sequences of the isolates AY274119.3-Tor2 and AY278741-Urbani. The sequence of the E protein of the SARS-CoV 031589 strain corresponds to the sequence SEQ ID NO: 14 in the sequence listing appended as an annex.

[0394] The plasmid, called SARS-E, was deposited under the No. I-3046, on May 28, 2003, at the Collection Nationale de Cultures de Microorganismes, 25 rue du Docteur Roux, 75724 Paris Cedex 15; it contains the cDNA sequence encoding the E protein of the SARS-CoV strain derived from the sample recorded under the No. 031589, as defined above, said sequence corresponding to the nucleotides at positions 26082 to 26413 (SEQ ID NO: 15), with reference to the Genbank sequence accession No. AY274119.3.

[0395] The sequence of the amplicon representing the cDNA encoding M (SEQ ID NO: 16) from the SARS-CoV strain derived from the sample No. 031589 does not contain differences in relation to the corresponding sequence of the isolate AY274119.3-Tor2. By contrast, at position 26857, the

isolate AY278741-Urbani contains a c and the sequence of the SARS-CoV strain derived from the sample recorded under the No. 031589 contains a t. This mutation results in a modification of the amino acid sequence of the corresponding protein: at position 154, a proline (AY278741-Urbani) is changed to serine in the SARS-CoV strain derived from the sample recorded under the No. 031589. The sequence of the M protein of the SARS-CoV strain derived from the sample recorded under the No. 031589 corresponds to the sequence SEQ ID NO: 17 in the sequence listing appended as an annex.

[0396] The plasmid, called SARS-M, was deposited under the No. I-3047, on May 28, 2003, at the Collection Nationale de Cultures de Microorganismes, 25 rue du Docteur Roux, 75724 Paris Cedex 15; it contains the cDNA sequence encoding the M protein of the SARS-CoV strain derived from the sample recorded under the No. 031589, as defined above; said sequence corresponding to the nucleotides at positions 26330 to 27098 (SEQ ID NO: 18), with reference to the Genbank sequence accession No. AY274119.3.

2.3) cDNA Corresponding to ORF3, ORF4, ORF7 to ORF11

[0397] The same amplification, cloning and sequencing strategy was used to obtain the cDNA fragments corresponding respectively to the following ORFs: ORF3, ORF4, ORF7, ORF8, ORF9, ORF10 and ORF11. The pairs of primers used for the first amplification are:

[0398] ORF3 and ORF4: S/SE/F1/+25069-25088 and S/SE/R1/-/26300-26281

[0399] ORF7 to ORF11: S/MN/F1/+26898-26917 and S/MN/R1/-/28287-28266

[0400] The pairs of primers used for the second amplification are:

[0401] ORF3 and ORF4: S/SE/F2/+25110-25129 and S/SE/R2/-/26244-26225

[0402] ORF7 to ORF11: S/MN/F2/+26977-26996 and S/MN/R2/-/28218-28199

[0403] The conditions for the first amplification (RT-PCR) are the following: 45 min at 42° C., 10 min at 55° C., 2 min at 94° C. followed by 40 cycles comprising a step of denaturation at 94° C. for 15 sec, a step of annealing at 58° C. for 30 sec and a step of extension at 68° C. for 1 min, with 5 sec increment per cycle and finally a step of terminal extension at 68° C. for 7 min. The conditions for the nested PCR are the following: a step of denaturation at 94° C. for 2 min was followed by 40 cycles comprising a step of denaturation at 94° C. for 20 sec, a step of annealing at 58° C. for 30 sec and a step of extension at 72° C. for 50 sec, with 4 sec increment per cycle and finally a step of terminal extension at 72° C. for 7 min.

[0404] The amplification products obtained corresponding to the cDNAs containing respectively ORF3 and 4 and ORF7 to 11 were sequenced with the aid of the primers: S/SE/+25363, S/SE/+25835, S/SE/-/25494, S/SE/-/25875, S/MN/+27839, S/MN/+27409, S/MN/-/27836, S/MN/-/27799 and cloned as above for the other ORFs, to give the plasmids called SARS-SE and SARS-MN. The DNA of these clones was isolated and sequenced with the aid of these same primers and of the universal primers M13 sense and M13 antisense.

[0405] The sequence of the amplicon representing the cDNA of the region containing ORF3 and ORF4 (SEQ ID NO: 7) of the SARS-CoV strain derived from the sample No.

031589 contains a nucleotide difference in relation to the corresponding sequence of the isolate AY274119-Tor2. This mutation at position 25298 results in a modification of the amino acid sequence of the corresponding protein (ORF3): at position 11, an arginine (AY274119-Tor2) is changed to glycine in the SARS-CoV strain derived from the sample No. 031589. By contrast, no mutation was identified in relation to the corresponding sequence of the isolate AY278741-Urbani. The sequences of ORF3 and 4 of the SARS-CoV strain derived from the sample No. 031589 correspond respectively to the sequences SEQ ID NO: 10 and 12 in the sequence listing appended as an annex.

[0406] The plasmid, called SARS-SE, was deposited under the No. I-3126, on Nov. 13, 2003, at the Collection Nationale de Cultures de Microorganismes, 25 rue du Docteur Roux, 75724 Paris Cedex 15; it contains the cDNA corresponding to the region situated between ORF-S and ORF-E and overlapping ORF-E of the SARS-CoV strain derived from the sample recorded under the No. 031589, as defined above, said region corresponding to the nucleotides at positions 25110 to 26244 (SEQ ID NO: 8), with reference to the Genbank sequence accession No. AY274119.3.

[0407] The sequence of the amplicon representing the cDNA corresponding to the region containing ORF7 to ORF11 (SEQ ID NO: 19) of the SARS-CoV strain derived from the sample No. 031589 does not contain differences in relation to the corresponding sequences of the isolates AY274119-Tor2 and AY278741-Urbani. The sequences of ORF7 to 11 of the SARS-CoV strain derived from the sample No. 031589 correspond respectively to the sequences SEQ ID NO: 22, 24, 26, 28 and 30 in the sequence listing appended as an annex.

[0408] The plasmid, called SARS-MN, was deposited under the No. I-3125, on Nov. 13, 2003, at the Collection Nationale de Cultures de Microorganismes, 25 rue du Docteur Roux, 75724 Paris Cedex 15; it contains the cDNA sequence corresponding to the region situated between ORF-M and ORF-N of the SARS-CoV strain derived from the sample recorded under the No. 031589 and collected in Hanoi, as defined above, said sequence corresponding to the nucleotides at positions 26977 to 28218 (SEQ ID NO: 20), with reference to the Genbank sequence accession No. AY274119.3.

[0409] The sequence of the amplicon representing the cDNA corresponding to the region containing ORF7 to ORF11 (SEQ ID NO: 19) of the SARS-CoV strain derived from the sample No. 031589 does not contain differences in relation to the corresponding sequences of the isolates AY274119-Tor2 and AY278741-Urbani. The sequences of ORF7 to 11 of the SARS-CoV strain derived from the sample No. 031589 correspond respectively to the sequences SEQ ID NO: 22, 24, 26, 28 and 30 in the sequence listing appended as an annex.

2.4) cDNA Encoding the N Protein and Including ORF13 and ORF14

[0410] The cDNA was synthesized and amplified as described above for the fragments Sa and Sb. More specifically, the reaction mixture containing: 5 µl of RNA, 5 µl of H₂O for injection, 4 µl of 5× reverse transcriptase buffer, 2 µl of dNTP (5 mM), 2 µl of oligo 20T (5 µM), 0.5 µl of RNasin (40 IU/µl) and 1.5 µl of AMV-RT (10 IU/µl Promega) was incubated in a thermocycler under the fol-

lowing conditions: 45 min at 42° C., 15 min at 55° C., 5 min at 95° C., and it was then kept at +4° C.

[0411] A first PCR amplification was performed with the pair of primers S/N/F3/+28023 and S/N/R3/-29480.

[0412] The reaction mixture as above for the amplification of the S1 and S2 fragments was incubated in a thermocycler, under the following conditions: an initial step of denaturation at 94° C. for 2 min was followed by 40 cycles comprising a step of denaturation at 94° C. for 20 sec, a step of annealing at 55° C. for 30 sec and then a step of extension at 72° C. for 1 min 30 sec with 10 sec of additional extension at each cycle, and then a final step of extension at 72° C. for 5 min.

[0413] The amplicon obtained at the first PCR amplification was subjected to a second PCR amplification step (nested PCR) with the pairs of primer S/N/F4/+28054 and S/N/R4/-29430 under conditions identical to those of the first amplification.

[0414] The amplification product obtained, corresponding to the cDNA encoding the N protein of the SARS-CoV strain derived from the sample No. 031589, was sequenced with the aid of the primers: S/N/F4/+28054, S/N/R4/-29430, S/N/+28468, S/N/+28918 and S/N/-28607 and cloned as above for the other ORFs, to give the plasmid called SARS-N. The DNA of these clones was isolated and sequenced with the aid of the universal primers M13 sense and M13 antisense, and the primers S/N/+28468, S/N/+28918 and S/N/-28607.

[0415] The sequence of the amplicon representing the cDNA corresponding to ORF-N and including ORF13 and ORF14 (SEQ ID NO: 36) of the SARS-CoV strain derived from the sample No. 031589 does not contain differences in relation to the corresponding sequences of the isolates AY274119.3-Tor2 and AY278741-Urbani. The sequence of the N protein of the SARS-CoV strain derived from the sample No. 031589 corresponds to the sequence SEQ ID NO: 37 in the sequence listing appended as an annex.

[0416] The sequences of ORF13 and 14 of the SARS-CoV strain derived from the sample No. 031589 correspond respectively to the sequences SEQ ID NO: 32 and 34 in the sequence listing appended as an annex.

[0417] The plasmid, called SARS-N, was deposited under the No. I-3048, on Jun. 5, 2003, at the Collection Nationale de Cultures de Microorganismes, 25 rue du Docteur Roux, 75724 Paris Cedex 15; it contains the cDNA encoding the N protein of the SARS-CoV strain derived from the sample recorded under the No. 031589, as defined above, said sequence corresponding to the nucleotides at positions 28054 to 29430 (SEQ ID NO: 38), with reference to the Genbank sequence accession No. AY274119.3.

2.5) Noncoding 5' and 3' Ends

a) Noncoding 5' end (5'NC)

a₁) Synthesis of the cDNA

[0418] The RNAs derived from the sample 031589, extracted as above, were subjected to reverse transcription under the following conditions:

[0419] The RNA (15 µl) and the primer S/L/-443 (3 µl at the concentration of 5 µM) were incubated for 10 min at 75° C.

[0420] Next, the 5× reverse transcriptase buffer (6 μl, INVITROGEN), 10 Mm dNTP (1 μl), 0.1 M DTT (3 μl) were added and the mixture was incubated at 50° C. for 3 min.

[0421] Finally, the reverse transcriptase (3 μl of Super-script®, INVITROGEN) was added to the preceding mixture which was incubated at 50° C. for 1 h 30 min and then at 90° C. for 2 min.

[0422] The cDNA thus obtained was purified with the aid of the QIAquick PCR purification kit (QIAGEN) according to the manufacturer's recommendations.

b₁) Terminal Transferase Reaction (TdT)

[0423] The cDNA (10 μl) is incubated for 2 min at 100° C., stored in ice, and the following are then added: H₂O (2.5 μl), 5× TdT buffer (4 μl, AMERSHAM), 5 mM dATP (2 μl) and TdT (1.5 μl, AMERSHAM). The mixture thus obtained is incubated for 45 min at 37° C. and then for 2 min at 65° C.

[0424] The product obtained is amplified by a first PCR reaction with the aid of the primers: S/L/-225-206 and anchor 14T: 5'-AGATGAATTCGGTAC-CTTTTTTTTTTTTTTTT-3' (SEQ ID NO: 68). The amplification conditions are the following: an initial step of denaturation at 94° C. for 2 min is followed by 10 cycles comprising a step of denaturation at 94° C. for 10 sec, a step of annealing at 45° C. for 30 sec and then a step of extension at 72° C. for 30 sec and then by 30 cycles comprising a step of denaturation at 94° C. for 10 sec, a step of annealing at 50° C. for 30 sec and then a step of extension at 72° C. for 30 sec, and then a final step of extension at 72° C. for 5 min.

[0425] The product of the first PCR amplification was subjected to a second amplification step with the aid of the primers: S/L/-204-185 and anchor 14T mentioned above under conditions identical to those of the first amplification. The amplicon thus obtained was purified, sequenced with the aid of the primer S/L/-182-163 and it was then cloned as above for the different ORFs, to give the plasmid called SARS-5'NC. The DNA of this clone was isolated and sequenced with the aid of the universal primers M13 sense and M13 antisense and the primer S/L/-182-163 mentioned above.

[0426] The amplicon representing the cDNA corresponding to the 5'NC end of the SARS-CoV strain derived from the sample recorded under the No. 031589 corresponds to the sequence SEQ ID NO: 72 in the sequence listing appended as an annex; this sequence does not contain differences in relation to the corresponding sequences of the isolates AY274119.3-Tor2 and AY278741-Urbani.

[0427] The plasmid, called SARS-5'NC, was deposited under the No. I-3124, on Nov. 7, 2003, at the Collection Nationale de Cultures de Microorganismes, 25 rue du Docteur Roux, 75724 Paris Cedex 15; it contains the cDNA corresponding to the noncoding 5' end of the genome of the SARS-CoV strain derived from the sample recorded under the No. 031589, as defined above, said sequence corresponding to the nucleotides at positions 1 to 204 (SEQ ID NO: 39), with reference to the Genbank sequence accession No. AY274119.3.

b) Noncoding 3' end (3'NC)

a₁) Synthesis of the cDNA

[0428] The RNAs derived from the sample 031589, extracted as above, were subjected to reverse transcription, according to the following protocol: the reaction mixture containing: RNA (5 μl), H₂O (5 μl), 5× reverse transcriptase buffer (4 μl), 5 mM dNTP (2 μl), 5 μM Oligo 20T (2 μl), 40 U/μl RNasin (0.5 μl) and 10 IU/μl RT-AMV (1.5 μl, PROMEGA) was incubated in a thermocycler, under the following conditions: 45 min at 42° C., 15 min at 55° C., 5 min at 95° C., and it was then kept at +4° C.

[0429] The cDNA obtained was amplified by a first PCR reaction with the aid of the primers S/N/+28468-28487 and anchor 14T mentioned above. The amplification conditions are the following: an initial step of denaturation at 94° C. for 2 min is followed by 10 cycles comprising a step of denaturation at 94° C. for 20 sec, a step of annealing at 45° C. for 30 sec and then a step of extension at 72° C. for 50 sec and then 30 cycles comprising a step of denaturation at 94° C. for 20 sec, a step of annealing at 50° C. for 30 sec and then a step of extension at 72° C. for 50 sec, and then a final step of extension at 72° C. for 5 min.

[0430] The product of the first PCR amplification was subjected to a second amplification step with the aid of the primers S/N/+28933-28952 and anchor 14T mentioned above, under conditions identical to those of the first amplification. The amplicon thus obtained was purified, sequenced with the aid of the primer S/N/+29257-29278 and cloned as above for the different ORFs, to give the plasmid called SARS-3'NC. The DNA of this clone was isolated and sequenced with the aid of the universal primers M13 sense and M13 antisense and the primer S/N/+29257-29278 mentioned above.

[0431] The amplicon representing the cDNA corresponding to the 3'NC end of the SARS-CoV strain derived from the sample recorded under the No. 031589 corresponds to the sequence SEQ ID NO: 73 in the sequence listing appended as an annex; this sequence does not contain differences in relation to the corresponding sequences of the isolates AY274119.3-Tor2 and AY278741-Urbani.

[0432] The plasmid called SARS-3'NC was deposited under the No. I-3123 on Nov. 7, 2003, at the Collection Nationale de Cultures de Microorganismes, 25 rue du Docteur Roux, 75724 Paris Cedex 15; it contains the cDNA sequence corresponding to the noncoding 3' end of the genome of the SARS-CoV strain derived from the sample recorded under the No. 031589, as defined above, said sequence corresponding to that situated between the nucleotide at positions 28933 to 29277 (SEQ ID NO: 40), with reference to the Genbank sequence accession No. AY274119.3, ends with a series of nucleotides a.

2.6) ORF1a and ORF1b

[0433] The amplification of the 5' region containing ORF1a and ORF1b of the SARS-CoV genome derived from the sample 031589 was performed by carrying out RT-PCR reactions followed by nested PCRs according to the same principles as those described above for the other ORFs. The amplified fragments overlap over several tenths of bases, thus allowing computer reconstruction of the complete sequence of this part of the genome. On average, the amplified fragments are of two kilobases.

[0434] 14 overlapping fragments, called L0 to L12, were thus amplified with the aid of the following primers:

TABLE II

Primers used for the amplification of the 5' region (ORF1a and ORF1b)				
REGION AMPLIFIED AND SEQUENCED (does not include the primers)	RT-PCR sense primer	RT-PCR antisense primer	Nested PCR sense primer	Nested PCR antisense primer
L0 50-480	S/L0/F1/+30	S/L0/R1/-481		
L1 231-2240	S/L1/F1/+147	S/L1/R1/-2336	S/L1/F2/+211	S/L1/R2/-2241
L2 2156-4167	S/L2/F1/+2033	S/L2/R1/-4192	S/L2/F2/+2136	S/L2/R2/-4168
L3 3913-5324	S/L3bis/F1+3850	S/L3bis/R1/-5365	S/L3bis/F2+3892	S/L3bis/R2/-5325
L4b 4952-6023	S/L4b/F1/+4878	S/L4b/R1/-6061	S/L4b/F2/+4932	S/L4b/R2/-6024
L4 5325-7318	S/L4/F1/+5272	S/L4/R1/-7392	S/L4/F2/+5305	S/L4/R2/-7323
L5 7296-9156	S/L5/F1/+7111	S/L5/R1/-9253	S/L5/F2/+7275	S/L5/R2/-9157
L6 9053-11066	S/L6/F1/+8975	S/L6/R1/-11151	S/L6/F2/+9032	S/L6/R2/-11067
L7 10928-12962	S/L7/F1/+10883	S/L7/R1/-13050	S/L7/F2/+10928	S/L7/R2/-12963
L8 12835-14834	S/L8/F1/+12690	S/L8/R1/-14857	S/L8/F2/+12815	S/L8/R2/-14835
L9 14765-16624	S/L9/F1/+14688	S/L9/R1/-16678	S/L9/F2/+14745	S/L9/R2/-16625
L10 16534-18570	S/L10/F1/+16451	S/L10/R1/-18594	S/L10/F2/+16514	S/L10/R2/-18571
L11 18521-20582	S/L11/F1/+18441	S/L11/R1/-20612	S/L11/F2/+18500	S/L11/R2/-20583
L12 20338-22205.	S/L12/F1/+20279	S/L12/R1/-22229	S/L12/F2/+20319	S/L12/R2/-22206

All the fragments were amplified under the following conditions, except fragment L0 which was amplified as described above for ORF-M:

RT-PCR: 30 min at 42° C., 15 min at 55° C., 2 min at 94° C., and then the cDNA obtained is amplified under the following conditions: 40 cycles comprising: a step of denaturation at 94° C. for 15 sec, a step of annealing at 58° C. for 30 sec and then a step of extension at 68° C. for 1 min 30 sec, with 5 sec additional extension at each cycle, and then a final step of extension at 68° C. for 7 min.

Nested PCR: An initial step of denaturation at 94° C. for 2 min is followed by 35 cycles comprising: a step of denaturation at 94° C. for 15 sec, a step of annealing at 60° C. for 30 sec and then a step of extension at 72° C. for 1 min 30 sec, with 5 sec of additional extension at each cycle, and then a final step of extension at 72° C. for 7 min.

[0435] The amplification products were sequenced with the aid of the primers defined in table III below:

TABLE III

Primers used for the sequencing of the 5' region (ORF1a and ORF1b)	
Names	Sequences (SEQ ID NO: 76 to 139)
S/L3/+4932	5'-CCACACACAGCTTGTGGATA-3'
S/L4/+6401	5'-CCGAAGTTGTAGGCAATGTC-3'
S/L4/+6964	5'-TTGGTGCTCCTTCTTATTG-3'
S/L4/-/6817	5'-CCGGCATCCAAACATAATTT-3'
S/L5/-/7633	5'-TGGTCACTAGGGTTGATTGG-3'
S/L5/-/8127	5'-CATCCTTTGTGTC AACATCG-3'
S/L5/-/8633	5'-GTCACGAGTGACACCAATCCT-3'
S/L5/+7839	5'-ATGCGACGAGTCTGCTTCTA-3'
S/L5/+8785	5'-TTCATAGTGCCTGGCTTACC-3'
S/L5/+8255	5'-ATCTTGGCGCATGTATTGAC-3'
S/L6/-/9422	5'-TGCATTAGCAGCAACAACAT-3'

TABLE III-continued

Primers used for the sequencing of the 5' region (ORF1a and ORF1b)		
Names	Sequences (SEQ ID NO: 76 to 139)	
S/L6/-/9966	5'-TCTGCAGAACAGCAGAAGTG-3'	
S/L6/-/10542	5'-CCTGTGTCAGTTTGTCTGTCA-3'	
S/L6/+10677	5'-CCTGTGTCAGTGAAGTACA-3'	
S/L6/+10106	5'-ATGTCATTTGCACAGCAGAA-3'	
S/L6/+9571	5'-CTTCAATGGTTTGCCATGTT-3'	
S/L7/-/11271	5'-TGCGAGCTGTCATGAGAATA-3'	
S/L7/-/11801	5'-AACCGAGAGCAGTACCACAG-3'	
S/L7/-/12383	5'-TTTGGCTGCTGTAGTCAATG-3'	
S/L7/+12640	5'-CTACGACAGATGCTCTGTGC-3'	
S/L7/+12088	5'-GAGCAGGCTGTAGTCAATG-3'	
S/L7/+11551	5'-TTAGGCTATTGTTGCTGCTG-3'	
S/L8/-13160	5'-CAGACAACATGAAGCACCAC-3'	
S/L8/-/13704	5'-CGTGACGTGATATATGTGG-3'	

TABLE III-continued

Primers used for the sequencing of the 5' region (ORF1a and ORF1b)	
Names	Sequences (SEQ ID NO: 76 to 139)
S/L8/-14284	5'-TGCACAATGAAGGATACACC-3'
S/L8/+14453	5'-ACATAGCTCGGCTCAGTT-3'
S/L8/+13968	5'-GGCATTGTAGGCGTACTGAC-3'
S/L8/+13401	5'-GTTTGCGGGTAAAGTGACAG-3'
S/L9/-15098	5'-TAGTGGCGGCTATTGACTTC-3'
S/L9/-15677	5'-CTAAACCTTGAGCCGCATAG-3'
S/L9/-16247	5'-CATGGTCATAGCAGCACTTG-3'
S/L9/+16323	5'-CCAGGTTGTGATGTCCTGAT-3'
S/L9/+15858	5'-CCTTACCCAGATCCATCAAG-3'
S/L9/+15288	5'-CGCAAACATAACACTTGCTG-3'
S/L10/-16914	5'-AGTGTGGGTACAAGCCAGT-3'
S/L10/-17466	5'-GTTCCAAGGAACATGTCTGG-3'
S/L10/-18022	5'-AGGTGCCTGTGTAGGATGAA-3'
S/L10/+18245	5'-GGGCTGTTCATGCAACTAGAG-3'
S/L10/+17663	5'-TCTTACACGCAATCTGCTT-3'
S/L10/+17061	5'-TACCCATCTGCTCGCATAGT-3'
S/L11/-18877	5'-GCAAGCAGAATTAACCCTCA-3'
S/L11/-19396	5'-AGCACCACCTAAATTGCATC-3'
S/L11/-20002	5'-TGGTCCCTTTGAAGGTGTTA-3'
S/L11/+20245	5'-TCGAACACATCGTTTATGGA-3'
S/L11/+19611	5'-GAAGCACCTGTTTCCATCAT-3'
S/L11/+19021	5'-ACGATGCTCAGCCATGTAGT-3'
SARS/L1/F3/+800	5'-GAGGTGCACTCACTCGCTAT-3'
SARS/L1/F4/+1391	5'-CAGAGATTGGACCTGAGCAT-3'
SARS/L1/F5/+1925	5'-CAGCAAACCACTCAATTCCT-3'
SARS/L1/R3/-1674	5'-AAATGATGGCAACCTCTCA-3'
SARS/L1/R4/-1107	5'-CACGTGGTTGAATGACTTTG-3'
SARS/L1/R5/-520	5'-ATTTCTGCAACCAGCTCAAC-3'
SARS/L2/F3/+2664	5'-CGCATTGTCTCCTGGTTTAC-3'
SARS/L2/F4/+3232	5'-GAGATTGAGCCAGAACCAGA-3'
SARS/L2/F5/+3746	5'-ATGAGCAGGTTGTCTATGGAT-3'
SARS/L2/R3/-3579	5'-CTGCCCTAAGAAGCTGGATG-3'
SARS/L2/R4/-2991	5'-TTTCTTACCAGCATCATCA-3'
SARS/L2/R5/-2529	5'-CACCGTCTTGAGAAACAACC-3'
SARS/L3/F3/+2664	5'-TCTTTGGCTGGCTTACAG-3'
SARS/L3/F4/+5305	5'-GCTGGTGATGCTGCTAATT-3'
SARS/L3/F5/+5822	5'-CCATCAAGCCTGTGTGCTAT-3'
SARS/L3/R3/-5610	5'-CAGGTGGTGCAGACATCATA-3'
SARS/L3/R4/-4988	5'-AACATCAGCACCATCAAGT-3'
SARS/L3/R5/-4437	5'-ATCGGACACCATAGTCAACG-3'

[0436] The sequences of the fragments L0 to L12 of the SARS-CoV strain derived from the sample recorded under the No. 031589 correspond respectively to the sequences SEQ ID NO: 41 to SEQ ID NO: 54 in the sequence listing appended as an annex. Among these sequences, only that corresponding to the fragments L5 contains a nucleotide difference in relation to the corresponding sequence of the isolate AY278741-Urbani. This t/c mutation at position 7919 results in a modification of the amino acid sequence of the corresponding protein, encoded by ORF1a: at position 2552, a valine (gtt codon; AY278741) is changed to alanine (gct codon) in the SARS-CoV strain 031589. By contrast, no mutation was identified in relation to the corresponding sequence of the isolate AY274119.3-Urbani. The other fragments do not exhibit differences in relation to the corresponding sequences of the isolates Tor2 and Urbani.

EXAMPLE 2

Production and Purification of the Recombinant N and S Proteins of the SARS-CoV Strain Derived from the Sample Recorded Under the Number 031589

[0437] The entire N protein and two polypeptide fragments of the S protein of the SARS-CoV strain derived from

the sample recorded under the number 031589 were produced in *E. coli*, in the form of fusion proteins comprising an N- or C-terminal polyhistidine tag. In the two S polypeptides, the N- and C-terminal hydrophobic sequences of the S protein (signal peptide: positions 1 to 13 and transmembrane helix: positions 1196 to 1218) were deleted whereas the β helix (positions 565 to 687) and the two motifs of the coiled-coil type (positions 895 to 980 and 1155 to 1186) of the S protein were preserved. These two polypeptides consist of: a long fragment (S_L) corresponding to positions 14 to 1193 of the amino acid sequence of the S protein and a short fragment (S_C) corresponding to positions 475 to 1193 of the amino acid sequence of the S protein.

1) Cloning of the cDNAs N, S_L and S_C into the Expression Vectors pIVEX2.3 and pIVEX2.4

[0438] The cDNAs corresponding to the N protein and to the S_L and S_C fragments were amplified by PCR under standard conditions, with the aid of the DNA polymerase Platinum Pfx® (INVITROGEN). The plasmids SRAS-N and SRAS-S were used as template and the following oligonucleotides as primers:

5'- <u>CCCATATG</u> TCTGATAATGGACCCCAATCAAA C-3'	(N sense, SEQ ID NO: 55)
5'- <u>CCCCGGG</u> TGCCTGAGTTGAATCAGCAGAAG C-3'	(N antisense, SEQ ID NO: 56)
5'- <u>CCCATATG</u> AGTGACCTTGACCGGTGCACCA C-3'	(S_C sense, SEQ ID NO: 57)
5'- <u>CCCATATG</u> AAACCTTGACCCACCTGCT C-3'	(S_L sense, SEQ ID NO: 58)
5'- <u>CCCCGGG</u> TTAATATATTGCTCATATTTTCC C-3'	(S_C and S_L antisense, SEQ ID NO: 29)

[0439] The sense primers introduce an NdeI site (underlined) while the antisense primers introduce an XmaI site (underlined). The 3 amplification products were column purified (QIAquick PCR Purification kit, QIAGEN) and cloned into an appropriate vector. The plasmid DNA purified from the 3 constructs (QIAfilter Midi Plasmid kit, QIAGEN) was verified by sequencing and digested with the enzymes NdeI and XmaI. The 3 fragments corresponding to the cDNAs N, S_L and S_C were purified on agarose gel and then inserted into the plasmids pIVEX2.3MCS (C-terminal polyhistidine tag) and pIVEX2.4d (N-terminal polyhistidine tag) digested beforehand with the same enzymes. After verification of the constructs, the 6 expression vectors thus obtained (pIV2.3N, pIV2.3 S_C , pIV2.3 S_L , pIV2.4N, pIV2.4 S_C also called pIV2.4 S_1 , pIV2.4 S_2) were then used, on the one hand to test the expression of the proteins in vitro, and on the other hand to transform the bacterial strain BL21(DE3)pDIA17 (NOVAGEN). These constructs encode proteins whose expected molecular mass is the following: pIV2.3N (47174 Da), pIV2.3 S_C (82897 Da), pIV2.3 S_L (132056 Da), pIV2.4N (48996 Da), pIV2.4 S_1 (81076 Da) and pIV2.4 S_2 (133877 Da). Bacteria transformed with pIV2.3N were deposited at the CNCM on Oct. 23, 2003, under the number I-3117, and bacteria transformed with pIV2.4 S_1 were deposited at the CNCM on Oct. 23, 2003, under the number I-3118.

2) Analysis of the Expression of the Recombinant Proteins In Vitro and In Vivo

[0440] The expression of recombinant proteins from the 6 recombinant vectors was tested, in a first instance, in a

system in vitro (RTS100, Roche). The proteins produced in vitro, after incubation of the recombinant vectors pIVEX for 4 h at 30° C., in the RTS100 system, were analyzed by Western blotting with the aid of an anti-(his)₆ antibody coupled to peroxidase. The result of expression in vitro (FIG. 1) shows that only the N protein is expressed in large quantities, regardless of the position, N- or C-terminal, of the polyhistidine tag. In a second step, the expression of the N and S proteins was tested in vivo at 30° C. in LB medium in the presence or in the absence of inducer (1 mM IPTG). The N protein is very well produced in this bacterial system (FIG. 2) and is found mainly in a soluble fraction after lysis of the bacteria. By contrast, the long version of S (S_L) is very weakly produced and is completely insoluble (FIG. 3). The short version (S_C) also exhibits a very weak solubility, but an expression level that is much higher than that of the long version. Moreover, the construct S_C fused with a polyhistidine tag at the C-terminal position has a smaller size than that expected. An immunodetection experiment with an anti-polyhistidine antibody has shown that this construct was incomplete. In conclusion, the two constructs, pIV2.3N and pIV2.4S₁, which express respectively the entire N protein fused with the C-terminal polyhistidine tag and the short S protein fused with the N-terminal polyhistidine tag, were selected in order to produce the two proteins in a large quantity so as to purify them. The plasmids pIV2.3N and pIV2.4S were deposited respectively under the No. I-3117 and I-3118 at the CNCM, 25 rue du Docteur Roux, 75724 PARIS 15, on Oct. 23, 2003.

3) Analysis of the Antigenic Activity of the Recombinant Proteins

[0441] The antigenic activity of the N, S_L and S_C proteins was tested by Western blotting with the aid of two serum samples, obtained from the same patient infected with SARS-CoV, collected 8 days (M12) and 29 days (M13) after the onset of the SARS symptoms. The experimental protocol is as described in example 3. The results illustrated by FIG. 4 show (i) the seroconversion of the patient, and (ii) that the N protein possesses a higher antigenic reactivity than the short S protein.

4) Purification of the N Protein from pIV2.3N

[0442] Several experiments for purifying the N protein, produced from the vector pIV2.3N, were carried out according to the following protocol. The bacteria BL21(DE3)pDIA17, transformed with the expression vector pIV2.3N, were cultured at 30° C. in 1 liter of culture medium containing 0.1 mg/ml of ampicillin, and induced with 1 mM IPTG when the cell density equivalent to A₆₀₀=0.8 is reached (about 3 hours). After 2 hours of culture in the presence of inducer, the cells were recovered by centrifugation (10 min at 5000 rpm), resuspended in the lysis buffer (50 mM NaH₂PO₄, 0.3 M NaCl, 20 mM imidazole, pH 8, containing the mixture of protease inhibitors Complete®, Roche), and lysed with the French press (12 000 psi). After centrifugation of the bacterial lysate (15 min at 12 000 rpm), the supernatant (50 ml) was deposited at a flow rate of 1 ml/min on a metal chelation column (15 ml) (Ni-NTA superflow, Qiagen), equilibrated with the lysis buffer. After washing the column with 200 ml of lysis buffer, the N protein was eluted with an imidazole gradient (20→250 mM) in 10 column volumes. The fractions containing the N protein were assembled and analyzed by polyacrylamide gel electrophoresis under denaturing conditions followed by staining with Coomassie blue. The results illus-

trated by FIG. 5 show that the protocol used makes it possible to purify the N protein with a very satisfactory homogeneity (95%) and a mean yield of 15 mg of protein per liter of culture.

5) Purification of the S_C Protein from pIV2.4S_C (pIV2.4S₁)

[0443] The protocol followed for purifying the short S protein is very different from that described above because the protein is highly aggregated in the bacterial system (inclusion bodies). The bacteria BL21(DE3)pDIA17, transformed with the expression vector pIV2.4S₁, were cultured at 30° C. in 1 liter of culture medium containing 0.1 mg/ml of ampicillin, and induced with 1 mM IPTG when the cell density equivalent to A₆₀₀=0.8 is reached (about 3 hours). After 2 hours of culture in the presence of inducer, the cells were recovered by centrifugation (10 min at 5000 rpm), resuspended in the lysis buffer (0.1 M Tris-HCl, 1 mM EDTA, pH 7.5), and lysed with the French press (1200 psi). After centrifugation of the bacterial lysate (15 min at 12 000 rpm), the pellet was resuspended in 25 ml of lysis buffer containing 2% Triton X100 and 10 mM β-mercaptoethanol, and then centrifuged for 20 min at 12 000 rpm. The pellet was resuspended in 10 mM Tris-HCl buffer containing 7 M urea, and gently stirred for 30 min at room temperature. This final washing of the inclusion bodies with 7 M urea is necessary in order to remove most of the *E. coli* membrane proteins which co-sediment with the aggregated S_C protein. After a final centrifugation for 20 min at 12 000 rpm, the final pellet is resuspended in the 10 mM Tris-HCl buffer. The electrophoretic analysis of this preparation (FIG. 6) shows that the short S protein may be purified with a satisfactory homogeneity (about 90%) from the inclusion bodies (insoluble extract).

EXAMPLE 3

Immunodominance of the N Protein

[0444] The reactivity of the antibodies present in the serum of patients suffering from atypical pneumopathy caused by the SARS-associated coronavirus (SARS-CoV), toward the various proteins of this virus, was analyzed by Western blotting under the conditions described below.

1) Materials

a) Lysate of Cells Infected with SARS-CoV

[0445] Vero E6 cells (2×10⁶) were infected with SARS-CoV (isolate recorded under the number FFM/MA104) at a multiplicity of infection (M.O.I.) of 10⁻¹ or 10⁻² and then incubated in DMEM medium containing 2% FCS, at 35° C. in an atmosphere containing 5% CO₂. 48 hours later, the cellular lawn was washed with PBS and then lysed with 500 μl of loading buffer prepared according to Laemmli and containing β-mercaptoethanol. The samples were then boiled for 10 minutes and then sonicated for 3 times 20 seconds.

b) Antibodies

b₁) Serum from a Patient Suffering from Atypical Pneumopathy

[0446] The serum designated by a reference at the National Reference Center for Influenza Viruses (Northern region) under the No. 20033168 is that from a French patient suffering from atypical pneumopathy caused by SARS-CoV

collected on day 38 after the onset of the symptoms; the diagnosis of SARS-CoV infection was performed by nested RT-PCR and quantitative PCR.

b₂) Monospecific Rabbit Polyclonal Sera Directed Against the N Protein or the S Protein

[0447] The sera are those produced from the recombinant N and S_C proteins (example 2), according to the immunization protocol described in example 4; they are the rabbit P13097 serum (anti-N serum) and the rabbit P11135 serum (anti-S serum).

2) Method

[0448] 20 μl of lysate of cells infected with SARS-CoV at M.O.I. values of 10⁻¹ and 10⁻² and, as a control, 20 μl of a lysate of noninfected cells (mock) were separated on 10% SDS polyacrylamide gel and then transferred onto a nitrocellulose membrane. After blocking in a solution of PBS/5% milk/0.1% Tween and washing in PBS/0.1% Tween, this membrane was hybridized overnight at 4° C. with: (i) the immune serum No. 20033168 diluted 1/300, 1/1000 and 1/3000 in the buffer PBS/1% BSA/0.1% Tween, (ii) the rabbit P13097 serum (anti-N serum) diluted 1/50 000 in the same buffer and (iii) the rabbit P11135 serum (anti-S serum) diluted 1/10 000 in the same buffer. After washing in PBS/Tween, a secondary hybridization was performed with the aid of either sheep polyclonal antibodies directed against the heavy and light chains of human G immunoglobulins and coupled with peroxidase (NA933V, Amersham), or of donkey polyclonal antibodies directed against the heavy and light chains of the rabbit G immunoglobulins and coupled with peroxidase (NA934V, Amersham). The bound antibodies were visualized with the aid of the ECL+ kit (Amersham) and of Hyperfilm MP autoradiography films (Amersham). A molecular mass ladder (kDa) is presented in the figure.

3) Results

[0449] FIG. 7 shows that three polypeptides of apparent molecular mass 35, 55 and 200 kDa are specifically detected in the extracts of cells infected with SARS-CoV.

[0450] In order to identify these polypeptides, two other immunoblots (FIG. 8) were prepared on the same samples and under the same conditions with rabbit polyclonal antibodies specific for the nucleoprotein N (rabbit P13097, FIG. 8A) and for the spicule protein S (rabbit P11135, FIG. 8B). This experiment shows that the 200 kDa polypeptide corresponds to the SARS-CoV spicule glycoprotein S, that the 55 kDa polypeptide corresponds to the nucleoprotein N while the 35 kDa polypeptide probably represents a truncated or degraded form of N.

[0451] The data presented in FIG. 7 therefore show that the serum 20033168 strongly reacts with N and a lot more weakly with the SARS-CoV S since the 35 and 55 kDa polypeptides are visualized in the form of intense bands for 1/300, 1/1000 and 1/3000 dilutions of the immunoserum whereas the 200 kDa polypeptide is only weakly visualized for a dilution of 1/300. It is also possible to note that no other SARS-CoV polypeptide is detected for dilutions greater than 1/300 of the serum 20033168.

[0452] This experiment indicates that the antibody response specific for the SARS-CoV N dominates the antibody responses specific for the other SARS-CoV polypeptides and in particular the antibody response directed against

the S glycoprotein. It indicates an immuno-dominance of the nucleoprotein N during human infections with SARS-CoV.

EXAMPLE 4

Preparation of Monospecific Polyclonal Antibodies Directed Against the SRAS-associated Coronavirus (SARS-CoV) N and S Proteins

1) Materials and Method

[0453] Three rabbits (P13097, P13081, P13031) were immunized with the purified recombinant polypeptide corresponding to the entire nucleoprotein (N), prepared according to the protocol described in example 2. After a first injection of 0.35 mg per rabbit of protein emulsified in complete Freund's adjuvant (intradermal route), the animals received 3 booster injections at 3 and then 4 weeks' interval, of 0.35 mg of recombinant protein emulsified in incomplete Freund's adjuvant.

[0454] Three rabbits (P11135, P13042, P14001) were immunized with the recombinant polypeptide corresponding to the short fragment of the S protein (S_C) produced as described in example 2. As this polypeptide is found mainly in the form of inclusion bodies in the bacterial cytoplasm, the animals received 4 intradermal injections at 3-4 weeks' interval of a preparation of inclusion bodies corresponding to 0.5 mg of recombinant protein emulsified in incomplete Freund's adjuvant. The first 3 injections were made with a preparation of inclusion bodies prepared according to the protocol described in example 2, while the fourth injection was made with a preparation of inclusion bodies which were prepared according to the protocol described in example 2 and then purified on sucrose gradient and washed in 2% Triton X100.

[0455] For each rabbit, a preimmune (p.i.) serum was prepared before the first immunization and an immune serum (I.S.) 5 weeks after the fourth immunization.

[0456] In a first instance, the reactivity of the sera was analyzed by ELISA test on preparations of recombinant proteins similar to those used for the immunizations; the ELISA tests were carried out according to the protocol and with the reagents as described in example 6.

[0457] In a second instance, the reactivity of the sera was analyzed by preparing an immunoblot (Western blot) of a lysate of cells infected with SARS-CoV, according to the protocol as described in example 3.

2) Results

[0458] The ELISA tests (FIG. 9) demonstrate that the preparations of recombinant N protein and of inclusion bodies of the short fragment of the S protein (S_C) are immunogenic in animals and that the titer of the immune sera is high (more than 1/25 000).

[0459] The immunoblot (FIG. 8) shows that the rabbit P13097 immune serum recognizes two polypeptides present in the lysates of cells infected with SARS-CoV: a polypeptide whose apparent molecular mass (50-55 kDa based on experiments) is compatible with that of the nucleoprotein N (422 residues, predicted molecular mass of 46 kDa) and a polypeptide of 35 kDa, which probably represents a truncated or degraded form of N.

[0460] This experiment also shows that the rabbit P11135 serum mainly recognizes a polypeptide whose apparent molecular mass (180-220 kDa based on experiments) is compatible with a glycosylated form of S (1255 residues, nonglycosylated polypeptide chain of 139 kDa), as well as lighter polypeptides, which probably represent truncated and/or nonglycosylated forms of S.

[0461] In conclusion, all these experiments demonstrate that the recombinant polypeptides expressed in *E. coli* and corresponding to the SARS-CoV N and S proteins make it possible to induce, in animals, polyclonal antibodies capable of recognizing the native forms of these proteins.

EXAMPLE 5

Preparation of Monospecific Polyclonal Antibodies Directed Against the SARS-associated Coronavirus (SARS-CoV) M and E Proteins

1) Analysis of the Structure of the M and E Proteins

a) E Protein

[0462] The structure of the SARS-CoV E protein (76 amino acids) was analyzed in silico, with the aid of various software packages such as signalP v1.1, NetNGlyc 1.0, THMM 1.0 and 2.0 (Krogh et al., 2001, *J. Mol. Biol.*, 305(3):567-580) or alternatively TOPPED (von Heijne, 1992, *J. Mol. Biol.* 225, 487-494). The analysis shows that this nonglycosylated polypeptide is a type 1 membrane protein, containing a single transmembrane helix (aa 12-34 according to THMM), and in which the majority of the hydrophilic domain (42 residues) is located at the C-terminal end and probably inside the viral particle (endodomain). It is possible to note an inversion in the topology predicted by versions 1.0 (N-ter is external) and 2.0 (N-ter is internal) of the THMM software, but that other algorithms, in particular TOPPED and THUMBUP (Zhou et Zhou, 2003, *Protein Science* 12:1547-1555) confirm an external location of the N-terminal end of E.

b) M Protein

[0463] A similar analysis carried out on the SARS-CoV M protein (221 amino acids) shows that this polypeptide does not possess a signal peptide (according to the software signalP v1.1) but three transmembrane domains (residues 15-37, 50-72, 77-99 according to THMM2.0) and a large hydrophilic domain (aa 100-221) located inside the viral particle (endodomain). It is probably glycosylated on the asparagine at position 4 (according to NetNGlyc 1.0).

[0464] Thus, in agreement with the experimental data known for the other coronaviruses, it is remarkable that the two M and E proteins exhibit endodomains corresponding to the majority of the polypeptides and of the ectodomains that are very small in size.

[0465] The ectodomain of E probably corresponds to residues 1 to 11 or 1 to 12 of the protein: MYSFVSEETGT(L), SEQ ID NO: 70. Indeed, the probability associated with the transmembrane location of residue 12 is intermediate (0.56 according to THMM 2.0).

[0466] The ectodomain of M probably corresponds to residues 2 to 14 of the protein: ADNGTITVEELKQ, SEQ ID NO: 69. Indeed, the N-terminal methionine of M is very probably cleaved from the mature polypep-

ptide because the residue at position 2 is an alanine (Varshavsky, 1996, 93:12142-12149).

[0467] Moreover, the analysis of the hydrophobicity (Kyte & Doolittle, Hopp & Woods) of the E protein demonstrates that the C-terminal end of the endodomain of E is hydrophilic and therefore probably exposed at the surface of this domain. Thus, a synthetic peptide corresponding to this end is a good immunogenic candidate for inducing, in animals, antibodies directed against the endodomain of E. Consequently, a peptide corresponding to 24 C-terminal residues of E was synthesized.

2) Preparation of Antibodies Directed Against the Ectodomain of the M and E Proteins and the Endodomain of the E Protein

[0468] The peptides M2-14 (ADNGTITVEELKQ, SEQ ID NO: 69), E1-12 (MYSFVSEETGTL, SEQ ID NO: 70) and E53-76 (KPTVYVYSRVKNLNSSEGVPDLLV, SEQ ID NO: 71) were synthesized by Neosystem. They were coupled with KLH (Keyhole Limpet Hemocyanin) with the aid of MBS (m-maleimido-benzoyl-N-hydroxysuccinimide ester) via a cysteine added during the synthesis either at the N-terminus of the peptide (case for E53-76) or at the C-terminus (case of M2-14 and E1-12).

[0469] Two rabbits were immunized with each of the conjugates, according to the following immunization protocol: after a first injection of 0.5 mg of peptide coupled with KLH and emulsified in complete Freund's adjuvant (intradermal route), the animals receive 2 to 4 booster injections at 3 or 4 weeks' interval of 0.25 mg of peptide coupled to KLH and emulsified in incomplete Freund's adjuvant.

[0470] For each rabbit, a preimmune (p.i.) serum was prepared before the first immunization and an immune serum (I.S.) is prepared 3 to 5 weeks after the booster injections.

[0471] The reactivity of the sera was analyzed by Western blotting with the aid of extracts of cells infected with SARS-CoV (FIG. 43B) or with the aid of extracts of cells infected with a recombinant vaccinia virus expressing the protein E (VV-TG-E, FIG. 43A) or M (VV-TN-M, FIG. 43C) of the SARS-CoV 031589 isolate.

[0472] The immune sera of the rabbits 22234 and 22240, immunized with the conjugate KLH-E53-76, recognize a polypeptide of about 9 to 10 kD, which is present in the extracts of cells infected with SARS-CoV but absent from the extracts of noninfected cells (FIG. 43B). The apparent mass of this polypeptide is compatible with the predicted mass of the E protein, which is 8.4 kD. Similarly, the immune serum of the rabbit 20047, immunized with the conjugate KLH-E1-12, recognizes a polypeptide present in the extracts of cells infected with the VV-TG-E virus, whose apparent molar mass is compatible with that of the E protein (FIG. 43A).

[0473] The immune serum of the rabbits 20013 and 20080, immunized with the conjugate KLH-M2-14, recognizes a polypeptide present in the extracts of cells infected with the VV-TN-M virus (FIG. 43C), whose apparent molar mass (about 18 kD) is compatible with that of the glycoprotein M, which is 25.1 kD and has a high iso-electric point (9.1 for the naked polypeptide).

[0474] These results demonstrate that the peptides E1-12 and E53-76, on the one hand, and the peptide M2-14, on the other hand, make it possible to induce, in animals, polyclonal antibodies capable of recognizing the native forms of the SARS-CoV E and M proteins, respectively.

EXAMPLE 6

Analysis of the ELISA Reactivity of the Recombinant N Protein Toward Sera from Patients Suffering from SARS

1) Materials

[0475] The antigen used to prepare the solid phases is the purified recombinant nucleoprotein N prepared according to the protocol described in example 2.

[0476] The sera to be tested (table IV) were chosen on the basis of the results of analysis of their reactivity by immunofluorescence (IF-SARS titer), toward cells infected with SARS-CoV.

TABLE IV

Sera tested by ELISA				
Reference	Serum No.	Type of serum	Date of the serum***	IF-SARS titer
3050	A	Control	na*	nt**
3048	B	Control	na	nt
033168	D	Patient 1-SARS	Apr. 27, 2003 (D38)	320
033397	E	Patient-1 SARS	May 11, 2005 (D52)	320
032632	F	Patient-2 SARS	Mar. 21, 2003 (D17)	2500
032791	G	Patient-3 SARS	Apr. 04, 2003 (D3)	<40
033258	H	Patient-3 SARS	Apr. 28, 2003 (D27)	160

*na: not applicable.

**nt: not tested.

***the dates indicated correspond to the number of days after the onset of the SARS symptoms.

2) Method

[0477] The N protein (100 μ l) diluted at various concentrations in 0.1 M carbonate buffer, pH 9.6 (1, 2 or 4 μ g/ml) is distributed into the wells of ELISA plates, and then the plates are incubated overnight at laboratory temperature. The plates are washed with PBS-Tween buffer saturated with PBS-skimmed milk-sucrose (5%) buffer. The test sera (100 μ l), diluted beforehand (1/50, 1/100, 1/200, 1/400, 1/800, 1/1600 and 1/3200) are added and then the plates are incubated for 1 h at 37° C. After 3 washings, the peroxidase-labeled anti-human IgG conjugate (reference 209-035-098, JACKSON) diluted 1/18 000 is added and then the plates are incubated for 1 h at 37° C. After 4 washings, the chromogen (TMB) and the substrate (H₂O₂) are added and the plates are incubated for 30 min at room temperature, protected from light. The reaction is then stopped and then the absorbance at 450 nm is measured with the aid of an automated reader.

3) Results

[0478] The ELISA tests (FIG. 10) demonstrate that the recombinant N protein preparation is specifically recognized by the antibodies of sera from patients suffering from SARS collected in the late phase of the infection (≥ 17 days after the onset of the symptoms) whereas it is not significantly recognized by the antibodies of a patient's serum collected

in the early phase of the infection (3 days after the onset of the symptoms) or by control sera from subjects not suffering from SARS.

EXAMPLE 7

ELISA Tests Prepared for a Very Specific and Sensitive Detection of a SARS-associated Coronavirus Infection, from Sera of Patients

1) Indirect ELISA IgG Test

a) Reagents

Preparation of the Plates

[0479] The plates are sensitized with a solution of N protein at 2 μ g/ml in a 10 mM PBS buffer, pH 7.2, phenol red at 0.25 ml/l. 100 μ l of solution are deposited in the wells and left to incubate at room temperature overnight. Saturation is obtained by prewashing in 10 mM PBS/0.1% Tween buffer, followed by washing with a saturation solution PBS, 25% milk/sucrose.

Diluent Sera

[0480] Buffer 0.48 g/l TRIS, 10 mM PBS, 3.7 g/l EDTA, 15% v/v milk, pH 6.7

Diluent Conjugate

[0481] Citrate buffer (15 g/l), 0.5% Tween, 25% bovine serum, 12% NaCl, 6% v/v skimmed milk pH 6.5

Conjugate

[0482] 50 \times anti-human IgG conjugate, marketed by Bio-Rad: *Platelia H. pylori* kit ref 72778

Other Solutions:

[0483] Washing solution R2, solutions for visualizing with TMB R8 diluent, R9 chromogen, R10 stopping solution: reagents marketed by Bio-Rad (e.g.: *Platelia pylori* kit, ref 72778)

b) Procedure

[0484] Dilute the sera 1/200 in the sample diluent

[0485] Distribute 100 μ l/well

[0486] Incubation 1 h at 37° C.

[0487] 3 washings in 10 \times WASHING solution R2 diluted beforehand 10-fold in demineralized water (i.e., 1 \times washing solution)

[0488] Distribute 100 μ l of conjugate (50 \times conjugate to be diluted immediately before use in the diluent conjugate provided)

[0489] Incubation 1 h at 37° C.

[0490] 4 washings in 1 \times washing solution

[0491] Distribute 200 μ l/well of visualization solution (to be diluted immediately before use e.g.: 1 ml of R9 in 10 ml of R8)

[0492] Incubation for 30 min at room temperature in the dark

[0493] Stop the reaction with 100 μ l/well of R10

[0494] READING at 450/620 nm

[0495] The results can be interpreted by taking a THRESHOLD serum giving a response above which the sera tested would be considered as positive. This serum is chosen and diluted so as to give a significantly higher signal than the background noise.

2) Double Epitope ELISA Test

a) Reagents

Preparation of the Plates

[0496] The plates are sensitized with a solution of N protein at 1 µg/ml in a 10 mM PBS buffer, pH 7.2, phenol red at 0.25 ml/l. 100 µl of solution are deposited in the wells and left to incubate at room temperature overnight. Saturation is obtained by prewashing in 10 mM PBS/0.1% Tween buffer, followed by washing with a saturation solution 10 mM PBS, 25% (V/V) milk.

Diluent Sera and Conjugate

[0497] Buffer 50 mM TRIS saline, pH 8, 2% milk

Conjugate

[0498] This is the purified recombinant N protein coupled with peroxidase according to the Nakane protocol (Nakane P. K. and Kawaoi A.; (1974): *Peroxydase-labeled antibody, a new method of conjugation. The Journal of Histochemistry and Cytochemistry* Vol. 22, N) 23, pp. 1084-1091), in respective molar ratios 1/2. This ProtN POD conjugate is used at a concentration of 2 µg/ml in serum/conjugate diluent.

Other Solutions:

[0499] Washing solution R2, solutions for visualization with TMB R8, diluent, R9 chromogen, R10 stopping solution: reagents marketed by Bio-Rad (e.g. *Platelia pylori* kit, ref 72778).

b) Procedure

1st Step in "Predilution" Plate

[0500] Dilute each serum 1/5 in the predilution plate (48 µl of diluent+12 µl of serum).

[0501] After having diluted all the sera, distribute 60 µl of conjugate.

[0502] Where appropriate, the serum+conjugate mix is left to incubate.

2nd Step in "Reaction" Plate

[0503] Transfer 100 µl of mixture/well into the reaction plate

[0504] Incubation 1 h 37° C.

[0505] 5 washings in 10× WASHING solution R2 diluted 10-fold beforehand in demineralized water (→1× washing solution)

[0506] Distribute 200 µl/well of visualization solution (to be diluted immediately before use e.g.: 1 ml of R9 in 10 ml of R8)

[0507] Incubation 30 min at room temperature and protected from light

[0508] Stop the reaction with 100 µl/well of R10

[0509] READING at 450/620 nm

[0510] Likewise as for the indirect ELISA test, the results can be interpreted using a "threshold value" serum. Any serum having a response greater than the threshold value serum will be considered as positive.

2) Results

[0511] The sera of patients classified as probable cases of SARS from the French hospital of Hanoi, Vietnam or in relation with the French hospital of Hanoi (JYK) were analyzed using the indirect IgG-N test and the double epitope N test.

[0512] The results of the indirect IgG-N test (FIGS. 14 and 15) and double epitope N test (FIGS. 16 and 17) show an excellent correlation between them and with an indirect ELISA test comparing the reactivity of the sera toward a lysate of VeroE6 cells infected or not infected with SARS-CoV (ELISA-SARS-CoV lysate; see table V below). All the sera collected 12 days or more after the onset of the symptoms were found to be positive, including in patients for whom it had not been possible to document the SARS-CoV virus infection by analyzing respiratory samples by RT-PCR, probably because of a sample being collected too late during the infection (\geq D12). In the case of the patient TTH for whom a nasal sample collected on D7 was found to be negative by RT-PCR, the quality of the sample may be in question.

[0513] Some sera were found to be negative whereas the presence of SARS-CoV was detected by RT-PCR. They are in all cases early sera collected less than 10 days after the onset of the symptoms (e.g.: serum # 032637). In the case of a patient PTH (serum # 032673), only a suspicion of SARS was raised at the time the samples were collected.

[0514] In conclusion, the indirect IgG-N and N-double epitope serological tests make it possible to document the SARS-CoV infection in all the patients for the sera collected 12 days or more after the infection.

TABLE V

Results of the ELISA tests						
Sample Num	Patient	Day	PCR-SARS (1)	ELISA SARS-CoV lysate (2)	IgG-N (2nd series)	2Xepitope (2nd series)
033168	JYK	38	POS	+++	>5000	NT
033597	JYK	74	POS	NT	~5000	NT
032552	VTT	8	NEG-D3&D8&D12	NEG	<200	<5
032544	CTP	16	NEG-D16&D20	++	>5000	>>20

TABLE V-continued

Results of the ELISA tests						
Sample Num	Patient	Day	PCR-SARS (1)	ELISA SARS-CoV lysate (2)	IgG-N (2nd series)	2Xepitope (2nd series)
032546	CJF	15	NEG D15&D19	++	>5000	>>20
032548	PTL	17	NEG D17&D21	++	>5000	>>20
032550	NTH	17	NEG-D17&D21	++	>5000	>>20
032553	VTT	8	NEG- D3&D8&D12	NEG	<200	<5
032554	NTBV	4	POS	NEG	<200	<5
032555	NTBV	4	POS	NEG	<200	<5
032564	NTP	15	POS	++	>5000	>>20
032629	NVH	4	POS	NEG	<200	<5
032631	BTIX	9	POS	NEG	<200	<5
032635	NHH	4	POS	NEG	<200	<5
032637	NHB	10	POS	NEG	<200	<5
032642	BTIX	9	POS	NEG	<200	<5
032643	LTDH	1	POS	NEG	<200	<5
032644	NTBV	4	POS	NEG	<200	<5
032646	TTH	12	NEG D7&D12&D16	++	>5000	>>20
032647	DTH	17	NEG D17&D21	++	>5000	>>20
032648	NNT	15	NEG D15&D19	++	>5000	>>20
032649	PTH	17	NEG D17&D21	++	>5000	>>20
032672	LVV	16	NEG D16&D20	+	>5000	>>20
032673	PTTH	NA	NEG	NEG	<200	<5
032674	PNB	17	NEG D17&D21	++	>5000	>>20
032682	VTH	12	NEG D12&D16	++	>5000	>>20
032683	DTV	17	NEG D17&D21	+	>1000	>>20

Remarks:

(1): The RT-PCR analyses were carried out by nested RT-PCR BNI, LC Artus and LC-N on nasal or pharyngeal swabs; POS means that at least one sample was found to be positive in this patient.

(2): The reactivity of the sera in the ELISA test using a lysate of cells infected with SARS-CoV was classified as very highly reactive (+++), highly reactive (++), reactive (+) and negative according to the OD value obtained at the dilutions tested.

EXAMPLE 8

Detection of SARS-associated Coronavirus
(SARS-CoV) by RT-PCR

1) Real Time Development of RT-PCR Conditions with the Aid of Primers Specific for the Gene for the Nucleocapsid Protein—"Light Cycler N" Test

a) Design of the Primers and Probes

[0515] The primers and probes were designed from the sequence of the genome of the SARS-CoV strain derived from the sample recorded under the number 031589, with the aid of the programme "Light Cycler Probe Design (Roche)". Thus, the following two series of primers and probes were selected:

series 1 (SEQ ID NO: 60, 61, 64, 65):

sense primer:

N/+ /28507:

5'-GGC ATC GTA TGG GTT G-3'

[28507-28522]

-continued

antisense primer:

N/- /28774:

5'-CAG TTT CAC CAC CTC C-3'

[28774-28759]

probe 1:

5'-GGC ACC CGC AAT CCT AAT AAC AAT

GC-fluorescein 3'

[28561-28586]

probe 2:

5' Red705-GCC ACC GTG CTA CAA CTT

CCT-phosphate

[28588-28608]

series 2 (SEQ ID NO: 62, 63, 66, 67)

sense primer:

N/+ /28375:

5'-GGC TAC TAC CGA AGA G-3'

[28375-28390]

antisense primer:

N/- /28702:

5'-AAT TAC CGC GAC TAC G-3'

[28702-28687]

-continued

probe 1:
SARS/N/FL:
5'-ATA CAC CCA AAG ACC ACA TTG GC- [28541-28563]
fluorescein 3'

probe 2:
SARS/N/LC705:
5' Red705-CCC GCA ATC CTA ATA ACA ATG [28565-28589]
CTG C-phosphate 3'

b) Analysis of the Efficacy of the Two Primer Pairs

[0516] In order to test the respective efficacy of the two pairs of primers, an RT-PCR amplification was carried out on a synthetic RNA corresponding to nucleotides 28054-29430 of the genome of the SARS-CoV strain derived from the sample recorded under the number 031589 and containing the sequence of the N gene.

[0517] More specifically:

[0518] This synthetic RNA was prepared by in vitro transcription with the aid of the T7 phage RNA polymerase, of a DNA template obtained by linearization of the plasmid SRAS-N with the enzyme Bam H1. After eliminating the DNA template by digestion with the aid of DNase I, the synthetic RNAs are purified by a phenol-chloroform extraction, followed by two successive precipitations in ammonium acetate and isopropanol. They are then quantified by measuring the absorbance at 260 nm and their quality is checked by the ratio of the absorbances at 260 and 280 nm and by agarose gel electrophoresis. Thus, the concentration of the synthetic RNA preparation used for these studies is 1.6 mg/ml, which corresponds to 2.1×10^{15} copies/ml of RNA.

[0519] Decreasing quantities of synthetic RNA were amplified by RT-PCR with the aid of the "Superscript™ One-Step RT-PCR with Platinum® Taq" kit and the pairs of primers No. 1 (N+/28507, N-/28774) (FIG. 1A) and No. 2 (N+/28375, N-/28702) (FIG. 1B), according to the supplier's instructions. The amplification conditions used are the following: the cDNA was synthesized by incubation for 30 min at 45° C., 15 min at 55° C. and then 2 min at 94° C. and it was then amplified by 5 cycles comprising: a step of denaturation at 94° C. for 15 sec, a step of annealing at 45° C. for 30 sec and then a step of extension at 72° C. for 30 sec, followed by 35 cycles comprising: a step of denaturation at 94° C. for 15 sec, a step of annealing at 55° C. for 30 sec and then a step of extension at 72° C. for 30 sec, with 2 sec of additional extension at each cycle, and a final step of extension at 72° C. for 5 min. The amplification products obtained were then kept at 10° C.

[0520] The results presented in FIG. 11 show that the pair of primers No. 2 (N+/28375, N-/28702) makes it possible to detect up to 10 copies of RNA (band of weak intensity) or 10^2 copies (band of good intensity) against 10^4 copies for the pair of primers No. 1 (N+/28507, N-/28774). The amplicons are respectively 268 bp (pair 1) and 328 bp (pair 2).

c) Development of Real Time RT-PCR

[0521] A real time RT-PCR was developed with the aid of the pair of primers No. 2 and of the pair of probes consisting of SRAS/N/FL and SRAS/N/LC705 (FIG. 2).

[0522] The amplification was carried out on a LightCycler™ (Roche) with the aid of the "Light Cycler RNA Amplification Kit Hybridization Probes" kit (reference 2 015 145, Roche) under the following optimized conditions. A reaction mixture containing: H₂O (6.8 µl), 25 mM MgCl₂ (0.8 µl, 4 µM Mg²⁺ final), 5× reaction mixture (4 µl), 3 µM probe SRAS/N/FL (0.5 µl, 0.075 µM final), 3 µM probe SRAS/N/LC705 (0.5 µl, 0.075 µM final), 10 µM primer N+/28375 (1 µl, 0.5 µM final), 10 µM primer N-/28702 (1 µl, 0.5 µM final), enzyme mixture (0.4 µl) and sample (viral RNA, 5 µl) was amplified according to the following program:

Reverse transcription:		
50° C.	10:00 min	analysis mode: none
Denaturation:		
95° C.	30 sec × 1	analysis mode: none
Amplification:		
95° C.	2 sec	} ×45
50° C.	15 sec	
72° C.	13 sec	
Annealing:		
40° C.	30 sec × 1	analysis mode: none

*The fluorescence is measured at the end of the annealing and at each cycle (in SINGLE mode).

[0523] The results presented in FIG. 12 show that this real time RT-PCR is very sensitive since it makes it possible to detect 10^2 copies of synthetic RNA in 100% of the 5 samples analyzed (29/29 samples in 8 experiments) and up to 10 copies of RNA in 100% of the 5 samples analyzed (40/45 samples in 8 experiments). It also shows that this RT-PCR makes it possible to detect the presence of the SARS-CoV genome in a sample and to quantify the number of genomes present. By way of example, the viral RNA of a SARS-CoV stock cultured on Vero E6 cells was extracted with the aid of the "Qiamp viral RNA extraction" kit (Qiagen), diluted to 0.05×10^{-14} and analyzed by real time RT-PCR according to the protocol described above; the analysis presented in FIG. 12 shows that this virus stock contains 6.5×10^9 genome-equivalents/ml (geq/ml), which is entirely similar to the 1.0×10^{10} geq/ml value measured with the aid of the "RealArt™ HPA-Coronavirus LC RT PCR Reagents" kit marketed by Artus.

2) Development of Nested RT-PCR Conditions Targeting the Gene for RNA Polymerase—"CDC (Centers for Disease Control and Prevention)/IP Nested RT-PCR" Test

a) Extraction of the Viral RNA

[0524] Clinical sample: QIamp viral RNA Mini Kit (QIAGEN) according to the manufacturer's instructions, or an equivalent technique. The RNA is eluted in a volume of 60 µl.

b) "SNE/SAR" Nested RT-PCR

First Step: "SNE" Coupled RT-PCR

[0525] The Invitrogen "Superscript™ One-Step RT-PCR with Platinum® Taq" kit was used, but the "Titan" kit from Roche Boehringer can be used in its place with similar results.

[0526] Oligonucleotides:

SNE-S1
5' GGT TGG GAT TAT CCA AAA TGT GA 3'

SNE-AS1
5' GCA TCA TCA GAA AGA ATC ATC ATG 3'

→ Expected size: 440 bp

[0527] 1. Prepare a mix:

H2O	6.5 µl
Reaction mix 2X	12.5 µl
Oligo SNE-S1 50 µM	0.2 µl
Oligo SNE-AS1 50 µM	0.2 µl
RNAsin 40 U/µl	0.12 µl
RT/Platinum Taq mix	0.5 µl

[0528] 2. To 20 µl of the mix, add 5 µl of RNA and carry out the amplification on a thermocycler (ABI 9600 conditions):

2.1	45° C.	30 min.		
	55° C.	15 min.		
	94° C.	2 min.		
2.2.	94° C.	15 sec.	}	×5 cycles
	45° C.	30 sec.		
	72° C.	30 sec.		
2.3.	94° C.	15 sec.	}	×35 cycles
	55° C.	30 sec.		
	72° C.	30 sec. + 2 sec./cycle		
2.4.	72° C.	5 min.		
2.5	10° C.	∞		

Storage at +4° C.

[0529] The RNAsin (N2511/N2515) from Promega was used as RNase inhibitors.

[0530] Synthetic RNAs served as positive control. As the control, 10³, 10² and 10 copies of synthetic RNA_{SNE} were amplified in each experiment.

Second Step: "SAR" Nested PCR

[0531] Oligonucleotides:

SAR1-S
5' CCT CTC TTG TTC TTG CTC GCA 3'

SAR1-AS
5' TAT AGT GAG CCG CCA CAC ATG 3'

→ Expected size: 121 bp

[0532] 1. Prepare a mix:

H2O	35.8 µl
Taq buffer 10X	5 µl
MgCl ₂ 25 mM	4 µl
Mix dNTPs 5 mM	2 µl
Oligo SAR1-S 50 µM	0.5 µl
Oligo SAR1-AS 50 µM	0.5 µl
Taq DNA pol 5 U/µl	0.25 µl

[0533] AmpliTaq DNA Pol from Applied Biosystems was used (10× buffer without MgCl₂, ref 27216601).

[0534] 2. To 48 µl of the mix, add 2 µl of the product from the first PCR and carry out the amplification (ABI 9600 conditions):

2.1.	94° C.	2 min.		
2.2.	94° C.	30 sec.	}	×5 cycles
	45° C.	45 sec.		
	72° C.	30 sec.		
2.3.	94° C.	30 sec.	}	×35 cycles
	55° C.	30 sec.		
	72° C.	30 sec. + 1 sec./cycle		
2.4.	72° C.	5 min.		
2.5	10° C.	∞		

[0535] 3. Analyze 10 µl of the reaction product on "low-melting" gel (Seakem GTG type) containing 3% agarose.

[0536] The sensitivity of the nested test is routinely, under the conditions described, 10 copies of RNA.

[0537] 4. The fragments can then be purified on QIAquick PCR kit (QIAGEN) and sequenced with the oligos SAR1-S and SAR1-AS.

3) Detection of the SARS-CoV RNA by PCR from Respiratory Samples

a) First Comparative Study

[0538] A comparative study was carried out on a series of respiratory samples received by the National Reference Center for the Influenza Virus (Northern region) and likely to contain SARS-CoV. To do this, the RNA was extracted from the samples with the aid of the "QIAMP viral RNA extraction" kit (Qiagen) and analyzed by real time RT-PCR, on the one hand with the aid of the pairs of primers and probes of the No. 2 series under the conditions described above on the one hand, and on the other hand with the aid of the kit "LightCycler SARS-CoV quantification kit" marketed by Roche (reference 03 604 438). The results are summarized in table VI below. They show that 18 of the 26 samples are negative and 5 of the 26 samples are positive for the two kits, while one sample is positive for the Roche kit alone and two for the "series 2" N reagents alone. Additionally, for 3 samples (20032701, 20032712, 20032714) the quantities of RNA detected are markedly higher with the reagents (probes and primers) of the No. 2 series. These results indicate that the "series 2" N primers and probes are more sensitive for the detection of the SARS-CoV genome in biological samples than those of the kit currently available.

TABLE VI

Real time RT-PCR analysis of the RNAs extracted from a series of samples from 5 patients with the aid of the pairs of primers and probes of the No. 2 series ("series 2" N) or of the kit "Lightcycler SARS-CoV quantification kit" (Roche). The type of sample is indicated as well as the number of copies of viral genome measured in each of the two tests. NEG: negative RT-PCR.				
Sample No.	Patient	Type of sample	ROCHE KIT	"Series 2" N
20033082	K	nasal	NEG	NEG
20033083	K	pharyngeal	NEG	NEG
20033086	K	nasal	NEG	NEG
20033087	K	pharyngeal	NEG	NEG
20032802	M	nasal	NEG	NEG
20032803	M	expectoration	NEG	NEG
20032806	M	nasal or pharyngeal	NEG	NEG
20031746ARN2	C	pharyngeal	NEG	NEG
20032711	C	nasal or pharyngeal	39	NEG
20032910	B	nasal	NEG	NEG
20032911	B	pharyngeal	NEG	NEG
20033356	V	expectoration	NEG	NEG
20033357	V	expectoration	NEG	NEG
20031725	K	endotracheal asp.	NEG	150
20032657	K	endotracheal asp.	NEG	NEG
20032698	K	endotracheal asp.	NEG	NEG
20032720	K	endotracheal asp.	3	5
20033074	K	stools	115	257
20032701	M	pharyngeal	443	1676
20032702	M	expectoration	NEG	249
20031747ARN2	C	pharyngeal	NEG	NEG
20032712	C	unknown	634	6914
20032714	C	pharyngeal	17	223
20032800	B	nasal	NEG	NEG
20033353	V	nasal	NEG	NEG
20033384	V	nasal	NEG	NEG

b) Second Comparative Study

[0539] The performance of various nested RT-PCR and real time RT-PCR methods were then compared for 121 respiratory samples from possible cases of SARS at the French hospital in Hanoi, Vietnam, taken between the 4th and the 17th day after the onset of the symptoms. Among these samples, 14 were found to be positive during a first test using the nested RT-PCR method targeting ORF1b (encoding replicase) as described initially by Bernhard Nocht Institute (BNI nested RT-PCR).

[0540] Information relating to this test is available on the internet, at the address <http://www15.bni-hamburg.de/bni2/neu2/getfile.cgi?area=engl&diagnostics&pid=4112>.

[0541] The various tests compared in this study are:

[0542] the quantitative RT-PCR method according to the invention, with the "series 2" N primers and probes described above (LightCycler N column),

[0543] the nested RT-PCR test targeting the RNA polymerase gene described above, developed by the CDC, BNI and Institut Pasteur <CDC/IP nested RT-PCR>),

[0544] the ARTUS kit with the reference "HPA Corona LC RT-PCR Kit # 5601-02", which is a real time RT-PCR test targeting the ORF1b gene,

[0545] the BNI nested RT-PCR test, also targeting the RNA polymerase gene mentioned above.

[0546] The inventors observed:

[0547] 1) an inter-test variability for the same technique, linked to the degradation of the RNA preparation during repeated thawing, in particular for the samples containing the lowest quantities of RNA,

[0548] 2) a reduced sensitivity of the CDC/IP nested RT-PCR compared with the BNI nested RT-PCR, and

[0549] 3) a comparable sensitivity of the quantitative RT-PCR test according to the invention (Lightcycler N) compared with the Artus LightCycler (LC) test.

[0550] These results, which are presented in table VII below, show that the quantitative RT-PCR test according to the invention constitutes an excellent addition—or an alternative—to the tests currently available. Indeed, the SARS-linked coronavirus is an emergent virus which is capable of changing rapidly. In particular, the gene for the RNA polymerase of the SARS-linked coronavirus, which is targeted in most of the tests currently available, can recombine with that of other coronaviruses not linked to SARS. The use of a test targeting this gene exclusively could then lead to the production of false-negatives.

[0551] The quantitative RT-PCR test according to the invention does not target the same genomic region as the ARTUS kit since it targets the gene encoding the N protein. By carrying out a diagnostic test targeting two different genes of the SARS-linked coronavirus, it can therefore be hoped to avoid false-negative type results which could be due to the genetic evolution of the virus.

[0552] Furthermore, it appears particularly advantageous to target the gene for the nucleocapsid protein because it is very stable because of the high selection pressure linked to the high structural constraints regarding this protein.

TABLE VII

Comparison of various methods of analysis by gene amplification, from 121 samples of probable cases of SARS at the French hospital in Hanoi, Vietnam (epidemic 2003)							
NRC No.	Sample type (1)	Sample collection day	Patient	CDC/IP nested RT-PCR	BNI nested RT-PCR	Artus LightCycler kit	LightCycler N (IP)
107 samples	N and P			Negative	Negative	Negative	Negative
032529	P	10	NHB	Negative	Positive	Negative	Negative
032530	N	10	NHB	Positive	Positive	3.10E+01	4.20E+01
032531	P	7	LP	Positive	Positive	7.70E+00	3.10E+00

TABLE VII-continued

Comparison of various methods of analysis by gene amplification, from 121 samples of probable cases of SARS at the French hospital in Hanoi, Vietnam (epidemic 2003)							
NRC No.	Sample type (1)	Sample collection day	Patient	CDC/IP nested RT-PCR	BNI nested RT-PCR	Artus Light Cycler kit	Light Cycler N (IP)
032534	N	15	BND	Positive	Positive	1.60E+00	Negative
032600	P	4	NHH	Negative	Positive	Negative	1.30E+02
032612	P	17	NTS	Negative	Positive	Negative	Negative
032688	P	9	BTX	Positive	Positive	Negative	Negative
032689	N	4	NVH	Positive	Positive	1.20E+01	2.30E+02
032690	P	4	NVH	Negative	Positive	1.60E+00	Negative
032727	P	8	NVH	Positive	Positive	2.30E+02	4.00E+02
032728	N	8	NVH	Positive	Positive	1.10E+03	1.60E+04
032729	P	14	NHB	Positive	Positive	5.90E+00	3.40E+01
032730	N	14	NHB	Positive	Positive	1.30E+02	4.80E+02
032741	P	8	NHH	Positive	Positive	2.10E+02	1.30E+02
	positives			10	14	10	9
	fraction detected from the 14 positives			71.4%	100.0%	71.4%	64.3%

(1) P = pharyngeal swab N = nasal swab

EXAMPLE 9

Production and Characterization of Monoclonal Antibodies Directed Against the N Protein

[0553] Balb C mice were immunized with the purified recombinant N protein and their spleen cells fused with an appropriate murine myeloma according to the Köhler and Milstein techniques.

[0554] Nineteen anti-N antibody secreting hybridomas were preselected and their immunoreactivities determined. These antibodies do indeed recognize the recombinant N protein (in ELISA) with variable intensities, and the natural viral N protein in ELISA and/or in Western blotting. FIGS. 18 to 20 show the results of these tests for 15 of these 19 monoclonal antibodies.

[0555] The highly reactive clones 12, 17, 28, 57, 72, 76, 86, 87, 98, 103, 146, 156, 166, 170, 199, 212, 218, 219 and 222 were subcloned. Specificity studies were carried out with the appropriate tools in order to determine the epitopes recognized and verify the absence of reactivity toward other human coronaviruses and certain respiratory viruses.

[0556] Epitope mapping studies (performed on spot membrane with the aid of overlapping peptides of 15 aa) and additional studies performed on the natural N protein in Western blotting revealed the existence of 4 groups of monoclonal antibodies:

[0557] 1. Monoclonal antibodies specific for a major linear epitope at the N-ter position (75-81, sequence: INTNSVP).

[0558] The representative of this group is antibody 156. The hybridoma producing this antibody was deposited at the Collection Nationale de Cultures de Microorganismes (CNCM) of the Institut Pasteur (Paris, France) on Dec. 1, 2004, under the number I-3331. This same epitope is also recognized by a rabbit serum (anti-N polyclonal) obtained by conventional immunization with the aid of this same N protein.

[0559] 2. Monoclonal antibodies specific for a major linear epitope located in a central position (position 217-224, sequence: ETALALL); the representatives of this group are the monoclonal antibodies 87 and 166. The hybridoma producing antibody 87 was deposited at the CNCM on Dec. 1, 2004, under the number I-3328.

[0560] 3. Monoclonal antibodies specific for a major linear epitope located at the C-terminal position (position 403-408, sequence: DFFRQL), the representatives of this group are the antibodies 28, 57 and 143. The hybridoma producing antibody 57 was deposited at the CNCM on Dec. 1, 2004, under the number I-3330.

[0561] 4. Monoclonal antibodies specific for a discontinuous conformational epitope. This group of antibodies does not recognize any of the peptides spanning the sequence of the N protein, but react strongly on the non-denatured natural protein. The representative of this final group is the antibody 86. The hybridoma producing this antibody was deposited at the CNCM on Dec. 1, 2004, under the number I-3329.

[0562] Table VIII below summarizes the epitope mapping results obtained:

TABLE VIII

Epitope mapping of the monoclonal antibodies			
Antibody	Epitope	Position	Region
28	DFSRQL Q	403 . . . 408	C-Ter.
143	DFSRQL Q		
76	DFSRQL Q		
57	DFSRQL Q		
	FFGMS RI	315 . . . 319	
146	LPQRQ	383 . . . 387	
166	ETALALLL	217 . . . 224	central
87	ETALALL	217 . . . 224	
156	INTNSGP	75 . . . 81	N-Ter.
86	Conformational		
212	Conformational		
170	Conformational		

EXAMPLE 10

Combinations of the Monoclonal Antibodies for the Development of a Sensitive Immunocapture Test Specific for the Viral N Antigen in the Serum or Biological Fluids of Patients Infected with the SARS-CoV Virus

[0563] The antibodies listed below were selected because of their very specific properties for an additional capture and detection study of the viral N protein, in the serum of the subjects or patients.

[0564] These antibodies were produced in ascites on mice, purified by affinity chromatography and used alone or in combination, as capture antibodies and as signal antibodies.

[0565] List of the antibodies selected:

[0566] Ab anti-C-ter region (No. 28, 57, 143)

[0567] Ab anti-central region (No. 87, 166)

[0568] Ab anti-N-ter region (No. 15-6)

[0569] Ab anti-discontinuous conformational epitope 486)

1) Preparation of the Reagents:

a) Immunocapture ELISA plates

[0570] The plates are sensitized with the antibody solutions at 5 µg/ml in 0.1 M carbonate buffer, pH 9.6. The (monovalent or plurivalent) solutions are deposited in a volume of 100 µl in the wells and incubated overnight at room temperature. These plates are then washed with PBS buffer (10 mM pH 7.4 supplemented with 0.1% Tween 20) and then saturated with a PBS solution supplemented with 0.3% BSA and 5% sucrose). The plates are then dried and then packaged in a bag in the presence of a desiccant. They are ready to use.

b) Conjugates

[0571] The purified antibodies were coupled with peroxidase according to the Nakane protocol (Nakane et al.—1974, J. of Histo and cytochemistry, vol. 22, pp. 1084-1091) in a ratio of one molecule of IgG per 3 molecules of peroxidase. These conjugates were purified by exclusion chromatography and stored concentrated (concentration between 1 and 2 mg/ml) in the presence of 50% glycerol and at -20° C. They are diluted for their use in the assays at the final concentration of 1 or 2 µg/ml in PBS buffer (pH 7.4) supplemented with 1% BSA.

c) Other Reagents

[0572] Human sera negative for all the serum markers for the HIV, HBV, HCV and THLV viruses

[0573] Pool of negative human sera supplemented with 0.5% Triton X 100

[0574] Inactivated viral Ag: viral culture supernatant inactivated by irradiation and inactivation verified after placing in culture on sensitive cells—titer of the suspension before inactivation about 10⁷ infectious particles per ml or alternatively about 5×10⁹ physical viral particles per ml of antigen

[0575] The Ag samples diluted in negative human serum: these samples were prepared by diluting 1:100 and then by 5-fold serial dilution.

[0576] These noninfectious samples mimic human samples thought to contain low to very low concentrations of viral nucleoprotein N. Such samples are not available for routine work.

[0577] Washing solution R2, solution for visualization TMB R8, chromogen R9 and stop solution R10, are the generic reagents marketed by Bio-Rad in its ELISA kits (e.g.: *Platelia pylori* kit ref. 72778).

2) Procedure

[0578] The samples of human sera overloaded with inactivated viral Ag are distributed in an amount of 100 µl per well, directly in the ready-to-use sensitized plates, and then incubated for 1 hour at 37° C. (Bio-Rad IPS incubation).

[0579] The material not bound to the solid phase is removed by 3 washings (washing with dilute R2 solution, automatic LP 35 washer).

[0580] The appropriate conjugates, diluted to the final concentration of 1 or 2 µg/ml, are distributed in an amount of 100 µl per well and the plates are again incubated for one hour at 37° C. (IPS incubation).

[0581] The excess conjugate is removed by 4 successive washings (dilute R2 solution—LP 35 washer).

[0582] The presence of conjugate attached to the plates is visualized after adding 100 µl of visualization solution prepared before use (1 ml of R9 and 10 ml of R8) and after incubation for 30 minutes, at room temperature and protected from light.

[0583] The enzymatic reaction is finally blocked by adding 100 µl of R10 reagent (1 N H₂SO₄) to all the wells.

[0584] The reading is carried out with the aid of an appropriate microplate reader at double wavelength (450/620 nm).

[0585] The results can be interpreted by using, as provisional threshold value, the mean of at least two negative controls multiplied by a factor of 2 or alternatively the mean of 100 negative sera supplemented with an increment corresponding to 6 SD (standard deviation calculated on the 100 individual measurements).

3) Results

[0586] Various capture antibody and signal antibody combinations were tested based on the properties of the antibodies selected, and avoiding the combinations of antibodies specific for the same epitopes in solid phase and as conjugates.

[0587] The best results were obtained with the 4 combinations listed below. These results are reproduced in table IX below.

1. Combination F/28

[0588] Solid phase (Ab 166+87 central region): conjugate antibody 28 (C-ter)

2. Combination G/28

[0589] Solid phase (Ab 86—conformational epitope): conjugate antibody 28 (C-ter)

3. Combination H/28

[0590] Solid phase (Ab 86, 166 and 87 central region and conformational epitope): conjugate antibody 28 (C-ter)

4. Combination H/28+87

[0591] Solid phase (Ab 86, 166 and 87 central region and conformational epitope): mixed conjugate antibodies 28 (C-ter) and 87 (central)

5. Combination G/87

[0592] Solid phase (Ab 86—conformational epitope): conjugate antibody 87 (central region)

[0593] The first 4 combinations exhibit equivalent and reproduced performance levels, greater than the other combinations used (such as for example the combination G/87). Of course, in these combinations, a monoclonal antibody may be replaced with another antibody recognizing the same epitope. Thus, the following variants may be mentioned:

6. Variant of the combination F/28

[0594] Solid phase (Ab 87 only): conjugate antibody 57 (C-ter)

7. Variant of the combination G/28

[0595] Solid phase (Ab 86—conformational epitope): conjugate antibody 57 (C-ter)

8. Variant of the combination H/28

[0596] Solid phase (Ab 86 and 87 central region and conformational epitope): conjugate antibody 57 (C-ter)

9. Variant of the combination H/28+87

[0597] Solid phase (Ab 86 and 87 central region and conformational epitope): mixed conjugate antibodies 57 (C-ter) and 87 (central)

TABLE IX

Test of immunoreactivity of the anti-SARS-CoV nucleoprotein Abs: optical densities measured with each combination of antibodies according to the dilutions of the inactivated viral antigen.						
No.	Dilution	F/28	G/28	G/87	H/28	H/28 + 87
0	1/100	5	5	3.495	3.900	5
1	1/500	3.795	3.814	1.379	3.702	3.804
2	1/2 500	2.815	2.950	0.275	3.268	2.680
3	1/12 500	0.987	1.038	0.135	1.374	0.865
4	1/62 500	0.404	0.348	0.125	0.480	0.328
5	1/312 500	0.285	0.211	0.123	0.240	0.215
6	Control	0.210	0.200	0.098	0.186	0.156
7	Control	0.269	0.153	0.104	0.193	0.202

[0598] The detection limit for these 4 experimental trials corresponds to the antigen dilution in negative serum 1:62 500. A rapid extrapolation suggests the detection of less than 10^3 infectious particles per ml of sera.

[0599] From this study, it is evident that the most appropriate antibodies for the capture of the native viral nucleoprotein are the antibodies specific for the central region and/or for a conformational epitope, both being antibodies also selected for their high affinity for the native antigen.

[0600] Having determined the best antibodies for the composition of the solid phase, the antibodies to be selected as a priority for the detection of the antigens attached to the solid phase are the complementary antibodies specific for a dominant epitope in the C-ter region. The use of any other complementary antibody specific for epitopes located in the N-ter region of the protein leads to average or poor results.

EXAMPLE 11

Eukaryotic Expression Systems for the SARS-associated Coronavirus (SARS-CoV) Spicule (S) Protein

1) Optimization of the Conditions for Expression of the SARS-CoV S in Mammalian Cells

[0601] The conditions for transient expression of the SARS-CoV spicule (S) protein were optimized in mammalian cells (293T, VeroE6).

[0602] For that, a DNA fragment containing the cDNA for SARS-CoV S was amplified by PCR with the aid of the oligonucleotides 5'-ATAGGATCCA CCAIGTTTAT TTTCTTATTA TTTCTTACTC TCACT-3' and 5'-ATACTC-GAGTT ATGTGTAATG TAATTTGACA CCCTTG-3' from the plasmid pSARS-S (C.N.C.M. No. I-3059) and then inserted between the BamH1 and Xho1 sites of the plasmid pTRIPAU3-CMV containing a lentiviral vector TRIP (Sirven, 2001, Mol. Ther., 3, 438-448) in order to obtain the plasmid pTRIP-S. The BamH1 and Xho1 fragment containing the cDNA for S was then subcloned between BamH1 and Xho1 of the eukaryotic expression plasmid pcDNA3.1(+) (Clontech) in order to obtain the plasmid pcDNA-S. The Nhe1 and Xho1 fragment containing the cDNA for S was then subcloned between the corresponding sites of the expression plasmid pCI (Promega) in order to obtain the plasmid pCI-S. The WPRE sequences of the woodchuck hepatitis virus ("Woodchuck Hepatitis Virus posttranscriptional regulatory element") and the CTE sequences ("constitutive transport element") of the simian retrovirus from Mason-Pfizer were inserted into each of the two plasmids pcDNA-S and pCI-S between the Xho1 and Xba1 sites in order to obtain respectively the plasmids pcDNA-S-CTE, pcDNA-S-WPRE, pCI-S-CTE and pCI-S-WPRE (FIG. 21). The plasmid pCI-S-WPRE was deposited at the CNM, on Nov. 22, 2004, under the number I-3323. All the inserts were sequenced with the aid of a BigDye Terminator v1.1 kit (Applied Biosystems) and an automated sequencer ABI377.

[0603] The capacity of the plasmid constructs to direct the expression of SARS-CoV S in mammalian cells was assessed after transfection of VeroE6 cells (FIG. 22). In this experiment, monolayers of 5×10^5 VeroE6 cells in 35 mm Petri dishes were transfected with 2 μ g of plasmids pcDNA (as control), pcDNA-S, pCI and pCI-S and 6 μ l of Fugene6 reagent according to the manufacturer's instructions (Roche). After 48 hours of incubation at 37° C. and under 5% CO₂, cellular extracts were prepared in loading buffer according to Laemmli, separated on 8% SDS polyacrylamide gel, and then transferred onto a PVDF membrane (BioRad). The detection of this immunoblot (Western blot) was carried out with the aid of an anti-S rabbit polyclonal serum (immune serum from the rabbit P11135: cf. example 4 above) and donkey polyclonal antibodies directed against rabbit IgGs and coupled with peroxidase (NA934V, Amersham). The bound antibodies were visualized by luminescence with the aid of the ECL+ kit (Amersham) and autoradiography films Hyperfilm MP (Amersham).

[0604] This experiment (FIG. 22) shows that the plasmid pcDNA-S does not make it possible to direct the expression of SARS-CoV S at detectable levels whereas the plasmid pCI-S allows a weak expression, close to the limit of detection, which may be detected when the film is overexposed. Similar results were obtained when the expression of S was sought by immunofluorescence (data not shown). This impossibility to detect effective expression of S cannot be

attributed to the detection techniques used since the S protein can be detected at the expected size (180 kDa) in an extract of cells infected with SARS-CoV or in an extract of VeroE6 cells infected with the recombinant vaccinia virus VV-TF7.3 and transfected with the plasmid pcDNA-S. In this latter experiment, the virus VV-TF7.3 expresses the RNA polymerase of the T7 phage and allows the cytoplasmic transcription of an uncapped RNA capable of being efficiently translated. This experiment suggests that the expression defects described above are due to an intrinsic inability of the cDNA for S to be efficiently expressed when the step for transcription to messenger RNA is carried out at the nuclear level.

[0605] In a second experiment, the effect of the CTE and WPRE signals on the expression of S was assessed after transfection of VeroE6 (FIG. 23A) and 293T (FIG. 23B) cells and according to a protocol similar to that described above. Whereas the expression of S cannot be detected after transfection of the plasmids pcDNA-S-CTE and pcDNA-S-WPRE derived from pcDNA-S, the insertion of the WPRE and CTE signals greatly improves the expression of S in the context of the expression plasmid pCI-S.

[0606] To specify this result, a second series of experiments were carried out where the immunoblot is quantitatively visualized by luminescence and acquisition on a digital imaging device (FluorS, BioRad). The analysis of the results obtained with the QuantityOne v4.2.3 software (BioRad) shows that the WPRE and CTE sequences increase respectively the expression of S by a factor of 20 to 42 and 10 to 26 in Vero E6 cells (table X). In 293T cells (table X), the effect of the CTE sequence is more moderate (4 to 5 times) whereas that of the WPRE sequence remains high (13 to 28 times).

TABLE X

Plasmid	cell	exp. 1	exp. 2
PCI	VeroE6	0.0	0.0
pCI-S	VeroE6	1.0 ± 0.1	1.0
pCI-S-CTE	VeroE6	9.8 ± 0.9	26.4
pCI-S-WPRE	VeroE6	20.1 ± 2.0	42.3
PCI	293T	0.0	0.0
pCI-S	293T	1.0	1.0
pCI-S-CTE	293T	4.6	4.0
pCI-S-WPRE	293T	27.6	12.8

[0607] In summary, all these results show that the expression, in mammalian cells, of the cDNA for the SARS-CoV S under the control of the RNA polymerase II promoter

sequences requires, to be efficient, the expression of a splice signal and of either of the sequences WPRE and CTE.

2) Production of Stable Lines Allowing the Expression of SARS-CoV S

[0608] The cDNA for the SARS-CoV S protein was cloned in the form of a BamHI-XhoI fragment into the plasmid pTRIPΔU3-CMV containing a defective lentiviral vector TRIP with central DNA flap (Sirven et al., 2001, Mol. Ther., 3: 438-448) in order to obtain the plasmid pTRIP-S (FIG. 24). Transient cotransfection according to Zennou et al. (2000, Cell, 101: 173-185) of this plasmid, of an encapsidation plasmid (p8.2) and of a plasmid for expression of the VSV envelope glycoprotein G (pHCMV-G) in 293T cells allowed the preparation of retroviral pseudoparticles containing the vector TRIP-S and pseudotyped with the envelope protein G. These pseudotyped TRIP-S vectors were used to transduce 293T and FRhK-4 cells: no expression of the S protein could be detected by Western blotting and immunofluorescence in the transduced cells (data not presented).

[0609] The optimum expression cassettes consisting of the CMV virus immediate/early promoter, a splice signal, cDNA for S and either of the posttranscriptional signals WPRE or CTE described above were then substituted for the EF1α-EGFP cassette of the defective lentiviral expression vector with central DNA flap TRIPΔU3-EF1α (Sirven et al., 2001, Mol. Ther., 3: 438-448) (FIG. 25). These substitutions were carried out by a series of successive subclonings of the S expression cassettes which were excised from the plasmids PCT-S-CTE (BglII-ApaI) or respectively pCI-S-WPRE (BglII-SalI) and then inserted between the MluI and KpnI sites or respectively MluI or XhoI sites of the plasmid TRIPΔU3-EF1α in order to obtain the plasmids pTRIP-SD/SA-S-CTE and pTRIP-SD/SA-S-WPRE, deposited at the CNM, on Dec. 1, 2004, under the numbers I-3336 and I-3334, respectively. Pseudotyped vectors were produced according to Zennou et al. (2000, Cell, 101: 173-185) and used to transduce 293T cells (10 000 cells) and FRhK-4 cells (15 000 cells) according to a series of 5 successive transduction cycles with a quantity of vectors corresponding to 25 ng (TRIP-SD/SA-S-CTE) or 22 ng (TRIP-SD/SA-S-WPRE) of p24 per cycle.

[0610] The transduced cells were cloned by limiting dilution and a series of clones were qualitatively analyzed for the expression of SARS-CoV S by immunofluorescence (data not shown), and then quantitatively by Western blotting (FIG. 25) with the aid of an anti-S rabbit polyclonal serum. The results presented in FIG. 25 show that clones 2 and 15 of FRhK4-s-CTE cells transduced with TRIP-SD/SA-S-CTE and clones 4, 9 and 12 of FRhK4-S-WPRE cells transduced with TRIP-SD/SA-S-WPRE allow the expression of the SARS-CoV S at respectively low or moderate levels if they are compared to those which can be observed during infection with SARS-CoV.

[0611] In summary, the vectors TRIP-SD/SA-S-CTE and TRIP-SD/SA-S-WPRE allow the production of stable clones of FRhK-4 cells and similarly 293T cells expressing SARS-CoV S, whereas the assays carried out with the "parent" vector TRIP-S remained unsuccessful, which demonstrates the need for a splice signal and for either of the sequences CTE and WPRE for the production of stable cell clones expressing the S protein.

[0612] In addition, these modifications of the vector TRIP (insertion of a splice signal and of a post-transcriptional signal like CTE and WPRE) could prove advantageous for improving the expression of other cDNAs than that for S.

3) Production of Stable Lines Allowing the Expression of a Soluble Form of SARS-CoV S. Purification of this Recombinant Antigen.

[0613] A cDNA encoding a soluble form of the S protein (Ssol) was obtained by fusing the sequences encoding the ectodomain of the protein (amino acids 1 to 1193) with those of a tag (FLAG: DYKDDDDK) via a BspE1 linker encoding the SG dipeptide. Practically, in order to obtain the plasmid pcDNA-Ssol, a DNA fragment encoding the ectodomain of SARS-CoV S was amplified by PCR with the aid of the oligonucleotides 5'-ATAGGATCCA CCAATGTTTAT TTTCTTATTA TTTCTTACTC TCACT-3' and 5'-ACCTC-CGGAT TTAATATATT GCTCATATTT TCCCAA-3' from the plasmid pcDNA-S, and then inserted between the unique BamH1 and BspE1 sites of a modified eukaryotic expression plasmid pcDNA3.1(+) (Clontech) containing the tag sequence FLAG between its BamH1 and Xho1 sites:

```
// GGATCC . . . nnn . . . TCC GGA GAT TAT AAA GAT
   BamH1                S G D Y K D
```

```
GAC GAC GAT AAA TAA CTCGAG //
   D D D K ter Xho1
```

[0614] The Nhe1-Xho1 and BamH1-Xho1 fragments, containing the cDNA for S, were then excised from the plasmid pcDNA-Ssol, and subcloned between the corresponding sites of the plasmid pTRIP-SD/SA-S-CTE and of the plasmid pTRIP-SD/SA-S-WPRE, respectively, in order to obtain the plasmids pTRIP-SD/SA-Ssol-CTE and pTRIP-SD/SA-Ssol-WPRE, deposited at the CNCM, on Dec. 1, 2004, under the numbers I-3337 and I-3335, respectively.

[0615] Pseudotyped vectors were produced according to Zennou et al. (2000, Cell, 101:173-185) and used to transduce FRhK-4 cells (15 000 cells) according to a series of 5 successive transduction cycles (15 000 cells) with a quantity of vector corresponding to 24 ng (TRIP-SD/SA-Ssol-CTE) or 40 ng (TRIP-SD/SA-Ssol-WPRE) of p24 per cycle. The transduced cells were cloned by limiting dilution and a series of 16 clones transduced with TRIP-SD/SA-Ssol-CTE and of 15 clones with TRIP-SD/SA-Ssol-WPRE were analyzed for the expression of the Ssol polypeptide by Western blotting visualized with an anti-FLAG monoclonal antibody (FIG. 26 and data not presented), and by capture ELISA specific for the Ssol polypeptide which was developed for this purpose (table XI and data not presented). Part of the process for selecting the best secretory clones is shown in FIG. 26. Capture ELISA is based on the use of solid phases coated with polyclonal antibodies of rabbits immunized with purified and inactivated SARS-CoV. These solid phases allow the capture of the Ssol polypeptide secreted into the cellular supernatants, whose presence is then visualized with a series of steps successively involving the attachment of an anti-FLAG monoclonal antibody (M2, SIGMA), of anti-mouse IgG(H+L) biotinylated rabbit polyclonal antibodies (Jackson) and of a streptavidin-peroxidase conjugate (Amersham) and then the addition of chromogen and substrate (TMB+H₂O₂, KPL)

TABLE XI

Vector	Clone	OD (450 nm)
Control	—	0.031
TRIP-SD/SA-Ssol-	CTE2	0.547
CTE	CTE3	0.668
	CTE9	0.171
	CTE12	0.208
	CTE13	0.133
TRIP-SD/SA-Ssol-	WPRE1	0.061
WPRE	WPRE10	0.134

[0616] The cell line secreting the highest quantities of Ssol polypeptide in the culture supernatant is the FRhK4-Ssol-CTE3 line. It was subjected to a second series of 5 cycles of transduction with the vector TRIP-SD/SA-Ssol-CTE under conditions similar to those described above and then cloned. The subclone secreting the highest quantities of Ssol was selected by a combination of Western blot and capture ELISA analysis: it is the subclone FRhK4-Ssol-30, which was deposited at the CNCM, on Nov. 22, 2004, under the name I-3325.

[0617] The FRhK4-Ssol-30 line allows the quantitative production and purification of the recombinant Ssol polypeptide. In a typical experiment where the experimental conditions for growth, production and purification were optimized, the cells of the FRhK4-Ssol-30 line are inoculated in standard culture medium (pyruvate-free DMEM containing 4.5 g/l of glucose and supplemented with 5% FCS, 100 U/ml of penicillin and 100 µg/ml of streptomycin) in the form of a subconfluent monolayer (1 million cells per each 100 cm² in 20 ml of medium). At confluence, the standard medium is replaced with the secretion medium where the quantity of FCS is reduced to 0.5% and the quantity of medium reduced to 16 ml per each 100 cm². The culture supernatant is removed after 4 to 5 days of incubation at 35° C. and under 5% CO₂. The recombinant polypeptide Ssol is purified from the supernatant by the succession of steps of filtration on 0.1 µm polyethersulfone (PES) membrane, concentration by ultrafiltration on a PES membrane with a 50 kD cut-off, affinity chromatography on anti-FLAG matrix with elution with a solution of FLAG peptide (DYKDDDDK) at 100 µg/ml in TBS (50 mM tris, pH 7.4, 150 mM NaCl) and then gel filtration chromatography in TBS on sephadex G-75 beads (Pharmacia). The concentration of the purified recombinant Ssol polypeptide was determined by micro-BCA test (Pierce) and then its biochemical characteristics analyzed.

[0618] Analysis by 8% SDS acrylamide gel stained with silver nitrate demonstrates a predominant polypeptide whose molecular mass is about 180 kD and whose degree of purity may be evaluated at 98% (FIG. 27A). Two main peaks are detected by SELDI-TOF mass spectrometry (Cypher-ger): they correspond to single and double charged forms of a predominant polypeptide whose molecular mass is thus determined at 182.6±3.7 kD (FIGS. 27B and C). After transfer onto Prosorb membrane and rinsing in 0.1% TFA,

the N-terminal end of the Ssol polypeptide was sequenced in liquid phase by Edman degradation on 5 residues (ABI494, Applied Biosystems) and determined as being SDLDR (FIG. 27D). This demonstrates that the signal peptide located at the N-terminal end of the SARS-CoV S protein, composed of aa 1 to 13 (MFIFLLFLTLTSG) according to an analysis carried out with the software signalP v2.0 (Nielsen et al., 1997, *Protein Engineering*, 10:1-6), is cleaved from the mature Ssol polypeptide. The recombinant Ssol polypeptide therefore consists of amino acids 14 to 1193 of the SARS-CoV S protein fused at the C-terminals with a sequence SGDYKDDDDK containing the sequence of the FLAG tag (underlined). The difference between the theoretical molar mass of the naked Ssol polypeptide (132.0 kD) and the real molar mass of the mature polypeptide (182.6 kD) suggests that the Ssol polypeptide is glycosylated.

[0619] A preparation of purified Ssol polypeptide, whose protein concentration was determined by micro-BCA test, makes it possible to prepare a calibration series in order to measure, with the aid of the capture ELISA test described above, the concentrations of Ssol present in the culture supernatants and to review the characteristics of the secretory lines. According to this test, the FRhK4-Ssol-CT3 line secretes 4 to 6 µg/ml of polypeptide Ssol while the FRhK4-Ssol-30 line secretes 9 to 13 µg/ml of Ssol after 4 to 5 days of culture at confluence. In addition, the purification scheme presented above makes it possible routinely to purify from 1 to 2 mg of Ssol polypeptide per liter of culture supernatant.

EXAMPLE 12

Gene Immunization Involving the SARS-associated Corona Virus (SARS-CoV) Spicule (S) Protein

[0620] The effect of a splice signal and of the posttranscriptional signals WPRE and CTE was analyzed after gene immunization of BALB/c mice (FIG. 28).

[0621] For that, BALB/c mice were immunized at intervals of 4 weeks by injecting into the tibialis anterior a saline solution of 50 µg of plasmid DNA of pcDNA-S and pCI-S and, as a control, 50 µg of plasmid DNA of pcDNA-N (directing the expression of SARS-CoV N) or of pCI-HA (directing the expression of the HA of the influenza virus A/PR/8/34) and the immune sera collected 3 weeks after the 2nd injection. The presence of antibodies directed against the SARS-CoV S was assessed by indirect ELISA using as antigen a lysate of VeroE6 cells infected with SARS-CoV and, as a control, a lysate of noninfected VeroE6 cells. The anti-SARS-CoV antibody titers (TI) are calculated as the reciprocal of the dilution producing a specific OD of 0.5 (difference between OD measured on a lysate of infected cells and OD measured on a lysate of noninfected cells) after visualization with an anti-mouse IgG polyclonal antibody coupled with peroxidase (NA931V, Amersham) and TMB supplemented with H₂O₂ (KPL) (FIG. 28A).

[0622] Under these conditions, the expression plasmid pcDNA-S only allows the induction of low antibody titers directed against SARS-CoV S in 3 mice out of 6 (LOG₁₀(TI)=1.9±0.6) whereas the plasmid pcDNA-N allows the induction of anti-N antibodies at high titers (LOG₁₀(TI)=3.9±0.3) in all the animals, and the control plasmids (pCI, pCI-HA) do not result in any detectable antibody (LOG₁₀(TI)<1.7). The plasmid pCI-S equipped with a splice signal allows the induction of antibodies at high titers (LOG₁₀(TI)=3.7±0.2), which are approximately 60 times higher than those observed after injection of the plasmid pcDNA-S (p<10⁻⁵).

[0623] The efficiency of the posttranscriptional signals was studied by carrying out a dose-response study of the anti-S antibody titers induced in the BALB/c mouse as a function of the quantity of plasmid DNA used as immunogen (2 µg, 10 µg and 50 µg). This study (FIG. 28B) demonstrates that the posttranscriptional signal WPRE greatly improves the efficiency of gene immunization when small doses of DNA are used (p<10⁻⁵ for a dose of 2 µg of DNA and p<10⁻² for a dose of 10 µg), whereas the effect of the CTE signal remains marginal (p=0.34 for a dose of 2 µg of DNA).

[0624] Finally, the antibodies induced in mice after gene immunization neutralize the infectivity of SARS-CoV in vitro (FIGS. 29A and 29B) at titers which are consistent with the titers measured by ELISA.

[0625] In summary, the use of a splice signal and of the posttranscriptional signal WPRE of the woodchuck hepatitis virus considerably improves the induction of neutralizing antibodies directed against SARS-CoV after gene immunization with the aid of plasmid DNA directing the expression of the cDNA for SARS-CoV S.

EXAMPLE 13

Diagnostic Applications of the S Protein

[0626] The ELISA reactivity of the recombinant Ssol polypeptide was analyzed with respect to sera from patients suffering from SARS.

[0627] The sera from probable cases of SARS tested were chosen on the basis of the results (positive or negative) of analysis of their specific reactivity toward the native antigens of SARS-CoV by immunofluorescence test on VeroE6 cells infected with SARS-CoV and/or by indirect ELISA test using as antigen a lysate of VeroE6 cells infected with SARS-CoV. The sera of these patients are identified by a serial number of the National Reference Center for Influenza Viruses and by the initials of the patient and the number of days elapsed since the onset of the symptoms. All the sera of probable cases (cf. Table XII) recognize the native antigens of SARS-CoV, with the exception of the serum 032552 of the patient VTT for whom infection with SARS-CoV could not be confirmed by RT-PCR performed on respiratory samples of days 3, 8 and 12. A panel of control sera was used as control (TV sera): they are sera collected in France before the SARS epidemic that occurred in 2003.

TABLE XII

Sera of probable cases of SARS		
Serum	Patient	Sample collection day
031724	JYK	7
033168	JYK	38
033597	JYK	74
032632	NTM	17
032634	THA	15
032541	PHV	10
032542	NIH	17
032552	VTT	8
032633	PTU	16
032791	JLB	3
033258	JLB	27
032703	JCM	8
033153	JCM	29

[0628] Solid phases sensitized with the recombinant Ssol polypeptide were prepared by adsorption of a solution of purified Ssol polypeptide at 2 µg/ml in PBS in the wells of an ELISA plate, and then the plates are incubated overnight at 4° C. and washed with PBS-Tween buffer (PBS, 0.1% Tween 20). After saturating the ELISA plates with a solution of PBS-10% skimmed milk (weight/volume) and washing in PBS-Tween, the sera to be tested (100 µl) are diluted 1/400 in PBS skimmed milk-Tween buffer (PBS, 3% skimmed milk, 0.1% Tween) and then added to the wells of the sensitized ELISA plate. The plates are incubated for 1 h at 37° C. After 3 washings with PBS-Tween buffer, the anti-human IgG conjugate labeled with peroxidase (ref. NA933V, Amersham) diluted 1/4000 in PBS-skimmed milk-Tween buffer is added, and then the plates are incubated for 1 hour at 37° C. After 6 washings with PBS-Tween buffer, the chromogen (TMB) and the substrate (H₂O₂) are added and the plates are incubated for 10 minutes protected from light. The reaction is stopped by adding a 1 N H₂PO₄ solution, and then the absorbance is measured at 450 nm with a reference at 620 nm.

[0629] The ELISA tests (FIG. 30) demonstrate that the recombinant Ssol polypeptide is specifically recognized by the serum antibodies of patients suffering from SARS collected at the medium or late phase of infection (≥10 days after the onset of the symptoms) whereas it is not significantly recognized by the serum antibodies of 2 patients (JLB and JCM) collected in the early phase of infection (3 to 8 days after the onset of the symptoms) or by control sera of subjects not suffering from SARS. The serum antibodies of patients JLB and JCM show a seroconversion between days 3 and 27 for the first and 8 and 29 for the second after the onset of the symptoms, which confirms the specificity of the reactivity of these sera toward the Ssol polypeptide.

[0630] In conclusion, these results demonstrate that the recombinant Ssol polypeptide may be used as an antigen for the development of an ELISA test for serological diagnosis of infection with SARS-CoV.

EXAMPLE 14

Vaccine Applications of the Recombinant Soluble S Protein

[0631] The immunogenicity of the recombinant Ssol polypeptide was studied in mice.

[0632] For that, a group of 6 mice was immunized at 3 weeks' interval with 10 µg of recombinant Ssol polypeptide adjuvanted with 1 mg of aluminum hydroxide (Alu-gel-S, Serva) diluted in PBS. Three successive immunizations were performed and the immune sera were collected 3 weeks after each of the immunizations (IS1, IS2, IS3). As a control, a group of mice (mock group) received aluminum hydroxide alone according to the same protocol.

[0633] The immune sera were analyzed per pool for each of the 2 groups by indirect ELISA using a lysate of VeroE6 cells infected with SARS-CoV as antigen and as a control a lysate of noninfected VeroE6 cells. The anti-SARS-CoV antibody titers are calculated as the reciprocal of the dilution producing a specific OD of 0.5 after visualization with an anti-mouse IgG (H+L) polyclonal antibody coupled with peroxidase (NA931V, Amersham) and TMB supplemented with H₂O₂ (KPL). This analysis (FIG. 31) shows that the

immunization with the Ssol polypeptide induces in mice, from the first immunization, antibodies directed against the native form of the SARS-CoV spicule protein present in the lysate of infected VeroE6 cells. After 2 then 3 immunizations, the anti-S antibody titers become very high.

[0634] The immune sera were analyzed per pool for each of the two groups for their capacity to seroneutralize the infectivity of SARS-CoV. 4 points of seroneutralization on FRhK-4 cells (100 TCID₅₀ of SARS-CoV) are produced for each of the 2-fold dilutions tested from 1/20. The seroneutralizing titer is calculated according to the Reed and Munsch method as the reciprocal of the dilution neutralizing the infectivity of 2 wells out of 4. This analysis shows that the antibodies induced in mice by the Ssol polypeptide are neutralizing: the titers observed are very high after 2 and then 3 immunizations (greater than 2560 and 5120 respectively, table XIII).

TABLE XIII

Induction of antibodies directed against SARS-CoV after immunization with the recombinant Ssol polypeptide. The immune sera were analyzed per pool for each of the two groups for their capacity to seroneutralize the infectivity of 100 TCID ₅₀ of SARS-CoV on FRhK-4 cells. 4 points are produced for each of the 2-fold dilutions tested from 1/20. The seroneutralizing titer is calculated according to the Reed and Munsch method as the reciprocal of the dilution neutralizing the infectivity of 2 wells out of 4.		
Group	Sera	Neutralizing Ab
Mock	pi	<20
	IS1	<20
	IS2	<20
	IS3	<20
Ssol	pi	<20
	IS1	57
	IS2	>2560
	IS3	>5120

[0635] The neutralizing titers observed in mice immunized with the Ssol polypeptide reach levels far greater than the titers observed by Yang et al. in mice (2004, Nature, 428:561-564) and those observed by Buchholz in the hamster (2004, PNAS 101:9804-9809) which protect respectively mice and hamsters from infection with SARS-CoV. It is therefore probable that the neutralizing antibodies induced in mice after immunization with the Ssol polypeptide protect these animals against infection with SARS-CoV.

EXAMPLE 15

Optimized Synthetic Gene for the Expression in Mammalian Cells of the SARS-associated Coronavirus (SARS-CoV) Spicule (S) Protein

1) Design of the Synthetic Gene

[0636] A synthetic gene encoding the SARS-CoV spicule protein was designed from the gene of the isolate 031589 (plasmid pSARS-S, C.N.C.M. No. I-3059) so as to allow high levels of expression in mammalian cells and in particular in cells of human origin.

[0637] For that:

[0638] the use of codons of the wild-type gene of the isolate 031589 was modified so as to become close to

the bias observed in humans and to improve the efficiency of translation of the corresponding mRNA

[0639] the overall GC content of the gene was increased so as to extend the half-life of the corresponding mRNA

[0640] the optionally cryptic motifs capable of interfering with an efficient expression of the gene were deleted (splice donor and acceptor sites, polyadenylation signals, sequences very rich (>80%) or very low (<30%) in GC, repeat sequences, sequences involved in the formation of secondary RNA structures, TATA boxes)

[0641] a second STOP codon was added to allow efficient termination of translation.

[0642] In addition, CpG motifs were introduced into the gene so as to increase its immunogenicity as DNA vaccine. In order to facilitate the manipulation of the synthetic gene, two BamH1 and Xho1 restriction sites were placed on either side of the open reading frame of the S protein, and the BamH1, Xho1, Nhe1, Kpn1, BspE1 and Sal1 restriction sites were avoided in the synthetic gene.

[0643] The sequence of the synthetic gene designed (gene 040530) is given in SEQ ID No: 140.

[0644] An alignment of the synthetic gene 040530 with the sequence of the wild-type gene of the isolate 031589 of SARS-CoV deposited at the C.N.C.M. under the number I-3059 (SEQ ID No: 4, plasmid pSRAS-S) is presented in FIG. 32.

2) Plasmid Constructs

[0645] The synthetic gene SEQ ID No: 140 was assembled from synthetic oligonucleotides and cloned between the Kpn1 and Sac1 sites of the plasmid pUC-Kana in order to give the plasmid 040530pUC-Kana. The nucleotide sequence of the insert of the plasmid 040530pUC-Kana was verified by automated sequencing (Applied).

[0646] A Kpn1-Xho1 fragment containing the synthetic gene 040530 was excised from the plasmid 040530pUC-Kana and subcloned between the Nhe1 and Xho1 sites of the expression plasmid pCI (Promega) in order to obtain the plasmid pCI-SSYNTH, deposited at the CNCM on Dec. 1, 2004, under the number I-3333.

[0647] A synthetic gene encoding the soluble form of the S protein was then obtained by fusing the synthetic sequences encoding the ectodomain of the S protein (amino acids 1 to 1193) with those of the tag (FLAG:DYKDDDDK) via a linker BspE1 encoding the dipeptide SG. Practically, a DNA fragment encoding the ectodomain of the SARS-CoV S was amplified by PCR with the aid of the oligonucleotides 5'-ACTAGCTAGCGGATCCACCATGTTTCATCTT CCTG-3' and 5'-AGTATCCGGAC TTG ATGIAC TCTCG-TACTTGC-3' from the plasmid 04053-0pUC-Kana, digested with Nhe1 and BspE1 and then inserted between the unique Nhe1 and BspE1 sites of the plasmid pCI-Ssol, to give the plasmid pCI-SCUBE, deposited at the CNCM on Dec. 1, 2004, under the number I-3332. The plasmids pCI-Ssol, pCI-Ssol-CTE, and pCI-Ssol-WPRE (deposited at the CNCM, on Nov. 22, 2004, under the number I-3324) had been previously obtained by subcloning the Kpn1-Xho1 fragment excised from the plasmid pcDNA-Ssol (see technical note of DI 2004-106) between the Nhe1 and Xho1 sites of the plasmids pCI, pCI-S-CTE and pCI-S-WPRE respectively.)

[0648] The plasmids pCI-Scube and pCI-Ssol encode the same recombinant Ssol polypeptide.

3) Results

[0649] The capacity of the synthetic gene encoding the S protein to efficiently direct the expression of the SARS-CoV S in mammalian cells was compared with that of the wild-type gene after transient transfection of primate cells (VeroE6) and of human cells (293T).

[0650] In the experiment presented in FIG. 33 and in table XIV, monolayers of 5×10^5 VeroE6 cells or 7×10^5 293T cells in 35 mm Petri dishes were transfected with 2 μ g of plasmids pCI (as control), pCI-S, pCI-S-CTE, pCI-S-WPRE and pCI-S-Synth and 6 μ l of Fugene6 reagent according to the manufacturer's instructions (Roche). After 48 hours of incubation at 37° C. and under 5% CO₂, cell extracts were prepared in loading buffer according to Laemmli, separated on 8% SDS polyacrylamide gel and then transferred onto a PVDF membrane (BioRad). The detection of this immunoblot (Western blot) was carried out with the aid of an anti-S rabbit polyclonal serum (immune serum of the rabbit P11135: cf example 4 above) and of donkey polyclonal antibodies directed against rabbit IgGs and coupled with peroxidase (NA934V, Amersham). The immunoblot was quantitatively visualized by luminescence with the aid of the ECL+ kit (Amersham) and acquisition on a digital imaging device (FluorS, BioRad).

[0651] The analysis of the results obtained with the software QuantityOne v4.2.3 (BioRad) shows that in this experiment, the plasmid pCI-Synth allows the transient expression of the S protein at high levels in the VeroE6 and 293T cells, whereas the plasmid pCI-S does not make it possible to induce expression at sufficient levels to be detected. The expression levels observed are of the order of twice as high as those observed with the plasmid pCI-S-WPRE.

TABLE XIV

Use of a synthetic gene for the expression of the SARS-CoV S. Cell extracts prepared 48 hours after transfection of VeroE6 or 293T cells with the plasmids pCI, pCI-S, pCI-S-CTE, pCI-S-WPRE and pCI-S-Synth were separated on 8% SDS acrylamide gel and analyzed by Western blotting with the aid of an anti-S rabbit polyclonal antibody and an anti-rabbit IgG(H + L) polyclonal antibody coupled with peroxidase (NA934V, Amersham). The Western blot is visualized by luminescence (ECL+, Amersham) and acquisition on a digital imaging device (FluorS, BioRad). The expression levels of the S protein were measured by quantifying the two predominant bands identified on the image (see FIG. 33) and are indicated according to an arbitrary scale where the value 1 represents the level measured after transfection of the plasmid pCI-S-WPRE.

Plasmid	VeroE6	293T
pCI	0.0	0.0
pCI-S	≤0.1	≤0.1
pCI-S-CTE	0.5	≤0.1
pCI-S-WPRE	1.0	1.0
pCI-Ssynth	1.8	1.9

[0652] In a second instance, the capacity of the synthetic gene Scube to efficiently direct the synthesis and the secretion of the Ssol polypeptide by mammalian cells was compared with that of the wild-type gene after transient transfection of hamster cells (BHK-21) and of human cells (293T).

[0653] In the experiment presented in table XV, monolayers of 6×10^5 BHK-21 cells and 7×10^5 293T cells in 35 mm Petri dishes were transfected with 2 μ g of plasmids pCI (as control), pCI-Ssol, pCI-Ssol-CTE, pCI-Ssol-WPRE and pCI-Scube and 6 μ l of Fugene6 reagent according to the manufacturer's instructions (Roche). After 48 hours of incubation at 37° C. and under 5% CO₂, the cellular supernatants were collected and quantitatively analyzed for the secretion of the Ssol polypeptide by a capture ELISA test specific for the Ssol polypeptide.

[0654] Analysis of the results shows that, in this experiment, the plasmid pCI-Scube allows the expression of the Ssol polypeptide at levels 8 times (BHK-21 cells) to 20 times (293T cells) higher than the plasmid pCI-Ssol. The levels of expression observed are of the order of twice (293T cells) to 5 times (BHK-21 cells) as high as those observed with the plasmid pCI-Ssol-WPRE.

TABLE XV

Plasmid	BHK	293T
pCI	<20	<20
pCI-Ssol	<20	56 ± 10
pCI-Ssol-CTE	<20	63 ± 8
pCI-Ssol-WPRE	28 ± 1	531 ± 15
pCI-Scube	152 ± 6	1140 ± 20

[0655] In summary, these results show that the expression, in mammalian cells, of the synthetic gene 040530 encoding SARS-CoV S under the control of RNA polymerase II promoter sequences is much more efficient than that of the wild-type gene of the 031589 isolate. This expression is even more efficient than that directed by the wild-type gene in the presence of the WPRE sequences of the woodchuck hepatitis virus.

4) Applications

[0656] The use of the synthetic gene 040530 encoding SARS-CoV S or its Scube variant encoding the polypeptide Ssol is capable of advantageously replacing the wild-type gene in numerous applications where the expression of S is necessary at high levels. In particular in order to:

[0657] improve the efficiency of gene immunization with plasmids of the pCI-Ssynth or even pCI-Ssynth-CTE or pCI-Ssynth-WPRE type

[0658] establish novel cell lines expressing higher quantities of the S protein or of the Ssol polypeptide with the aid of recombinant lentiviral vectors carrying the Ssynth gene or the Scube gene respectively

[0659] improve the immunogenicity of the recombinant lentiviral vectors allowing the expression of the S protein or of the Ssol polypeptide

[0660] improve the immunogenicity of live vectors allowing the expression of the S protein or of the Ssol polypeptide like recombinant vaccinia viruses or recombinant measles viruses (see examples 16 and 17 below)

EXAMPLE 16

Expression of the SARS-associated Coronavirus (SARS-CoV) Spicule (S) Protein with the Aid of Recombinant Vaccinia Viruses

Vaccine Application

Application to the Production of a Soluble Form of the Spicule (S) Protein and Design of a Serological Test for SARS

1) Introduction

[0661] The aim of this example is to evaluate the capacity of recombinant vaccinia viruses (VV) expressing various SARS-associated coronavirus (SARS-CoV) antigens to constitute novel vaccine candidates against SARS and a means of producing recombinant antigens in mammalian cells.

[0662] For that, the inventors focused on the SARS-CoV spicule (S) protein which makes it possible to induce, after gene immunization in animals, antibodies neutralizing the infectivity of SARS-CoV, and a soluble and secreted form of this protein, the Ssol polypeptide, which is composed of the ectodomain (aa 1-1193) of S fused at its C-ter end with a tag FLAG (DYKDDDDK) via a BspE1 linker encoding the SG dipeptide. This Ssol polypeptide exhibits an antigenicity similar to that of the S protein and allows, after injection into mice in the form of a purified protein adjuvanted with aluminum hydroxide, the induction of high neutralizing antibody titers against SARS-CoV.

[0663] The various forms of the S gene were placed under the control of the promoter of the 7.5K gene and then introduced into the thymidine kinase (TK) locus of the Copenhagen strain of the vaccinia virus by double homologous recombination in vivo. In order to improve the immunogenicity of the recombinant vaccinia viruses, a synthetic late promoter was chosen in place of the 7.5K promoter, in order to increase the production of S and Ssol during the late phases of the viral cycle.

[0664] After having isolated the recombinant vaccinia viruses and verified their capacity to express the SARS-CoV S antigen, their capacity to induce in mice an immune response against SARS was tested. After having purified the Ssol antigen from the supernatant of infected cells, an ELISA test for serodiagnosis of SARS was designed, and its efficiency was evaluated with the aid of sera from probable cases of SARS.

2) Construction of the Recombinant Viruses

[0665] Recombinant vaccinia viruses directing the expression of the S glycoprotein of the 031589 isolate of SARS-CoV and of a soluble and secreted form of this protein, the Ssol polypeptide, under the control of the 7.5K promoter were obtained. With the aim of increasing the levels of expression of S and Ssol, recombinant viruses in which the cDNAs for S and for Ssol are placed under the control of a late synthetic promoter were also obtained.

[0666] The plasmid pTG186poly is a transfer plasmid for the construction of recombinant vaccinia viruses (Kieny, 1986, *Biotechnology*, 4:790-795). As such, it contains the VV thymidine kinase gene into which the promoter of the 7.5K gene has been inserted followed by a multiple cloning site allowing the insertion of heterologous genes (FIG. 34A). The promoter of the 7.5K gene in fact contains a tandem of two promoter sequences that are respectively active during the early (P_E) and late (P_L) phases of the vaccinia virus replication cycle. The BamHI-XhoI fragments were excised from the plasmids pTRIP-S and pcDNA-Ssol respectively and inserted between the BamHI and SmaI sites of the plasmid pTG186poly in order to give the plasmids pTG-S and pTG-Ssol (FIG. 34A). The plasmids pTG-S and pTG-Ssol were deposited at the CNCM, on Dec. 2, 2004, under the numbers I-3338 and I-3339, respectively.

[0667] The plasmids pTN480, pTN-S and pTN-Ssol were obtained from the plasmids pTG186poly, pTG-S and pTG-Ssol respectively, by substituting the NdeI-PstI fragment containing the 7.5K promoter by a DNA fragment containing the synthetic late promoter 480, which was obtained by hybridization of the oligonucleotides 5'-TATGAGCTTT TTTTTTTTTT TTTTTTTGGC ATATAAATAG ACTCG-
CGCG CCATCTGCA-3' and 5'-GATGGCGCGC-
CGAGTCTATT TATATGCCAA AAAAAAAAAA
AAAAAAAAAGC TCA-3' (FIG. 34B). The insert was sequenced with the aid of a BigDye Terminator v1.1 kit (Applied Biosystems) and an automated sequencer ABI377. The sequence of the late synthetic promoter 480 as cloned into the transfer plasmids of the pTN series is indicated in FIG. 34C. The plasmids pTN-S and pTN-Ssol were deposited at the CNCM, on Dec. 2, 2004, under the numbers I-3340 and I-3341, respectively.

[0668] The recombinant vaccinia viruses were obtained by double homologous recombination in vivo between the TK cassette of the transfer plasmids of the series pTG and pTN and the TK gene of the Copenhagen strain of the vaccinia virus according to a procedure described by Kieny et al. (1984, *Nature*, 312:163-166). Briefly, CV-1 cells are transfected with the aid of DOTAP (Roche) with genomic DNA of the Copenhagen strain of the vaccinia virus and each of the transfer plasmids of the pTG and pTN series described above, and then superinfected with the helper vaccinia virus VV-ts7 for 24 hours at 33° C. The helper virus is counter-selected by incubation at 40° C. for 2 days and then the recombinant viruses (TK- phenotype) selected by two cloning cycles under agar medium on 143Btk- cells in the presence of BuDr (25 µg/ml). The 6 viruses VV-TG, VV-TG-S, VV-TG-Ssol, VV-TN, VV-TN-S, and VV-TN-Ssol are respectively obtained with the aid of the transfer plasmids pTG186poly, pT-G-S, pTG-Ssol, pTN480, pTN-S, pTN-Ssol. The viruses VV-TG and VV-TN do not express any heterologous gene and were used as TK- control in the experiments. The preparations of recombinant viruses were performed on monolayers of CV-1 or BHK-21 cells and the titer in plaque forming units (p.f.u) determined on CV-1 cells according to Earl and Moss (1998, *Current Protocols in Molecular Biology*, 16.16.1-16.16.13).

3) Characterization of the Recombinant Viruses

[0669] The expression of the transgenes encoding the S protein and the Ssol polypeptide was assessed by Western blotting.

[0670] Monolayers of CV-1 cells were infected at a multiplicity of 2 with various recombinant vaccinia viruses VV-TG, VV-TG-S, VV-TG-Ssol, VV-TN, VV-TN-S and VV-TN-Ssol. After 18 hours of incubation at 37° C. and under 5% CO₂, cellular extracts were prepared in loading buffer according to Laemmli, separated on 8% SDS polyacrylamide gel and then transferred onto a PVDF membrane (BioRad). The detection of this immunoblot (Western blot) was performed with the aid of an anti-S rabbit polyclonal serum (immune serum from the rabbit P11135: cf. example 4) and donkey polyclonal antibodies directed against rabbit IgGs and coupled with peroxidase (NA934V, Amersham). The bound antibodies were visualized by luminescence with the aid of the ECL+kit (Amersham) and autoradiography films Hyperfilm MP (Amersham).

[0671] As shown in FIG. 35A, the recombinant virus VV-TN-S directs the expression of the S protein at levels which are comparable to those which can be observed 8 h after infection with SARS-CoV but which are much higher than those which can be observed after infection with VV-TG-S. In a second experiment (FIG. 35B), the analysis of variable quantities of cellular extracts shows that the levels of expression observed after infection with viruses of the TN series (VV-TN-S and VV-TN-Ssol) are about 10 times as high as those observed with the viruses of the TG series (VV-TG-S and VV-TG-Ssol, respectively); In addition, the Ssol polypeptide is secreted into the supernatant of CV-1 cells infected with the VV-TN-Ssol virus more efficiently than in the supernatant of cells infected with VV-TG-Ssol (FIG. 36A). In this experiment, the VV-TN-Sflag virus was used as a control because it expresses the membrane form of the S protein fused at its C-ter end with the FLAG tag. The Sflag protein is not detected in the supernatant of cells infected with VV-TN-Sflag, demonstrating that the Ssol polypeptide is indeed actively secreted after infection with VV-TN-Ssol.

[0672] These results demonstrate that the recombinant vaccinia viruses are indeed carriers of the transgenes and allow the expression of the SRAS glycoprotein in its membrane form (S) or in a soluble or secreted form (Ssol). The vaccinia viruses carrying the synthetic promoter 480 allow the expression of S and the secretion of Ssol at levels much higher than the viruses carrying the promoter of the 7.5K gene.

4) Application to the Production of a Soluble Form of SARS-CoV S. Purification of this Recombinant Antigen and Diagnostic Applications

[0673] The BHK-21 line is the cell line which secretes the highest quantities of Ssol polypeptide after infection with the VV-TN-Ssol virus among the lines tested (BHK-21, CV1, 293T and FrhK-4, FIG. 36B); it allows the quantitative production and purification of the recombinant Ssol polypeptide. In a typical experiment where the experimental conditions for infection, production and purification were optimized, the BHK-21 cells are inoculated in standard culture medium (pyruvate-free DMEM containing 4.5 g/l of glucose and supplemented with 5% TPB, 5% FCS, 100 U/ml of penicillin and 100 µg/ml of streptomycin) in the form of a subconfluent monolayer (10 million cells for each 100 cm² in 25 ml of medium). After 24 h of incubation at 37° C. under 5% CO₂, the cells are infected at an M.O.I. of 0.03 and the standard medium replaced with the secretion medium

where the quantity of FCS is reduced to 0.5% and the TPB eliminated. The culture supernatant is removed after 2.5 days of incubation at 35° C. and under 5% CO₂ and the vaccinia virus inactivated by addition of Triton X-100 (0.1%). After filtration on 0.1 µm polyethersulfone (PES) membrane, the recombinant Ssol polypeptide is purified by affinity chromatography on an anti-FLAG matrix with elution with a solution of FLAG peptide (DYKDDDDK) at 100 µg/ml in TBS (50 mM Tris, pH 7.4, 150 mM NaCl).

[0674] The analysis by 8% SDS acrylamide gel stained with silver nitrate identified a predominant polypeptide whose molecular mass is about 180 kD and whose degree of purity is greater than 90% (FIG. 37). The concentration of the purified Ssol recombinant polypeptide was determined by comparison with molecular mass markers and estimated at 24 ng/µl.

[0675] This purified Ssol polypeptide preparation makes it possible to produce a calibration series in order to measure, with the aid of a capture ELISA test, the Ssol concentrations present in the culture supernatants. According to this test, the BHK-21 line secretes about 1 µg/ml of Ssol polypeptide under the production conditions described above. In addition, the purification scheme presented makes it possible to purify of the order of 160 µg of Ssol polypeptide per liter of culture supernatant.

[0676] The ELISA reactivity of the recombinant Ssol polypeptide was analyzed toward sera from patients suffering from SARS.

[0677] The sera of probable cases of SARS tested were chosen on the basis of the results (positive or negative) of analysis of their specific reactivity toward the native antigens of SARS-CoV by immunofluorescence test on VeroE6 cells infected with SARS-CoV and/or by indirect ELISA test using, as antigen, a lysate of VeroE6 cells infected with SARS-CoV. The sera of these patients are identified by a serial number of the National Reference Center for Influenza Viruses and by the patient's initials and the number of days elapsed since the onset of the symptoms. All the sera of probable cases (cf. table XVI) recognize the native antigens of SARS-CoV with the exception of the serum 032552 of the patient VTT, for which infection with SARS-CoV could not be confirmed by RT-PCR performed on respiratory samples of days 3, 8 and 12. A panel of control sera was used as control (TV sera): they are sera collected in France before the SARS epidemic which occurred in 2003.

TABLE XVI

Sera of probable cases of SARS		
Serum	Patient	Sample collection day
033168	JYK	38
033597	JYK	74
032632	NTM	17
032634	THA	15
032541	PHV	10
032542	NIH	17
032552	VTT	8
032633	PTU	16

[0678] Solid phases sensitized with the recombinant Ssol polypeptide were prepared by adsorption of a solution of purified Ssol polypeptide at 4 µg/ml in PBS in the wells of

an ELISA plate. The plates are incubated overnight at 4° C. and then washed with PBS-Tween buffer (PBS, 0.1% Tween 20). After washing with PBS-Tween, the sera to be tested (100 µl) are diluted 1/100 and 1/400 in PBS-skimmed milk-Tween buffer (PBS, 3% skimmed milk, 0.1% Tween) and then added to the wells of the sensitized ELISA plate. The plates are then incubated for 1 h at 37° C. After 3 washings with PBS-Tween buffer, the anti-human IgG conjugate labeled with peroxidase (ref. NA933V, Amersham) diluted 1/4000 in PBS-skimmed milk-Tween buffer is added and then the plates are incubated for one hour at 37° C. After 6 washings with PBS-Tween buffer, the chromogen (TMB) and the substrate (H₂O₂) are added and the plates are incubated for 10 minutes protected from light. The reaction is stopped by adding a 1M solution of H₃PO₄ and then the absorbance is measured at 450 nm with a reference at 620 nm.

[0679] The ELISA tests (FIG. 38) demonstrate that the recombinant Ssol polypeptide is specifically recognized by the serum antibodies of patients suffering from SARS, collected at the middle or late phase of infection (≥ 10 days after the onset of the symptoms), whereas it is not significantly recognized by the serum antibodies of the control sera of subjects not suffering from SARS.

[0680] In conclusion, these results demonstrate that the recombinant Ssol polypeptide can be purified from the supernatant of mammalian cells infected with the recombinant vaccinia virus VV-TN-Ssol and can be used as antigen for developing an ELISA test for serological diagnosis of infection with SARS-CoV.

5. Vaccine Applications

[0681] The immunogenicity of the recombinant vaccinia viruses was studied in mice.

[0682] For that, groups of 7 BALB/c mice were immunized by the i.v. route twice at 4 weeks' interval with 10⁶ p.f.u. of recombinant vaccinia viruses VV-TG, VV-T-G-S, VV-TG-Ssol, VV-TN, VV-TN-S and VV-TN-Ssol and, as a control, VV-TG-HA which directs the expression of hemagglutinin of the A/PR/8/34 strain of the influenza virus. The immune sera were collected 3 weeks after each of the immunizations (IS1, IS2).

[0683] The immune sera were analyzed per pool for each of the groups by indirect ELISA using a lysate of VeroE6 cells infected with SARS-CoV as antigen and, as control, a lysate of noninfected VeroE6 cells. The anti-SARS-CoV antibody titers (TI) are calculated as the reciprocal of the dilution producing a specific OD of 0.5 after visualization with an anti-mouse IgG(H+L) polyclonal antibody coupled with peroxidase (NA931V, Amersham) and TMB supplemented with H₂O₂ (KPL). This analysis (FIG. 39A) shows that immunization with the virus VV-TG-S and VV-TN-S induces in mice, from the first immunization, antibodies directed against the native form of the SARS-CoV spicule protein present in the lysate of infected VeroE6 cells. The responses induced by the VV-TN-S virus are higher than those induced by the VV-TG-S virus after the first (TI=740 and TI=270 respectively) and the second (TI=3230 and TI=600 respectively) immunization. The VV-TN-Ssol virus induces high anti-SARS-CoV antibody titers after two immunizations (TI=640), whereas the virus VV-TG-Ssol induces a response at the detection limit (TI=40).

[0684] The immune sera were analyzed per pool for each of the groups for their capacity to seroneutralize the infectivity of SARS-CoV. 4 seroneutralization points on FRhK-4 cells (100 TCID₅₀ of SARS-CoV) are produced for each of the 2-fold dilutions tested from 1/20. The seroneutralizing titer is calculated according to the Reed and Munsch method as the reciprocal of the dilution neutralizing the infectivity of 2 wells out of 4. This analysis shows that the antibodies induced in mice by the vaccinia viruses expressing the S protein or the Ssol polypeptide are neutralizing and that the viruses with synthetic promoters are more efficient immunogens than the viruses carrying the 7.5K promoter: the highest titers (640) are observed after 2 immunizations with the virus VV-TN-S (FIG. 39B).

[0685] The protective power of the neutralizing antibodies induced in mice after immunization with the recombinant vaccinia viruses is evaluated with the aid of a challenge infection with SARS-CoV.

6) Other Applications

[0686] Third generation recombinant vaccinia viruses are constructed by substituting the wild-type sequences of the S and Ssol genes by synthetic genes optimized for the expression in mammalian cells, described above. These recombinant vaccinia viruses are capable of expressing larger quantities of S and Ssol antigens and therefore of exhibiting increased immunogenicity.

[0687] The recombinant vaccinia virus VV-TN-Ssol can be used for the quantitative production and purification of the Ssol antigen for diagnostic (serology by ELISA) and vaccine (subunit vaccine) applications.

EXAMPLE 17

Recombinant Measles Virus Expressing the SARS-associated Coronavirus (SARS-CoV) Spicule (S) Protein. Vaccine Applications.

1) Introduction

[0688] The measles vaccine (MV) induces a lasting protective immunity in humans after a single injection (Hilleman, 2002, Vaccine, 20: 651-665). The protection conferred is very robust and is based on the induction of an antibody response and of a CD4 and CD8 cell response. The MV genome is very stable and no reversion of the vaccine strains to virulence has ever been observed. The measles virus belongs to the genus *Morbillivirus* of the Paramyxoviridae family; it is an enveloped virus whose genome is a 16 kb single-stranded RNA of negative polarity (FIG. 40A) and whose exclusively cytoplasmic replication cycle excludes any possibility of integration into the genome of the host. The measles vaccine is thus one of the most effective and one of the safest live vaccines used in the human population. Frederic Tangy's team recently developed an expression vector on the basis of the Schwarz strain of the measles virus, which is the safest attenuated strain and the most widely used in humans as vaccine against measles. This vaccine strain may be isolated from an infectious molecular clone while preserving its immunogenicity in primates and in mice that are sensitive to the infection. It constitutes, after insertion of additional transcription units, a vector for the expression of heterologous sequences (Combredet, 2003, J. Virol. 77: 11546-11554). In addition, a recombinant MV Schwarz expressing the envelope glycoprotein of the West Nile virus (WNV) induces an effective and lasting antibody response which protects mice from a lethal challenge infec-

tion with WNV (Despres et al., 2004, J. Infect. Dis., in press). All these characteristics make the attenuated Schwarz strain of the measles virus an extremely promising candidate vector for the construction of novel recombinant live vaccines.

[0689] The aim of this example is to evaluate the capacity of recombinant measles viruses (MV) expressing various SARS-associated coronavirus (SARS-CoV) antigens to constitute novel candidate vaccines against SARS.

[0690] The inventors focused on the SARS-CoV spicule (S) protein, which makes it possible to induce, after gene immunization in animals, antibodies neutralizing the infectivity of SARS-CoV, and on a soluble and secreted form of this protein, the Ssol polypeptide, which is composed of the ectodomain (aa 1-1193) of S fused at its C-ter end with a FLAG tag (DYKDDDDK) via a BspE1 linker encoding the SG dipeptide. This Ssol polypeptide exhibits a similar antigenicity to that of the S protein and allows, after injection into mice in the form of a purified protein adjuvanted with aluminum hydroxide, the induction of high neutralizing antibody titers against SARS-CoV.

[0691] The various forms of the S gene were introduced in the form of an additional transcription unit between the P (phosphoprotein) and M (matrix) genes into the cDNA of the Schwarz strain of MV previously described (Combredet, 2003, J. Virol. 77: 11546-11554; EP application No. 02291551.6 of Jun. 20, 2002, and EP application No. 02291550.8 of Jun. 20, 2002). After having isolated the recombinant viruses MVSchw2-SARS-S and MVSchw2-SARS-Ssol and checked their capacity to express the SARS-CoV S antigen, their capacity to induce a protective immune response against SARS in mice and then in monkeys was tested.

2) Construction of the Recombinant Viruses

[0692] The plasmid pTM-MVSchw-ATU2 (FIG. 40B) contains an infectious cDNA corresponding to the antigenome of the Schwarz vaccine strain of the measles virus (MV) into which an additional transcription unit (ATU) has been introduced between the P (phosphoprotein) and M (matrix) genes (Combredet, 2003, Journal of Virology, 77: 11546-11554). Recombinant genomes MVSchw2-SARS-S and MVSchw2-SARS-Ssol of the measles virus were constructed by inserting ORFs of the S protein and of the Ssol polypeptide into the additional transcription unit of the MVSchw-ATU2 vector.

[0693] For that, a DNA fragment containing the SARS-CoV S cDNA was amplified by PCR with the aid of the oligonucleotides 5'-ATACGTACGA CCAATGTTTAT TTTCTAATA TTTCTTACTC TCACT-3' and 5'-AT-AGCCGCT CATTATGTGT AATGTAATTT GACAC-CCTTG-3' using the plasmid pcDNA-S as template and then inserted into the plasmid pCR®2.1-TOPO (Invitrogen) in order to obtain the plasmid pTOPO-S-MV. The two oligonucleotides used contain restriction sites BsiWI and BssHIII, so as to allow subsequent insertion into the measles vector, and were designed so as to generate a sequence of 3774 nt including the codons for initiation and termination, so as to observe the rule of 6 which stipulates that the length of the genome of a measles virus must be divisible by 6 (Calain & Roux, 1993, J. Virol., 67: 4822-4830; Schneider et al., 1997, Virology, 227: 314-322). The insert was sequenced with the aid of a BigDye Terminator v1.1 kit (Applied Biosystems) and an automated sequencer ABI377.

[0694] To express a soluble and secreted form of SARS-CoV S, a plasmid containing the cDNA of the Ssol polypep-

tide corresponding to the ectodomain (aa 1-1193) of SARS-CoV S fused at its C-ter end with the sequence of a FLAG tag (DYKDDDDK) via a BspE1 linker encoding the SG dipeptide was then obtained. For that, a DNA fragment was amplified with the aid of the oligonucleotides 5'-CCATTTCAAC AATTTGGCCG-3' and 5'-ATAGGATC-CGCGCGCTCAIT ATTTATCGTC GTCATCTTTA TAATC-3' from the plasmid pcDNA-Ssol and then inserted into the plasmid pTOPO-S-MV between the SalI and BamHI sites in order to obtain the plasmid pTOPO-S-MV-SF. The sequence generated is 3618 nt long between the BsiW1 and BssHII sites and observes the rule of 6. The insert was sequenced as indicated above.

[0695] The BsiW1-BssHII fragments containing the cDNAs for the S protein and the Ssol polypeptide were then excised by digestion of the plasmids pTOPO-S-MV and pTOPO-S-MV-SF and then subcloned between the corresponding sites of the plasmid pTM-MV Schw-ATU2 in order to give the plasmids pTM-MV Schw2-SARS-S and pTM-MV Schw2-SARS-Ssol (FIG. 40B). These two plasmids were deposited at the C.N.C.M. on Dec. 1, 2004, under the numbers I-3326 and I-3327, respectively.

[0696] The recombinant measles viruses corresponding to the plasmids pTM-MV Schw2-SARS-S and pTM-MV Schw2-SARS-Ssol were obtained by reverse genetics according to the system based on the use of a helper cell line, described by Radecke et al. (1995, *Embo J.*, 14: 5773-5784) and modified by Parks et al. (1999, *J. Virol.*, 73: 3560-3566). Briefly, the helper cells 293-3-46 are transfected according to the calcium phosphate method with 5 µg of the plasmids pTM-MV Schw2-SARS-S or pTM-MV Schw2-SARS-Ssol and 0.02 µg of the plasmid pEMC-La directing the expression of the MV L polymerase (gift from M. A. Billeter). After incubating overnight at 37° C., a heat shock is produced for 2 hours at 43° C. and the transfected cells are transferred onto a monolayer of Vero cells. For each of the two plasmids, syncytia appeared after 2 to 3 days of coculture and were transferred successively onto monolayers of Vero cells at 70% confluence in 35 mm Petri dishes and then in 25 and 75 cm² flasks. When the syncytia have reached 80-90% confluence, the cells are recovered with the aid of a scraper and then frozen and thawed once. After low-speed centrifugation, the supernatant containing the virus is stored in aliquots at -80° C. The titers of the recombinant viruses MV Schw2-SARS-S and MV Schw2-SARS-Ssol were determined by limiting dilution on Vero cells and the titer as dose infecting 50% of the wells (TCID₅₀) calculated according to the Kärber method.

3) Characterization of the Recombinant Viruses

[0697] The expression of the transgenes encoding the S protein and the Ssol polypeptide was assessed by Western blotting and immunofluorescence.

[0698] Monolayers of Vero cells in T-25 flasks were infected at a multiplicity of 0.05 by various passages of the two viruses MV Schw2-SARS-S and MV Schw2-SARS-Ssol and the wild-type virus MWSchw as a control. When the syncytia had reached 80 to 90% confluence, cytoplasmic extracts were prepared in an extraction buffer (150 mM NaCl, 50 mM Tris-HCl, pH 7.2, 1% Triton X-100, 0.1% SDS, 1% DOC) and then diluted in loading buffer according to Laemmli, separated on 8% SDS polyacrylamide gel and transferred onto a PVDF membrane (BioRad). The detection

of this immunoblot (Western blot) was carried out with the aid of an anti-S rabbit polyclonal serum (immune serum of the rabbit P11135: cf. example 4 above) and donkey polyclonal antibodies directed against rabbit IgGs and coupled with peroxidase (NA934V, Amersham). The bound antibodies were visualized by luminescence with the aid of the ECL+kit (Amersham) and Hyperfilm MP autoradiography films (Amersham).

[0699] Vero cells in monolayers on glass slides were infected with the two viruses MV Schw2-SARS-S and MV Schw2-SARS-Ssol and the wild-type virus MWSchw as a control at multiplicities of infection of 0.05. When the syncytia had reached 90 to 100% (MV Schw2-SARS-Ssol virus) or 30 to 40% (MV Schw2-SARS-S, MWSchw) confluence, the cells were fixed in a 4% PBS-PFA solution, permeabilized with a PBS solution containing 0.2% Triton and then labeled with rabbit polyclonal antibodies hyperimmunized with purified and inactivated SARS-CoV virions and with an anti-rabbit IgG(H+L) goat antibody conjugate coupled with FITC (Jackson).

[0700] As shown in FIGS. 41 and 42, the recombinant viruses MV Schw2-SARS-S and MV Schw2-SARS-Ssol direct the expression of the S protein and the Ssol polypeptide respectively at levels comparable to those which can be observed 8 h after infection with SARS-CoV. The expression of these polypeptides is stable after 3 passages of the recombinant viruses in cell culture. These results demonstrate that the recombinant measles viruses are indeed carriers of the transgenes and allow the expression of the SARS glycoprotein in its membrane form (S) or in a soluble form (Ssol). The Ssol polypeptide is expected to be secreted by cells infected with the MV Schw2-SARS-Ssol virus as is the case when this same polypeptide is expressed in mammalian cells after transient transfection of the corresponding sequences (cf. example 11 above).

4) Applications

[0701] Having shown that the viruses MV Schw2-SARS-S and MV Schw2-SARS-Ssol allow the expression of the SARS-CoV S, their capacity to induce a protective immune response against SARS-CoV in CD46^{+/-} IFN-αβR^{-/-} mice, which is sensitive to infection by MV, is evaluated. The antibody response of the immunized mice is evaluated by ELISA test against the native antigens of SARS-CoV and for their capacity to neutralize the infectivity of SARS-CoV in vitro, using the methodologies described above. The protective power of the response will be evaluated by measuring the reduction in the pulmonary viral load 2 days after a nonlethal challenge infection with SARS-CoV.

[0702] Second generation recombinant measles viruses are constructed by substituting the wild-type sequences of the S and Sol genes by synthetic genes optimized for expression in mammalian cells, described in example 15 above. These recombinant measles viruses are capable of expressing larger quantities of the S and Ssol antigens and therefore of exhibiting increased immunogenicity.

[0703] Alternatively, the wild-type or synthetic genes encoding the S protein or the Ssol polypeptide may be inserted into the measles vector MV Schw-ATU3 in the form of an additional transcription unit located between the H and L genes, and then the recombinant viruses produced and characterized in a similar manner. This insertion is capable

of generating recombinant viruses possessing different characteristics (multiplication of the virus, level of expression of the transgene) and possibly an improved immunogenicity compared with those obtained after insertion of the transgenes between the P and N genes.

[0704] The recombinant measles virus MVSchw2-SARS-Ssol may be used for the quantitative production and the purification of the Ssol antigen for diagnostic and vaccine applications.

EXAMPLE 18

Other Applications Linked to the S Protein

[0705] a) The lentiviral vectors allowing the expression of S or Ssol (or even of fragments of S) can constitute a

recombinant vaccine against SARS-CoV, to be used in human or veterinary prophylaxis. In order to demonstrate the feasibility of such a vaccine, the immunogenicity of the recombinant lentiviral vectors TRIP-SD/SA-S-WPRE and TRIP-SD/SA-Ssol-WPRE is studied in mice.

[0706] b) Monoclonal antibodies are produced with the aid of the recombinant Ssol polypeptide. According to the results presented in example 14 above, these antibodies or at least the majority of them will recognize the native form of the SARS-CoV S and will be capable of diagnostic and/or prophylactic applications.

[0707] c) A serological test for SARS is developed with the Ssol polypeptide used as antigen and the double epitope methodology.

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Phe	Asn	Ala	Thr	Lys	Phe	Pro	Ser	Val	Tyr	Ala	Trp	Glu	Arg	Lys	Lys		
	330					335						340					
att	tct	aat	tgt	gtt	gct	gat	tac	tct	gtg	ctc	tac	aac	tca	aca	ttt		1168
Ile	Ser	Asn	Cys	Val	Ala	Asp	Tyr	Ser	Val	Leu	Tyr	Asn	Ser	Thr	Phe		
	345				350					355					360		
ttt	tca	acc	ttt	aag	tgc	tat	ggc	gtt	tct	gcc	act	aag	ttg	aat	gat		1216
Phe	Ser	Thr	Phe	Lys	Cys	Tyr	Gly	Val	Ser	Ala	Thr	Lys	Leu	Asn	Asp		
				365					370					375			
ctt	tgc	ttc	tcc	aat	gtc	tat	gca	gat	tct	ttt	gta	gtc	aag	gga	gat		1264
Leu	Cys	Phe	Ser	Asn	Val	Tyr	Ala	Asp	Ser	Phe	Val	Val	Lys	Gly	Asp		
				380				385						390			
gat	gta	aga	caa	ata	gcg	cca	gga	caa	act	ggt	ggt	att	gct	gat	tat		1312

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Asp Val Arg Gln Ile Ala Pro Gly Gln Thr Gly Val Ile Ala Asp Tyr	
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aat tat aaa ttg cca gat gat ttc atg ggt tgt gtc ctt gct tgg aat	1360
Asn Tyr Lys Leu Pro Asp Asp Phe Met Gly Cys Val Leu Ala Trp Asn	
410 415 420	
act agg aac att gat gct act tca act ggt aat tat aat tat aaa tat	1408
Thr Arg Asn Ile Asp Ala Thr Ser Thr Gly Asn Tyr Asn Tyr Lys Tyr	
425 430 435 440	
agg tat ctt aga cat ggc aag ctt agg ccc ttt gag aga gac ata tct	1456
Arg Tyr Leu Arg His Gly Lys Leu Arg Pro Phe Glu Arg Asp Ile Ser	
445 450 455	
aat gtg cct ttc tcc cct gat ggc aaa cct tgc acc cca cct gct ctt	1504
Asn Val Pro Phe Ser Pro Asp Gly Lys Pro Cys Thr Pro Pro Ala Leu	
460 465 470	
aat tgt tat tgg cca tta aat gat tat ggt ttt tac acc act act ggc	1552
Asn Cys Tyr Trp Pro Leu Asn Asp Tyr Gly Phe Tyr Thr Thr Thr Gly	
475 480 485	
att ggc tac caa cct tac aga gtt gta gta ctt tct ttt gaa ctt tta	1600
Ile Gly Tyr Gln Pro Tyr Arg Val Val Val Leu Ser Phe Glu Leu Leu	
490 495 500	
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Asn Ala Pro Ala Thr Val Cys Gly Pro Lys Leu Ser Thr Asp Leu Ile	
505 510 515 520	
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Lys Asn Gln Cys Val Asn Phe Asn Phe Asn Gly Leu Thr Gly Thr Gly	
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gtg tta act cct tct tca aag aga ttt caa cca ttt caa caa ttt ggc	1744
Val Leu Thr Pro Ser Ser Lys Arg Phe Gln Pro Phe Gln Gln Phe Gly	
540 545 550	
cgt gat gtt tct gat ttc act gat tcc gtt cga gat cct aaa aca tct	1792
Arg Asp Val Ser Asp Phe Thr Asp Ser Val Arg Asp Pro Lys Thr Ser	
555 560 565	
gaa ata tta gac att tca cct tgc tct ttt ggg ggt gta agt gta att	1840
Glu Ile Leu Asp Ile Ser Pro Cys Ser Phe Gly Gly Val Ser Val Ile	
570 575 580	
aca cct gga aca aat gct tca tct gaa gtt gct gtt cta tat caa gat	1888
Thr Pro Gly Thr Asn Ala Ser Ser Glu Val Ala Val Leu Tyr Gln Asp	
585 590 595 600	
gtt aac tgc act gat gtt tct aca gca att cat gca gat caa ctc aca	1936
Val Asn Cys Thr Asp Val Ser Thr Ala Ile His Ala Asp Gln Leu Thr	
605 610 615	
cca gct tgg cgc ata tat tct act gga aac aat gta ttc cag act caa	1984
Pro Ala Trp Arg Ile Tyr Ser Thr Gly Asn Asn Val Phe Gln Thr Gln	
620 625 630	
gca ggc tgt ctt ata gga gct gag cat gtc gac act tct tat gag tgc	2032
Ala Gly Cys Leu Ile Gly Ala Glu His Val Asp Thr Ser Tyr Glu Cys	
635 640 645	
gac att cct att gga gct ggc att tgt gct agt tac cat aca gtt tct	2080
Asp Ile Pro Ile Gly Ala Gly Ile Cys Ala Ser Tyr His Thr Val Ser	
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tta tta cgt agt act agc caa aaa tct att gtg gct tat act atg tct	2128
Leu Leu Arg Ser Thr Ser Gln Lys Ser Ile Val Ala Tyr Thr Met Ser	
665 670 675 680	
tta ggt gct gat agt tca att gct tac tct aat aac acc att gct ata	2176
Leu Gly Ala Asp Ser Ser Ile Ala Tyr Ser Asn Asn Thr Ile Ala Ile	
685 690 695	
cct act aac ttt tca att agc att act aca gaa gta atg cct gtt tct	2224

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Pro Thr Asn Phe Ser Ile Ser Ile Thr Thr Glu Val Met Pro Val Ser	
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Met Ala Lys Thr Ser Val Asp Cys Asn Met Tyr Ile Cys Gly Asp Ser	
715 720 725	
act gaa tgt gct aat ttg ctt ctc caa tat ggt agc ttt tgc aca caa	2320
Thr Glu Cys Ala Asn Leu Leu Gln Tyr Gly Ser Phe Cys Thr Gln	
730 735 740	
cta aat cgt gca ctc tca ggt att gct gct gaa cag gat cgc aac aca	2368
Leu Asn Arg Ala Leu Ser Gly Ile Ala Ala Glu Gln Asp Arg Asn Thr	
745 750 755 760	
cgt gaa gtg ttc gct caa gtc aaa caa atg tac aaa acc cca act ttg	2416
Arg Glu Val Phe Ala Gln Val Lys Gln Met Tyr Lys Thr Pro Thr Leu	
765 770 775	
aaa tat ttt ggt ggt ttt aat ttt tca caa ata tta cct gac cct cta	2464
Lys Tyr Phe Gly Gly Phe Asn Phe Ser Gln Ile Leu Pro Asp Pro Leu	
780 785 790	
aag cca act aag agg tct ttt att gag gac ttg ctc ttt aat aag gtg	2512
Lys Pro Thr Lys Arg Ser Phe Ile Glu Asp Leu Leu Phe Asn Lys Val	
795 800 805	
aca ctc gct gat gct ggc ttc atg aag caa tat ggc gaa tgc cta ggt	2560
Thr Leu Ala Asp Ala Gly Phe Met Lys Gln Tyr Gly Glu Cys Leu Gly	
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gat att aat gct aga gat ctc att tgt gcg cag aag ttc aat gga ctt	2608
Asp Ile Asn Ala Arg Asp Leu Ile Cys Ala Gln Lys Phe Asn Gly Leu	
825 830 835 840	
aca gtg ttg cca cct ctg ctc act gat gat atg att gct gcc tac act	2656
Thr Val Leu Pro Pro Leu Leu Thr Asp Asp Met Ile Ala Ala Tyr Thr	
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Gly Ala Ala Leu Gln Ile Pro Phe Ala Met Gln Met Ala Tyr Arg Phe	
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Asn Gly Ile Gly Val Thr Gln Asn Val Leu Tyr Glu Asn Gln Lys Gln	
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atc gcc aac caa ttt aac aag gcg att agt caa att caa gaa tca ctt	2848
Ile Ala Asn Gln Phe Asn Lys Ala Ile Ser Gln Ile Gln Glu Ser Leu	
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Thr Thr Thr Ser Thr Ala Leu Gly Lys Leu Gln Asp Val Val Asn Gln	
925 930 935	
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Asn Ala Gln Ala Leu Asn Thr Leu Val Lys Gln Leu Ser Ser Asn Phe	
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ggt gca att tca agt gtg cta aat gat atc ctt tcg cga ctt gat aaa	2992
Gly Ala Ile Ser Ser Val Leu Asn Asp Ile Leu Ser Arg Leu Asp Lys	
955 960 965	
gtc gag gcg gag gta caa att gac agg tta att aca ggc aga ctt caa	3040
Val Glu Ala Glu Val Gln Ile Asp Arg Leu Ile Thr Gly Arg Leu Gln	
970 975 980	
agc ctt caa acc tat gta aca caa caa cta atc agg gct gct gaa atc	3088
Ser Leu Gln Thr Tyr Val Thr Gln Gln Leu Ile Arg Ala Ala Glu Ile	
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Leu Gly Gln Ser Lys	Arg Val Asp Phe Cys	Gly Lys Gly Tyr His	
	1020	1025	1030
ctt atg tcc ttc cca	caa gca gcc ccg cat	ggt gtt gtc ttc cta	3223
Leu Met Ser Phe Pro	Gln Ala Ala Pro His	Gly Val Val Phe Leu	
	1035	1040	1045
cat gtc acg tat gtg	cca tcc cag gag agg	aac ttc acc aca gcg	3268
His Val Thr Tyr Val	Pro Ser Gln Glu Arg	Asn Phe Thr Thr Ala	
	1050	1055	1060
cca gca att tgt cat	gaa ggc aaa gca tac	ttc cct cgt gaa ggt	3313
Pro Ala Ile Cys His	Glu Gly Lys Ala Tyr	Phe Pro Arg Glu Gly	
	1065	1070	1075
ggt ttt gtg ttt aat	ggc act tct tgg ttt	att aca cag agg aac	3358
Val Phe Val Phe Asn	Gly Thr Ser Trp Phe	Ile Thr Gln Arg Asn	
	1080	1085	1090
ttc ttt tct cca caa	ata att act aca gac	aat aca ttt gtc tca	3403
Phe Phe Ser Pro Gln	Ile Ile Thr Thr Asp	Asn Thr Phe Val Ser	
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gga aat tgt gat gtc	ggt att ggc atc att	aac aac aca gtt tat	3448
Gly Asn Cys Asp Val	Val Ile Gly Ile Ile	Asn Asn Thr Val Tyr	
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gat cct ctg caa cct	gag ctt gac tca ttc	aaa gaa gag ctg gac	3493
Asp Pro Leu Gln Pro	Glu Leu Asp Ser Phe	Lys Glu Glu Leu Asp	
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Lys Tyr Phe Lys Asn	His Thr Ser Pro Asp	Val Asp Leu Gly Asp	
	1140	1145	1150
att tca ggc att aac	gct tct gtc gtc aac	att caa aaa gaa att	3583
Ile Ser Gly Ile Asn	Ala Ser Val Val Asn	Ile Gln Lys Glu Ile	
	1155	1160	1165
gac cgc ctc aat gag	gtc gct aaa aat tta	aat gaa tca ctc att	3628
Asp Arg Leu Asn Glu	Val Ala Lys Asn Leu	Asn Glu Ser Leu Ile	
	1170	1175	1180
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Asp Leu Gln Glu Leu	Gly Lys Tyr Glu Gln	Tyr Ile Lys Trp Pro	
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Trp Tyr Val Trp Leu	Gly Phe Ile Ala Gly	Leu Ile Ala Ile Val	
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Met Val Thr Ile Leu	Leu Cys Cys Met Thr	Ser Cys Cys Ser Cys	
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Leu Lys Gly Ala Cys	Ser Cys Gly Ser Cys	Cys Lys Phe Asp Glu	
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Asp Asp Ser Glu Pro	Val Leu Lys Gly Val	Lys Leu His Tyr Thr	
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<210> SEQ ID NO 3

<211> LENGTH: 1255

<212> TYPE: PRT

<213> ORGANISM: CORONAVIRUS

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His Thr Ser Ser Met Arg Gly Val Tyr Tyr Pro Asp Glu Ile Phe Arg
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50 55 60
Asn Val Thr Gly Phe His Thr Ile Asn His Thr Phe Gly Asn Pro Val
65 70 75 80
Ile Pro Phe Lys Asp Gly Ile Tyr Phe Ala Ala Thr Glu Lys Ser Asn
85 90 95
Val Val Arg Gly Trp Val Phe Gly Ser Thr Met Asn Asn Lys Ser Gln
100 105 110
Ser Val Ile Ile Ile Asn Asn Ser Thr Asn Val Val Ile Arg Ala Cys
115 120 125
Asn Phe Glu Leu Cys Asp Asn Pro Phe Phe Ala Val Ser Lys Pro Met
130 135 140
Gly Thr Gln Thr His Thr Met Ile Phe Asp Asn Ala Phe Asn Cys Thr
145 150 155 160
Phe Glu Tyr Ile Ser Asp Ala Phe Ser Leu Asp Val Ser Glu Lys Ser
165 170 175
Gly Asn Phe Lys His Leu Arg Glu Phe Val Phe Lys Asn Lys Asp Gly
180 185 190
Phe Leu Tyr Val Tyr Lys Gly Tyr Gln Pro Ile Asp Val Val Arg Asp
195 200 205
Leu Pro Ser Gly Phe Asn Thr Leu Lys Pro Ile Phe Lys Leu Pro Leu
210 215 220
Gly Ile Asn Ile Thr Asn Phe Arg Ala Ile Leu Thr Ala Phe Ser Pro
225 230 235 240
Ala Gln Asp Ile Trp Gly Thr Ser Ala Ala Ala Tyr Phe Val Gly Tyr
245 250 255
Leu Lys Pro Thr Thr Phe Met Leu Lys Tyr Asp Glu Asn Gly Thr Ile
260 265 270
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275 280 285
Ser Val Lys Ser Phe Glu Ile Asp Lys Gly Ile Tyr Gln Thr Ser Asn
290 295 300
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Val Tyr Ala Trp Glu Arg Lys Lys Ile Ser Asn Cys Val Ala Asp Tyr
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Ser Val Leu Tyr Asn Ser Thr Phe Phe Ser Thr Phe Lys Cys Tyr Gly
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Asp Ser Phe Val Val Lys Gly Asp Asp Val Arg Gln Ile Ala Pro Gly

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Glu Asp Leu Leu Phe Asn Lys Val Thr Leu Ala Asp Ala Gly Phe Met
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Lys Gln Tyr Gly Glu Cys Leu Gly Asp Ile Asn Ala Arg Asp Leu Ile
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Cys Ala Gln Lys Phe Asn Gly Leu Thr Val Leu Pro Pro Leu Leu Thr
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Asp Asp Met Ile Ala Ala Tyr Thr Ala Ala Leu Val Ser Gly Thr Ala
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Thr Ala Gly Trp Thr Phe Gly Ala Gly Ala Ala Leu Gln Ile Pro Phe
 865 870 875 880

Ala Met Gln Met Ala Tyr Arg Phe Asn Gly Ile Gly Val Thr Gln Asn
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Val Leu Tyr Glu Asn Gln Lys Gln Ile Ala Asn Gln Phe Asn Lys Ala
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Ile Ser Gln Ile Gln Glu Ser Leu Thr Thr Thr Ser Thr Ala Leu Gly
 915 920 925

Lys Leu Gln Asp Val Val Asn Gln Asn Ala Gln Ala Leu Asn Thr Leu
 930 935 940

Val Lys Gln Leu Ser Ser Asn Phe Gly Ala Ile Ser Ser Val Leu Asn
 945 950 955 960

Asp Ile Leu Ser Arg Leu Asp Lys Val Glu Ala Glu Val Gln Ile Asp
 965 970 975

Arg Leu Ile Thr Gly Arg Leu Gln Ser Leu Gln Thr Tyr Val Thr Gln
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Gln Leu Ile Arg Ala Ala Glu Ile Arg Ala Ser Ala Asn Leu Ala Ala
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Thr Lys Met Ser Glu Cys Val Leu Gly Gln Ser Lys Arg Val Asp
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Phe Cys Gly Lys Gly Tyr His Leu Met Ser Phe Pro Gln Ala Ala
 1025 1030 1035

Pro His Gly Val Val Phe Leu His Val Thr Tyr Val Pro Ser Gln
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Glu Arg Asn Phe Thr Thr Ala Pro Ala Ile Cys His Glu Gly Lys
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Trp Phe Ile Thr Gln Arg Asn Phe Phe Ser Pro Gln Ile Ile Thr
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Thr Asp Asn Thr Phe Val Ser Gly Asn Cys Asp Val Val Ile Gly
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Ile Ile Asn Asn Thr Val Tyr Asp Pro Leu Gln Pro Glu Leu Asp
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Ser Phe Lys Glu Glu Leu Asp Lys Tyr Phe Lys Asn His Thr Ser
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Pro Asp Val Asp Leu Gly Asp Ile Ser Gly Ile Asn Ala Ser Val
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Val Asn Ile Gln Lys Glu Ile Asp Arg Leu Asn Glu Val Ala Lys
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Ala Gly Leu Ile Ala Ile Val Met Val Thr Ile Leu Leu Cys Cys
 1205 1210 1215

Met Thr Ser Cys Cys Ser Cys Leu Lys Gly Ala Cys Ser Cys Gly
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<210> SEQ ID NO 4

<211> LENGTH: 3943

<212> TYPE: DNA

<213> ORGANISM: CORONAVIRUS

<400> SEQUENCE: 4

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gtggttcttg ctgcaagttt gatgaggatg actctgagcc agttctcaag ggtgtcaaat 3840
tacattacac ataaacgaac ttatggattt gtttatgaga ttttttactc ttggatcaat 3900

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 tactgcacag ccagtaaaaa ttgacaatgc ttctcctgca agt 3943

<210> SEQ ID NO 5

<211> LENGTH: 2049

<212> TYPE: DNA

<213> ORGANISM: CORONAVIRUS

<400> SEQUENCE: 5

ctcttctgga aaaaggtagg cttatcatta gagaaaacaa cagagttgtg gtttcaagtg 60
 atattcttgt taacaactaa acgaacatgt ttattttcctt attatttctt actctcacta 120
 gtggtagtga ccttgaccgg tgcaccactt ttgatgatgt tcaagctcct aattacactc 180
 aacatacttc atctatgagg ggggtttact atcctgatga aatttttaga tcagacactc 240
 tttatttaac tcaggattta tttcttccat tttattctaa tggtagcaggg tttcactacta 300
 ttaatcatac gtttgccaac cctgtcatac cttttaagga tggattttat tttgctgcca 360
 cagagaaatc aaatgttgtc cgtgggtggg tttttggttc taccatgaac aacaagtcac 420
 agtcggtgat tattattaac aattctacta atgttggtat acgagcatgt aactttgaat 480
 tegtgtgacaa ccctttcttt gctgtttcta aacctatggg tacacagaca catactatga 540
 tattcgataa tgcatttaat tgcactttcg agtacatatac tgatgccttt tcgcttgatg 600
 tttcagaaaa gtcaggtaat tttaaacact tacgagagtt tegtgtttaa aataaagatg 660
 ggtttctcta tgtttataag ggctatcaac ctatagatgt agttcgtgat ctaccttctg 720
 gttttaacac tttgaaacct atttttaagt tgcctcttgg tattaacatt acaaatttta 780
 gagccattct tacagccttt tcacctgctc aagacatttg gggcacgtca gctgcagcct 840
 attttgttg ctattttaa ccaactacat ttatgctcaa gtatgatgaa aatggtacaa 900
 tcacagatgc tgttgattgt tctcaaaatc cacttgctga actcaaatgc tctgttaaga 960
 gctttgagat tgacaaagga atttaccaga cctctaattt cagggttggt ccctcaggag 1020
 atgttgtgag attccctaatt attacaaact tgtgtccttt tggagagggt tttaatgcta 1080
 ctaaattccc ttctgtctat gcatgggaga gaaaaaaaaa ttctaattgt gttgctgatt 1140
 actctgtgct ctacaactca acattttttt caacctttaa gtgctatggc gtttctgcca 1200
 ctaagttgaa tgatctttgc ttctccaatg tctatgcaga ttcttttga gtcaggggag 1260
 atgatgtaag acaaatagcg ccaggacaaa ctggtgttat tgctgattat aattataaat 1320
 tgccagatga tttcatgggt tgtgtccttg cttggaatac taggaacatt gatgctactt 1380
 caactggtaa ttataattat aaatataggt atcttagaca tggcaagcct aggccttttg 1440
 agagagacat atctaattgt cctttctccc ctgatggcaa accttgcacc ccacctgctc 1500
 ttaattgtta ttggcatta aatgattatg gtttttacac cactactggc attggctacc 1560
 aaccttacag agttgtagta ctttcttttg aactttttaa tgcaccggcc acggtttgtg 1620
 gaccaaaatt atccactgac cttattaaga accagtgtgt caattttaat tttaatggac 1680
 tcaactgtac tgggtgtgta actccttctt caaagagatt tcaaccattt caacaatttg 1740
 gccgtgatgt ctctgatttc actgattccg ttcgagatcc taaaacatct gaaatattag 1800
 acatttcacc ttgctctttt ggggtgtgaa gtgtaattac acctggaaca atgcttcat 1860
 ctgaagttgc tgttctatat caagatgta actgcactga tgtttctaca gcaatccatg 1920
 cagatcaact cacaccagct tggcgcatac attctactgg aaacaatgta ttccagactc 1980

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aagcaggctg tcttatagga gctgagcatg tcgacacttc ttatgagtgc gacattccta 2040
ttggagctg 2049

<210> SEQ ID NO 6
<211> LENGTH: 2027
<212> TYPE: DNA
<213> ORGANISM: CORONAVIRUS

<400> SEQUENCE: 6
catgcagatc aactcacacc agcttggcgc atatattcta ctggaaacaa tgtattccag 60
actcaagcag gctgtcttat aggagctgag catgtcgaca cttcttatga gtgcgacatt 120
cctattggag ctggcatttg tgctagttac catacagttt ctttattacg tagtactagc 180
caaaaatcta ttgtggctta tactatgtct ttagggtgctg atagttcaat tgcttactct 240
aataacacca ttgctatacc tactaacttt tcaattagca ttactacaga agtaatgcct 300
gtttctatgg ctaaacctc cgtagattgt aatatgtaca tctgcgaga ttctactgaa 360
tgtgctaatt tgcttctcca atatggtagc ttttgcacac aactaaatcg tgcactctca 420
ggatttgctg ctgaacagga tcgcaacaca cgtgaagtgt tcgctcaagt caaacaatg 480
tacaaaaccc caactttgaa atattttggt ggttttaatt tttcacaat attacctgac 540
cctctaaagc caactaagag gtcttttatt gaggacttgc tctttaataa ggtgacactc 600
gctgatgctg gttctatgaa gcaatatggc gaatgcctag gtgatattaa tgctagagat 660
ctcatttgty cgagaagtt caatgggctt acagtgttgc cacctctgct cactgatgat 720
atgattgctg cctacactgc tgctctagtt agtggtagct ccactgctgg atggacattt 780
gggtgctggc ctgctcttca aatacctttt gctatgcaaa tggcatatag gttcaatggc 840
attggagtta cccaaaatgt tctctatgag aacccaaaac aaatcgccaa ccaatttaac 900
aaggcgatta gtcaaatca agaatcactt acaacaacat caactgcatt gggcaagctg 960
caagacgttg ttaaccagaa tgctcaagca ttaaacacac ttgttaaaca acttagctct 1020
aatTTTggtg caatttcaag tTgTctaaat gatatccttt cgcgacttga taaagtcgag 1080
gcgaggtac aaattgacag gTtaattaca ggcagacttc aaagccttca aacctatgta 1140
acacaacaac taatcagggc Tgctgaaatc agggcttctg ctaatcttgc Tgctactaaa 1200
atgtctgagt TgTtcttTg acaatcaaaa agagttgact tTtTgTgaaa gggctaccac 1260
cttatgtcct tcccacaagc agcccgcgat ggtgttgtct tcctacatgt cacgtatgtg 1320
ccatcccag agaggaactt caccacagcg ccagcaattt gTcatgaagg caaagcatac 1380
ttccctcgtg aaggTgtTtt TgtgtTtaat ggcacttctt ggtttattac acagaggaac 1440
ttcttttctc cacaaataat tactacagac aatacatttg tctcaggaaa ttgtgatgtc 1500
gttattggcg tcattaacaa cacagtttat gatcctctgc aacctgagct Tgactcattc 1560
aaagaagagc Tggacaagta cttcaaaaat catacatcac cagatgttga tctTgTcgac 1620
atttcaggca ttaacgcttc Tgtcgtcaac attcaaaaag aaattgaccg cctcaatgag 1680
gtcgtcaaaa atttaaatga atcactcatt gaccttcaag aattgggaaa atatgagcaa 1740
tatattaaat ggccttggtg Tgtttggctc ggcttcattg ctggactaat Tgccatcgtc 1800
atggttacaa tcttgctttg ttgcatgact agttgttgca gttgcctcaa gggTgcatgc 1860
tctTgtggtt cttgctgcaa gttTgatgag gatgactctg agccagtTct caagggtgTc 1920

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aaattacatt acacataaac gaacttatgg atttgtttat gagatTTTTT actcttggat 1980
caattactgc acagccagta aaaattgaca atgcttctcc tgcaagt 2027

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<210> SEQ ID NO 7
<211> LENGTH: 1096
<212> TYPE: DNA
<213> ORGANISM: CORONAVIRUS

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<400> SEQUENCE: 7

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tcttgctttg ttgcatgact agttgttgca gttgcctcaa gggtgcatgc tcttgtgggt 60
cttgcctgca gtttgatgag gatgactctg agccagttct caaggggtgc aaattacatt 120
acacataaac gaacttatgg atttgtttat gagatTTTTT actcttggat caattactgc 180
acagccagta aaaattgaca atgcttctcc tgcaagtact gttcatgcta cagcaacgat 240
accgctacaa gcctcactcc ctttcggatg gcttgttatt ggcgttgcat ttcttgctgt 300
ttttcagagc gctacaaaa taattgcgct caataaaaaga tggcagctag ccctttataa 360
gggcttccag ttcatttgca atttactgct gctatttggg accatctatt cacatctttt 420
gcttgcctgc gcaggtatgg aggcgcaatt tttgtacctc tatgcctga tatatcttct 480
acaatgcatc aacgcatgta gaattattat gagatgttgg ctttgttggg agtgcaaatc 540
caagaaccca ttactttatg atgccaacta ctttgttggc tggcacacac ataactatga 600
ctactgtata ccatataaca gtgtcacaga tacaattgct gttactgaag gtgacggcat 660
ttcaacacca aaactcaaag aagactacca aattgggtgg tattctgagg atagccactc 720
aggtgttaaa gactatgtcg ttgtacatgg ctatttcacc gaagtttact accagcttga 780
gtctacacaa attactacag aactgtgat tgaaaatgct acattcttca tctttaacaa 840
gcttgttaaa gaccaccga atgtgcaaat acacacaatc gacggctctt caggagtgtc 900
taatccagca atggatccaa tttatgatga gccgacgacg actactagcg tgcctttgta 960
agcacaagaa agtgagtacg aacttatgta ctcatctggt tcggaagaaa caggtagctt 1020
aatagttaat agcgtacttc ttttcttgc tttcgtggta ttcttgctag tcacactagc 1080
catccttact gcgctt 1096

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<210> SEQ ID NO 8
<211> LENGTH: 1135
<212> TYPE: DNA
<213> ORGANISM: CORONAVIRUS

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<400> SEQUENCE: 8

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attgccatcg tcatggttac aatcttgctt tgttgcatga ctagtgttg cagttgcctc 60
aagggtgcat gctcttggg ttcttgcctg aagtttgatg aggatgactc tgagccagtt 120
ctcaaggggtg tcaattaca ttacacataa acgaacttat ggatttgttt atgagatttt 180
ttactcttgg atcaattact gcacagccag taaaaattga caatgcttct cctgcaagta 240
ctgttcatgc tacagcaacg ataccgctac aagcctcact ccctttcgga tggcttgtta 300
ttggcgttgc atttcttgcg gttttcaga gogctaccaa aataattgcg ctcaataaaa 360
gatggcagct agccctttat aagggtctcc agttcatttg caatttactg ctgctatttg 420
ttaccatcta ttcacatctt ttgcttgcg ctgcaggtat ggaggcga tttttgtacc 480
tctatgcctt gatataatct ctacaatgca tcaacgcatg tagaattatt atgagatgtt 540

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ggctttggtg gaagtgcaaa tccaagaacc cactacttta tgatgccaac tactttggtt 600
gctggcacac acataactat gactactgta taccatataa cagtgtcaca gatacaattg 660
tcgttactga aggtgacggc atttcaacac caaaactcaa agaagactac caaattggtg 720
gttattctga ggataggcac tcagggtgta aagactatgt cgttgtacat ggctatttca 780
ccgaagtta ctaccagctt gagtctacac aaattactac agacactggt attgaaaatg 840
ctacattctt catctttaac aagcttgta aagaccacc gaatgtgcaa atacacacaa 900
tcgacggctc ttcaggagtt gctaataccag caatggatcc aatttatgat gagccgacga 960
cgactactag cgtgcctttg taagcacaag aaagtgagta cgaacttatg tactcattcg 1020
tttcggaaga aacaggtacg ttaatagta atagcgtact tctttttctt gctttcgtgg 1080
tattcttgct agtcacacta gccatcctta ctgcgcttcg attgtgtgcg tactg 1135

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<210> SEQ ID NO 9
<211> LENGTH: 1096
<212> TYPE: DNA
<213> ORGANISM: CORONAVIRUS
<220> FEATURE:
<221> NAME/KEY: CDS
<222> LOCATION: (137)..(958)
<223> OTHER INFORMATION:

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<400> SEQUENCE: 9

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cttgctgcaa gtttgatgag gatgactctg agccagttct caagggtgtc aaattacatt 120
acacataaac gaactt atg gat ttg ttt atg aga ttt ttt act ctt gga tca 172
      Met Asp Leu Phe Met Arg Phe Phe Thr Leu Gly Ser
      1             5             10
att act gca cag cca gta aaa att gac aat gct tct cct gca agt act 220
Ile Thr Ala Gln Pro Val Lys Ile Asp Asn Ala Ser Pro Ala Ser Thr
      15             20             25
gtt cat gct aca gca acg ata ccg cta caa gcc tca ctc cct ttc gga 268
Val His Ala Thr Ala Thr Ile Pro Leu Gln Ala Ser Leu Pro Phe Gly
      30             35             40
tgg ctt gtt att ggc gtt gca ttt ctt gct gtt ttt cag agc gct acc 316
Trp Leu Val Ile Gly Val Ala Phe Leu Ala Val Phe Gln Ser Ala Thr
      45             50             55             60
aaa ata att gcg ctc aat aaa aga tgg cag cta gcc ctt tat aag ggc 364
Lys Ile Ile Ala Leu Asn Lys Arg Trp Gln Leu Ala Leu Tyr Lys Gly
      65             70             75
ttc cag ttc att tgc aat tta ctg ctg cta ttt gtt acc atc tat tca 412
Phe Gln Phe Ile Cys Asn Leu Leu Leu Leu Phe Val Thr Ile Tyr Ser
      80             85             90
cat ctt ttg ctt gtc gct gca ggt atg gag gcg caa ttt ttg tac ctc 460
His Leu Leu Leu Val Ala Ala Gly Met Glu Ala Gln Phe Leu Tyr Leu
      95             100             105
tat gcc ttg ata tat ttt cta caa tgc atc aac gca tgt aga att att 508
Tyr Ala Leu Ile Tyr Phe Leu Gln Cys Ile Asn Ala Cys Arg Ile Ile
      110             115             120
atg aga tgt tgg ctt tgt tgg aag tgc aaa tcc aag aac cca tta ctt 556
Met Arg Cys Trp Leu Cys Trp Lys Cys Lys Ser Lys Asn Pro Leu Leu
      125             130             135             140
tat gat gcc aac tac ttt gtt tgc tgg cac aca cat aac tat gac tac 604
Tyr Asp Ala Asn Tyr Phe Val Cys Trp His Thr His Asn Tyr Asp Tyr
      145             150             155

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tgt ata cca tat aac agt gtc aca gat aca att gtc gtt act gaa ggt	652
Cys Ile Pro Tyr Asn Ser Val Thr Asp Thr Ile Val Val Thr Glu Gly	
160 165 170	
gac ggc att tca aca cca aaa ctc aaa gaa gac tac caa att ggt ggt	700
Asp Gly Ile Ser Thr Pro Lys Leu Lys Glu Asp Tyr Gln Ile Gly Gly	
175 180 185	
tat tct gag gat agg cac tca ggt gtt aaa gac tat gtc gtt gta cat	748
Tyr Ser Glu Asp Arg His Ser Gly Val Lys Asp Tyr Val Val Val His	
190 195 200	
ggc tat ttc acc gaa gtt tac tac cag ctt gag tct aca caa att act	796
Gly Tyr Phe Thr Glu Val Tyr Tyr Gln Leu Glu Ser Thr Gln Ile Thr	
205 210 215 220	
aca gac act ggt att gaa aat gct aca ttc ttc atc ttt aac aag ctt	844
Thr Asp Thr Gly Ile Glu Asn Ala Thr Phe Phe Ile Phe Asn Lys Leu	
225 230 235	
gtt aaa gac cca ccg aat gtg caa ata cac aca atc gac ggc tct tca	892
Val Lys Asp Pro Pro Asn Val Gln Ile His Thr Ile Asp Gly Ser Ser	
240 245 250	
gga gtt gct aat cca gca atg gat cca att tat gat gag ccg acg acg	940
Gly Val Ala Asn Pro Ala Met Asp Pro Ile Tyr Asp Glu Pro Thr Thr	
255 260 265	
act act agc gtg cct ttg taagcacaag aaagtgagta cgaacttatg	988
Thr Thr Ser Val Pro Leu	
270	
tactcattcg tttcgaaga aacaggtacg ttaatagtta atagcgtact tctttttctt	1048
gctttcgtgg tattcttgct agtcacacta gccatcctta ctgcgctt	1096

<210> SEQ ID NO 10

<211> LENGTH: 274

<212> TYPE: PRT

<213> ORGANISM: CORONAVIRUS

<400> SEQUENCE: 10

Met Asp Leu Phe Met Arg Phe Phe Thr Leu Gly Ser Ile Thr Ala Gln	
1 5 10 15	
Pro Val Lys Ile Asp Asn Ala Ser Pro Ala Ser Thr Val His Ala Thr	
20 25 30	
Ala Thr Ile Pro Leu Gln Ala Ser Leu Pro Phe Gly Trp Leu Val Ile	
35 40 45	
Gly Val Ala Phe Leu Ala Val Phe Gln Ser Ala Thr Lys Ile Ile Ala	
50 55 60	
Leu Asn Lys Arg Trp Gln Leu Ala Leu Tyr Lys Gly Phe Gln Phe Ile	
65 70 75 80	
Cys Asn Leu Leu Leu Leu Phe Val Thr Ile Tyr Ser His Leu Leu Leu	
85 90 95	
Val Ala Ala Gly Met Glu Ala Gln Phe Leu Tyr Leu Tyr Ala Leu Ile	
100 105 110	
Tyr Phe Leu Gln Cys Ile Asn Ala Cys Arg Ile Ile Met Arg Cys Trp	
115 120 125	
Leu Cys Trp Lys Cys Lys Ser Lys Asn Pro Leu Leu Tyr Asp Ala Asn	
130 135 140	
Tyr Phe Val Cys Trp His Thr His Asn Tyr Asp Tyr Cys Ile Pro Tyr	
145 150 155 160	
Asn Ser Val Thr Asp Thr Ile Val Val Thr Glu Gly Asp Gly Ile Ser	

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	165		170		175	
Thr Pro Lys	Leu Lys Glu Asp Tyr Gln Ile Gly Gly Tyr Ser Glu Asp					
	180		185		190	
Arg His Ser	Gly Val Lys Asp Tyr Val Val Val His Gly Tyr Phe Thr					
	195		200		205	
Glu Val Tyr	Tyr Gln Leu Glu Ser Thr Gln Ile Thr Thr Asp Thr Gly					
	210		215		220	
Ile Glu Asn	Ala Thr Phe Phe Ile Phe Asn Lys Leu Val Lys Asp Pro					
	225		230		235	240
Pro Asn Val	Gln Ile His Thr Ile Asp Gly Ser Ser Gly Val Ala Asn					
		245		250		255
Pro Ala Met	Asp Pro Ile Tyr Asp Glu Pro Thr Thr Thr Thr Ser Val					
	260		265		270	
Pro Leu						
<210> SEQ ID NO 11						
<211> LENGTH: 1096						
<212> TYPE: DNA						
<213> ORGANISM: CORONAVIRUS						
<220> FEATURE:						
<221> NAME/KEY: CDS						
<222> LOCATION: (558)..(1019)						
<223> OTHER INFORMATION:						
<400> SEQUENCE: 11						
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acacataaac	gaacttatgg	at ttgtttat	gagat ttttt	actcttgat	caattactgc	180
acagccagta	aaaattgaca	atgcttctcc	tgcaagtact	gttcatgcta	cagcaacgat	240
accgctacaa	gcctcactcc	ctttcggatg	gcttgttatt	ggcgttgcat	ttcttgctgt	300
ttttcagagc	gctacaaaa	taattgcgct	caataaaaaga	tggcagctag	ccctttataa	360
gggcttccag	ttcatttgca	at ttactgct	gctat ttgtt	accatctatt	cacatctttt	420
gcttgctgct	gcaggtatgg	aggcgcaatt	tttgtacctc	tatgccttga	tatattttct	480
acaatgcatc	aacgcatgta	gaattattat	gagatg ttgg	ctttgttgga	agtgcaaatc	540
caagaaccca	ttactttt	atg atg oca	act act ttg	ttt gct ggc	aca cac	590
	Met Met Pro	Thr Thr Leu	Phe Ala Gly	Thr His		
	1		5		10	
ata act atg	act act gta	tac cat ata	aca gtg tca	cag ata caa	ttg	638
Ile Thr Met	Thr Thr Val	Tyr His Ile	Thr Val Ser	Gln Ile Gln	Leu	
	15		20		25	
tcg tta ctg	aag gtg acg	gca ttt caa	cac caa aac	tca aag aag	act	686
Ser Leu Leu	Lys Val Thr	Ala Phe Gln	His Gln Asn	Ser Lys Lys	Thr	
	30		35		40	
acc aaa ttg	gtg gtt att	ctg agg ata	ggc act cag	gtg tta aag	act	734
Thr Lys Leu	Val Val Ile	Leu Arg Ile	Gly Thr Gln	Val Leu Lys	Thr	
	45		50		55	
atg tcg ttg	tac atg gct	att tca ccg	aag ttt act	acc agc ttg	agt	782
Met Ser Leu	Tyr Met Ala	Ile Ser Pro	Lys Phe Thr	Thr Ser Leu	Ser	
	60		65		70	75
cta cac aaa	tta cta cag	aca ctg gta	ttg aaa atg	cta cat tct	tca	830
Leu His Lys	Leu Leu Gln	Thr Leu Val	Leu Lys Met	Leu His Ser	Ser	
	80		85		90	

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tct tta aca agc ttg tta aag acc cac cga atg tgc aaa tac aca caa      878
Ser Leu Thr Ser Leu Leu Lys Thr His Arg Met Cys Lys Tyr Thr Gln
          95                      100                      105

tcg acg gct ctt cag gag ttg cta atc cag caa tgg atc caa ttt atg      926
Ser Thr Ala Leu Gln Glu Leu Leu Ile Gln Gln Trp Ile Gln Phe Met
          110                      115                      120

atg agc cga cga cga cta cta gcg tgc ctt tgt aag cac aag aaa gtg      974
Met Ser Arg Arg Arg Leu Leu Ala Cys Leu Cys Lys His Lys Lys Val
          125                      130                      135

agt acg aac tta tgt act cat tcg ttt cgg aag aaa cag gta cgt      1019
Ser Thr Asn Leu Cys Thr His Ser Phe Arg Lys Lys Gln Val Arg
          140                      145                      150

taatagttaa tagcgtactt ctttttcttg ctttcgtggg attccttgcta gtcacactag  1079

ccatccttac tgcgctt                                          1096

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<210> SEQ ID NO 12
<211> LENGTH: 154
<212> TYPE: PRT
<213> ORGANISM: CORONAVIRUS

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<400> SEQUENCE: 12

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Met Met Pro Thr Thr Leu Phe Ala Gly Thr His Ile Thr Met Thr Thr
1          5          10          15

Val Tyr His Ile Thr Val Ser Gln Ile Gln Leu Ser Leu Leu Lys Val
20        25        30

Thr Ala Phe Gln His Gln Asn Ser Lys Lys Thr Thr Lys Leu Val Val
35        40        45

Ile Leu Arg Ile Gly Thr Gln Val Leu Lys Thr Met Ser Leu Tyr Met
50        55        60

Ala Ile Ser Pro Lys Phe Thr Thr Ser Leu Ser Leu His Lys Leu Leu
65        70        75        80

Gln Thr Leu Val Leu Lys Met Leu His Ser Ser Ser Leu Thr Ser Leu
85        90        95

Leu Lys Thr His Arg Met Cys Lys Tyr Thr Gln Ser Thr Ala Leu Gln
100       105       110

Glu Leu Leu Ile Gln Gln Trp Ile Gln Phe Met Met Ser Arg Arg Arg
115       120       125

Leu Leu Ala Cys Leu Cys Lys His Lys Lys Val Ser Thr Asn Leu Cys
130       135       140

Thr His Ser Phe Arg Lys Lys Gln Val Arg
145       150

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<210> SEQ ID NO 13
<211> LENGTH: 332
<212> TYPE: DNA
<213> ORGANISM: CORONAVIRUS
<220> FEATURE:
<221> NAME/KEY: CDS
<222> LOCATION: (36)..(263)
<223> OTHER INFORMATION:

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<400> SEQUENCE: 13

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tgcctttgta agcacaagaa agtgagtacg aactt atg tac tca ttc gtt tcg      53
          Met Tyr Ser Phe Val Ser
          1                      5

gaa gaa aca ggt acg tta ata gtt aat agc gta ctt ctt ttt ctt gct      101
Glu Glu Thr Gly Thr Leu Ile Val Asn Ser Val Leu Leu Phe Leu Ala

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10	15	20	
ttc gtg gta ttc ttg cta gtc aca cta gcc atc ctt act gcg ctt cga			149
Phe Val Val Phe Leu Leu Val Thr Leu Ala Ile Leu Thr Ala Leu Arg			
25	30	35	
ttg tgt gcg tac tgc tgc aat att gtt aac gtg agt tta gta aaa cca			197
Leu Cys Ala Tyr Cys Cys Asn Ile Val Asn Val Ser Leu Val Lys Pro			
40	45	50	
acg gtt tac gtc tac tcg cgt gtt aaa aat ctg aac tct tct gaa gga			245
Thr Val Tyr Val Tyr Ser Arg Val Lys Asn Leu Asn Ser Ser Glu Gly			
55	60	65	70
gtt cct gat ctt ctg gtc taaacgaact aactattatt attattctgt			293
Val Pro Asp Leu Leu Val			
75			
ttggaacttt aacattgctt atcatggcag acaacggta			332
<210> SEQ ID NO 14			
<211> LENGTH: 76			
<212> TYPE: PRT			
<213> ORGANISM: CORONAVIRUS			
<400> SEQUENCE: 14			
Met Tyr Ser Phe Val Ser Glu Glu Thr Gly Thr Leu Ile Val Asn Ser			
1	5	10	15
Val Leu Leu Phe Leu Ala Phe Val Val Phe Leu Leu Val Thr Leu Ala			
20	25	30	
Ile Leu Thr Ala Leu Arg Leu Cys Ala Tyr Cys Cys Asn Ile Val Asn			
35	40	45	
Val Ser Leu Val Lys Pro Thr Val Tyr Val Tyr Ser Arg Val Lys Asn			
50	55	60	
Leu Asn Ser Ser Glu Gly Val Pro Asp Leu Leu Val			
65	70	75	
<210> SEQ ID NO 15			
<211> LENGTH: 332			
<212> TYPE: DNA			
<213> ORGANISM: CORONAVIRUS			
<400> SEQUENCE: 15			
tgccctttgta agcacaagaa agtgagtacg aacttatgta ctcattcgtt tcggaagaaa			60
caggtacggtt aatagttaat agcgtacttc ttttcttgc tttcgtggta ttcttgctag			120
tcacactagc catccttact gcgcttcgat tgtgtgcgta ctgctgcaat attgttaacg			180
tgagtttagt aaaaccaacg gtttaagtct actcgcgtgt taaaaatctg aactcttctg			240
aaggagtcc tgatcttctg gtctaacaaga actaactatt attattattc tgtttggaac			300
tttaacattg cttatcatgg cagacaacgg ta			332
<210> SEQ ID NO 16			
<211> LENGTH: 708			
<212> TYPE: DNA			
<213> ORGANISM: CORONAVIRUS			
<220> FEATURE:			
<221> NAME/KEY: CDS			
<222> LOCATION: (41)..(703)			
<223> OTHER INFORMATION:			
<400> SEQUENCE: 16			
tattattatt attctgtttg gaactttaac attgcttacc atg gca gac aac ggt			55

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	Met	Ala	Asp	Asn	Gly	
	1				5	
act att acc gtt gag gag ctt aaa caa ctc ctg gaa caa tgg aac cta						103
Thr Ile Thr Val Glu Glu Leu Lys Gln Leu Leu Glu Gln Trp Asn Leu						
	10				20	
gta ata ggt ttc cta ttc cta gcc tgg att atg tta cta caa ttt gcc						151
Val Ile Gly Phe Leu Phe Leu Ala Trp Ile Met Leu Leu Gln Phe Ala						
	25				30	35
tat tct aat cgg aac agg ttt ttg tac ata ata aag ctt gtt ttc ctc						199
Tyr Ser Asn Arg Asn Arg Phe Leu Tyr Ile Ile Lys Leu Val Phe Leu						
	40			45		50
tgg ctc ttg tgg cca gta aca ctt gct tgt ttt gtg ctt gct gct gtc						247
Trp Leu Leu Trp Pro Val Thr Leu Ala Cys Phe Val Leu Ala Ala Val						
	55			60		65
tac aga att aat tgg gtg act ggc ggg att gcg att gca atg gct tgt						295
Tyr Arg Ile Asn Trp Val Thr Gly Gly Ile Ala Ile Ala Met Ala Cys						
	70			75		80
att gta ggc ttg atg tgg ctt agc tac ttc gtt gct tcc ttc agg ctg						343
Ile Val Gly Leu Met Trp Leu Ser Tyr Phe Val Ala Ser Phe Arg Leu						
		90			95	100
ttt gct cgt acc cgc tca atg tgg tca ttc aac cca gaa aca aac att						391
Phe Ala Arg Thr Arg Ser Met Trp Ser Phe Asn Pro Glu Thr Asn Ile						
	105			110		115
ctt ctc aat gtg cct ctc cgg ggg aca att gtg acc aga ccg ctc atg						439
Leu Leu Asn Val Pro Leu Arg Gly Thr Ile Val Thr Arg Pro Leu Met						
	120			125		130
gaa agt gaa ctt gtc att ggt gct gtg atc att cgt ggt cac ttg cga						487
Glu Ser Glu Leu Val Ile Gly Ala Val Ile Ile Arg Gly His Leu Arg						
	135			140		145
atg gcc gga cac tcc cta ggg cgc tgt gac att aag gac ctg cca aaa						535
Met Ala Gly His Ser Leu Gly Arg Cys Asp Ile Lys Asp Leu Pro Lys						
	150			155		160
gag atc act gtg gct aca tca cga acg ctt tct tat tac aaa tta gga						583
Glu Ile Thr Val Ala Thr Ser Arg Thr Leu Ser Tyr Tyr Lys Leu Gly						
	170				175	180
gcg tcg cag cgt gta ggc act gat tca ggt ttt gct gca tac aac cgc						631
Ala Ser Gln Arg Val Gly Thr Asp Ser Gly Phe Ala Ala Tyr Asn Arg						
	185			190		195
tac cgt att gga aac tat aaa tta aat aca gac cac gcc ggt agc aac						679
Tyr Arg Ile Gly Asn Tyr Lys Leu Asn Thr Asp His Ala Gly Ser Asn						
	200			205		210
gac aat att gct ttg cta gta cag taagt						708
Asp Asn Ile Ala Leu Leu Val Gln						
	215			220		

<210> SEQ ID NO 17

<211> LENGTH: 221

<212> TYPE: PRT

<213> ORGANISM: CORONAVIRUS

<400> SEQUENCE: 17

Met	Ala	Asp	Asn	Gly	Thr	Ile	Thr	Val	Glu	Glu	Leu	Lys	Gln	Leu	Leu
1				5					10					15	
Glu	Gln	Trp	Asn	Leu	Val	Ile	Gly	Phe	Leu	Phe	Leu	Ala	Trp	Ile	Met
			20				25						30		
Leu	Leu	Gln	Phe	Ala	Tyr	Ser	Asn	Arg	Asn	Arg	Phe	Leu	Tyr	Ile	Ile
		35					40				45				

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Lys Leu Val Phe Leu Trp Leu Leu Trp Pro Val Thr Leu Ala Cys Phe
 50 55 60

Val Leu Ala Ala Val Tyr Arg Ile Asn Trp Val Thr Gly Gly Ile Ala
 65 70 75 80

Ile Ala Met Ala Cys Ile Val Gly Leu Met Trp Leu Ser Tyr Phe Val
 85 90 95

Ala Ser Phe Arg Leu Phe Ala Arg Thr Arg Ser Met Trp Ser Phe Asn
 100 105 110

Pro Glu Thr Asn Ile Leu Leu Asn Val Pro Leu Arg Gly Thr Ile Val
 115 120 125

Thr Arg Pro Leu Met Glu Ser Glu Leu Val Ile Gly Ala Val Ile Ile
 130 135 140

Arg Gly His Leu Arg Met Ala Gly His Ser Leu Gly Arg Cys Asp Ile
 145 150 155 160

Lys Asp Leu Pro Lys Glu Ile Thr Val Ala Thr Ser Arg Thr Leu Ser
 165 170 175

Tyr Tyr Lys Leu Gly Ala Ser Gln Arg Val Gly Thr Asp Ser Gly Phe
 180 185 190

Ala Ala Tyr Asn Arg Tyr Arg Ile Gly Asn Tyr Lys Leu Asn Thr Asp
 195 200 205

His Ala Gly Ser Asn Asp Asn Ile Ala Leu Leu Val Gln
 210 215 220

<210> SEQ ID NO 18
 <211> LENGTH: 769
 <212> TYPE: DNA
 <213> ORGANISM: CORONAVIRUS

<400> SEQUENCE: 18

cctgatcttc tggctaaac gaactaacta ttattattat tctgtttga actttaacat 60
 tgcttatcat gccagacaac ggtactatta ccggtgagga gcttaaacia ctcttggaaac 120
 aatggaacct agtaaatggt ttcctattcc tagcctggat tatgttacta caatttgcct 180
 attctaactg gaacagggtt ttgtacataa taaagcttgt ttcctctgg ctcttgtggc 240
 cagtaaacct tgcttgtttt gtgcttgctg ctgtctacag aattaattgg gtgactggcg 300
 ggattgcatg tgcaatggct tgtattgtag gcttgatgtg gcttagctac ttcggttgcct 360
 ccttcaggct gtttgcctgt acccgctcaa tgtggtcatt caaccagaa acaaacattc 420
 ttctcaatgt gcctctccgg gggacaattg tgaccagacc gctcatggaa agtgaacttg 480
 tcattggtgc tgtgatcatt cgtggtcact tgcgaatggc cggacactcc ctagggcgct 540
 gtgacattaa ggacctgcca aaagagatca ctgtggctac atcacgaacg ctttcttatt 600
 acaaattagg agcgtgcgag cgtgtaggca ctgattcagg ttttgcgtca tacaaccgct 660
 accgtattgg aaactataaa ttaaatacag accacgccgg tagcaacgac aatattgctt 720
 tgctagtaca gtaagtgaca acagatgttt catcttgttg acttocagg 769

<210> SEQ ID NO 19
 <211> LENGTH: 1231
 <212> TYPE: DNA
 <213> ORGANISM: CORONAVIRUS

<400> SEQUENCE: 19

taccgtattg gaaactataa attaaataca gaccacgccg gttagcaaga caatattgct 60

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ttgctagtac agtaagtgac aacagatggt tcatcttggt gacttccagg ttacaatagc 120
agagatattg attatcatta tgaggacttt caggattgct atttggaaac ttgacgttat 180
aataagttca atagtggagac aattatthaa gcctctaact aagaagaatt attcggagtt 240
agatgatgaa gaacctatgg agttagatta tccataaaac gaacatgaaa attattctct 300
tcctgacatt gattgtattht acatcttgcg agctatatca ctatcaggag tgtgttagag 360
gtacgactgt actactaaaa gaaccttgcc catcaggaaac atacgagggc aattcaccat 420
ttcaccctct tgctgacaat aaatttgac taacttgac tagcacacac ttgctttttg 480
cttgctgta cggtaactga catacctatc agctgctgac aagatcagtt tcacaaaaac 540
ttttcatcag acaagaggag gttcaacaag agctctactc gccacttttt ctctattgtt 600
ctgctctagt attttataata ctttgcttca ccattaagag aaagacagaa tgaatgagct 660
cactttaatt gacttctatt tgtgcttttt agcctttctg ctattccttg ttttaataat 720
gcttattata ttttggtttt cactcgaaat ccaggatcta gaagaacctt gtaccaaagt 780
ctaaacgaac atgaaacttc tcattgtttt gacttgattt tctctatgca gttgcatatg 840
cactgtagta cagcgctgtg catctaataa acctcatgtg cttgaagatc cttgtaaggt 900
acaacactag gggtaatact tatagcactg cttggctttg tgctctagga aaggttttac 960
cttttcatag atggcacact atggttcaaa catgcacacc taatgttact atcaactgtc 1020
aagatccagc tgggtgtgag cttatagcta ggtgttgta ccttcatgaa ggtcaccaaa 1080
ctgctgcatt tagagacgta cttgtgtttt taaataaacg aacaaattaa aatgtctgat 1140
aatggacccc aatcaaacca acgtagtgcc ccccgatta catttgggtg acccacagat 1200
tcaactgaca ataaccagaa tggaggacgc a 1231

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<210> SEQ ID NO 20
<211> LENGTH: 1242
<212> TYPE: DNA
<213> ORGANISM: CORONAVIRUS

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<400> SEQUENCE: 20

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gcatacaacc gctaccggtat tggaaactat aaattaaata cagaccacgc cggtagcaac 60
gacaatattg ctttgctagt acagtaagt acaacagatg tttcatcttg ttgacttcca 120
ggttacaata gcagagatat tgattatcat tatgaggact ttcaggattg ctatttggaa 180
tcttgacggt ataataagtt caatagttag acagttattht aagcctctaa ctaagaagaa 240
ttattcggag ttagatgatg aagaacctat ggagtttagat tatccataaa acgaacatga 300
aaattattct cttcctgaca ttgattgtat ttacatcttg cgagctatat cactatcagg 360
agtgtgttag aggtacgact gtactactaa aagaaccttg cccatcagga acatcagagg 420
gcaattcacc atttcaccct cttgctgaca ataaatttgc actaacttgc actagcacac 480
actttgcttt tgcttgtgct gacggtaact gacataccta tcagctgctg gcaagatcag 540
tttcacaaa acttttcatc agacaagagg aggttcaaca agagctctac tcgccacttt 600
ttctcattgt tgctgctcta gtatttttaa tactttgctt caccattaag agaagacag 660
aatgaatgag ctcaactthaa ttgacttcta tttgtgcttt ttgaccttct tgctattcct 720
tgttttaata atgcttatta tttttgtgtt ttcactcgaac atccaggatc tagaagaacc 780
ttgtacaaa gtctaaacga acatgaaact tctcattgtt ttgacttcta tttctctatg 840

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cagttgcata tgcaactgtag tacagcgctg tgcatactaat aaacctcatg tgcttgaaga 900
tccttgtaag gtacaacact aggggtaata cttatagcac tgcttggtt tgtgctctag 960
gaaaggtttt accttttcat agatggcaca ctatggttca aacatgcaca cctaattgta 1020
ctatcaactg tcaagatcca gctgggtggtg cgcttatagc taggtgttgg taccttcatg 1080
aaggtcacca aactgctgca tttagagacg tacttgttgt tttaaataaa cgaacgaatt 1140
aaaatgtctg ataatggacc ccaatcaaac caacgtagtg ccccccgcac tacatttggg 1200
ggacccacag attcaactga caataaccag aatggaggac gc 1242

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<210> SEQ ID NO 21
<211> LENGTH: 1231
<212> TYPE: DNA
<213> ORGANISM: CORONAVIRUS
<220> FEATURE:
<221> NAME/KEY: CDS
<222> LOCATION: (86)..(274)
<223> OTHER INFORMATION:

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<400> SEQUENCE: 21

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taccgtattg gaaactataa attaaataca gaccacgccg gtagcaacga caatattgct 60
ttgctagtac agtaagtgac aacag atg ttt cat ctt gtt gac ttc cag gtt 112
Met Phe His Leu Val Asp Phe Gln Val
1 5
aca ata gca gag ata ttg att atc att atg agg act ttc agg att gct 160
Thr Ile Ala Glu Ile Leu Ile Ile Ile Met Arg Thr Phe Arg Ile Ala
10 15 20 25
att tgg aat ctt gac gtt ata ata agt tca ata gtg aga caa tta ttt 208
Ile Trp Asn Leu Asp Val Ile Ile Ser Ser Ile Val Arg Gln Leu Phe
30 35 40
aag cct cta act aag aag aat tat tcg gag tta gat gat gaa gaa cct 256
Lys Pro Leu Thr Lys Lys Asn Tyr Ser Glu Leu Asp Asp Glu Glu Pro
45 50 55
atg gag tta gat tat cca taaaacgaac atgaaaatta ttctcttcct 304
Met Glu Leu Asp Tyr Pro
60
gacattgatt gtatttcat cttgcgagct atatcactat caggagtgtg ttagaggtac 364
gactgtacta ctaaaagaac cttgccatc aggaacatac gagggcaatt caccatttca 424
ccctcttgct gacaataaat ttgcactaac ttgcactagc acacactttg cttttgcttg 484
tgctgacggt actcgacata cctatcagct gcgtgcaaga tcagtttcac caaaactttt 544
catcagacaa gaggagggtc aacaagagct ctactcgcca ctttttctca ttgttgctgc 604
tctagtattt ttaatacttt gcttcacat taagagaaag acagaatgaa tgagctcact 664
ttaattgact tctatttggc ctttttagcc tttctgctat tccttgttt aataatgctt 724
attatatttt ggttttctact cgaatccag gatctagaag aacctgtac caaagtctaa 784
acgaacatga aacttctcat tgttttgact tgtatttctc tatgcagttg catatgcact 844
gtagtacagc gctgtgcatc taataaacct catgtgcttg aagatccttg taaggtaaca 904
cactaggggt aatacttata gcaactgctg gotttgtgct ctaggaaagg ttttaccttt 964
tcatagatgg cacactatgg ttcaaacatg cacacctaata gttactatca actgtcaaga 1024
tccagctggt ggtgcgctta tagctagggt ttggtacett catgaaggtc accaaaactgc 1084
tgcatctaga gacgtacttg ttgttttaaa taaacgaaca aattaaaatg tctgataatg 1144

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gacccaatc aaaccaacgt agtgccccc gcattacatt tggtaggaccc acagattcaa 1204
 ctgacaataa ccagaatgga ggacgca 1231

<210> SEQ ID NO 22
 <211> LENGTH: 63
 <212> TYPE: PRT
 <213> ORGANISM: CORONAVIRUS

<400> SEQUENCE: 22

Met Phe His Leu Val Asp Phe Gln Val Thr Ile Ala Glu Ile Leu Ile
 1 5 10 15
 Ile Ile Met Arg Thr Phe Arg Ile Ala Ile Trp Asn Leu Asp Val Ile
 20 25 30
 Ile Ser Ser Ile Val Arg Gln Leu Phe Lys Pro Leu Thr Lys Lys Asn
 35 40 45
 Tyr Ser Glu Leu Asp Asp Glu Pro Met Glu Leu Asp Tyr Pro
 50 55 60

<210> SEQ ID NO 23
 <211> LENGTH: 1231
 <212> TYPE: DNA
 <213> ORGANISM: CORONAVIRUS
 <220> FEATURE:
 <221> NAME/KEY: CDS
 <222> LOCATION: (285)..(650)
 <223> OTHER INFORMATION:

<400> SEQUENCE: 23

taccgtattg gaaactataa attaaataca gaccacgccg gttagcaacga caatattgct 60
 ttgctagtagc agtaagtgc aacagatggt tcatcttggt gacttccagg ttacaatagc 120
 agagatattg attatcatta tgaggacttt caggattgct atttgaatc ttgacgttat 180
 aataagtcca atagtgcac aattatctaa gcctctaact aagaagaatt attcggagtt 240
 agatgatgaa gaacctatgg agttagatta tccataaaac gaac atg aaa att att 296
 Met Lys Ile Ile
 1
 ctc ttc ctg aca ttg att gta ttt aca tct tgc gag cta tat cac tat 344
 Leu Phe Leu Thr Leu Ile Val Phe Thr Ser Cys Glu Leu Tyr His Tyr
 5 10 15 20
 cag gag tgt gtt aga ggt acg act gta cta cta aaa gaa cct tgc cca 392
 Gln Glu Cys Val Arg Gly Thr Thr Val Leu Leu Lys Glu Pro Cys Pro
 25 30 35
 tca gga aca tac gag ggc aat tca cca ttt cac cct ctt gct gac aat 440
 Ser Gly Thr Tyr Glu Gly Asn Ser Pro Phe His Pro Leu Ala Asp Asn
 40 45 50
 aaa ttt gca cta act tgc act agc aca cac ttt gct ttt gct tgt gct 488
 Lys Phe Ala Leu Thr Cys Thr Ser Thr His Phe Ala Phe Ala Cys Ala
 55 60 65
 gac ggt act cga cat acc tat cag ctg cgt gca aga tca gtt tca cca 536
 Asp Gly Thr Arg His Thr Tyr Gln Leu Arg Ala Arg Ser Val Ser Pro
 70 75 80
 aaa ctt ttc atc aga caa gag gag gtt caa caa gag ctc tac tcg cca 584
 Lys Leu Phe Ile Arg Gln Glu Glu Val Gln Gln Glu Leu Tyr Ser Pro
 85 90 95 100
 ctt ttt ctc att gtt gct gct cta gta ttt tta ata ctt tgc ttc acc 632
 Leu Phe Leu Ile Val Ala Ala Leu Val Phe Leu Ile Leu Cys Phe Thr
 105 110 115

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att aag aga aag aca gaa tgaatgagct cactttaatt gacttctatt      680
Ile Lys Arg Lys Thr Glu
      120

tgtgcttttt agcctttctg ctattccttg ttttaataat gcttattata ttttggtttt      740
cactcgaaat ccaggatcta gaagaacctt gtaccaaagt ctaaacgaac atgaaacttc      800
tcattgtttt gacttgatt tctctatgca gttgcatatg cactgtagta cagcgctgtg      860
catctaataa acctcatgtg cttgaagatc cttgtaaggt acaacactag gggaataact      920
tatagcactg cttggctttg tgctctagga aaggttttac cttttcatag atggcacact      980
atggttcaaa catgcacacc taatgttact atcaactgtc aagatccagc tgggtgtgcg     1040
cttatagcta ggtgttgga ccttcatgaa ggtcacaaa ctgctgcatt tagagacgta     1100
cttgtgtgtt taaataaacg aacaaattaa aatgtctgat aatggacccc aatcaaacca     1160
acgtagtgcc ccccgatta catttgggtg acccacagat tcaactgaca ataaccagaa     1220
tggaggacgc a                                                           1231

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<210> SEQ ID NO 24
<211> LENGTH: 122
<212> TYPE: PRT
<213> ORGANISM: CORONAVIRUS

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<400> SEQUENCE: 24

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Met Lys Ile Ile Leu Phe Leu Thr Leu Ile Val Phe Thr Ser Cys Glu
 1           5           10          15

Leu Tyr His Tyr Gln Glu Cys Val Arg Gly Thr Thr Val Leu Leu Lys
      20           25           30

Glu Pro Cys Pro Ser Gly Thr Tyr Glu Gly Asn Ser Pro Phe His Pro
      35           40           45

Leu Ala Asp Asn Lys Phe Ala Leu Thr Cys Thr Ser Thr His Phe Ala
      50           55           60

Phe Ala Cys Ala Asp Gly Thr Arg His Thr Tyr Gln Leu Arg Ala Arg
      65           70           75           80

Ser Val Ser Pro Lys Leu Phe Ile Arg Gln Glu Glu Val Gln Gln Glu
      85           90           95

Leu Tyr Ser Pro Leu Phe Leu Ile Val Ala Ala Leu Val Phe Leu Ile
      100          105          110

Leu Cys Phe Thr Ile Lys Arg Lys Thr Glu
      115          120

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<210> SEQ ID NO 25
<211> LENGTH: 1231
<212> TYPE: DNA
<213> ORGANISM: CORONAVIRUS
<220> FEATURE:
<221> NAME/KEY: CDS
<222> LOCATION: (650)..(781)
<223> OTHER INFORMATION:

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<400> SEQUENCE: 25

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taccgtattg gaaactataa attaaataca gaccacgccc gtagcaacga caatattgct      60
ttgctagtac agtaagtgc aacagatggt tcatcttggt gacttccagg ttacaatagc     120
agagatattg attatcatta tgaggacttt caggattgct atttgaatc ttgacgttat     180
aataagttca atagtgcagc aattatntaa gcctctaact aagaagaatt attcggagtt     240

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agatgatgaa gaacctatgg agttagatta tccataaaac gaacatgaaa attattctct 300
tcctgacatt gattgtatatt acatcttgcc agctatatca ctatcaggag tgtgtagag 360
gtacgactgt actactaaaa gaaccttgcc catcaggaaac atacgagggc aattcaccat 420
ttaccctct tgctgacaat aaatttgcc taacttgcc tagcacacac ttgcttttg 480
cttgctgta cggtaactga catacctatc agctgctgac aagatcagtt tcacaaaac 540
tttcatcag acaagaggag gttcaacaag agctctactc gccacttttt ctctattgtg 600
ctgctctagt attttaata ctttgcttca ccattaagag aaagacaga atg aat gag 658
                                     Met Asn Glu
                                     1
ctc act tta att gac ttc tat ttg tgc ttt tta gcc ttt ctg cta ttc 706
Leu Thr Leu Ile Asp Phe Tyr Leu Cys Phe Leu Ala Phe Leu Leu Phe
   5                10                15
ctt gtt tta ata atg ctt att ata ttt tgg ttt tca ctc gaa atc cag 754
Leu Val Leu Ile Met Leu Ile Ile Phe Trp Phe Ser Leu Glu Ile Gln
  20                25                30                35
gat cta gaa gaa cct tgt acc aaa gtc taaacgaaca tgaaacttct 801
Asp Leu Glu Glu Pro Cys Thr Lys Val
   40
cattgttttg acttgtatatt ctctatgac ttgcatatgc actgtagtac agcgtgtgac 861
atctaataaa cctcatgtgc ttgaagatcc ttgtaaggta caacactagg ggtaactactt 921
atagcactgc ttggctttgt gctctaggaa aggttttacc ttttcataga tggcacacta 981
tggttcaaac atgcacacct aatgttacta tcaactgtca agatccagct ggtgggtgccc 1041
ttatagctag gtgttggtac cttcatgaag gtcaccaaac tgctgcattt agagacgtac 1101
ttgtgttttt aaataaacga acaaatataa atgtctgata atggacccca atcaaaccaa 1161
cgtagtgtccc cccgatttac atttggtgga cccacagatt caactgacaa taaccagaat 1221
ggaggacgca 1231

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<210> SEQ ID NO 26
<211> LENGTH: 44
<212> TYPE: PRT
<213> ORGANISM: CORONAVIRUS

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<400> SEQUENCE: 26

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Met Asn Glu Leu Thr Leu Ile Asp Phe Tyr Leu Cys Phe Leu Ala Phe
 1                5                10                15
Leu Leu Phe Leu Val Leu Ile Met Leu Ile Ile Phe Trp Phe Ser Leu
 20                25                30
Glu Ile Gln Asp Leu Glu Glu Pro Cys Thr Lys Val
 35                40

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<210> SEQ ID NO 27
<211> LENGTH: 1231
<212> TYPE: DNA
<213> ORGANISM: CORONAVIRUS
<220> FEATURE:
<221> NAME/KEY: CDS
<222> LOCATION: (791)..(907)
<223> OTHER INFORMATION:

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<400> SEQUENCE: 27

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taccgtattg gaaactataa attaaatata gaccacgccg gtagcaacga caatattgct 60

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Ser Cys Ile Cys Thr Val Val Gln Arg Cys Ala Ser Asn Lys Pro His
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Val Leu Glu Asp Pro Cys Lys Val Gln His
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<210> SEQ ID NO 28
<211> LENGTH: 39
<212> TYPE: PRT
<213> ORGANISM: CORONAVIRUS

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<400> SEQUENCE: 28

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Asp Pro Cys Lys Val Gln His
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<210> SEQ ID NO 29
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<212> TYPE: DNA
<213> ORGANISM: CORONAVIRUS
<220> FEATURE:
<221> NAME/KEY: CDS
<222> LOCATION: (876)..(1127)
<223> OTHER INFORMATION:

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<400> SEQUENCE: 29

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                               1                               5

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Val Arg Tyr Asn Thr Arg Gly Asn Thr Tyr Ser Thr Ala Trp Leu Cys
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Ala Leu Gly Lys Val Leu Pro Phe His Arg Trp His Thr Met Val Gln
                25                30                35

aca tgc aca cct aat gtt act atc aac tgt caa gat cca gct ggt ggt    1037
Thr Cys Thr Pro Asn Val Thr Ile Asn Cys Gln Asp Pro Ala Gly Gly
                40                45                50

gcg ctt ata gct agg tgt tgg tac ctt cat gaa ggt cac caa act gct    1085
Ala Leu Ile Ala Arg Cys Trp Tyr Leu His Glu Gly His Gln Thr Ala
55                60                65                70

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Ala Phe Arg Asp Val Leu Val Val Leu Asn Lys Arg Thr Asn
                75                80

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<210> SEQ ID NO 30

<211> LENGTH: 84

<212> TYPE: PRT

<213> ORGANISM: CORONAVIRUS

<400> SEQUENCE: 30

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Ser Thr Ala Trp Leu Cys Ala Leu Gly Lys Val Leu Pro Phe His Arg
                20                25                30

Trp His Thr Met Val Gln Thr Cys Thr Pro Asn Val Thr Ile Asn Cys
                35                40                45

Gln Asp Pro Ala Gly Gly Ala Leu Ile Ala Arg Cys Trp Tyr Leu His
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Lys Arg Thr Asn

<210> SEQ ID NO 31
<211> LENGTH: 21221
<212> TYPE: DNA
<213> ORGANISM: CORONAVIRUS

<400> SEQUENCE: 31

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Gln Gly Gln Asn Ser Ala Asp Pro Lys Val Tyr Pro Ile Ile Leu Arg
          35           40           45
Leu Gly Ser Gln Leu Ser Leu Ser Met Ala Arg Arg Asn Leu Asp Ser
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Lys Leu Ala Thr Thr Glu Glu Leu Pro Asp Glu Phe Val Val Val Thr
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Ala Lys

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35 40 45
Val Gly Glu Ile Leu Leu Leu Glu Trp Leu Ala Glu Val Val Lys Leu
50 55 60
Pro Ser Arg Tyr Cys Cys
65 70

<210> SEQ ID NO 36
<211> LENGTH: 1377
<212> TYPE: DNA
<213> ORGANISM: CORONAVIRUS
<220> FEATURE:
<221> NAME/KEY: CDS
<222> LOCATION: (67)..(1335)
<223> OTHER INFORMATION:

<400> SEQUENCE: 36

atgaaggtca ccaaactgct gcatttagag acgtacttgt tgttttaaat aaacgaacaa 60
attaaa atg tct gat aat gga ccc caa tca aac caa cgt agt gcc ccc 108
Met Ser Asp Asn Gly Pro Gln Ser Asn Gln Arg Ser Ala Pro
1 5 10
cgc att aca ttt ggt gga ccc aca gat tca act gac aat aac cag aat 156
Arg Ile Thr Phe Gly Gly Pro Thr Asp Ser Thr Asp Asn Asn Gln Asn
15 20 25 30
gga gga cgc aat ggg gca agg cca aaa cag cgc cga ccc caa ggt tta 204
Gly Gly Arg Asn Gly Ala Arg Pro Lys Gln Arg Arg Pro Gln Gly Leu
35 40 45
ccc aat aat act gcg tct tgg ttc aca gct ctc act cag cat ggc aag 252
Pro Asn Asn Thr Ala Ser Trp Phe Thr Ala Leu Thr Gln His Gly Lys
50 55 60

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gag gaa ctt aga ttc cct cga ggc cag ggc gtt cca atc aac acc aat Glu Glu Leu Arg Phe Pro Arg Gly Gln Gly Val Pro Ile Asn Thr Asn 65 70 75	300
agt ggt cca gat gac caa att ggc tac tac cga aga gct acc cga cga Ser Gly Pro Asp Asp Gln Ile Gly Tyr Tyr Arg Arg Ala Thr Arg Arg 80 85 90	348
gtt cgt ggt ggt gac ggc aaa atg aaa gag ctc agc ccc aga tgg tac Val Arg Gly Gly Asp Gly Lys Met Lys Glu Leu Ser Pro Arg Trp Tyr 95 100 105 110	396
ttc tat tac cta gga act ggc cca gaa gct tca ctt ccc tac ggc gct Phe Tyr Tyr Leu Gly Thr Gly Pro Glu Ala Ser Leu Pro Tyr Gly Ala 115 120 125	444
aac aaa gaa ggc atc gta tgg gtt gca act gag gga gcc ttg aat aca Asn Lys Glu Gly Ile Val Trp Val Ala Thr Glu Gly Ala Leu Asn Thr 130 135 140	492
ccc aaa gac cac att ggc acc cgc aat cct aat aac aat gct gcc acc Pro Lys Asp His Ile Gly Thr Arg Asn Pro Asn Asn Ala Ala Thr 145 150 155	540
gtg cta caa ctt cct caa gga aca aca ttg cca aaa ggc ttc tac gca Val Leu Gln Leu Pro Gln Gly Thr Thr Leu Pro Lys Gly Phe Tyr Ala 160 165 170	588
gag gga agc aga ggc ggc agt caa gcc tct tct cgc tcc tca tca cgt Glu Gly Ser Arg Gly Gly Ser Gln Ala Ser Ser Arg Ser Ser Ser Arg 175 180 185 190	636
agt cgc ggt aat tca aga aat tca act cct ggc agc agt agg gga aat Ser Arg Gly Asn Ser Arg Asn Ser Thr Pro Gly Ser Ser Arg Gly Asn 195 200 205	684
tct cct gct cga atg gct agc gga ggt ggt gaa act gcc ctc gcg cta Ser Pro Ala Arg Met Ala Ser Gly Gly Glu Thr Ala Leu Ala Leu 210 215 220	732
ttg ctg cta gac aga ttg aac cag ctt gag agc aaa gtt tct ggt aaa Leu Leu Leu Asp Arg Leu Asn Gln Leu Glu Ser Lys Val Ser Gly Lys 225 230 235	780
ggc caa caa caa caa ggc caa act gtc act aag aaa tct gct gct gag Gly Gln Gln Gln Gln Gly Gln Thr Val Thr Lys Lys Ser Ala Ala Glu 240 245 250	828
gca tct aaa aag cct cgc caa aaa cgt act gcc aca aaa cag tac aac Ala Ser Lys Lys Pro Arg Gln Lys Arg Thr Ala Thr Lys Gln Tyr Asn 255 260 265 270	876
gtc act caa gca ttt ggg aga cgt ggt cca gaa caa acc caa gga aat Val Thr Gln Ala Phe Gly Arg Arg Gly Pro Glu Gln Thr Gln Gly Asn 275 280 285	924
ttc ggg gac caa gac cta atc aga caa gga act gat tac aaa cat tgg Phe Gly Asp Gln Asp Leu Ile Arg Gln Gly Thr Asp Tyr Lys His Trp 290 295 300	972
ccg caa att gca caa ttt gct cca agt gcc tct gca ttc ttt gga atg Pro Gln Ile Ala Gln Phe Ala Pro Ser Ala Ser Ala Phe Phe Gly Met 305 310 315	1020
tca cgc att ggc atg gaa gtc aca cct tcg gga aca tgg ctg act tat Ser Arg Ile Gly Met Glu Val Thr Pro Ser Gly Thr Trp Leu Thr Tyr 320 325 330	1068
cat gga gcc att aaa ttg gat gac aaa gat cca caa ttc aaa gac aac His Gly Ala Ile Lys Leu Asp Asp Lys Asp Pro Gln Phe Lys Asp Asn 335 340 345 350	1116
gtc ata ctg ctg aac aag cac att gac gca tac aaa aca ttc cca cca Val Ile Leu Leu Asn Lys His Ile Asp Ala Tyr Lys Thr Phe Pro Pro 355 360 365	1164

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aca gag cct aaa aag gac aaa aag aaa aag act gat gaa gct cag cct      1212
Thr Glu Pro Lys Lys Asp Lys Lys Lys Lys Thr Asp Glu Ala Gln Pro
          370                      375                      380

ttg ccg cag aga caa aag aag cag ccc act gtg act ctt ctt cct gcg      1260
Leu Pro Gln Arg Gln Lys Lys Gln Pro Thr Val Thr Leu Leu Pro Ala
          385                      390                      395

gct gac atg gat gat ttc tcc aga caa ctt caa aat tcc atg agt gga      1308
Ala Asp Met Asp Asp Phe Ser Arg Gln Leu Gln Asn Ser Met Ser Gly
          400                      405                      410

gct tct gct gat tca act cag gca taa acactcatga tgaccacaca      1355
Ala Ser Ala Asp Ser Thr Gln Ala
415                      420

aggcagatgg gctatgtaaa cg      1377

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<210> SEQ ID NO 37
<211> LENGTH: 422
<212> TYPE: PRT
<213> ORGANISM: CORONAVIRUS

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<400> SEQUENCE: 37

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Met Ser Asp Asn Gly Pro Gln Ser Asn Gln Arg Ser Ala Pro Arg Ile
1          5          10          15

Thr Phe Gly Gly Pro Thr Asp Ser Thr Asp Asn Asn Gln Asn Gly Gly
          20          25          30

Arg Asn Gly Ala Arg Pro Lys Gln Arg Arg Pro Gln Gly Leu Pro Asn
          35          40          45

Asn Thr Ala Ser Trp Phe Thr Ala Leu Thr Gln His Gly Lys Glu Glu
          50          55          60

Leu Arg Phe Pro Arg Gly Gln Gly Val Pro Ile Asn Thr Asn Ser Gly
          65          70          75          80

Pro Asp Asp Gln Ile Gly Tyr Tyr Arg Arg Ala Thr Arg Arg Val Arg
          85          90          95

Gly Gly Asp Gly Lys Met Lys Glu Leu Ser Pro Arg Trp Tyr Phe Tyr
          100          105          110

Tyr Leu Gly Thr Gly Pro Glu Ala Ser Leu Pro Tyr Gly Ala Asn Lys
          115          120          125

Glu Gly Ile Val Trp Val Ala Thr Glu Gly Ala Leu Asn Thr Pro Lys
          130          135          140

Asp His Ile Gly Thr Arg Asn Pro Asn Asn Asn Ala Ala Thr Val Leu
          145          150          155          160

Gln Leu Pro Gln Gly Thr Thr Leu Pro Lys Gly Phe Tyr Ala Glu Gly
          165          170          175

Ser Arg Gly Gly Ser Gln Ala Ser Ser Arg Ser Ser Ser Arg Ser Arg
          180          185          190

Gly Asn Ser Arg Asn Ser Thr Pro Gly Ser Ser Arg Gly Asn Ser Pro
          195          200          205

Ala Arg Met Ala Ser Gly Gly Gly Glu Thr Ala Leu Ala Leu Leu Leu
          210          215          220

Leu Asp Arg Leu Asn Gln Leu Glu Ser Lys Val Ser Gly Lys Gly Gln
          225          230          235          240

Gln Gln Gln Gly Gln Thr Val Thr Lys Lys Ser Ala Ala Glu Ala Ser
          245          250          255

Lys Lys Pro Arg Gln Lys Arg Thr Ala Thr Lys Gln Tyr Asn Val Thr

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	260		265		270	
Gln Ala Phe Gly Arg Arg Gly Pro Glu Gln Thr Gln Gly Asn Phe Gly	275		280		285	
Asp Gln Asp Leu Ile Arg Gln Gly Thr Asp Tyr Lys His Trp Pro Gln	290		295		300	
Ile Ala Gln Phe Ala Pro Ser Ala Ser Ala Phe Phe Gly Met Ser Arg	305		310		315	320
Ile Gly Met Glu Val Thr Pro Ser Gly Thr Trp Leu Thr Tyr His Gly		325		330		335
Ala Ile Lys Leu Asp Asp Lys Asp Pro Gln Phe Lys Asp Asn Val Ile		340		345		350
Leu Leu Asn Lys His Ile Asp Ala Tyr Lys Thr Phe Pro Pro Thr Glu		355		360		365
Pro Lys Lys Asp Lys Lys Lys Lys Thr Asp Glu Ala Gln Pro Leu Pro		370		375		380
Gln Arg Gln Lys Lys Gln Pro Thr Val Thr Leu Leu Pro Ala Ala Asp		385		390		395
Met Asp Asp Phe Ser Arg Gln Leu Gln Asn Ser Met Ser Gly Ala Ser		405		410		415
Ala Asp Ser Thr Gln Ala		420				

<210> SEQ ID NO 38
 <211> LENGTH: 1377
 <212> TYPE: DNA
 <213> ORGANISM: CORONAVIRUS

<400> SEQUENCE: 38

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atgaaggtca ccaaactgct gcatttagag acgtacttgt tgttttaaat aaacgaacaa    60
attaaaaatg ctgataatgg accccaatca aaccaacgta gtgccccccg cattacattt    120
ggtggaccca cagattcaac tgacaataac cagaatggag gacgcaatgg ggcaaggcca    180
aaacagcgcc gacccaagg tttacccaat aatactgctt cttgggtcac agctctcaact    240
cagcatggca aggaggaact tagattccct cgaggccagg gcgttccaat caacaccaat    300
agtgggtccag atgaccaaata tggctactac cgaagagcta cccgacgagt tcgtggtggt    360
gacggcaaaa tgaaagagct cagccccaga tggctacttct attacctagg aactggccca    420
gaagcttcac ttccctacgg cgctaacaaa gaaggcatcg tatgggttgc aactgagggga    480
gccttgaata cacccaaga ccacattgac acccgcaatc ctaataacaa tgctgccacc    540
gtgctacaac ttctcaagg aacaacattg ccaaaaggct tctacgcaga ggaagcaga    600
ggcggcagtc aagcctcttc tcgctoctca tcacgtagtc gcgtaattc aagaaattca    660
actcctggca gcagtagggg aaattctcct gctcgaatgg ctacgaggag tggtgaaact    720
gccctcgcgc tattgctgct agacagattg aaccagcttg agagcaaagt ttctggtaaa    780
ggccaacaac aacaaggcca aactgtcact aagaatctg ctgctgaggc atctaaaaag    840
cctcgccaaa aacgtactgc cacaaaacag tacaacgtca ctcaagcatt tgggagacgt    900
ggtccagaac aaaccaagg aaatttcggg gaccaagacc taatcagaca aggaactgat    960
tacaacatt ggccgcaaat tgcacaattt gctccaagtg cctctgcatt ctttggaatg    1020
tcacgcattg gcatggaagt cacaccttcg ggaacatggc tgacttatca tggagccatt    1080
    
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aaattggatg acaaagatcc acaattcaaa gacaacgtca tactgctgaa caagcacatt 1140
gacgcataca aaacattccc accaacagag cctaaaaagg acaaaaagaa aaagactgat 1200
gaagctcagc ctttgccgca gagacaaaag aagcagccca ctgtgactct tcttcctgcg 1260
gctgacatgg atgattttctc cagacaactt caaaattcca tgagtggagc ttctgctgat 1320
tcaactcagg cataaacact catgatgacc acacaaggca gatgggctat gtaaacg 1377

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<210> SEQ ID NO 39
<211> LENGTH: 204
<212> TYPE: DNA
<213> ORGANISM: CORONAVIRUS

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<400> SEQUENCE: 39

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atattaggtt ttacctacc caggaaaagc caaccaacct cgatctcttg tagatctggt 60
ctctaaacga actttaaaat ctgtgtagct gtcgctcggc tgcatgccta gtgcacctac 120
gcagtataaa caataataaa ttttactgtc gttgacaaga aacgagtaac tcgtccctct 180
tctgcagact gcttacggtt tcgt 204

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<210> SEQ ID NO 40
<211> LENGTH: 809
<212> TYPE: DNA
<213> ORGANISM: CORONAVIRUS

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<400> SEQUENCE: 40

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actcaagcat ttgggagacg tgggtccagaa caaacccaag gaaatttcgg ggaccaagac 60
ctaatacagc aaggaactga ttacaacatc tggccgcaaa ttgcacaatt tgctccaagt 120
gcctctgcat tctttggaat gtcacgcatt ggcatggaag tcacaccttc ggaacatgg 180
ctgacttatac atggagccat taaattggat gacaaagatc cacaaattcaa agacaacgtc 240
atactgctga acaagcacat tgacgcatac aaaacattcc caccaacaga gcctaaaaag 300
gacaaaaaga aaaagactga tgaagctcag cctttgccgc agagacaaaa gaagcagccc 360
actgtgactc ttcttctctc ggctgacatg gatgatttct ccagacaact tcaaaattcc 420
atgagtggag cttctgctga ttcaactcag gcataaacac tcatgatgac cacacaaggc 480
agatgggcta tgtaaactgt ttcgcaattc cgtttacgat acatagtcta ctcttggtgca 540
gaatgaattc tcgtaactaa acagcacaag taggtttagt taactttaat ctcacatagc 600
aatctttaat caatgtgtaa cattagggag gacttgaaag agccaccaca ttttcatcga 660
ggccacgcgg agtacgatc aggggtacag gaataatgct agggagagct gcctatatgg 720
aagagcccta atgtgtaaaa ttaattttag tagtgctatc cccatgtgat ttaaatagct 780
tcttaggaga atgacaaaaa aaaaaaaaa 809

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<210> SEQ ID NO 41
<211> LENGTH: 448
<212> TYPE: DNA
<213> ORGANISM: CORONAVIRUS

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<400> SEQUENCE: 41

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aatgaacaca tagggctggt caagctgggg cagtacgctt tttccagct ctactagacc 60
acaagtgcc a tttttagggt gttcaogtgc ctccgatagg gcctcttcca cagagtcccc 120
gaagccacgc actagcacgt ctctaacctg aaggacaggc aaactgagtt ggacgtgtgt 180

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tttctcgttg acaccaagaa caaggctctc catcttacct ttcggtcaca cccggacgaa	240
acctaggtat gctgatgatc gactgcaaca cggacgaaac cgtaagcagt ctgcagaaga	300
gggacgagtt actcgtttct tgtcaacgac agtaaaattht attattgttt atactgcgta	360
ggtgactag gcatgcagcc gagcgacagc tacacagatt ttaaagttcg tttagagaac	420
agatctacaa gagatcgagg ttggttgg	448

<210> SEQ ID NO 42

<211> LENGTH: 2033

<212> TYPE: DNA

<213> ORGANISM: CORONAVIRUS

<400> SEQUENCE: 42

atacctaggt ttcgtccggg tgtgaccgaa aggtaagatg gagagccttg ttcttggtgt	60
caacgagaaa acacacgtcc aactcagttt gcctgtcctt caggttagag acgtgctagt	120
gcgtggcttc ggggactctg tggaaagagc cctatcggag gcacgtgaac acctcaaaaa	180
tggcacttgt ggtctagtag agctggaaaa aggcgtactg cccagcttg aacagcccta	240
tggtttcatt aaacgttctg atgccttaag caccaatcac ggccacaagg tcgttgagct	300
ggttgacgaa atggacggca ttcagtacgg tcgtagcggg ataactctgg gactactcgt	360
gccacatgtg ggcgaaaccc caattgcata cgcgatgtt cttcttcgta agaacggtaa	420
taaggagacc ggtggtcata gctatggcat cgatctaaag tcttatgact taggtgacga	480
gcttgccact gatcccattg aagattatga acaaaactgg aactactaagc atggcagtgg	540
tgcactccgt gaactcactc gtgagctcaa tggaggtgca gtcactcgtc atgtcgacaa	600
caatttctgt ggcccagatg ggtaccctct tgattgcatc aaagattttc tcgcacgcgc	660
gggcaagtca atgtgactc tttccgaaca acttgattac atcgagtcga agagaggtgt	720
ctactctgct cgtgaccatg agcatgaaat tgcctggttc actgagcgtc ctgataagag	780
ctacgagcac cagacaccct tcgaaattaa gagtgccaag aaatttgaca ctttcaaagg	840
ggaatgcccc aagtttgtgt ttcctcttaa ctcaaaagtc aaagtcattc aaccacgtgt	900
tgaaaagaaa aagactgagg gtttcatggg gogtatacgc tctgtgtacc ctgttgcac	960
tccacaggag tgaacaata tgcacttgtc taccttgatg aaatgtaatc attgcatga	1020
agtttcatg cagacgtgag actttctgaa agccacttgt gaacattgtg gactgaaaa	1080
tttagttatt gaagaccta ctacatgtg gtacctacct actaatgctg tagtgaat	1140
gccatgtcct gcctgtcaag acccagagat tggacctgag catagtgttg cagattatca	1200
caaccactca aacattgaaa ctgcactcag caaggagggt aggactagat gttttggagg	1260
ctgtgtgttt gcctatgttg gctgctataa taagcgtgcc tactgggttc ctctgtctag	1320
tgctgatatt ggctcaggcc atactggcat tactggtgac aatgtggaga ccttgaatga	1380
ggatctcctt gagatactga gtcgtgaaag tgtaaacatt aacattgttg gcgattttca	1440
tttgaatgaa gaggttgcca tcattttggc atctttctct gcttctacaa gtgoccttat	1500
tgacactata aagagtcttg attacaagtc tttcaaaacc attgttgagt cctgcggtaa	1560
ctataaagtt accaagggaa agcccgtaaa aggtgcttgg aacattggac aacagagatc	1620
agttttaaca cactgtgtg gttttccctc acaggctgct ggtgttatca gatcaatttt	1680
tgcgcgacaa cttagtcgag caaacactc aattcctgat ttgcaaagag cagctgtcac	1740

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catacttgat ggtatttctg aacagtcatt acgtcttgtc gacgccatgg tttatacttc	1800
agacctgctc accaacagtg tcattattat ggcatatgta actggtggtc ttgtacaaca	1860
gacttctcag tggttgtcta atcttttggg cactactgtt gaaaaactca ggcctatcct	1920
tgaatggatt gaggcgaaac ttagtgcag agttgaattt ctcaaggatg cttgggagat	1980
tctcaaattt ctcattacag gtgtttttga catcgtcaag ggtcaaatac agg	2033

<210> SEQ ID NO 43

<211> LENGTH: 2018

<212> TYPE: DNA

<213> ORGANISM: CORONAVIRUS

<400> SEQUENCE: 43

ggattgaggc gaaacttagt gcaggagttg aatttctcaa ggatgcttgg gagattctca	60
aatttctcat tacagggtgtt ttgacatcg tcaagggtca aatacagggt gcttcagata	120
acatcaagga ttgtgtaaaa tgcttcattg atgttgtaa caaggcactc gaaatgtgca	180
ttgatcaagt cactatcgct ggcgcaaagt tgcgatcact caacttaggt gaagtcttca	240
tcgctcaaag caagggactt taccgtcagt gtatactggt caaggagcag ctgcaactac	300
tcatgcctct taaggcacca aaagaagtaa cctttcttga aggtgattca catgacacag	360
tacttacctc tgaggaggtt gttctcaaga acggtgaact cgaagcactc gagacgcccg	420
ttgatagctt cacaaatgga gctatcgttg gcacaccagt ctgtgtaaat ggcctcatgc	480
tcttagagat taaggacaaa gaacaatact gcgcattgtc tcctggttta ctggctacaa	540
acaaatgtct tcgcttaaaa gggggtgcac caattaaagg tgtaaccttt ggagaagata	600
ctgtttggga agttcaaggt tacaagaatg tgagaatcac atttgagctt gatgaacgtg	660
ttgacaaagt gcttaatgaa aagtgcctcg tctacactgt tgaatccggg accgaagtta	720
ctgagtttgc atgtgttgta gcagaggctg ttgtgaagac tttacaacca gtttctgatc	780
tccttaccaa catgggtatt gatcttgatg agtggagtgt agctacattc tacttatttg	840
atgatgctgg tgaagaaaac ttttcatcac gtatgtattg ttccttttac cctccagatg	900
aggaagaaga ggacgatgca gagtgtgagg aagaagaaat tgatgaaacc tgtgaacatg	960
agtacggtac agaggatgat tatcaaggtc tccctctgga atttggtgcc tcagctgaaa	1020
cagttcgagt tgaggaaaga gaagaggaag actggctgga tgatactact gagcaatcag	1080
agattgagcc agaaccagaa cctacacctg aagaaccagt taatcagttt actggttatt	1140
taaaacttac tgacaatgtt gccattaat gtgttgacat cgtaaggag gcacaaagtg	1200
ctaactctat ggtgattgta aatgctgcta acatacacct gaaacatggg ggtgggtgag	1260
cagggtcact caacaaggca accaatggtg ccatgcaaaa ggagagtgat gattacatta	1320
agctaaatgg ccctcttaca gtaggaggtt cttgtttgct ttctggacat aatcttgcta	1380
agaagtgtct gcatgtgttt ggacctaac taaatgcagg tgaggacatc cagcttctta	1440
aggcagcata tgaaaatttc aattcacagg acatcttact tgcaccattg ttgtcagcag	1500
gcataatttg tgctaaacca cttcagctct tacaagtgtg cgtgcagacg gttcgtacac	1560
aggtttatat tgcagtcaat gacaaagctc tttatgagca ggttgatcatg gattatcttg	1620
ataacctgaa gcctagagtg gaagcaccta aacaagagga gccaccaaac acagaagatt	1680
ccaaaactga ggagaaatct gtcgtacaga agcctgtcga tgtgaagcca aaaattaagg	1740

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cctgcattga tgaggttacc acaacactgg aagaaactaa gtttcttacc aataagttac 1800
tcttgtttgc tgatatcaat ggtaagcttt accatgattc tcagaacatg cttagaggtg 1860
aagatatgtc tttccttgag aaggatgcac cttacatggt aggtgatggt atcactagtg 1920
gtgatatac tttgtttgta ataccctcca aaaaggctgg tggcactact gagatgctct 1980
caagagcttt gaagaaagtg ccagttgatg agtatata 2018

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<210> SEQ ID NO 44
<211> LENGTH: 1442
<212> TYPE: DNA
<213> ORGANISM: CORONAVIRUS

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<400> SEQUENCE: 44

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ttgatgaggt taccacaaca ctggaagaaa ctaagtttct taccaataag ttactcttgt 60
ttgctgatat caatggtaag ctttaccatg attctcagaa catgcttaga ggtgaagata 120
tgtctttcct tgagaaggat gcaccttaca tggtaggtga tgttatcact agtggtgata 180
tcactttgtg tgtaataccc tccaaaaagg ctggtggcac tactgagatg ctctcaagag 240
cttgaagaa agtgccagtt gatgagtata taaccacgta ccctggacaa ggatgtgctg 300
gttatacact tgaggaagct aagactgctc ttaagaaatg caaatctgca ttttatgtac 360
taccttcaga agcacctaag gctaaggaag agattctagg aactgtatcc tggaaattga 420
gagaaatgct tgctcatgct gaagagacaa gaaaattaat gcctatatgc atggatgta 480
gagccataat ggcaaccatc caacgtaagt ataaaggaat taaaattcaa gagggcatcg 540
ttgactatgg tgtccgatcc ttcttttata ctagttaaaga gcctgtagct tctattatta 600
cgaagctgaa ctctctaaat gagccgcttg tcacaatgcc aattggttat gtgacacatg 660
gttttaatct tgaagaggct gcgctgctga tgcgttctct taaagctcct gccgtagtgt 720
cagtatcatc accagatgct gttactacat ataatggata cctcacttcg tcatcaaaga 780
catctgagga gcactttgta gaaacagttt ctttggctgg ctcttacaga gattgttcct 840
attcaggaca gcgtacagag ttagggtgtg aatttcttaa gcgtggtgac aaaattgtgt 900
accacactct ggagagcccc gtcgagtttc atcttgacgg tgaggttctt tcacttgaca 960
aactaaagag tctcttatcc ctgctggagg ttaagactat aaaagtgttc acaactgtgg 1020
acaacactaa tctccacaca cagcttggg atatgtctat gacatagga cagcagttt 1080
gtccaacata cttggatggt gctgatgta caaaaattaa acctcatgta aatcatgagg 1140
gtaagacttt ctttgtacta cctagtgatg acacactacg tagtgaagct ttcgagtact 1200
accatactct tgatgagagt tttcttgta ggtacatgct tgctttaaac cacacaaaga 1260
aatgaaatt tcctcaagtt ggtggtttaa cttcaattaa atgggctgat acaattgtt 1320
atgtgtctag tgttttatta gcacttcaac agcttgaagt caaattcaat gcaccagcac 1380
ttcaagaggc ttattataga gccctgctg gtgatgctgc taacttttgt gcaactatac 1440
tc 1442

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<210> SEQ ID NO 45
<211> LENGTH: 1050
<212> TYPE: DNA
<213> ORGANISM: CORONAVIRUS

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<400> SEQUENCE: 45

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atatgtctat gacatatgga cagcagtttg gtccaacata cttggatggt gctgatgta	60
caaaaattaa acctcatgta aatcatgagg gtaagacttt ctttgtaacta cctagtgatg	120
acacactacg tagtgaagct ttcgagtact accatactct tgatgagagt tttcttggtgta	180
ggtacatgtc tgctttaaac cacacaaaga aatggaaatt tcctcaagtt ggtggtttaa	240
cttcaattaa atgggctgat aacaattggt atttgtctag tgttttatta gcacttcaac	300
agcttgaagt caaattcaat gcaccagcac ttcaagaggc ttattataga gccctgctg	360
gtgatgctgc taacttttgt gcactcatac tcgcttacag taataaaact gttggcgagc	420
ttggtgatgt cagagaaact atgaccatc ttctacagca tgctaatttg gaatctgcaa	480
agcgagttct taatgtggtg tgtaaacatt gtggtcagaa aactactacc ttaacgggtg	540
tagaagctgt gatgtatatg ggtactctat cttatgataa tcttaagaca ggtgtttcca	600
ttccatggtg gtgtggctcg gatgctacac aatatctagt acaacaagag tcttcttttg	660
ttatgatgtc tgcaccacct gctgagtata aattacagca aggtacattc ttatgtgcga	720
atgagtacac tggaactat cagtgtggtc attacactca tataactgct aaggagacc	780
tctatcgtat tgacggagct caccttaca agatgtcaga gtacaaagga ccagtgactg	840
atgttttcta caaggaaca tcttacacta caaccatcaa gcctgtgctg tataaactcg	900
atggagttac ttacacagag attgaacaa aattggatgg gtattataaa aaggataatg	960
cttactatac agagcgcct atagacctg taccaactca accattacca aatgcgagtt	1020
ttgataattt caaactcaca tgttctaaca	1050

<210> SEQ ID NO 46

<211> LENGTH: 1995

<212> TYPE: DNA

<213> ORGANISM: CORONAVIRUS

<400> SEQUENCE: 46

tttgtgcact catactcgct tacagtaata aaactggttg cgagcttggg gatgtcagag	60
aaactatgac ccatcttcta cagcatgcta atttgaatc tgcaaagcga gttcttaatg	120
tggtgtgtaa acattgtggt cagaaaacta ctaccttaac ggggtgtaga gctgtgatgt	180
atatgggtac tctatcttat gataatctta agacaggtgt ttccattcca tgtgtgtgtg	240
gtcgtgatgc tacacaatat ctagtacaac aagagtcttc ttttgttatg atgtctgcac	300
cacctgctga gtataaatta cagcaaggta cattcttatg tgcgaatgag tacactggta	360
actatcagtg tggtcattac actcatataa ctgctaagga gacctctat cgtattgacg	420
gagctcacct tacaagatg tcagagtaca aaggaccagt gactgatggt ttctacaagg	480
aaacatctta cactacaacc atcaagcctg tgtcgtataa actcagatgga gttacttaca	540
cagagattga accaaaattg gatgggtatt ataaaaagga taatgcttac tatacagagc	600
agcctataga cttgtacca actcaacat taccaaatgc gagttttgat aatttcaaac	660
tcacatgttc taacacaaaa tttgctgatg atttaaatca aatgacaggc ttcacaagc	720
cagcttcacg agagctatct gtcacattct tcccagactt gaatggcgat gtagtggcta	780
ttgactatag aactatttca gcgagtttca agaaagggtgc taaattactg cataagccaa	840
ttgtttggca cattaaccag gctacaacca agacaacgtt caaaccaaac acttgggtgtt	900
tacgttctct ttggagtaca aagccagtag atacttcaaa ttcattttaa gttctggcag	960

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tagaagacac acaaggaatg gacaatcttg cttgtgaaag tcaacaaccc acctctgaag	1020
aagtagtgga aaatcctacc atacagaagg aagtcataga gtgtgacgtg aaaactaccg	1080
aagttgtagg caatgtcata cttaaaccat cagatgaagg tgtaaagta acacaagagt	1140
taggtcatga ggatcttatg gctgcttatg tggaaaacac aagcattacc attaagaaac	1200
ctaatgagct ttcactagcc ttaggtttaa aaacaattgc cactcatggt attgctgcaa	1260
ttaatagtgt tccttgagggt aaaatcttgg cttatgtcaa accattctta ggacaagcag	1320
caattacaac atcaaatgac gctaagagat tagcacaacg tgtgtttaac aattatatgc	1380
cttatgtggt tacattattg ttccaattgt gtacttttac taaaagtacc aattctagaa	1440
ttagagcttc actacctaca actattgcta aaaatagtgt taagagtgtt gctaaattat	1500
gtttgatgc cggcattaat tatgtgaagt cacccaaatt ttctaaattg ttcacaatcg	1560
ctatgtggct attgttggtta agtatttgct taggttctct aatctgtgta actgctgctt	1620
ttggtgtact cttatctaatt tttggtgctc cttcttattg taatggcggt agagaattgt	1680
atcttaattc gtctaacggt actactatgg atttctgtga aggttctttt ccttcagca	1740
ttgtttaag tggattagac tcccttgatt cttatccagc tcttgaacc attcaggtga	1800
cgattcatc gtacaagcta gacttgacaa ttttaggtct ggccgctgag tgggttttg	1860
catatatggt gttcacaaaa ttcttttatt tattaggtct ttcagctata atgcaggtgt	1920
tctttggcta ttttgctagt catttcatca gcaattcttg gctcatgtgg tttatcatta	1980
gtattgtaca aatgg	1995

<210> SEQ ID NO 47

<211> LENGTH: 1884

<212> TYPE: DNA

<213> ORGANISM: CORONAVIRUS

<400> SEQUENCE: 47

aattcttggc tcatgtgggt tatcattagt attgtacaaa tggcaccogt ttctgcaatg	60
gtaggatgt acatcttctt tgcttcttct tactacatat ggaagagcta tgttcatatc	120
atggatgggt gcacctcttc gacttgcatg atgtgctata agcgcaatcg tgccacacgc	180
gtagagtga caactattgt taatggcatg aagagatctt tctatgtcta tgcaaatgga	240
ggccgtggct tctgcaagac tcacaattgg aattgtctca attgtgacac attttgcaact	300
gtagtagcat tcattagtga tgaagttgct cgtgatttgt cactccagtt taaaagacca	360
atcaacccta ctgaccagtc atcgtatatt gttgatagtg ttgctgtgaa aaatggcgcg	420
cttcacctct actttgacaa ggctgggtcaa aagacctatg agagacatcc gctctcccat	480
tttgcattt tagacaattt gagagctaac aacctaaag gttcactgcc tattaatgtc	540
atagtttttg atggcaagtc caaatgagac gagtctgctt ctaagtctgc ttctgtgtac	600
tacagtcagc tgatgtgcca acctattctg ttgcttgacc aagctcttgt atcagacggt	660
ggagatagta ctgaagtttc cgttaagatg tttgatgctt atgtcgacac cttttcagca	720
acttttagtg ttctcatgga aaaacttaag gcacttgttg ctacagctca cagcgagtta	780
gcaaaggggt tagctttaga tgggtgcctt tctacattcg tgtcagctgc ccgacaaggt	840
gtagttgata ccgatgttga cacaaaggat gttattgaat gtctcaaact ttcacatcac	900
tctgacttag aagtgacagc tgacagttgt aacaatttca tgctcaccta taataagggt	960

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gaaaacatga cgcccagaga tcttgccgca tgtattgact gtaatgcaag gcatatcaat 1020
gcccaagtag caaaaagtca caatgtttca ctcatctgga atgtaaaaga ctacatgtct 1080
ttatctgaac agctgcgtaa acaaattcgt agtgctgcca agaagaacaa catacctttt 1140
agactaactt gtgtacaac tagacagggt gtcaatgtca taactactaa aatctcactc 1200
aagggtggta agattgtag tacttgtttt aaacttatgc ttaaggccac attattgtgc 1260
gttcttgctg cattggtttg ttatatcggt atgccagtac atacattgtc aatccatgat 1320
ggttacacaa atgaaatcat tggttacaaa gccattcagg atggtgtcac tcgtgacatc 1380
atctctactg atgattgttt tgcaataaaa catgctgggt ttgacgcatg gtttagccag 1440
cgtggtgggt catacaaaaa tgacaaaagc tgccctgtag tagctgctat cattacaaga 1500
gagattgggt tcatagtgcc tggcttaccg ggtactgtgc tgagagcaat caatggtgac 1560
ttcttgcaat ttctacctcg tgtttttagt gctgttgca acatttgcta cacaccttc 1620
aaactcattg agtatagtga ttttgctacc tctgcttgcg ttcttgctgc tgagtgtaca 1680
atttttaag atgctatggg caaacctgtg ccatattggt atgacactaa tttgctagag 1740
ggttctatct cttatagtga gcttcgtcca gacactcgtt atgtgcttat ggatggttcc 1800
atcatacagt ttccatacac ttacctggag ggttctgtta gagtagtaac aacttttgat 1860
gctgagtact gtagacatgg taca 1884

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<210> SEQ ID NO 48

<211> LENGTH: 2020

<212> TYPE: DNA

<213> ORGANISM: CORONAVIRUS

<400> SEQUENCE: 48

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cactcgttat gtgcttatgg atggttccat catacagttt cctaacactt acctggaggg 60
ttctgttaga gtagtaacaa cttttgatgc tgagtactgt agacatggta catgcgaaag 120
gtcagaagta ggtatttgcc tatctaccag tggtagatgg gttcttaata atgagcatta 180
cagagctcta tcaggagttt tctgtggtgt tgatgctgat aatctcatag ctaacatctt 240
tactcctctt gtcaacctg tgggtgcttt agatgtgtct gcttcagtag tggctgggtg 300
tattattgcc atattggtga cttgtgctgc ctactacttt atgaaattca gacgtgtttt 360
tggtagtagc aacctgttg ttgctgctaa tgcacttttg tttttgatgt ctttactat 420
actctgtctg gtaccagctt acagctttct gccgggagtc tactcagtct tttacttgta 480
cttgacattc tatttcacca atgatgttct attcttggt caccttcaat ggtttgccat 540
gttttctcct attgtgcctt tttggataac agcaatctat gtattctgta tttctctgaa 600
gcaactgcat tggttcttta acaactatct taggaaaaga gtcattgtta atggagttac 660
atntagtagc ttcgaggagg ctgctttgtg tacctttttg ctcaacaagg aaatgtacct 720
aaaaattgct agcgagacac tgttgccact tacacagtat aacaggtatc ttgctctata 780
taacaagtag aagtatttca gtggagcctt agatactacc agctatcgtg aagcagcttg 840
ctgccactta gaaaaggctc taaatgactt tagcaactca ggtgctgatg ttctctacca 900
accaccacag acatcaatca cttctgctgt tctgcagagt ggttttagga aaatggcatt 960
cccgtcagcg aaagtgaag ggtgcatggt acaagtaacc tgtggaacta caactttaa 1020
tggattgtgg ttggatgaca cagtatactg tocaagacat gtcatttgca cagcagaaga 1080

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catgcttaat cctaactatg aagatctgct cattcgcaa tccaaccata gctttcttgt 1140
tcaggctggc aatgttcaac ttcgtgttat tggccattct atgcaaaatt gtctgcttag 1200
gcttaaagtt gatacttcta accctaagac acccaagtat aaatttgtcc gtatccaacc 1260
tggtaaaca ttttcagttc tagcatgcta caatggttca ccatctggtg tttatcagtg 1320
tgccatgaga cctaatacata ccattaaagg ttctttcctt aatggatcat gtggtagtgt 1380
tggttttaac attgattatg attgctgtgc tttctgctat atgcatcata tggagcttcc 1440
aacaggagta cacgctggta ctgacttaga aggtaaattc tatggtccat ttgttgacag 1500
acaaactgca caggctgcag gtacagacac aaccataaca ttaaatgttt tggcatggct 1560
gtatgctgct gttatcaatg gtgataggtg gtttcttaat agattcacca ctactttgaa 1620
tgactttaac cttgtggcaa tgaagtacaa ctatgaacct ttgacacaag atcatgttga 1680
catattggga cctctttctg ctcaaacagg aattgccgtc ttagatatgt gtgctgcttt 1740
gaaagagctg ctgcagaatg gtatgaatgg tcgtactatc cttggtagca ctattttaga 1800
agatgagttt acaccatttg atgttgttag acaatgctct ggtgttacct tccaaggtaa 1860
gttcaagaaa attgttaagg gcaactcatca ttggatgctt ttaactttct tgacatcact 1920
attgattcct gttcaaaagta cacagtggtc actgttttct tttgtttacg agaatgcttt 1980
cttgccattt actcttggtg ttatggcaat tgctgcatgt 2020

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<210> SEQ ID NO 49

<211> LENGTH: 2040

<212> TYPE: DNA

<213> ORGANISM: CORONAVIRUS

<400> SEQUENCE: 49

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agcatttcca gcctgaagac gtactgtagc agctaaactg cccagcacca tacctctatt 60
taggttggtt aagcctttga tgaagtacaa gtatttcact ttaggccctt ttggtgtgtc 120
tgtaacaaac ctacaagggt gttccagttc tgtgtaaatt gtacctgtac catcactcct 180
agggaatcta gcccttttga gatcttgggt gtctgatagt aatgccagca caaacctacc 240
tcccctcgaa ttgttatagt aggcaagtgc attgtcatca gtacaagctg tttgtgtggt 300
accagccgca caggacatct gtcgtagtgc tactggactc agttcattat tctgtagtgt 360
aacagctgag ttggctctta gagctgtaac aataagaggc caagccaaat ttggtgaatt 420
gtccatgtta atttcaacta gtggaacaat cttgctatcc gcatcaacaa cttgctggat 480
ttcccagagt gcagatgcat atgtaaagggt gttaccatca caagtgttct tgtaggtacc 540
ataatcaggg acaacaacca tgagtttggc tgctgtagtc aatggatga tgttgagtgg 600
aacacaacca tcacgctgat tgttgataat gttgttaagt gcatcattat caagcttcct 660
aagcatagtg aagagcattg tttgcatagc actagttact tttgccctct tgcctcaga 720
tcttgctgtt ttgtacattt gggctatagc ctgatctgcc atcttttoca acttgcgttg 780
catggcagca tcacggctca actcagattt agccacattc aaagatttct ttaacttttt 840
gagaacgact tcagaatcac cattagctac agcctgctca taggcctcct gggcagtggc 900
ataagcggca tatgatggta aagaactaaa ttctgaagca atagcctgaa gagtagcag 960
gttatcgagc atttctctgc acaacctatt aatgtctaca gcaccctgca tggatagcaa 1020
aacagacaaa agagaaacca tcttctcgaa agcttcagtt gtgtcttttg caagaagaat 1080

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atcattgtgg agttgtacac attgtgcca caatttagaa gatgactcta ctctaagttg 1140
ttgaagaacc gagagcagta ccacagatgt gcactttacg tcagacattt tagactgtac 1200
agtagcaacc ttgatacatg gtttacctcc aatacccaac aacttaatgt taagcttgaa 1260
agcatcaata ctactcttag gaggcaaaag cccctgggag ttcataatacc taaattcttg 1320
tgtagagacc aagtagtcat aaacaccaag agtaagcctg aagtaacggt tgagtaaaca 1380
gaaaaggcca aagtagcagc agcaacaata gcctaagaaa caataaaca gcatgatata 1440
ctgtaagggtg ttgccagtaa taaataaaca tgggtaatac tcaacacaca caaacactat 1500
agctctagct aaaaacatga tagtcgtaac gacaccagaa tagttagagg ttacagaaat 1560
aactaaggcc cacatggaaa tagcttgatc taaagcatta ccatagtaga ctttgtaaac 1620
aagtgtaatg acattcatca gtgtccaac acgtctagca gcatcatcat aaacagtgcg 1680
agctgtcatg agaataagca aaactaaagc tgaagcatac ataacacaat ccttaagcct 1740
ataaccagac aagctagtgt cagccaattc aagccatgtc atgatacga tcaccagct 1800
agcaggcatg tagaccatat taaagtaagc aactgttgca agagaaggta acagaaacaa 1860
gcacaagaat gcgtgcttat gcttaacaag cagcatagca catgcagcaa ttgccataat 1920
accaagagta aatggcaaga aagcattctc gtaacaaaag aaaaacagtg accactgtgt 1980
actttgaaca agaatcaata gtgatgtcaa gaaagttaa agcatccaat gatgagtga 2040

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<210> SEQ ID NO 50

<211> LENGTH: 2012

<212> TYPE: DNA

<213> ORGANISM: CORONAVIRUS

<400> SEQUENCE: 50

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cttgtaggtt tttacagac acacaaaag ggcctaaagt gaaatacttg tacttcatca 60
aaggcttaa caacctaat agaggtagt tgctgggagc tttagctgct acagtacgct 120
ttcaggctgg aaatgctaca gaagtacctg ccaattcaac tgtgctttcc ttctgtgctt 180
ttgcagtaga ccctgctaaa gcatataagg attacctagc aagtggagga caaccaatca 240
ccaactgtgt gaagatgttg tgtacacaca ctggtacagg acaggcaatt actgtaacac 300
cagaagctaa catggacca gagtcctttg gtggtgcttc atgttgtctg tattgtagat 360
gccacattga ccatccaaat cctaaaggat tctgtgactt gaaaggtaa tacgtccaaa 420
tacctaccac ttgtgcta atgacctagg gttttact tagaacaca gtctgtaccg 480
tctgcggaat gtggaagggt tatggctgta gttgtgacca actccgcaa ccctgatgc 540
agtctcgga tgcatacaac tttttaaagc ggtttgcggt gtaagtgcag cccgtcttac 600
accgtgcggc acaggcacta gtactgatgt cgtctacagg gcttttgata ttacaacga 660
aaaagttgct ggttttgcaa agttcctaaa aactaattgc tgtcgcttcc aggagaagga 720
tgaggaaggc aatttattag actcttactt tgtagttaag aggcatacta tgtctaacta 780
ccaacatgaa gagactattt ataacttggt taaagattgt ccagcggttg ctgtccatga 840
ctttttcaag tttagagtag atggtgacat ggtaccacat atatcacgtc agcgtctaac 900
taaatacaca atggctgatt tagtctatgc tctacgtcat tttgatgagg gtaattgtga 960
tacattaaaa gaaatactcg tcacatacaa ttgctgtgat gatgattatt tcaataagaa 1020
ggattgggat gacttcgtag agaactccta catcttacgc gtatatgcta acttaggtga 1080

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gcgtgtacgc caatcattat taaagactgt acaattctgc gatgctatgc gtgatgcagg 1140
cattgttaggc gtactgacat tagataatca ggatcttaat gggaaactggg acgatttcgg 1200
tgatttcgta caagtagcac caggctgcgg agttcctatt gtggattcat attactcatt 1260
gctgatgccc atcctcactt tgactagggc attggctgct gagtcccata tggatgctga 1320
tctcgcaaaa coacttatta agtgggattt gctgaaatat gattttacgg aagagagact 1380
ttgtctcttc gaccgttatt ttaaattatt ggaccagaca taccatccca attgtattaa 1440
ctgtttggat gataggtgta tccttcattg tgcaaaacttt aatgtgttat tttctactgt 1500
gtttccacct acaagttttg gaccactagt aagaaaaata tttgtagatg gtgttccttt 1560
tgttgtttca actggatacc attttcgta gttaggagtc gtacataatc aggatgtaa 1620
cttacaatagc tcgctctca gtttcaagga acttttagtg tatgctgctg atccagctat 1680
gcatgcagct tctggcaatt tattgctaga taaacgcact acatgctttt cagtagctgc 1740
actaacaaac aatgttgctt tcaaaactgt caaacccggg aattttaata aagactttta 1800
tgactttgct gtgtctaaag gtttctttaa ggaaggaagt tctgttgaac taaaacactt 1860
cttctttgct caggatggca acgctgctat cagtgattat gactattatc gttataatct 1920
gccaacaatg tgtgatatca gacaactcct attcgtagtt gaagtgttg ataaactctt 1980
tgattgttac gatggtggct gtattaatgc ca 2012

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<210> SEQ ID NO 51

<211> LENGTH: 1877

<212> TYPE: DNA

<213> ORGANISM: CORONAVIRUS

<400> SEQUENCE: 51

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gtacttcgcg tacagtgga ataccatag acagcttaa tgtttcctca gtggctttga 60
gcgtttctgc tgcgaaaagc ttgagtctct cagtacaagt gttggcaagt atgtaatcgc 120
cagcattagt ccaatcacat gttgctatcg cattgaagtc agtgacattg tcaactgccta 180
cacatgtggt tttgtataaa ccaaaaacct gaccattagc acataatgga aaactaatgg 240
gaggcttatg tgacttgcaa taatagctca tacctcctag atacagttgt gtcacatcag 300
tgacatcaca acctggggca ttgcaaacat agggattaac agacaacact aatttgtgtg 360
atgttgaaat gacatggta tagcagcact tgcaacatag gaatggtctc ctaatacagg 420
cacgcgaacg aagtgaagtc tgtgaattgc acaatacaca agcacctaca gcctgcaaga 480
ctgtatgtgg tgtgtacata gcctcataaa actcaggttc ccagtaccgt gaggtgttat 540
cattagttag cattacggaa tacatgtcca acatgtggcc agtaagctca tcatgtaact 600
ttctaatagta ttgtaataac aagtgaaga catcagcata ctctgatta ggatgttttg 660
taagtgggta agcatcaata gccagtgaca cgaacctttc aatcataagt gtaccatctg 720
ttttgacaat atcatcgaca aaacagcctg cgcctaatat tcttgatgga tctgggtaag 780
gcaggtacac gtaatcatct ccttgtttaa ctagcattgt atgctgtgag caaaattcgt 840
gaggtccttt agtaaggta gtctcagtc aacattttgc ctacagatg aacacattat 900
tttgataata aagaactgcc ttaaagtctt taatgctagc tactaaaacct tgagccgcat 960
agttactggt atagcacaca acggcatcat cagaagaat catcatggag aaatgtttac 1020
gcaggaagc gtaaaactca tccacgaatt catgatcaac atccctatct ctatagagac 1080

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actcatagag cctgtgttgt agattgcgga catacttgtc agctatctta ttaccatcag 1140
ttgaaagaag tgcatttaca ttggctgtaa cagcttgaca aatgttaaag aactattag 1200
cataagcagt tntagatca cggatgatg ttccacctgg ttaacatat agtgagccgc 1260
cacacatgac catctcactt aatacttgcg cacactcgtt agctaacctg tagaaacggt 1320
gtgataagtt acagcaagtg ttatgtttgc gagcaagaac aagagaggcc attatcctaa 1380
gcatgttagg catggctctg tcacattttg gataatccca acccataagg tgtggagttt 1440
ctacatcact gtaaacagtt ttaacatat tatgccagcc accgtaaac ttgcttgctc 1500
caattaccac agtagctcct ctagtggcgg ctattgactt caataatttc tgatgaaact 1560
gtctatttgt catagtacta cagatagaga caccagctac ggtgcgagct ctattctttg 1620
cactaatggc atacttaaga ttcatttgag ttatagtagg gatgacatta cgcttagtat 1680
acgcgaaaag tgcactctga tcctcataac tcattgagtc ataataaagt ctagccttac 1740
cccatttatt aaatgggaaa ccagctgatt tatccagatt gttaacgatt acttggttg 1800
cattaataca gccaccatcg taacaatcaa agtatttatc aacaacttca actacgaata 1860
ggagttgtct gatatca 1877

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<210> SEQ ID NO 52

<211> LENGTH: 2051

<212> TYPE: DNA

<213> ORGANISM: CORONAVIRUS

<400> SEQUENCE: 52

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tcaggtccaa tcttgacaaa gtacttcatt gatgtaagct caaagccatg cgcccaaagg 60
acgaacacga ctctgtctga caatccttcc agtgtatcac tgagcatttg tactatctta 120
atacgcacta cattccaggg caagccttta tacatgagtg gtataagatg tttaaactgg 180
tcacctgggt gaggttttgc attaactctg gtgaattctg tgttattttc agtgtcaaca 240
taaccagtcg gtacagctac taagttaaca cctgtagaaa atcctagctg gagaggtagg 300
ttagtaccca cagcatctct agttgcatga cagccctcta catcaaagcc aatccacgca 360
cgaacgtgac gaatagcttc ttgcgggtg ataaacatat tagggtaacc attgacttgg 420
taattcattt tgaaacccat catagagatg agtctacggt aggtcatgtc ctttggtagt 480
cctggtagtg caacacataa tccttcagtc ttgaacttta tatcaacgct gaggtgtgta 540
ggtgcctgtg taggatgaag accagtaatg atcttactac agtccttaaa aagtccagtt 600
acattttctg cttgtaatgt agccacattg cgacgtggta tttctagact tgtaaattgc 660
agtttgcatt aaagatctct atcagacatt atgcacaaaa tgccaatttt tgcccttgtg 720
atagccacat tgaagcgggt gacattacaa gagtgtgctg tttcagtagt ttgtgtgaat 780
atgacatagt catattcaga accctgtgat gaatcaacag tctgcgtagg caatcctaag 840
atTTTTgaag ctacagcgtt ctgtgaatta taagtgaga taaaaacagc ttttctccaa 900
gcaggattgc gtgtaagaaa ttctcttaca acgcctatTT gaggtctggt gattgcagat 960
gaaacatcat gtgtaataac accctttagt aacattttga agcattgagc tgacttatcc 1020
ttgtgtgctt ttagcttatt gtcataaact aaagcactca cagtgtcaac aatttcagca 1080
ggacaacggc gacaagtTcc aaggaacatg tctggaccta ttgttttcat aagtctgcac 1140
actgaattaa aatattcttg ttctagtggt ccttagtca gcaatgtgcg gggggctggt 1200

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aattgagcag gatcgccaat atagacgtag tgttttgcac gaagtctagc attgacaaca 1260
ctcaagtcac aattagtagc catagagatt tcatcaaaga ctacaatgtc agcagttggt 1320
tctggcaatg catttacagt gcagaaaaca tactgttcta gtgttgaaat cactttgaa 1380
ttatcaaaac actctacgcg cgcacgcgca ggtatgattc tactacattt atctatgggc 1440
aaatatttta atgccttttc acatagggca tcaacagctg catgagagca tgcggtatac 1500
actatgcgag cagatgggta atagagagca agtccgatgg caaaatgact cttaccagta 1560
ccaggtggtc cttggagtgt agagtacttt tgcacgcca ccttttgata atttgcaaca 1620
ttgctagaaa actcatctga gatgttgagt gttgggtaca agccagtaat tctcacatag 1680
tgctcttgty gcactagagt aggtgcacta agtggcatta cagtgtgaga tgtcaacaca 1740
aagtaatcac caacattcaa cttgtatgtc gtagtacctc tgtacacaac agcatcacca 1800
tagtcacctt tttcaaaggt gtactctcca atctgtactt tactattttt agttacacgg 1860
taaccagtaa agacatagtt tctgttcaat ggtggtctag gttttccaac ctcccatgaa 1920
agatgcaatt ctctgtcaga gagtacttcg cgtacagtgg caataccata tgacagctta 1980
aatgtttcct cagtggcttt gagcgtttct gctgcgaaaa gcttgagtct ctcagtacaa 2040
gtgttgcaaa g 2051

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<210> SEQ ID NO 53

<211> LENGTH: 2075

<212> TYPE: DNA

<213> ORGANISM: CORONAVIRUS

<400> SEQUENCE: 53

```

tgctttagt tttgggtaga aggtttcaac atgtccatcc ttacaccaa gcatgaatga 60
aatttcagca tagtcaattg taaccttgac cacttttgaa atcactgaca aatcttgtga 120
ctttattatc tcgacaaagt catcaagtaa aagatcaatc acagaacaca cacattttga 180
tgaacctggt tgcgcatctg ttatgaagta atttttcact gtgctgtcca tagggataaa 240
atcctcta at taaagtggg aatcttgtga gcgcttggt aagcctatca ttaaatgaag 300
accgccaagt tgtccatgac tgaatctcc ataaacgatg tgttcgaagg catagccctc 360
gagcttatat cgctgtatga attcatccat agcgagctcg agaaagtcag tttccatttg 420
tgatctgggc ttaaaatcct ctaagtctct gctctgagta aagtagggtt caggcaactg 480
ttgaataatg ccgtctactt tcttaaagta gttaaactgt gtttttactg attctccaat 540
taatgtgact ccattgacgc tagcttgtgc tgggcccttt gaagggtgta gacctttgac 600
tgaaccttct gttattaaaa caccattacg ggcgtttcta aaaaggctta cctgtccttc 660
cactctacca tcaacaaga cagtaagtga agaacaagca ctctcagtag gtttcttggc 720
aatgtcagtc attgtgcaga cacctattgt agatacatgt gctggggctt ctcttttgta 780
gtcccagatt acagtattag cagcgatata aacacccaaa ttattgagta tcttaatctc 840
tggcactggt ttaatgttac gcttagccca aagctcaaat gcaacattaa caggaagtgt 900
tgtcttattt tcaaagatct ccacatcaat accatctacc tttgtgtaaa cagcattatt 960
aatgatggaa acaggtgctt cgccggcgtg tccatcaaag tgcctttat taacaacatt 1020
ataagccaca ttttctaaac tctgtaacct ggtaaatgta ttccacaggt tataagtatc 1080
aaattgtttg taaatccata ggctaaatcc agcagaaatc atcatattat atgcatccaa 1140

```

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gtactgtcgg tactcatttg catggtgtct gcaaacagca ccacctaaat tgcacgtgt	1200
aatacacgta gcagatttga gtggaacata atcaatatcc gacctactt gtttgccatg	1260
agactcacia ggactatcag aatagtaaaa gaaaggcaat tgctttaaata tagtaaatgc	1320
acttttatcg aaagctggag tgtggaatgc atgcttattc acatacaaac taccaccatc	1380
acagcctggt aagtccaagt ttgacaagac tcttgtgtca aacctacaca caattgcatt	1440
ggctgggtaa cgatcaacgt tacaattcca aaacaaacia acaccatcag tgaatttatc	1500
gtgatgtgta gcataagaat agaagagttc ctctattttg taagctttgt cactacatgg	1560
ctgagcatcg tagaacttcc attctacttc agcctgaggc acacactga tagcctttgg	1620
atccaatg tcatgaaga ctggaactt atcagcaagc aatgcagact tcacaaccat	1680
gtgtgttact tttctgcaag cagaattaac cctcagttca tctctataa tagggattc	1740
aacagaccia tcaacgcgct taacaaagca ctcatggact gctaaacatc tagtcatgat	1800
agcatcacia ctagccacat gtgcatttcc atgtacctgg caatgttggg catggttact	1860
ctgaaggta cccgtaaagc cccactgctg aacatcaatc ataaatgggt tatagacata	1920
gtcaaaacc acagaatgat tccagcagc ataagtatct gatgaagtag aaaagcaagt	1980
tgacgtttg tcacacagac aacacgttct ttcaggtcca atcttgacia agtacttcat	2040
tgatgtaagc tcaaagccat gcgccaaag gacga	2075

<210> SEQ ID NO 54

<211> LENGTH: 1891

<212> TYPE: DNA

<213> ORGANISM: CORONAVIRUS

<400> SEQUENCE: 54

aagattcacc acttaaatga gaggatttta tccctatgga cagcacagtg aaaaattact	60
tcataacaga tgcgcaaaca ggttcatcaa aatgtgtgtg ttctgtgatt gatcttttac	120
ttgatgactt tgtcagata ataaagtcac aagatttgtc agtgatttca aaagtggta	180
aggttacaat tgactatgct gaaatttcat tcatgctttg gtgtaaggat ggacatggtg	240
aaacttcta cccaaaacta caagcaagtc aagcgtggca accaggtgtt gcgatgccta	300
acttgtaaa gatgcaaaga atgcttcttg aaaagtgtga ccttcagaat tatggtgaaa	360
atgctgttat accaaaagga ataatgatga atgtcgaaa gtatactcaa ctgtgtcaat	420
acttaaatc acttacttta gctgtaccct acaacatgag agttattcac tttggtgctg	480
gctctgataa aggagttgca ccaggtacag ctgtgctcag acaatggttg ccaactggca	540
cactacttgt cgattcagat cttaatgact tctctccga cgcagattct actttaattg	600
gagactgtgc aacagtcac acggctaata aatgggacct tattattagc gatatgtatg	660
accctaggac caaacatgtg acaaaagaga atgactctaa agaagggttt ttcacttatc	720
tgtgtggatt tataaagcaa aaactagccc tgggtggttc tatagctgta aagataacag	780
agcattcttg gaatcgtgac ctttacaagc ttatgggcca tttctcatgg tggacagctt	840
ttgttaciaa tgtaaatgca tcatcatcgg aagcattttt aattggggct aactatcttg	900
gcaagcggaa ggaacaaatt gatggctata ccatgcatgc taactacatt ttctggagga	960
acacaaatcc tatccagttg tcttctatt cactctttga catgagcaaa tttctctta	1020
aattaagagg aactgctgta atgtctctta aggagaatca aatcaatgat atgatttatt	1080

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ctcttctgga aaaaggtagc cttatcatta gagaaaacaa cagagttgtg gtttcaagtg 1140
atattcttgt taacaactaa acgaacatgt ttattttctt attatttctt actctcacta 1200
gtggtagtga ccttgaccgg tgcaccactt ttgatgatgt tcaagctcct aattacactc 1260
aacatacttc atctatgagg ggggtttact atcctgatga aattttttaga tcagacactc 1320
tttatttaac tcaggattta tttcttccat tttattctaa tgettacaggg tttcactacta 1380
ttaatcatac gtttggaac cctgtcatac cttttaagga tggatttat tttgctgcca 1440
cagagaaatc aaatgtgtgc cgtgggtggg tttttggttc taccatgaac aacaagtcac 1500
agtcggtgat tattattaac aattctacta atgttggtat acgagcatgt aactttgaat 1560
tgtgtgacaa cctttctttt gctgtttcta aacctatggg tacacagaca cactactatga 1620
tattcgataa tgcatttaac tgcactttcg agtacatata tgatgccttt tcgcttgatg 1680
tttcagaaaa gtcaggtaac tttaaacact tacgagagtt tgtgtttaaa aataaagatg 1740
ggtttctcta tgtttataag ggctatcaac ctatagatgt agttcgtgat ctaccttctg 1800
gttttaaacac ttgaaacct atttttaagt tgcctcttgg tattaacatt acaaatttta 1860
gagccattct tacagccttt tcacctgctc a 1891

```

```

<210> SEQ ID NO 55
<211> LENGTH: 32
<212> TYPE: DNA
<213> ORGANISM: artificial sequence
<220> FEATURE:
<223> OTHER INFORMATION: N sens primer

```

```
<400> SEQUENCE: 55
```

```
cccatatgtc tgataatgga cccaatcaa ac 32
```

```

<210> SEQ ID NO 56
<211> LENGTH: 32
<212> TYPE: DNA
<213> ORGANISM: artificial sequence
<220> FEATURE:
<223> OTHER INFORMATION: N antisens primer

```

```
<400> SEQUENCE: 56
```

```
cccccggtg cctgagttga atcagcagaa gc 32
```

```

<210> SEQ ID NO 57
<211> LENGTH: 31
<212> TYPE: DNA
<213> ORGANISM: artificial sequence
<220> FEATURE:
<223> OTHER INFORMATION: Sc sens primer

```

```
<400> SEQUENCE: 57
```

```
cccatatgag tgaccttgac cggtgacca c 31
```

```

<210> SEQ ID NO 58
<211> LENGTH: 30
<212> TYPE: DNA
<213> ORGANISM: artificial sequence
<220> FEATURE:
<223> OTHER INFORMATION: SL sens primer

```

```
<400> SEQUENCE: 58
```

```
cccatatgaa accttgacc ccacctgctc 30
```

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<210> SEQ ID NO 59
<211> LENGTH: 33
<212> TYPE: DNA
<213> ORGANISM: Sc and SL antisens primer

<400> SEQUENCE: 59

cccccggtt taatatattg ctcatatattt ccc 33

<210> SEQ ID NO 60
<211> LENGTH: 16
<212> TYPE: DNA
<213> ORGANISM: Sens set 1 primer

<400> SEQUENCE: 60

ggcatcgtat gggttg 16

<210> SEQ ID NO 61
<211> LENGTH: 16
<212> TYPE: DNA
<213> ORGANISM: Antisens set 2 (28774-28759) primer

<400> SEQUENCE: 61

cagtttcacc acctcc 16

<210> SEQ ID NO 62
<211> LENGTH: 16
<212> TYPE: DNA
<213> ORGANISM: Sens set 2 (28375-28390) primer

<400> SEQUENCE: 62

ggctactacc gaagag 16

<210> SEQ ID NO 63
<211> LENGTH: 16
<212> TYPE: DNA
<213> ORGANISM: Antisens set 2 (28702-28687)primer

<400> SEQUENCE: 63

aattaccgcg actacg 16

<210> SEQ ID NO 64
<211> LENGTH: 26
<212> TYPE: DNA
<213> ORGANISM: Probe 1/set 1 (28561-28586)

<400> SEQUENCE: 64

ggcaccgca atcctaataa caatgc 26

<210> SEQ ID NO 65
<211> LENGTH: 21
<212> TYPE: DNA
<213> ORGANISM: Probe 2/set 1 (28588-28608)

<400> SEQUENCE: 65

gccaccgtgc tacaacttcc t 21

<210> SEQ ID NO 66
<211> LENGTH: 23
<212> TYPE: DNA
<213> ORGANISM: Probe 1/set 2 /probe N/FL (28541-28563)

-continued

<400> SEQUENCE: 66

atacacccaa agaccacatt ggc 23

<210> SEQ ID NO 67

<211> LENGTH: 25

<212> TYPE: DNA

<213> ORGANISM: Probe 2/set 2/probe SARS/N/LC705 (28565-28589)

<400> SEQUENCE: 67

cccgcaatcc taataacaat gctgc 25

<210> SEQ ID NO 68

<211> LENGTH: 30

<212> TYPE: DNA

<213> ORGANISM: artificial sequence

<220> FEATURE:

<223> OTHER INFORMATION: Anchor primer 14T

<400> SEQUENCE: 68

agatgaattc ggtacctttt tttttttttt 30

<210> SEQ ID NO 69

<211> LENGTH: 13

<212> TYPE: PRT

<213> ORGANISM: artificial sequence

<220> FEATURE:

<223> OTHER INFORMATION: M2-14 peptide

<400> SEQUENCE: 69

Ala Asp Asn Gly Thr Ile Thr Val Glu Glu Leu Lys Gln
1 5 10

<210> SEQ ID NO 70

<211> LENGTH: 12

<212> TYPE: PRT

<213> ORGANISM: artificial sequence

<220> FEATURE:

<223> OTHER INFORMATION: E1-12 peptide

<400> SEQUENCE: 70

Met Tyr Ser Phe Val Ser Glu Glu Thr Gly Thr Leu
1 5 10

<210> SEQ ID NO 71

<211> LENGTH: 24

<212> TYPE: PRT

<213> ORGANISM: artificial sequence

<220> FEATURE:

<223> OTHER INFORMATION: E53-72 peptide

<400> SEQUENCE: 71

Lys Pro Thr Val Tyr Val Tyr Ser Arg Val Lys Asn Leu Asn Ser Ser
1 5 10 15

Glu Gly Val Pro Asp Leu Leu Val
20

<210> SEQ ID NO 72

<211> LENGTH: 153

<212> TYPE: DNA

<213> ORGANISM: CORONAVIRUS

<400> SEQUENCE: 72

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```

gatattaggt ttttacctac ccaggaaaag ccaaccaacc tcgattctctt gtagatctgt    60
tctctaaacg aactttaaaa tctgtgtagc tgctgctcgg ctgcatgcct agtgcaccta    120
cgcagtataa acaataataa attttactgt cgt                                  153

```

```

<210> SEQ ID NO 73
<211> LENGTH: 410
<212> TYPE: DNA
<213> ORGANISM: CORONAVIRUS

```

```

<400> SEQUENCE: 73

```

```

ttctccagac aacttcaaaa ttccatgagt ggagcttctg ctgattcaac tcaggcataa    60
acactcatga tgaccacaca aggcagatgg gctatgtaaa cgttttcgcg attccgttta    120
cgatacatag tctactcttg tgcagaatga attctcgtaa ctaaacagca caagtagggt    180
tagttaactt taatctcaca tagcaatctt taatcaatgt gtaacattag ggaggacttg    240
aaagagccac cacattttca tcgaggccac gcggagtacg atcgagggta cagtgaataa    300
tgctaggggag agctgcctat atggaagagc cctaattgtgt aaaattaatt ttagtagtgc    360
tatccccatg tgattttaat agcttcttag gagaatgaca aaaaaaaaaa                410

```

```

<210> SEQ ID NO 74
<211> LENGTH: 4382
<212> TYPE: PRT
<213> ORGANISM: CORONAVIRUS

```

```

<400> SEQUENCE: 74

```

```

Met Glu Ser Leu Val Leu Gly Val Asn Glu Lys Thr His Val Gln Leu
 1           5           10          15
Ser Leu Pro Val Leu Gln Val Arg Asp Val Leu Val Arg Gly Phe Gly
 20          25          30
Asp Ser Val Glu Glu Ala Leu Ser Glu Ala Arg Glu His Leu Lys Asn
 35          40          45
Gly Thr Cys Gly Leu Val Glu Leu Glu Lys Gly Val Leu Pro Gln Leu
 50          55          60
Glu Gln Pro Tyr Val Phe Ile Lys Arg Ser Asp Ala Leu Ser Thr Asn
 65          70          75          80
His Gly His Lys Val Val Glu Leu Val Ala Glu Met Asp Gly Ile Gln
 85          90          95
Tyr Gly Arg Ser Gly Ile Thr Leu Gly Val Leu Val Pro His Val Gly
100         105         110
Glu Thr Pro Ile Ala Tyr Arg Asn Val Leu Leu Arg Lys Asn Gly Asn
115         120         125
Lys Gly Ala Gly Gly His Ser Tyr Gly Ile Asp Leu Lys Ser Tyr Asp
130         135         140
Leu Gly Asp Glu Leu Gly Thr Asp Pro Ile Glu Asp Tyr Glu Gln Asn
145         150         155         160
Trp Asn Thr Lys His Gly Ser Gly Ala Leu Arg Glu Leu Thr Arg Glu
165         170         175
Leu Asn Gly Gly Ala Val Thr Arg Tyr Val Asp Asn Asn Phe Cys Gly
180         185         190
Pro Asp Gly Tyr Pro Leu Asp Cys Ile Lys Asp Phe Leu Ala Arg Ala
195         200         205
Gly Lys Ser Met Cys Thr Leu Ser Glu Gln Leu Asp Tyr Ile Glu Ser

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210					215					220									
Lys	Arg	Gly	Val	Tyr	Cys	Cys	Arg	Asp	His	Glu	His	Glu	Ile	Ala	Trp	225	230	235	240
Phe	Thr	Glu	Arg	Ser	Asp	Lys	Ser	Tyr	Glu	His	Gln	Thr	Pro	Phe	Glu	245	250	255	
Ile	Lys	Ser	Ala	Lys	Lys	Phe	Asp	Thr	Phe	Lys	Gly	Glu	Cys	Pro	Lys	260	265	270	
Phe	Val	Phe	Pro	Leu	Asn	Ser	Lys	Val	Lys	Val	Ile	Gln	Pro	Arg	Val	275	280	285	
Glu	Lys	Lys	Lys	Thr	Glu	Gly	Phe	Met	Gly	Arg	Ile	Arg	Ser	Val	Tyr	290	295	300	
Pro	Val	Ala	Ser	Pro	Gln	Glu	Cys	Asn	Asn	Met	His	Leu	Ser	Thr	Leu	305	310	315	320
Met	Lys	Cys	Asn	His	Cys	Asp	Glu	Val	Ser	Trp	Gln	Thr	Cys	Asp	Phe	325	330	335	
Leu	Lys	Ala	Thr	Cys	Glu	His	Cys	Gly	Thr	Glu	Asn	Leu	Val	Ile	Glu	340	345	350	
Gly	Pro	Thr	Thr	Cys	Gly	Tyr	Leu	Pro	Thr	Asn	Ala	Val	Val	Lys	Met	355	360	365	
Pro	Cys	Pro	Ala	Cys	Gln	Asp	Pro	Glu	Ile	Gly	Pro	Glu	His	Ser	Val	370	375	380	
Ala	Asp	Tyr	His	Asn	His	Ser	Asn	Ile	Glu	Thr	Arg	Leu	Arg	Lys	Gly	385	390	395	400
Gly	Arg	Thr	Arg	Cys	Phe	Gly	Gly	Cys	Val	Phe	Ala	Tyr	Val	Gly	Cys	405	410	415	
Tyr	Asn	Lys	Arg	Ala	Tyr	Trp	Val	Pro	Arg	Ala	Ser	Ala	Asp	Ile	Gly	420	425	430	
Ser	Gly	His	Thr	Gly	Ile	Thr	Gly	Asp	Asn	Val	Glu	Thr	Leu	Asn	Glu	435	440	445	
Asp	Leu	Leu	Glu	Ile	Leu	Ser	Arg	Glu	Arg	Val	Asn	Ile	Asn	Ile	Val	450	455	460	
Gly	Asp	Phe	His	Leu	Asn	Glu	Glu	Val	Ala	Ile	Ile	Leu	Ala	Ser	Phe	465	470	475	480
Ser	Ala	Ser	Thr	Ser	Ala	Phe	Ile	Asp	Thr	Ile	Lys	Ser	Leu	Asp	Tyr	485	490	495	
Lys	Ser	Phe	Lys	Thr	Ile	Val	Glu	Ser	Cys	Gly	Asn	Tyr	Lys	Val	Thr	500	505	510	
Lys	Gly	Lys	Pro	Val	Lys	Gly	Ala	Trp	Asn	Ile	Gly	Gln	Gln	Arg	Ser	515	520	525	
Val	Leu	Thr	Pro	Leu	Cys	Gly	Phe	Pro	Ser	Gln	Ala	Ala	Gly	Val	Ile	530	535	540	
Arg	Ser	Ile	Phe	Ala	Arg	Thr	Leu	Asp	Ala	Ala	Asn	His	Ser	Ile	Pro	545	550	555	560
Asp	Leu	Gln	Arg	Ala	Ala	Val	Thr	Ile	Leu	Asp	Gly	Ile	Ser	Glu	Gln	565	570	575	
Ser	Leu	Arg	Leu	Val	Asp	Ala	Met	Val	Tyr	Thr	Ser	Asp	Leu	Leu	Thr	580	585	590	
Asn	Ser	Val	Ile	Ile	Met	Ala	Tyr	Val	Thr	Gly	Gly	Leu	Val	Gln	Gln	595	600	605	
Thr	Ser	Gln	Trp	Leu	Ser	Asn	Leu	Leu	Gly	Thr	Thr	Val	Glu	Lys	Leu	610	615	620	

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Arg Pro Ile Phe Glu Trp Ile Glu Ala Lys Leu Ser Ala Gly Val Glu
 625 630 635 640
 Phe Leu Lys Asp Ala Trp Glu Ile Leu Lys Phe Leu Ile Thr Gly Val
 645 650 655
 Phe Asp Ile Val Lys Gly Gln Ile Gln Val Ala Ser Asp Asn Ile Lys
 660 665 670
 Asp Cys Val Lys Cys Phe Ile Asp Val Val Asn Lys Ala Leu Glu Met
 675 680 685
 Cys Ile Asp Gln Val Thr Ile Ala Gly Ala Lys Leu Arg Ser Leu Asn
 690 695 700
 Leu Gly Glu Val Phe Ile Ala Gln Ser Lys Gly Leu Tyr Arg Gln Cys
 705 710 715 720
 Ile Arg Gly Lys Glu Gln Leu Gln Leu Leu Met Pro Leu Lys Ala Pro
 725 730 735
 Lys Glu Val Thr Phe Leu Glu Gly Asp Ser His Asp Thr Val Leu Thr
 740 745 750
 Ser Glu Glu Val Val Leu Lys Asn Gly Glu Leu Glu Ala Leu Glu Thr
 755 760 765
 Pro Val Asp Ser Phe Thr Asn Gly Ala Ile Val Gly Thr Pro Val Cys
 770 775 780
 Val Asn Gly Leu Met Leu Leu Glu Ile Lys Asp Lys Glu Gln Tyr Cys
 785 790 795 800
 Ala Leu Ser Pro Gly Leu Leu Ala Thr Asn Asn Val Phe Arg Leu Lys
 805 810 815
 Gly Gly Ala Pro Ile Lys Gly Val Thr Phe Gly Glu Asp Thr Val Trp
 820 825 830
 Glu Val Gln Gly Tyr Lys Asn Val Arg Ile Thr Phe Glu Leu Asp Glu
 835 840 845
 Arg Val Asp Lys Val Leu Asn Glu Lys Cys Ser Val Tyr Thr Val Glu
 850 855 860
 Ser Gly Thr Glu Val Thr Glu Phe Ala Cys Val Val Ala Glu Ala Val
 865 870 875 880
 Val Lys Thr Leu Gln Pro Val Ser Asp Leu Leu Thr Asn Met Gly Ile
 885 890 895
 Asp Leu Asp Glu Trp Ser Val Ala Thr Phe Tyr Leu Phe Asp Asp Ala
 900 905 910
 Gly Glu Glu Asn Phe Ser Ser Arg Met Tyr Cys Ser Phe Tyr Pro Pro
 915 920 925
 Asp Glu Glu Glu Asp Asp Ala Glu Cys Glu Glu Glu Ile Asp
 930 935 940
 Glu Thr Cys Glu His Glu Tyr Gly Thr Glu Asp Asp Tyr Gln Gly Leu
 945 950 955 960
 Pro Leu Glu Phe Gly Ala Ser Ala Glu Thr Val Arg Val Glu Glu Glu
 965 970 975
 Glu Glu Glu Asp Trp Leu Asp Asp Thr Thr Glu Gln Ser Glu Ile Glu
 980 985 990
 Pro Glu Pro Glu Pro Thr Pro Glu Glu Pro Val Asn Gln Phe Thr Gly
 995 1000 1005
 Tyr Leu Lys Leu Thr Asp Asn Val Ala Ile Lys Cys Val Asp Ile
 1010 1015 1020

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Val Lys	Glu Ala Gln Ser	Ala Asn Pro Met Val	Ile Val Asn Ala
1025		1030	1035
Ala Asn	Ile His Leu Lys His	Gly Gly Gly Val Ala	Gly Ala Leu
1040		1045	1050
Asn Lys	Ala Thr Asn Gly Ala	Met Gln Lys Glu Ser	Asp Asp Tyr
1055		1060	1065
Ile Lys	Leu Asn Gly Pro Leu	Thr Val Gly Gly Ser	Cys Leu Leu
1070		1075	1080
Ser Gly	His Asn Leu Ala Lys	Lys Cys Leu His Val	Val Gly Pro
1085		1090	1095
Asn Leu	Asn Ala Gly Glu Asp	Ile Gln Leu Leu Lys	Ala Ala Tyr
1100		1105	1110
Glu Asn	Phe Asn Ser Gln Asp	Ile Leu Leu Ala Pro	Leu Leu Ser
1115		1120	1125
Ala Gly	Ile Phe Gly Ala Lys	Pro Leu Gln Ser Leu	Gln Val Cys
1130		1135	1140
Val Gln	Thr Val Arg Thr Gln	Val Tyr Ile Ala Val	Asn Asp Lys
1145		1150	1155
Ala Leu	Tyr Glu Gln Val Val	Met Asp Tyr Leu Asp	Asn Leu Lys
1160		1165	1170
Pro Arg	Val Glu Ala Pro Lys	Gln Glu Glu Pro Pro	Asn Thr Glu
1175		1180	1185
Asp Ser	Lys Thr Glu Glu Lys	Ser Val Val Gln Lys	Pro Val Asp
1190		1195	1200
Val Lys	Pro Lys Ile Lys Ala	Cys Ile Asp Glu Val	Thr Thr Thr
1205		1210	1215
Leu Glu	Glu Thr Lys Phe Leu	Thr Asn Lys Leu Leu	Leu Phe Ala
1220		1225	1230
Asp Ile	Asn Gly Lys Leu Tyr	His Asp Ser Gln Asn	Met Leu Arg
1235		1240	1245
Gly Glu	Asp Met Ser Phe Leu	Glu Lys Asp Ala Pro	Tyr Met Val
1250		1255	1260
Gly Asp	Val Ile Thr Ser Gly	Asp Ile Thr Cys Val	Val Ile Pro
1265		1270	1275
Ser Lys	Lys Ala Gly Gly Thr	Thr Glu Met Leu Ser	Arg Ala Leu
1280		1285	1290
Lys Lys	Val Pro Val Asp Glu	Tyr Ile Thr Thr Tyr	Pro Gly Gln
1295		1300	1305
Gly Cys	Ala Gly Tyr Thr Leu	Glu Glu Ala Lys Thr	Ala Leu Lys
1310		1315	1320
Lys Cys	Lys Ser Ala Phe Tyr	Val Leu Pro Ser Glu	Ala Pro Asn
1325		1330	1335
Ala Lys	Glu Glu Ile Leu Gly	Thr Val Ser Trp Asn	Leu Arg Glu
1340		1345	1350
Met Leu	Ala His Ala Glu Glu	Thr Arg Lys Leu Met	Pro Ile Cys
1355		1360	1365
Met Asp	Val Arg Ala Ile Met	Ala Thr Ile Gln Arg	Lys Tyr Lys
1370		1375	1380
Gly Ile	Lys Ile Gln Glu Gly	Ile Val Asp Tyr Gly	Val Arg Phe
1385		1390	1395
Phe Phe	Tyr Thr Ser Lys Glu	Pro Val Ala Ser Ile	Ile Thr Lys

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1400	1405	1410
Leu Asn Ser Leu Asn Glu Pro 1415	Leu Val Thr Met 1420	Pro Ile Gly Tyr 1425
Val Thr His Gly Phe Asn Leu 1430	Glu Glu Ala Ala 1435	Arg Cys Met Arg 1440
Ser Leu Lys Ala Pro Ala Val 1445	Val Ser Val Ser 1450	Ser Pro Asp Ala 1455
Val Thr Thr Tyr Asn Gly Tyr 1460	Leu Thr Ser Ser 1465	Ser Lys Thr Ser 1470
Glu Glu His Phe Val Glu Thr 1475	Val Ser Leu Ala 1480	Gly Ser Tyr Arg 1485
Asp Trp Ser Tyr Ser Gly Gln 1490	Arg Thr Glu Leu 1495	Gly Val Glu Phe 1500
Leu Lys Arg Gly Asp Lys Ile 1505	Val Tyr His Thr 1510	Leu Glu Ser Pro 1515
Val Glu Phe His Leu Asp Gly 1520	Glu Val Leu Ser 1525	Leu Asp Lys Leu 1530
Lys Ser Leu Leu Ser Leu Arg 1535	Glu Val Lys Thr 1540	Ile Lys Val Phe 1545
Thr Thr Val Asp Asn Thr Asn 1550	Leu His Thr Gln 1555	Leu Val Asp Met 1560
Ser Met Thr Tyr Gly Gln Gln 1565	Phe Gly Pro Thr 1570	Tyr Leu Asp Gly 1575
Ala Asp Val Thr Lys Ile Lys 1580	Pro His Val Asn 1585	His Glu Gly Lys 1590
Thr Phe Phe Val Leu Pro Ser 1595	Asp Asp Thr Leu 1600	Arg Ser Glu Ala 1605
Phe Glu Tyr Tyr His Thr Leu 1610	Asp Glu Ser Phe 1615	Leu Gly Arg Tyr 1620
Met Ser Ala Leu Asn His Thr 1625	Lys Lys Trp Lys 1630	Phe Pro Gln Val 1635
Gly Gly Leu Thr Ser Ile Lys 1640	Trp Ala Asp Asn 1645	Asn Cys Tyr Leu 1650
Ser Ser Val Leu Leu Ala Leu 1655	Gln Gln Leu Glu 1660	Val Lys Phe Asn 1665
Ala Pro Ala Leu Gln Glu Ala 1670	Tyr Tyr Arg Ala 1675	Arg Ala Gly Asp 1680
Ala Ala Asn Phe Cys Ala Leu 1685	Ile Leu Ala Tyr 1690	Ser Asn Lys Thr 1695
Val Gly Glu Leu Gly Asp Val 1700	Arg Glu Thr Met 1705	Thr His Leu Leu 1710
Gln His Ala Asn Leu Glu Ser 1715	Ala Lys Arg Val 1720	Leu Asn Val Val 1725
Cys Lys His Cys Gly Gln Lys 1730	Thr Thr Thr Leu 1735	Thr Gly Val Glu 1740
Ala Val Met Tyr Met Gly Thr 1745	Leu Ser Tyr Asp 1750	Asn Leu Lys Thr 1755
Gly Val Ser Ile Pro Cys Val 1760	Cys Gly Arg Asp 1765	Ala Thr Gln Tyr 1770
Leu Val Gln Gln Glu Ser Ser 1775	Phe Val Met Met 1780	Ser Ala Pro Pro 1785

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Ala	Glu	Tyr	Lys	Leu	Gln	Gln	Gly	Thr	Phe	Leu	Cys	Ala	Asn	Glu
1790						1795					1800			
Tyr	Thr	Gly	Asn	Tyr	Gln	Cys	Gly	His	Tyr	Thr	His	Ile	Thr	Ala
1805					1810						1815			
Lys	Glu	Thr	Leu	Tyr	Arg	Ile	Asp	Gly	Ala	His	Leu	Thr	Lys	Met
1820					1825						1830			
Ser	Glu	Tyr	Lys	Gly	Pro	Val	Thr	Asp	Val	Phe	Tyr	Lys	Glu	Thr
1835					1840						1845			
Ser	Tyr	Thr	Thr	Thr	Ile	Lys	Pro	Val	Ser	Tyr	Lys	Leu	Asp	Gly
1850					1855						1860			
Val	Thr	Tyr	Thr	Glu	Ile	Glu	Pro	Lys	Leu	Asp	Gly	Tyr	Tyr	Lys
1865					1870						1875			
Lys	Asp	Asn	Ala	Tyr	Tyr	Thr	Glu	Gln	Pro	Ile	Asp	Leu	Val	Pro
1880					1885						1890			
Thr	Gln	Pro	Leu	Pro	Asn	Ala	Ser	Phe	Asp	Asn	Phe	Lys	Leu	Thr
1895					1900						1905			
Cys	Ser	Asn	Thr	Lys	Phe	Ala	Asp	Asp	Leu	Asn	Gln	Met	Thr	Gly
1910					1915						1920			
Phe	Thr	Lys	Pro	Ala	Ser	Arg	Glu	Leu	Ser	Val	Thr	Phe	Phe	Pro
1925					1930						1935			
Asp	Leu	Asn	Gly	Asp	Val	Val	Ala	Ile	Asp	Tyr	Arg	His	Tyr	Ser
1940					1945						1950			
Ala	Ser	Phe	Lys	Lys	Gly	Ala	Lys	Leu	Leu	His	Lys	Pro	Ile	Val
1955					1960						1965			
Trp	His	Ile	Asn	Gln	Ala	Thr	Thr	Lys	Thr	Thr	Phe	Lys	Pro	Asn
1970					1975						1980			
Thr	Trp	Cys	Leu	Arg	Cys	Leu	Trp	Ser	Thr	Lys	Pro	Val	Asp	Thr
1985					1990						1995			
Ser	Asn	Ser	Phe	Glu	Val	Leu	Ala	Val	Glu	Asp	Thr	Gln	Gly	Met
2000					2005						2010			
Asp	Asn	Leu	Ala	Cys	Glu	Ser	Gln	Gln	Pro	Thr	Ser	Glu	Glu	Val
2015					2020						2025			
Val	Glu	Asn	Pro	Thr	Ile	Gln	Lys	Glu	Val	Ile	Glu	Cys	Asp	Val
2030					2035						2040			
Lys	Thr	Thr	Glu	Val	Val	Gly	Asn	Val	Ile	Leu	Lys	Pro	Ser	Asp
2045					2050						2055			
Glu	Gly	Val	Lys	Val	Thr	Gln	Glu	Leu	Gly	His	Glu	Asp	Leu	Met
2060					2065						2070			
Ala	Ala	Tyr	Val	Glu	Asn	Thr	Ser	Ile	Thr	Ile	Lys	Lys	Pro	Asn
2075					2080						2085			
Glu	Leu	Ser	Leu	Ala	Leu	Gly	Leu	Lys	Thr	Ile	Ala	Thr	His	Gly
2090					2095						2100			
Ile	Ala	Ala	Ile	Asn	Ser	Val	Pro	Trp	Ser	Lys	Ile	Leu	Ala	Tyr
2105					2110						2115			
Val	Lys	Pro	Phe	Leu	Gly	Gln	Ala	Ala	Ile	Thr	Thr	Ser	Asn	Cys
2120					2125						2130			
Ala	Lys	Arg	Leu	Ala	Gln	Arg	Val	Phe	Asn	Asn	Tyr	Met	Pro	Tyr
2135					2140						2145			
Val	Phe	Thr	Leu	Leu	Phe	Gln	Leu	Cys	Thr	Phe	Thr	Lys	Ser	Thr
2150					2155						2160			

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Asn Ser 2165	Arg Ile Arg Ala Ser 2170	Leu Pro Thr Thr 2175	Ile Ala Lys Asn 2175
Ser Val 2180	Lys Ser Val Ala Lys 2185	Leu Cys Leu Asp 2190	Ala Gly Ile Asn 2190
Tyr Val 2195	Lys Ser Pro Lys Phe 2200	Ser Lys Leu Phe Thr 2205	Ile Ala Met 2205
Trp Leu 2210	Leu Leu Leu Ser Ile 2215	Cys Leu Gly Ser Leu 2220	Ile Cys Val 2220
Thr Ala 2225	Ala Phe Gly Val Leu 2230	Leu Ser Asn Phe Gly 2235	Ala Pro Ser 2235
Tyr Cys 2240	Asn Gly Val Arg Glu 2245	Leu Tyr Leu Asn Ser 2250	Ser Asn Val 2250
Thr Thr 2255	Met Asp Phe Cys Glu 2260	Gly Ser Phe Pro Cys 2265	Ser Ile Cys 2265
Leu Ser 2270	Gly Leu Asp Ser Leu 2275	Asp Ser Tyr Pro Ala 2280	Leu Glu Thr 2280
Ile Gln 2285	Val Thr Ile Ser Ser 2290	Tyr Lys Leu Asp Leu 2295	Thr Ile Leu 2295
Gly Leu 2300	Ala Ala Glu Trp Val 2305	Leu Ala Tyr Met Leu 2310	Phe Thr Lys 2310
Phe Phe 2315	Tyr Leu Leu Gly Leu 2320	Ser Ala Ile Met Gln 2325	Val Phe Phe 2325
Gly Tyr 2330	Phe Ala Ser His Phe 2335	Ile Ser Asn Ser Trp 2340	Leu Met Trp 2340
Phe Ile 2345	Ile Ser Ile Val Gln 2350	Met Ala Pro Val Ser 2355	Ala Met Val 2355
Arg Met 2360	Tyr Ile Phe Phe Ala 2365	Ser Phe Tyr Tyr Ile 2370	Trp Lys Ser 2370
Tyr Val 2375	His Ile Met Asp Gly 2380	Cys Thr Ser Ser Thr 2385	Cys Met Met 2385
Cys Tyr 2390	Lys Arg Asn Arg Ala 2395	Thr Arg Val Glu Cys 2400	Thr Thr Ile 2400
Val Asn 2405	Gly Met Lys Arg Ser 2410	Phe Tyr Val Tyr Ala 2415	Asn Gly Gly 2415
Arg Gly 2420	Phe Cys Lys Thr His 2425	Asn Trp Asn Cys Leu 2430	Asn Cys Asp 2430
Thr Phe 2435	Cys Thr Gly Ser Thr 2440	Phe Ile Ser Asp Glu 2445	Val Ala Arg 2445
Asp Leu 2450	Ser Leu Gln Phe Lys 2455	Arg Pro Ile Asn Pro 2460	Thr Asp Gln 2460
Ser Ser 2465	Tyr Ile Val Asp Ser 2470	Val Ala Val Lys Asn 2475	Gly Ala Leu 2475
His Leu 2480	Tyr Phe Asp Lys Ala 2485	Gly Gln Lys Thr Tyr 2490	Glu Arg His 2490
Pro Leu 2495	Ser His Phe Val Asn 2500	Leu Asp Asn Leu Arg 2505	Ala Asn Asn 2505
Thr Lys 2510	Gly Ser Leu Pro Ile 2515	Asn Val Ile Val Phe 2520	Asp Gly Lys 2520
Ser Lys 2525	Cys Asp Glu Ser Ala 2530	Ser Lys Ser Ala Ser 2535	Val Tyr Tyr 2535
Ser Gln 2540	Leu Met Cys Gln Pro 2545	Ile Leu Leu Leu Asp 2550	Gln Ala Leu 2550

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Thr	Arg	Tyr	Val	Leu	Met	Asp	Gly	Ser	Ile	Ile	Gln	Phe	Pro	Asn
2930						2935					2940			
Thr	Tyr	Leu	Glu	Gly	Ser	Val	Arg	Val	Val	Thr	Thr	Phe	Asp	Ala
2945						2950					2955			
Glu	Tyr	Cys	Arg	His	Gly	Thr	Cys	Glu	Arg	Ser	Glu	Val	Gly	Ile
2960						2965					2970			
Cys	Leu	Ser	Thr	Ser	Gly	Arg	Trp	Val	Leu	Asn	Asn	Glu	His	Tyr
2975						2980					2985			
Arg	Ala	Leu	Ser	Gly	Val	Phe	Cys	Gly	Val	Asp	Ala	Met	Asn	Leu
2990						2995					3000			
Ile	Ala	Asn	Ile	Phe	Thr	Pro	Leu	Val	Gln	Pro	Val	Gly	Ala	Leu
3005						3010					3015			
Asp	Val	Ser	Ala	Ser	Val	Val	Ala	Gly	Gly	Ile	Ile	Ala	Ile	Leu
3020						3025					3030			
Val	Thr	Cys	Ala	Ala	Tyr	Tyr	Phe	Met	Lys	Phe	Arg	Arg	Val	Phe
3035						3040					3045			
Gly	Glu	Tyr	Asn	His	Val	Val	Ala	Ala	Asn	Ala	Leu	Leu	Phe	Leu
3050						3055					3060			
Met	Ser	Phe	Thr	Ile	Leu	Cys	Leu	Val	Pro	Ala	Tyr	Ser	Phe	Leu
3065						3070					3075			
Pro	Gly	Val	Tyr	Ser	Val	Phe	Tyr	Leu	Tyr	Leu	Thr	Phe	Tyr	Phe
3080						3085					3090			
Thr	Asn	Asp	Val	Ser	Phe	Leu	Ala	His	Leu	Gln	Trp	Phe	Ala	Met
3095						3100					3105			
Phe	Ser	Pro	Ile	Val	Pro	Phe	Trp	Ile	Thr	Ala	Ile	Tyr	Val	Phe
3110						3115					3120			
Cys	Ile	Ser	Leu	Lys	His	Cys	His	Trp	Phe	Phe	Asn	Asn	Tyr	Leu
3125						3130					3135			
Arg	Lys	Arg	Val	Met	Phe	Asn	Gly	Val	Thr	Phe	Ser	Thr	Phe	Glu
3140						3145					3150			
Glu	Ala	Ala	Leu	Cys	Thr	Phe	Leu	Leu	Asn	Lys	Glu	Met	Tyr	Leu
3155						3160					3165			
Lys	Leu	Arg	Ser	Glu	Thr	Leu	Leu	Pro	Leu	Thr	Gln	Tyr	Asn	Arg
3170						3175					3180			
Tyr	Leu	Ala	Leu	Tyr	Asn	Lys	Tyr	Lys	Tyr	Phe	Ser	Gly	Ala	Leu
3185						3190					3195			
Asp	Thr	Thr	Ser	Tyr	Arg	Glu	Ala	Ala	Cys	Cys	His	Leu	Ala	Lys
3200						3205					3210			
Ala	Leu	Asn	Asp	Phe	Ser	Asn	Ser	Gly	Ala	Asp	Val	Leu	Tyr	Gln
3215						3220					3225			
Pro	Pro	Gln	Thr	Ser	Ile	Thr	Ser	Ala	Val	Leu	Gln	Ser	Gly	Phe
3230						3235					3240			
Arg	Lys	Met	Ala	Phe	Pro	Ser	Gly	Lys	Val	Glu	Gly	Cys	Met	Val
3245						3250					3255			
Gln	Val	Thr	Cys	Gly	Thr	Thr	Thr	Leu	Asn	Gly	Leu	Trp	Leu	Asp
3260						3265					3270			
Asp	Thr	Val	Tyr	Cys	Pro	Arg	His	Val	Ile	Cys	Thr	Ala	Glu	Asp
3275						3280					3285			
Met	Leu	Asn	Pro	Asn	Tyr	Glu	Asp	Leu	Leu	Ile	Arg	Lys	Ser	Asn
3290						3295					3300			

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His Ser	Phe Leu Val Gln	Ala Gly Asn Val Gln	Leu Arg Val Ile
3305		3310	3315
Gly His	Ser Met Gln Asn Cys	Leu Leu Arg Leu Lys	Val Asp Thr
3320		3325	3330
Ser Asn	Pro Lys Thr Pro Lys	Tyr Lys Phe Val Arg	Ile Gln Pro
3335		3340	3345
Gly Gln	Thr Phe Ser Val Leu	Ala Cys Tyr Asn Gly	Ser Pro Ser
3350		3355	3360
Gly Val	Tyr Gln Cys Ala Met	Arg Pro Asn His Thr	Ile Lys Gly
3365		3370	3375
Ser Phe	Leu Asn Gly Ser Cys	Gly Ser Val Gly Phe	Asn Ile Asp
3380		3385	3390
Tyr Asp	Cys Val Ser Phe Cys	Tyr Met His His Met	Glu Leu Pro
3395		3400	3405
Thr Gly	Val His Ala Gly Thr	Asp Leu Glu Gly Lys	Phe Tyr Gly
3410		3415	3420
Pro Phe	Val Asp Arg Gln Thr	Ala Gln Ala Ala Gly	Thr Asp Thr
3425		3430	3435
Thr Ile	Thr Leu Asn Val Leu	Ala Trp Leu Tyr Ala	Ala Val Ile
3440		3445	3450
Asn Gly	Asp Arg Trp Phe Leu	Asn Arg Phe Thr Thr	Thr Leu Asn
3455		3460	3465
Asp Phe	Asn Leu Val Ala Met	Lys Tyr Asn Tyr Glu	Pro Leu Thr
3470		3475	3480
Gln Asp	His Val Asp Ile Leu	Gly Pro Leu Ser Ala	Gln Thr Gly
3485		3490	3495
Ile Ala	Val Leu Asp Met Cys	Ala Ala Leu Lys Glu	Leu Leu Gln
3500		3505	3510
Asn Gly	Met Asn Gly Arg Thr	Ile Leu Gly Ser Thr	Ile Leu Glu
3515		3520	3525
Asp Glu	Phe Thr Pro Phe Asp	Val Val Arg Gln Cys	Ser Gly Val
3530		3535	3540
Thr Phe	Gln Gly Lys Phe Lys	Lys Ile Val Lys Gly	Thr His His
3545		3550	3555
Trp Met	Leu Leu Thr Phe Leu	Thr Ser Leu Leu Ile	Leu Val Gln
3560		3565	3570
Ser Thr	Gln Trp Ser Leu Phe	Phe Phe Val Tyr Glu	Asn Ala Phe
3575		3580	3585
Leu Pro	Phe Thr Leu Gly Ile	Met Ala Ile Ala Ala	Cys Ala Met
3590		3595	3600
Leu Leu	Val Lys His Lys His	Ala Phe Leu Cys Leu	Phe Leu Leu
3605		3610	3615
Pro Ser	Leu Ala Thr Val Ala	Tyr Phe Asn Met Val	Tyr Met Pro
3620		3625	3630
Ala Ser	Trp Val Met Arg Ile	Met Thr Trp Leu Glu	Leu Ala Asp
3635		3640	3645
Thr Ser	Leu Ser Gly Tyr Arg	Leu Lys Asp Cys Val	Met Tyr Ala
3650		3655	3660
Ser Ala	Leu Val Leu Leu Ile	Leu Met Thr Ala Arg	Thr Val Tyr
3665		3670	3675
Asp Asp	Ala Ala Arg Arg Val	Trp Thr Leu Met Asn	Val Ile Thr

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3680	3685	3690
Leu Val Tyr Lys Val Tyr Tyr 3695	Gly Asn Ala Leu Asp 3700	Gln Ala Ile 3705
Ser Met Trp Ala Leu Val Ile 3710	Ser Val Thr Ser Asn 3715	Tyr Ser Gly 3720
Val Val Thr Thr Ile Met Phe 3725	Leu Ala Arg Ala Ile 3730	Val Phe Val 3735
Cys Val Glu Tyr Tyr Pro Leu 3740	Leu Phe Ile Thr Gly 3745	Asn Thr Leu 3750
Gln Cys Ile Met Leu Val Tyr 3755	Cys Phe Leu Gly Tyr 3760	Cys Cys Cys 3765
Cys Tyr Phe Gly Leu Phe Cys 3770	Leu Leu Asn Arg Tyr 3775	Phe Arg Leu 3780
Thr Leu Gly Val Tyr Asp Tyr 3785	Leu Val Ser Thr Gln 3790	Glu Phe Arg 3795
Tyr Met Asn Ser Gln Gly Leu 3800	Leu Pro Pro Lys Ser 3805	Ser Ile Asp 3810
Ala Phe Lys Leu Asn Ile Lys 3815	Leu Leu Gly Ile Gly 3820	Gly Lys Pro 3825
Cys Ile Lys Val Ala Thr Val 3830	Gln Ser Lys Met Ser 3835	Asp Val Lys 3840
Cys Thr Ser Val Val Leu Leu 3845	Ser Val Leu Gln Gln 3850	Leu Arg Val 3855
Glu Ser Ser Ser Lys Leu Trp 3860	Ala Gln Cys Val Gln 3865	Leu His Asn 3870
Asp Ile Leu Leu Ala Lys Asp 3875	Thr Thr Glu Ala Phe 3880	Glu Lys Met 3885
Val Ser Leu Leu Ser Val Leu 3890	Leu Ser Met Gln Gly 3895	Ala Val Asp 3900
Ile Asn Arg Leu Cys Glu Glu 3905	Met Leu Asp Asn Arg 3910	Ala Thr Leu 3915
Gln Ala Ile Ala Ser Glu Phe 3920	Ser Ser Leu Pro Ser 3925	Tyr Ala Ala 3930
Tyr Ala Thr Ala Gln Glu Ala 3935	Tyr Glu Gln Ala Val 3940	Ala Asn Gly 3945
Asp Ser Glu Val Val Leu Lys 3950	Lys Leu Lys Lys Ser 3955	Leu Asn Val 3960
Ala Lys Ser Glu Phe Asp Arg 3965	Asp Ala Ala Met Gln 3970	Arg Lys Leu 3975
Glu Lys Met Ala Asp Gln Ala 3980	Met Thr Gln Met Tyr 3985	Lys Gln Ala 3990
Arg Ser Glu Asp Lys Arg Ala 3995	Lys Val Thr Ser Ala 4000	Met Gln Thr 4005
Met Leu Phe Thr Met Leu Arg 4010	Lys Leu Asp Asn Asp 4015	Ala Leu Asn 4020
Asn Ile Ile Asn Asn Ala Arg 4025	Asp Gly Cys Val Pro 4030	Leu Asn Ile 4035
Ile Pro Leu Thr Thr Ala Ala 4040	Lys Leu Met Val Val 4045	Val Pro Asp 4050
Tyr Gly Thr Tyr Lys Asn Thr 4055	Cys Asp Gly Asn Thr 4060	Phe Thr Tyr 4065

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Ala Ser Ala Leu Trp Glu Ile Gln Gln Val Val Asp Ala Asp Ser
4070 4075 4080

Lys Ile Val Gln Leu Ser Glu Ile Asn Met Asp Asn Ser Pro Asn
4085 4090 4095

Leu Ala Trp Pro Leu Ile Val Thr Ala Leu Arg Ala Asn Ser Ala
4100 4105 4110

Val Lys Leu Gln Asn Asn Glu Leu Ser Pro Val Ala Leu Arg Gln
4115 4120 4125

Met Ser Cys Ala Ala Gly Thr Thr Gln Thr Ala Cys Thr Asp Asp
4130 4135 4140

Asn Ala Leu Ala Tyr Tyr Asn Asn Ser Lys Gly Gly Arg Phe Val
4145 4150 4155

Leu Ala Leu Leu Ser Asp His Gln Asp Leu Lys Trp Ala Arg Phe
4160 4165 4170

Pro Lys Ser Asp Gly Thr Gly Thr Ile Tyr Thr Glu Leu Glu Pro
4175 4180 4185

Pro Cys Arg Phe Val Thr Asp Thr Pro Lys Gly Pro Lys Val Lys
4190 4195 4200

Tyr Leu Tyr Phe Ile Lys Gly Leu Asn Asn Leu Asn Arg Gly Met
4205 4210 4215

Val Leu Gly Ser Leu Ala Ala Thr Val Arg Leu Gln Ala Gly Asn
4220 4225 4230

Ala Thr Glu Val Pro Ala Asn Ser Thr Val Leu Ser Phe Cys Ala
4235 4240 4245

Phe Ala Val Asp Pro Ala Lys Ala Tyr Lys Asp Tyr Leu Ala Ser
4250 4255 4260

Gly Gly Gln Pro Ile Thr Asn Cys Val Lys Met Leu Cys Thr His
4265 4270 4275

Thr Gly Thr Gly Gln Ala Ile Thr Val Thr Pro Glu Ala Asn Met
4280 4285 4290

Asp Gln Glu Ser Phe Gly Gly Ala Ser Cys Cys Leu Tyr Cys Arg
4295 4300 4305

Cys His Ile Asp His Pro Asn Pro Lys Gly Phe Cys Asp Leu Lys
4310 4315 4320

Gly Lys Tyr Val Gln Ile Pro Thr Thr Cys Ala Asn Asp Pro Val
4325 4330 4335

Gly Phe Thr Leu Arg Asn Thr Val Cys Thr Val Cys Gly Met Trp
4340 4345 4350

Lys Gly Tyr Gly Cys Ser Cys Asp Gln Leu Arg Glu Pro Leu Met
4355 4360 4365

Gln Ser Ala Asp Ala Ser Thr Phe Leu Asn Gly Phe Ala Val
4370 4375 4380

<210> SEQ ID NO 75

<211> LENGTH: 2695

<212> TYPE: PRT

<213> ORGANISM: CORONAVIRUS

<400> SEQUENCE: 75

Arg Val Cys Gly Val Ser Ala Ala Arg Leu Thr Pro Cys Gly Thr Gly
1 5 10 15

Thr Ser Thr Asp Val Val Tyr Arg Ala Phe Asp Ile Tyr Asn Glu Lys

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20		25		30											
Val	Ala	Gly	Phe	Ala	Lys	Phe	Leu	Lys	Thr	Asn	Cys	Cys	Arg	Phe	Gln
	35						40					45			
Glu	Lys	Asp	Glu	Glu	Gly	Asn	Leu	Leu	Asp	Ser	Tyr	Phe	Val	Val	Lys
	50					55					60				
Arg	His	Thr	Met	Ser	Asn	Tyr	Gln	His	Glu	Glu	Thr	Ile	Tyr	Asn	Leu
	65				70					75					80
Val	Lys	Asp	Cys	Pro	Ala	Val	Ala	Val	His	Asp	Phe	Phe	Lys	Phe	Arg
				85					90						95
Val	Asp	Gly	Asp	Met	Val	Pro	His	Ile	Ser	Arg	Gln	Arg	Leu	Thr	Lys
			100					105					110		
Tyr	Thr	Met	Ala	Asp	Leu	Val	Tyr	Ala	Leu	Arg	His	Phe	Asp	Glu	Gly
		115					120					125			
Asn	Cys	Asp	Thr	Leu	Lys	Glu	Ile	Leu	Val	Thr	Tyr	Asn	Cys	Cys	Asp
	130					135					140				
Asp	Asp	Tyr	Phe	Asn	Lys	Lys	Asp	Trp	Tyr	Asp	Phe	Val	Glu	Asn	Pro
	145				150					155					160
Asp	Ile	Leu	Arg	Val	Tyr	Ala	Asn	Leu	Gly	Glu	Arg	Val	Arg	Gln	Ser
				165					170					175	
Leu	Leu	Lys	Thr	Val	Gln	Phe	Cys	Asp	Ala	Met	Arg	Asp	Ala	Gly	Ile
			180						185					190	
Val	Gly	Val	Leu	Thr	Leu	Asp	Asn	Gln	Asp	Leu	Asn	Gly	Asn	Trp	Tyr
		195					200					205			
Asp	Phe	Gly	Asp	Phe	Val	Gln	Val	Ala	Pro	Gly	Cys	Gly	Val	Pro	Ile
	210					215					220				
Val	Asp	Ser	Tyr	Tyr	Ser	Leu	Leu	Met	Pro	Ile	Leu	Thr	Leu	Thr	Arg
	225				230					235					240
Ala	Leu	Ala	Ala	Glu	Ser	His	Met	Asp	Ala	Asp	Leu	Ala	Lys	Pro	Leu
				245					250					255	
Ile	Lys	Trp	Asp	Leu	Leu	Lys	Tyr	Asp	Phe	Thr	Glu	Glu	Arg	Leu	Cys
			260					265						270	
Leu	Phe	Asp	Arg	Tyr	Phe	Lys	Tyr	Trp	Asp	Gln	Thr	Tyr	His	Pro	Asn
		275					280					285			
Cys	Ile	Asn	Cys	Leu	Asp	Asp	Arg	Cys	Ile	Leu	His	Cys	Ala	Asn	Phe
	290					295					300				
Asn	Val	Leu	Phe	Ser	Thr	Val	Phe	Pro	Pro	Thr	Ser	Phe	Gly	Pro	Leu
	305				310					315					320
Val	Arg	Lys	Ile	Phe	Val	Asp	Gly	Val	Pro	Phe	Val	Val	Ser	Thr	Gly
				325					330					335	
Tyr	His	Phe	Arg	Glu	Leu	Gly	Val	Val	His	Asn	Gln	Asp	Val	Asn	Leu
			340					345					350		
His	Ser	Ser	Arg	Leu	Ser	Phe	Lys	Glu	Leu	Leu	Val	Tyr	Ala	Ala	Asp
		355					360					365			
Pro	Ala	Met	His	Ala	Ala	Ser	Gly	Asn	Leu	Leu	Leu	Asp	Lys	Arg	Thr
	370					375					380				
Thr	Cys	Phe	Ser	Val	Ala	Ala	Leu	Thr	Asn	Asn	Val	Ala	Phe	Gln	Thr
	385				390					395					400
Val	Lys	Pro	Gly	Asn	Phe	Asn	Lys	Asp	Phe	Tyr	Asp	Phe	Ala	Val	Ser
				405					410					415	
Lys	Gly	Phe	Phe	Lys	Glu	Gly	Ser	Ser	Val	Glu	Leu	Lys	His	Phe	Phe
			420					425					430		

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Phe Ala Gln Asp Gly Asn Ala Ala Ile Ser Asp Tyr Asp Tyr Tyr Arg
 435 440 445
 Tyr Asn Leu Pro Thr Met Cys Asp Ile Arg Gln Leu Leu Phe Val Val
 450 455 460
 Glu Val Val Asp Lys Tyr Phe Asp Cys Tyr Asp Gly Gly Cys Ile Asn
 465 470 475 480
 Ala Asn Gln Val Ile Val Asn Asn Leu Asp Lys Ser Ala Gly Phe Pro
 485 490 495
 Phe Asn Lys Trp Gly Lys Ala Arg Leu Tyr Tyr Asp Ser Met Ser Tyr
 500 505 510
 Glu Asp Gln Asp Ala Leu Phe Ala Tyr Thr Lys Arg Asn Val Ile Pro
 515 520 525
 Thr Ile Thr Gln Met Asn Leu Lys Tyr Ala Ile Ser Ala Lys Asn Arg
 530 535 540
 Ala Arg Thr Val Ala Gly Val Ser Ile Cys Ser Thr Met Thr Asn Arg
 545 550 555 560
 Gln Phe His Gln Lys Leu Leu Lys Ser Ile Ala Ala Thr Arg Gly Ala
 565 570 575
 Thr Val Val Ile Gly Thr Ser Lys Phe Tyr Gly Gly Trp His Asn Met
 580 585 590
 Leu Lys Thr Val Tyr Ser Asp Val Glu Thr Pro His Leu Met Gly Trp
 595 600 605
 Asp Tyr Pro Lys Cys Asp Arg Ala Met Pro Asn Met Leu Arg Ile Met
 610 615 620
 Ala Ser Leu Val Leu Ala Arg Lys His Asn Thr Cys Cys Asn Leu Ser
 625 630 635 640
 His Arg Phe Tyr Arg Leu Ala Asn Glu Cys Ala Gln Val Leu Ser Glu
 645 650 655
 Met Val Met Cys Gly Gly Ser Leu Tyr Val Lys Pro Gly Gly Thr Ser
 660 665 670
 Ser Gly Asp Ala Thr Thr Ala Tyr Ala Asn Ser Val Phe Asn Ile Cys
 675 680 685
 Gln Ala Val Thr Ala Asn Val Asn Ala Leu Leu Ser Thr Asp Gly Asn
 690 695 700
 Lys Ile Ala Asp Lys Tyr Val Arg Asn Leu Gln His Arg Leu Tyr Glu
 705 710 715 720
 Cys Leu Tyr Arg Asn Arg Asp Val Asp His Glu Phe Val Asp Glu Phe
 725 730 735
 Tyr Ala Tyr Leu Arg Lys His Phe Ser Met Met Ile Leu Ser Asp Asp
 740 745 750
 Ala Val Val Cys Tyr Asn Ser Asn Tyr Ala Ala Gln Gly Leu Val Ala
 755 760 765
 Ser Ile Lys Asn Phe Lys Ala Val Leu Tyr Tyr Gln Asn Asn Val Phe
 770 775 780
 Met Ser Glu Ala Lys Cys Trp Thr Glu Thr Asp Leu Thr Lys Gly Pro
 785 790 795 800
 His Glu Phe Cys Ser Gln His Thr Met Leu Val Lys Gln Gly Asp Asp
 805 810 815
 Tyr Val Tyr Leu Pro Tyr Pro Asp Pro Ser Arg Ile Leu Gly Ala Gly
 820 825 830

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Cys Phe Val Asp Asp Ile Val Lys Thr Asp Gly Thr Leu Met Ile Glu
 835 840 845

Arg Phe Val Ser Leu Ala Ile Asp Ala Tyr Pro Leu Thr Lys His Pro
 850 855 860

Asn Gln Glu Tyr Ala Asp Val Phe His Leu Tyr Leu Gln Tyr Ile Arg
 865 870 875 880

Lys Leu His Asp Glu Leu Thr Gly His Met Leu Asp Met Tyr Ser Val
 885 890 895

Met Leu Thr Asn Asp Asn Thr Ser Arg Tyr Trp Glu Pro Glu Phe Tyr
 900 905 910

Glu Ala Met Tyr Thr Pro His Thr Val Leu Gln Ala Val Gly Ala Cys
 915 920 925

Val Leu Cys Asn Ser Gln Thr Ser Leu Arg Cys Gly Ala Cys Ile Arg
 930 935 940

Arg Pro Phe Leu Cys Cys Lys Cys Cys Tyr Asp His Val Ile Ser Thr
 945 950 955 960

Ser His Lys Leu Val Leu Ser Val Asn Pro Tyr Val Cys Asn Ala Pro
 965 970 975

Gly Cys Asp Val Thr Asp Val Thr Gln Leu Tyr Leu Gly Gly Met Ser
 980 985 990

Tyr Tyr Cys Lys Ser His Lys Pro Pro Ile Ser Phe Pro Leu Cys Ala
 995 1000 1005

Asn Gly Gln Val Phe Gly Leu Tyr Lys Asn Thr Cys Val Gly Ser
 1010 1015 1020

Asp Asn Val Thr Asp Phe Asn Ala Ile Ala Thr Cys Asp Trp Thr
 1025 1030 1035

Asn Ala Gly Asp Tyr Ile Leu Ala Asn Thr Cys Thr Glu Arg Leu
 1040 1045 1050

Lys Leu Phe Ala Ala Glu Thr Leu Lys Ala Thr Glu Glu Thr Phe
 1055 1060 1065

Lys Leu Ser Tyr Gly Ile Ala Thr Val Arg Glu Val Leu Ser Asp
 1070 1075 1080

Arg Glu Leu His Leu Ser Trp Glu Val Gly Lys Pro Arg Pro Pro
 1085 1090 1095

Leu Asn Arg Asn Tyr Val Phe Thr Gly Tyr Arg Val Thr Lys Asn
 1100 1105 1110

Ser Lys Val Gln Ile Gly Glu Tyr Thr Phe Glu Lys Gly Asp Tyr
 1115 1120 1125

Gly Asp Ala Val Val Tyr Arg Gly Thr Thr Thr Tyr Lys Leu Asn
 1130 1135 1140

Val Gly Asp Tyr Phe Val Leu Thr Ser His Thr Val Met Pro Leu
 1145 1150 1155

Ser Ala Pro Thr Leu Val Pro Gln Glu His Tyr Val Arg Ile Thr
 1160 1165 1170

Gly Leu Tyr Pro Thr Leu Asn Ile Ser Asp Glu Phe Ser Ser Asn
 1175 1180 1185

Val Ala Asn Tyr Gln Lys Val Gly Met Gln Lys Tyr Ser Thr Leu
 1190 1195 1200

Gln Gly Pro Pro Gly Thr Gly Lys Ser His Phe Ala Ile Gly Leu
 1205 1210 1215

Ala Leu Tyr Tyr Pro Ser Ala Arg Ile Val Tyr Thr Ala Cys Ser

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1220	1225	1230
His Ala 1235	Ala Val Asp Ala 1240	Leu Cys Glu Lys Ala 1245
Pro Ile 1250	Asp Lys Cys Ser 1255	Arg Ile Ile Pro Ala 1260
Glu Cys 1265	Phe Asp Lys Phe 1270	Lys Val Asn Ser Thr 1275
Val Phe 1280	Cys Thr Val Asn 1285	Ala Leu Pro Glu Thr 1290
Val Val 1295	Phe Asp Glu Ile 1300	Ser Met Ala Thr Asn 1305
Val Val 1310	Asn Ala Arg Leu 1315	Arg Ala Lys His Tyr 1320
Asp Pro 1325	Ala Gln Leu Pro 1330	Ala Pro Arg Thr Leu 1335
Thr Leu 1340	Glu Pro Glu Tyr 1345	Phe Asn Ser Val Cys 1350
Thr Ile 1355	Gly Pro Asp Met 1360	Phe Leu Gly Thr Cys 1365
Ala Glu 1370	Ile Val Asp Thr 1375	Val Ser Ala Leu Val 1380
Leu Lys 1385	Ala His Lys Asp 1390	Lys Ser Ala Gln Cys 1395
Tyr Lys 1400	Gly Val Ile Thr 1405	His Asp Val Ser Ser 1410
Pro Gln 1415	Ile Gly Val Val 1420	Arg Glu Phe Leu Thr 1425
Trp Arg 1430	Lys Ala Val Phe 1435	Ile Ser Pro Tyr Asn 1440
Val Ala 1445	Ser Lys Ile Leu 1450	Gly Leu Pro Thr Gln 1455
Ser Gln 1460	Gly Ser Glu Tyr 1465	Asp Tyr Val Ile Phe 1470
Glu Thr 1475	Ala His Ser Cys 1480	Asn Val Asn Arg Phe 1485
Thr Arg 1490	Ala Lys Ile Gly 1495	Ile Leu Cys Ile Met 1500
Leu Tyr 1505	Asp Lys Leu Gln 1510	Phe Thr Ser Leu Glu 1515
Asn Val 1520	Ala Thr Leu Gln 1525	Ala Glu Asn Val Thr 1530
Asp Cys 1535	Ser Lys Ile Ile 1540	Thr Gly Leu His Pro 1545
Thr His 1550	Leu Ser Val Asp 1555	Ile Lys Phe Lys Thr 1560
Val Asp 1565	Ile Pro Gly Ile 1570	Pro Lys Asp Met Thr 1575
Ile Ser 1580	Met Met Gly Phe 1585	Lys Met Asn Tyr Gln 1590
Pro Asn 1595	Met Phe Ile Thr 1600	Arg Glu Glu Ala Ile 1605

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Ala Trp	Ile Gly Phe Asp	Val Glu Gly Cys His	Ala Thr Arg Asp
1610		1615	1620
Ala Val	Gly Thr Asn Leu	Pro Leu Gln Leu Gly	Phe Ser Thr Gly
1625		1630	1635
Val Asn	Leu Val Ala Val	Pro Thr Gly Tyr Val	Asp Thr Glu Asn
1640		1645	1650
Asn Thr	Glu Phe Thr Arg	Val Asn Ala Lys Pro	Pro Pro Gly Asp
1655		1660	1665
Gln Phe	Lys His Leu Ile	Pro Leu Met Tyr Lys	Gly Leu Pro Trp
1670		1675	1680
Asn Val	Val Arg Ile Lys	Ile Val Gln Met Leu	Ser Asp Thr Leu
1685		1690	1695
Lys Gly	Leu Ser Asp Arg	Val Val Phe Val Leu	Trp Ala His Gly
1700		1705	1710
Phe Glu	Leu Thr Ser Met	Lys Tyr Phe Val Lys	Ile Gly Pro Glu
1715		1720	1725
Arg Thr	Cys Cys Leu Cys	Asp Lys Arg Ala Thr	Cys Phe Ser Thr
1730		1735	1740
Ser Ser	Asp Thr Tyr Ala	Cys Trp Asn His Ser	Val Gly Phe Asp
1745		1750	1755
Tyr Val	Tyr Asn Pro Phe	Met Ile Asp Val Gln	Gln Trp Gly Phe
1760		1765	1770
Thr Gly	Asn Leu Gln Ser	Asn His Asp Gln His	Cys Gln Val His
1775		1780	1785
Gly Asn	Ala His Val Ala	Ser Cys Asp Ala Ile	Met Thr Arg Cys
1790		1795	1800
Leu Ala	Val His Glu Cys	Phe Val Lys Arg Val	Asp Trp Ser Val
1805		1810	1815
Glu Tyr	Pro Ile Ile Gly	Asp Glu Leu Arg Val	Asn Ser Ala Cys
1820		1825	1830
Arg Lys	Val Gln His Met	Val Val Lys Ser Ala	Leu Leu Ala Asp
1835		1840	1845
Lys Phe	Pro Val Leu His	Asp Ile Gly Asn Pro	Lys Ala Ile Lys
1850		1855	1860
Cys Val	Pro Gln Ala Glu	Val Glu Trp Lys Phe	Tyr Asp Ala Gln
1865		1870	1875
Pro Cys	Ser Asp Lys Ala	Tyr Lys Ile Glu Glu	Leu Phe Tyr Ser
1880		1885	1890
Tyr Ala	Thr His His Asp	Lys Phe Thr Asp Gly	Val Cys Leu Phe
1895		1900	1905
Trp Asn	Cys Asn Val Asp	Arg Tyr Pro Ala Asn	Ala Ile Val Cys
1910		1915	1920
Arg Phe	Asp Thr Arg Val	Leu Ser Asn Leu Asn	Leu Pro Gly Cys
1925		1930	1935
Asp Gly	Gly Ser Leu Tyr	Val Asn Lys His Ala	Phe His Thr Pro
1940		1945	1950
Ala Phe	Asp Lys Ser Ala	Phe Thr Asn Leu Lys	Gln Leu Pro Phe
1955		1960	1965
Phe Tyr	Tyr Ser Asp Ser	Pro Cys Glu Ser His	Gly Lys Gln Val
1970		1975	1980

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Val 1985	Ser	Asp	Ile	Asp	Tyr	Val 1990	Pro	Leu	Lys	Ser	Ala 1995	Thr	Cys	Ile
Thr 2000	Arg	Cys	Asn	Leu	Gly	Gly 2005	Ala	Val	Cys	Arg	His 2010	His	Ala	Asn
Glu 2015	Tyr	Arg	Gln	Tyr	Leu	Asp 2020	Ala	Tyr	Asn	Met	Met 2025	Ile	Ser	Ala
Gly 2030	Phe	Ser	Leu	Trp	Ile	Tyr 2035	Lys	Gln	Phe	Asp	Thr 2040	Tyr	Asn	Leu
Trp 2045	Asn	Thr	Phe	Thr	Arg	Leu 2050	Gln	Ser	Leu	Glu	Asn 2055	Val	Ala	Tyr
Asn 2060	Val	Val	Asn	Lys	Gly	His 2065	Phe	Asp	Gly	His	Ala 2070	Gly	Glu	Ala
Pro 2075	Val	Ser	Ile	Ile	Asn	Asn 2080	Ala	Val	Tyr	Thr	Lys 2085	Val	Asp	Gly
Ile 2090	Asp	Val	Glu	Ile	Phe	Glu 2095	Asn	Lys	Thr	Thr	Leu 2100	Pro	Val	Asn
Val 2105	Ala	Phe	Glu	Leu	Trp	Ala 2110	Lys	Arg	Asn	Ile	Lys 2115	Pro	Val	Pro
Glu 2120	Ile	Lys	Ile	Leu	Asn	Asn 2125	Leu	Gly	Val	Asp	Ile 2130	Ala	Ala	Asn
Thr 2135	Val	Ile	Trp	Asp	Tyr	Lys 2140	Arg	Glu	Ala	Pro	Ala 2145	His	Val	Ser
Thr 2150	Ile	Gly	Val	Cys	Thr	Met 2155	Thr	Asp	Ile	Ala	Lys 2160	Lys	Pro	Thr
Glu 2165	Ser	Ala	Cys	Ser	Ser	Leu 2170	Thr	Val	Leu	Phe	Asp 2175	Gly	Arg	Val
Glu 2180	Gly	Gln	Val	Asp	Leu	Phe 2185	Arg	Asn	Ala	Arg	Asn 2190	Gly	Val	Leu
Ile 2195	Thr	Glu	Gly	Ser	Val	Lys 2200	Gly	Leu	Thr	Pro	Ser 2205	Lys	Gly	Pro
Ala 2210	Gln	Ala	Ser	Val	Asn	Gly 2215	Val	Thr	Leu	Ile	Gly 2220	Glu	Ser	Val
Lys 2225	Thr	Gln	Phe	Asn	Tyr	Phe 2230	Lys	Lys	Val	Asp	Gly 2235	Ile	Ile	Gln
Gln 2240	Leu	Pro	Glu	Thr	Tyr	Phe 2245	Thr	Gln	Ser	Arg	Asp 2250	Leu	Glu	Asp
Phe 2255	Lys	Pro	Arg	Ser	Gln	Met 2260	Glu	Thr	Asp	Phe	Leu 2265	Glu	Leu	Ala
Met 2270	Asp	Glu	Phe	Ile	Gln	Arg 2275	Tyr	Lys	Leu	Glu	Gly 2280	Tyr	Ala	Phe
Glu 2285	His	Ile	Val	Tyr	Gly	Asp 2290	Phe	Ser	His	Gly	Gln 2295	Leu	Gly	Gly
Leu 2300	His	Leu	Met	Ile	Gly	Leu 2305	Ala	Lys	Arg	Ser	Gln 2310	Asp	Ser	Pro
Leu 2315	Lys	Leu	Glu	Asp	Phe	Ile 2320	Pro	Met	Asp	Ser	Thr 2325	Val	Lys	Asn
Tyr 2330	Phe	Ile	Thr	Asp	Ala	Gln 2335	Thr	Gly	Ser	Ser	Lys 2340	Cys	Val	Cys
Ser 2345	Val	Ile	Asp	Leu	Leu	Leu 2350	Asp	Asp	Phe	Val	Glu 2355	Ile	Ile	Lys
Ser	Gln	Asp	Leu	Ser	Val	Ile	Ser	Lys	Val	Val	Lys	Val	Thr	Ile

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2360	2365	2370
Asp Tyr Ala Glu Ile Ser Phe Met Leu Trp Cys Lys Asp Gly His 2375 2380 2385		
Val Glu Thr Phe Tyr Pro Lys Leu Gln Ala Ser Gln Ala Trp Gln 2390 2395 2400		
Pro Gly Val Ala Met Pro Asn Leu Tyr Lys Met Gln Arg Met Leu 2405 2410 2415		
Leu Glu Lys Cys Asp Leu Gln Asn Tyr Gly Glu Asn Ala Val Ile 2420 2425 2430		
Pro Lys Gly Ile Met Met Asn Val Ala Lys Tyr Thr Gln Leu Cys 2435 2440 2445		
Gln Tyr Leu Asn Thr Leu Thr Leu Ala Val Pro Tyr Asn Met Arg 2450 2455 2460		
Val Ile His Phe Gly Ala Gly Ser Asp Lys Gly Val Ala Pro Gly 2465 2470 2475		
Thr Ala Val Leu Arg Gln Trp Leu Pro Thr Gly Thr Leu Leu Val 2480 2485 2490		
Asp Ser Asp Leu Asn Asp Phe Val Ser Asp Ala Asp Ser Thr Leu 2495 2500 2505		
Ile Gly Asp Cys Ala Thr Val His Thr Ala Asn Lys Trp Asp Leu 2510 2515 2520		
Ile Ile Ser Asp Met Tyr Asp Pro Arg Thr Lys His Val Thr Lys 2525 2530 2535		
Glu Asn Asp Ser Lys Glu Gly Phe Phe Thr Tyr Leu Cys Gly Phe 2540 2545 2550		
Ile Lys Gln Lys Leu Ala Leu Gly Gly Ser Ile Ala Val Lys Ile 2555 2560 2565		
Thr Glu His Ser Trp Asn Ala Asp Leu Tyr Lys Leu Met Gly His 2570 2575 2580		
Phe Ser Trp Trp Thr Ala Phe Val Thr Asn Val Asn Ala Ser Ser 2585 2590 2595		
Ser Glu Ala Phe Leu Ile Gly Ala Asn Tyr Leu Gly Lys Pro Lys 2600 2605 2610		
Glu Gln Ile Asp Gly Tyr Thr Met His Ala Asn Tyr Ile Phe Trp 2615 2620 2625		
Arg Asn Thr Asn Pro Ile Gln Leu Ser Ser Tyr Ser Leu Phe Asp 2630 2635 2640		
Met Ser Lys Phe Pro Leu Lys Leu Arg Gly Thr Ala Val Met Ser 2645 2650 2655		
Leu Lys Glu Asn Gln Ile Asn Asp Met Ile Tyr Ser Leu Leu Glu 2660 2665 2670		
Lys Gly Arg Leu Ile Ile Arg Glu Asn Asn Arg Val Val Val Ser 2675 2680 2685		
Ser Asp Ile Leu Val Asn Asn 2690 2695		

<210> SEQ ID NO 76
 <211> LENGTH: 20
 <212> TYPE: DNA
 <213> ORGANISM: Artificial sequence
 <220> FEATURE:
 <223> OTHER INFORMATION: S/L3/+4932 primer

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<400> SEQUENCE: 76

ccacacacag cttgtggata

20

<210> SEQ ID NO 77

<211> LENGTH: 20

<212> TYPE: DNA

<213> ORGANISM: Artificial sequence

<220> FEATURE:

<223> OTHER INFORMATION: S/L4/+6401 primer

<400> SEQUENCE: 77

ccgaagtgt aggcaatgtc

20

<210> SEQ ID NO 78

<211> LENGTH: 20

<212> TYPE: DNA

<213> ORGANISM: Artificial sequence

<220> FEATURE:

<223> OTHER INFORMATION: S/L4/+6964 primer

<400> SEQUENCE: 78

tttggtgctc cttcttattg

20

<210> SEQ ID NO 79

<211> LENGTH: 20

<212> TYPE: DNA

<213> ORGANISM: Artificial sequence

<220> FEATURE:

<223> OTHER INFORMATION: S/L4/-6817 primer

<400> SEQUENCE: 79

ccggcatcca aacataattt

20

<210> SEQ ID NO 80

<211> LENGTH: 20

<212> TYPE: DNA

<213> ORGANISM: Artificial sequence

<220> FEATURE:

<223> OTHER INFORMATION: S/L5/-7633 primer

<400> SEQUENCE: 80

tggtcagtag ggttgattgg

20

<210> SEQ ID NO 81

<211> LENGTH: 20

<212> TYPE: DNA

<213> ORGANISM: Artificial sequence

<220> FEATURE:

<223> OTHER INFORMATION: S/L5/-8127 primer

<400> SEQUENCE: 81

catcctttgt gtcaacatcg

20

<210> SEQ ID NO 82

<211> LENGTH: 20

<212> TYPE: DNA

<213> ORGANISM: Artificial sequence

<220> FEATURE:

<223> OTHER INFORMATION: S/L5/-8633 primer

<400> SEQUENCE: 82

gtcacgagtg acaccatcct

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<210> SEQ ID NO 83
<211> LENGTH: 20
<212> TYPE: DNA
<213> ORGANISM: Artificial sequence
<220> FEATURE:
<223> OTHER INFORMATION: S/L5/+7839 primer

<400> SEQUENCE: 83

atgcgacgag tctgcttcta 20

<210> SEQ ID NO 84
<211> LENGTH: 20
<212> TYPE: DNA
<213> ORGANISM: Artificial sequence
<220> FEATURE:
<223> OTHER INFORMATION: S/L5/+8785 primer

<400> SEQUENCE: 84

ttcatagtgc ctggcttacc 20

<210> SEQ ID NO 85
<211> LENGTH: 20
<212> TYPE: DNA
<213> ORGANISM: Artificial sequence
<220> FEATURE:
<223> OTHER INFORMATION: S/L5/+8255 primer

<400> SEQUENCE: 85

atcttgcgcg atgtattgac 20

<210> SEQ ID NO 86
<211> LENGTH: 20
<212> TYPE: DNA
<213> ORGANISM: Artificial sequence
<220> FEATURE:
<223> OTHER INFORMATION: S/L6/-9422 primer

<400> SEQUENCE: 86

tgcattagca gcaacaacat 20

<210> SEQ ID NO 87
<211> LENGTH: 20
<212> TYPE: DNA
<213> ORGANISM: Artificial sequence
<220> FEATURE:
<223> OTHER INFORMATION: S/L6/-9966 primer

<400> SEQUENCE: 87

tctgcagaac agcagaagtg 20

<210> SEQ ID NO 88
<211> LENGTH: 20
<212> TYPE: DNA
<213> ORGANISM: Artificial sequence
<220> FEATURE:
<223> OTHER INFORMATION: S/L6/-10542 primer

<400> SEQUENCE: 88

cctgtgcagt ttgtctgtca 20

<210> SEQ ID NO 89
<211> LENGTH: 20
<212> TYPE: DNA

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<213> ORGANISM: Artificial sequence
<220> FEATURE:
<223> OTHER INFORMATION: S/L6+/10677 primer

<400> SEQUENCE: 89

ccttgtggca atgaagtaca 20

<210> SEQ ID NO 90
<211> LENGTH: 20
<212> TYPE: DNA
<213> ORGANISM: Artificial sequence
<220> FEATURE:
<223> OTHER INFORMATION: S/L6+/10106 primer

<400> SEQUENCE: 90

atgtcatttg cacagcagaa 20

<210> SEQ ID NO 91
<211> LENGTH: 20
<212> TYPE: DNA
<213> ORGANISM: Artificial sequence
<220> FEATURE:
<223> OTHER INFORMATION: S/L6+/9571 primer

<400> SEQUENCE: 91

cttcaatggt ttgccatggt 20

<210> SEQ ID NO 92
<211> LENGTH: 20
<212> TYPE: DNA
<213> ORGANISM: Artificial sequence
<220> FEATURE:
<223> OTHER INFORMATION: S/L7-/11271 primer

<400> SEQUENCE: 92

tgcgagctgt catgagaata 20

<210> SEQ ID NO 93
<211> LENGTH: 20
<212> TYPE: DNA
<213> ORGANISM: Artificial sequence
<220> FEATURE:
<223> OTHER INFORMATION: S/L7-/11801 primer

<400> SEQUENCE: 93

aaccgagagc agtaccacag 20

<210> SEQ ID NO 94
<211> LENGTH: 20
<212> TYPE: DNA
<213> ORGANISM: Artificial sequence
<220> FEATURE:
<223> OTHER INFORMATION: S/L7-/12383 primer

<400> SEQUENCE: 94

tttgctgct gtagtcaatg 20

<210> SEQ ID NO 95
<211> LENGTH: 20
<212> TYPE: DNA
<213> ORGANISM: Artificial sequence
<220> FEATURE:
<223> OTHER INFORMATION: S/L7+/12640 primer

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<400> SEQUENCE: 95

ctacgacaga tgcctgtgc

20

<210> SEQ ID NO 96

<211> LENGTH: 20

<212> TYPE: DNA

<213> ORGANISM: Artificial sequence

<220> FEATURE:

<223> OTHER INFORMATION: S/L7/+12088 primer

<400> SEQUENCE: 96

gagcaggctg tagctaattg

20

<210> SEQ ID NO 97

<211> LENGTH: 20

<212> TYPE: DNA

<213> ORGANISM: Artificial sequence

<220> FEATURE:

<223> OTHER INFORMATION: S/L7/+11551 primer

<400> SEQUENCE: 97

ttaggctatt gttgctgctg

20

<210> SEQ ID NO 98

<211> LENGTH: 20

<212> TYPE: DNA

<213> ORGANISM: Artificial sequence

<220> FEATURE:

<223> OTHER INFORMATION: S/L8/-13160 primer

<400> SEQUENCE: 98

cagacaacat gaagcaccac

20

<210> SEQ ID NO 99

<211> LENGTH: 20

<212> TYPE: DNA

<213> ORGANISM: Artificial sequence

<220> FEATURE:

<223> OTHER INFORMATION: S/L8/-13704 primer

<400> SEQUENCE: 99

cgctgacgtg atatatgtgg

20

<210> SEQ ID NO 100

<211> LENGTH: 20

<212> TYPE: DNA

<213> ORGANISM: Artificial sequence

<220> FEATURE:

<223> OTHER INFORMATION: S/L8/-14284 primer

<400> SEQUENCE: 100

tgcaaatga aggatacacc

20

<210> SEQ ID NO 101

<211> LENGTH: 20

<212> TYPE: DNA

<213> ORGANISM: Artificial sequence

<220> FEATURE:

<223> OTHER INFORMATION: S/L8/+14453 primer

<400> SEQUENCE: 101

acatagctcg cgtctcagtt

20

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<210> SEQ ID NO 102
<211> LENGTH: 20
<212> TYPE: DNA
<213> ORGANISM: Artificial sequence
<220> FEATURE:
<223> OTHER INFORMATION: S/L8/+/13968 primer

<400> SEQUENCE: 102

ggcattgtag gcgtactgac 20

<210> SEQ ID NO 103
<211> LENGTH: 19
<212> TYPE: DNA
<213> ORGANISM: Artificial sequence
<220> FEATURE:
<223> OTHER INFORMATION: S/L8/+/13401 primer

<400> SEQUENCE: 103

gtttgcggtg taagtgcag 19

<210> SEQ ID NO 104
<211> LENGTH: 20
<212> TYPE: DNA
<213> ORGANISM: Artificial sequence
<220> FEATURE:
<223> OTHER INFORMATION: S/L9/-/15098 primer

<400> SEQUENCE: 104

tagtggcggc tattgacttc 20

<210> SEQ ID NO 105
<211> LENGTH: 20
<212> TYPE: DNA
<213> ORGANISM: Artificial sequence
<220> FEATURE:
<223> OTHER INFORMATION: S/L9/-/15677 primer

<400> SEQUENCE: 105

ctaaaccttg agccgcatag 20

<210> SEQ ID NO 106
<211> LENGTH: 20
<212> TYPE: DNA
<213> ORGANISM: Artificial sequence
<220> FEATURE:
<223> OTHER INFORMATION: S/L9/-/16247 primer

<400> SEQUENCE: 106

catggtcata gcagcacttg 20

<210> SEQ ID NO 107
<211> LENGTH: 21
<212> TYPE: DNA
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<400> SEQUENCE: 107

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<210> SEQ ID NO 108
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<212> TYPE: DNA

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<213> ORGANISM: Artificial sequence
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<223> OTHER INFORMATION: S/L9/+/15858 primer

<400> SEQUENCE: 108

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<210> SEQ ID NO 109
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<212> TYPE: DNA
<213> ORGANISM: Artificial sequence
<220> FEATURE:
<223> OTHER INFORMATION: S/L9/+/15288 primer

<400> SEQUENCE: 109

cgcaaacata acacttgctg 20

<210> SEQ ID NO 110
<211> LENGTH: 20
<212> TYPE: DNA
<213> ORGANISM: Artificial sequence
<220> FEATURE:
<223> OTHER INFORMATION: S/L10/-/16914 primer

<400> SEQUENCE: 110

agtgttgggt acaagccagt 20

<210> SEQ ID NO 111
<211> LENGTH: 20
<212> TYPE: DNA
<213> ORGANISM: Artificial sequence
<220> FEATURE:
<223> OTHER INFORMATION: S/L10/-/17466 primer

<400> SEQUENCE: 111

gttccaagga acatgtctgg 20

<210> SEQ ID NO 112
<211> LENGTH: 20
<212> TYPE: DNA
<213> ORGANISM: Artificial sequence
<220> FEATURE:
<223> OTHER INFORMATION: S/L10/-/18022 primer

<400> SEQUENCE: 112

aggtgcctgt gtaggatgaa 20

<210> SEQ ID NO 113
<211> LENGTH: 20
<212> TYPE: DNA
<213> ORGANISM: Artificial sequence
<220> FEATURE:
<223> OTHER INFORMATION: S/L10/+/18245 primer

<400> SEQUENCE: 113

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<210> SEQ ID NO 114
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<210> SEQ ID NO 115

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<212> TYPE: DNA

<213> ORGANISM: Artificial sequence

<220> FEATURE:

<223> OTHER INFORMATION: S/L10/+/17061 primer

<400> SEQUENCE: 115

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<210> SEQ ID NO 116

<211> LENGTH: 20

<212> TYPE: DNA

<213> ORGANISM: Artificial sequence

<220> FEATURE:

<223> OTHER INFORMATION: S/L11/-/18877 primer

<400> SEQUENCE: 116

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<210> SEQ ID NO 117

<211> LENGTH: 20

<212> TYPE: DNA

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<220> FEATURE:

<223> OTHER INFORMATION: S/L11/-/19396 primer

<400> SEQUENCE: 117

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<210> SEQ ID NO 118

<211> LENGTH: 20

<212> TYPE: DNA

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<220> FEATURE:

<223> OTHER INFORMATION: S/L11/-/20002 primer

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<210> SEQ ID NO 119

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<212> TYPE: DNA

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<220> FEATURE:

<223> OTHER INFORMATION: S/L11/+/20245 primer

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<210> SEQ ID NO 122
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<400> SEQUENCE: 122

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<210> SEQ ID NO 123
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<212> TYPE: DNA
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<400> SEQUENCE: 123

cagagattgg acctgagcat 20

<210> SEQ ID NO 124
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<212> TYPE: DNA
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<400> SEQUENCE: 124

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<210> SEQ ID NO 125
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<210> SEQ ID NO 126
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<210> SEQ ID NO 127
<211> LENGTH: 20
<212> TYPE: DNA

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<213> ORGANISM: Artificial sequence
<220> FEATURE:
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<210> SEQ ID NO 128
<211> LENGTH: 20
<212> TYPE: DNA
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<400> SEQUENCE: 128

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<210> SEQ ID NO 129
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<400> SEQUENCE: 129

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<223> OTHER INFORMATION: SARS/L2/F5/+3746 primer

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<210> SEQ ID NO 131
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<210> SEQ ID NO 132
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<400> SEQUENCE: 132

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<223> OTHER INFORMATION: SARS/L2/R5/-/2529 primer

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<223> OTHER INFORMATION: SARS/L3/F3/+ /4708 primer

<400> SEQUENCE: 134
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<210> SEQ ID NO 135
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<212> TYPE: DNA
<213> ORGANISM: Artificial sequence
<220> FEATURE:
<223> OTHER INFORMATION: SRAS/L3/F4/+ /5305 primer

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<210> SEQ ID NO 136
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<212> TYPE: DNA
<213> ORGANISM: Artificial sequence
<220> FEATURE:
<223> OTHER INFORMATION: SARS/L3/F5/+ /5822 primer

<400> SEQUENCE: 136
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<210> SEQ ID NO 137
<211> LENGTH: 20
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<213> ORGANISM: Artificial sequence
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<223> OTHER INFORMATION: SARS/L3/R3/- /5610 primer

<400> SEQUENCE: 137
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<210> SEQ ID NO 138
<211> LENGTH: 20
<212> TYPE: DNA
<213> ORGANISM: Artificial sequence
<220> FEATURE:
<223> OTHER INFORMATION: SARS/L3/R4/- /4988 primer

<400> SEQUENCE: 138
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<210> SEQ ID NO 139
<211> LENGTH: 20
<212> TYPE: DNA
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<220> FEATURE:
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<400> SEQUENCE: 139
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<210> SEQ ID NO 140
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<212> TYPE: DNA
<213> ORGANISM: Artificial sequence
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<223> OTHER INFORMATION: synthetic S gene

<400> SEQUENCE: 140

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<213> ORGANISM: Artificial sequence
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<400> SEQUENCE: 141

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<210> SEQ ID NO 142
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<212> TYPE: DNA
<213> ORGANISM: Artificial sequence
<220> FEATURE:
<223> OTHER INFORMATION: SNE-AS1 primer

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<400> SEQUENCE: 142

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<210> SEQ ID NO 145
 <211> LENGTH: 45
 <212> TYPE: DNA
 <213> ORGANISM: Artificial sequence
 <220> FEATURE:
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 <400> SEQUENCE: 145

ataggatcca ccatgtttat tttcttatta tttcttactc tcaact 45

<210> SEQ ID NO 146
 <211> LENGTH: 37
 <212> TYPE: DNA
 <213> ORGANISM: Artificial sequence
 <220> FEATURE:
 <223> OTHER INFORMATION: PCR primer
 <400> SEQUENCE: 146

atactcgagt tatgtgtaat gtaatttgac acccttg 37

<210> SEQ ID NO 147
 <211> LENGTH: 45
 <212> TYPE: DNA
 <213> ORGANISM: Artificial sequence
 <220> FEATURE:
 <223> OTHER INFORMATION: PCR primer
 <400> SEQUENCE: 147

ataggatcca ccatgtttat tttcttatta tttcttactc tcaact 45

<210> SEQ ID NO 148
 <211> LENGTH: 36
 <212> TYPE: DNA
 <213> ORGANISM: Artificial sequence
 <220> FEATURE:
 <223> OTHER INFORMATION: PCR primer
 <400> SEQUENCE: 148

acctccggat ttaatatatt gctcatatth tcccaa 36

<210> SEQ ID NO 149
 <211> LENGTH: 13
 <212> TYPE: PRT
 <213> ORGANISM: Artificial sequence
 <220> FEATURE:
 <223> OTHER INFORMATION: N-terminal end of SRAS-CoV S protein (amino-acids 1 to 13)
 <400> SEQUENCE: 149

Met Phe Ile Phe Leu Leu Phe Leu Thr Leu Thr Ser Gly
 1 5 10

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<210> SEQ ID NO 150
<211> LENGTH: 10
<212> TYPE: PRT
<213> ORGANISM: Artificial sequence
<220> FEATURE:
<223> OTHER INFORMATION: oligopeptide

<400> SEQUENCE: 150

Ser Gly Asp Tyr Lys Asp Asp Asp Lys
1 5 10

<210> SEQ ID NO 151
<211> LENGTH: 34
<212> TYPE: DNA
<213> ORGANISM: Artificial sequence
<220> FEATURE:
<223> OTHER INFORMATION: PCR primer

<400> SEQUENCE: 151

actagctagc ggatccacca tgttcatctt cctg 34

<210> SEQ ID NO 152
<211> LENGTH: 33
<212> TYPE: DNA
<213> ORGANISM: Artificial sequence
<220> FEATURE:
<223> OTHER INFORMATION: PCR primer

<400> SEQUENCE: 152

agtatccgga ctgtatgtac tgctcgtact tgc 33

<210> SEQ ID NO 153
<211> LENGTH: 59
<212> TYPE: DNA
<213> ORGANISM: Artificial sequence
<220> FEATURE:
<223> OTHER INFORMATION: oligonucleotid

<400> SEQUENCE: 153

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<210> SEQ ID NO 154
<211> LENGTH: 53
<212> TYPE: DNA
<213> ORGANISM: Artificial sequence
<220> FEATURE:
<223> OTHER INFORMATION: oligonucleotid

<400> SEQUENCE: 154

gatggcgcgc cgagtctatt tatatgccaa aaaaaaaaaa aaaaaaaagc tca 53

<210> SEQ ID NO 155
<211> LENGTH: 45
<212> TYPE: DNA
<213> ORGANISM: Artificial sequence
<220> FEATURE:
<223> OTHER INFORMATION: PCR primer

<400> SEQUENCE: 155

atacgtacga ccatgtttat tttcttatta tttcttactc tca 45

<210> SEQ ID NO 156
<211> LENGTH: 40

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<212> TYPE: DNA
<213> ORGANISM: Artificial sequence
<220> FEATURE:
<223> OTHER INFORMATION: PCR primer

<400> SEQUENCE: 156

atagcgcgct cattatgtgt aatgtaattt gacacccttg                40

<210> SEQ ID NO 157
<211> LENGTH: 20
<212> TYPE: DNA
<213> ORGANISM: Artificial sequence
<220> FEATURE:
<223> OTHER INFORMATION: PCR primer

<400> SEQUENCE: 157

ccatttcaac aatttgcccg                20

<210> SEQ ID NO 158
<211> LENGTH: 45
<212> TYPE: DNA
<213> ORGANISM: Artificial sequence
<220> FEATURE:
<223> OTHER INFORMATION: PCR primer

<400> SEQUENCE: 158

ataggatccg cgcgctcatt atttatcgtc gtcaccttta taatc        45

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1. An isolated or purified strain of severe acute respiratory syndrome-associated human coronavirus, characterized in that its genome has, in the form of complementary DNA, a serine codon at position 23220-23222 of the gene for the S protein or a glycine codon at position 25298-25300 of the gene for ORF3, and an alanine codon at position 7918-7920 of ORF1a or a serine codon at position 26857-26859 of the gene for the M protein, said positions being indicated in terms of reference to the Genbank sequence AY274119.3.

2. The isolated or purified coronavirus strain as claimed in claim 1, characterized in that the DNA equivalent of its genome has a sequence corresponding to the sequence SEQ ID NO: 1.

3. An isolated or purified polynucleotide, characterized in that its sequence is that of the genome of the isolated coronavirus strain as claimed in claim 1 or claim 2.

4. The isolated or purified polynucleotide as claimed in claim 3, characterized in that its sequence is SEQ ID NO: 1.

5. A pair of primers capable of amplifying a fragment of the sequence of the genome of a SARS-associated coronavirus or of its DNA equivalent, characterized in that it is selected from the group consisting of:

the pair of primers No. 1 corresponding respectively to positions 28507 to 28522 (sense primer, SEQ ID NO: 60) and 28774 to 28759 (antisense primer, SEQ ID NO: 61) of the sequence of the polynucleotide as claimed in claim 3 or claim 4,

the pair of primers No. 2 corresponding respectively to positions 28375 to 28390 (sense primer, SEQ ID NO: 62) and 28702 to 28687 (antisense primer, SEQ ID NO: 63) of the sequence of the polynucleotide as claimed in claim 3 or claim 4, and

the pair of primers consisting of the primers SEQ ID Nos: 55 and 56.

6. A probe capable of detecting the presence of the genome of a SARS-associated coronavirus or of a fragment thereof, characterized in that it is selected from the group consisting of the fragments corresponding to the following positions of the polynucleotide sequence as claimed in claim 3 or claim 4: 28561 to 28586, 28588 to 28608, 28541 to 28563 and 28565 to 28589 (SEQ ID NO: 64 to 67).

7. A recombinant cloning and/or expression vector, characterized in that it comprises an insert having the sequence SEQ ID NO: 38 and it is contained in a bacterial strain and it was deposited under the No. I-3048, on Jun. 5, 2003, at the Collection Nationale de Cultures de Microorganismes, 25 rue du Docteur Roux, 75724 Paris Cedex 15.

8. A recombinant cloning and/or expression vector, characterized in that it contains a cDNA fragment selected from the group consisting of:

a cDNA fragment encoding a C-terminal fusion of the N protein (SEQ ID NO: 37) with a polyhistidine tag, and

a cDNA fragment encoding an N-terminal fusion of the N protein (SEQ ID NO: 37) with a polyhistidine tag.

9. The recombinant expression vector as claimed in claim 8, characterized in that it is contained in a bacterial strain which was deposited under the No. I-3117, on Oct. 23, 2003, at the Collection Nationale de Cultures de Microorganismes, 25 rue du Docteur Roux, 75724 Paris Cedex 15.

10. A cell modified with a vector as claimed in any one of claims 7 to 9.

11. A hybridoma producing a monoclonal antibody against the N protein, characterized in that it is chosen from the following hybridomas:

- the hybridoma producing the monoclonal antibody 87, deposited at the CNCM on Dec. 1, 2004 under the number I-3328,
- the hybridoma producing the monoclonal antibody 86, deposited at the CNCM on Dec. 1, 2004 under the number I-3329,
- the hybridoma producing the monoclonal antibody 57, deposited at the CNCM on Dec. 1, 2004 under the number I-3330, and
- the hybridoma producing the monoclonal antibody 156, deposited at the CNCM on Dec. 1, 2004 under the number I-3331.
- 12.** A polyclonal or monoclonal antibody or antibody fragment directed against the N protein, characterized in that it is produced by a hybridoma as claimed in claim 11.
- 13.** A chip or filter, characterized in that it comprises an antibody or an antibody fragment as claimed in claim 12.
- 14.** An immunocapture test intended to detect a SARS-associated coronavirus infection, characterized in that it uses a monoclonal antibody specific for the native viral nucleoprotein (N protein).
- 15.** The immunocapture test as claimed in claim 14, characterized in that the antibody used for the capture of the native viral nucleoprotein is a monoclonal antibody specific for the central region and/or for a conformational epitope.
- 16.** The immunocapture test as claimed in claim 14 or 15, characterized in that the antibody used for the capture of the N protein is the monoclonal antibody mAb87, produced by the hybridoma deposited at the CNCM on Dec. 1, 2004 under the number I-3328.
- 17.** The immunocapture test as claimed in claim 14 or 15, characterized in that the antibody used for the capture of the N protein is the monoclonal antibody mAb86, produced by the hybridoma deposited at the CNCM on Dec. 1, 2004 under the number I-3329.
- 18.** The immunocapture test as claimed in claim 14 or 15, characterized in that the monoclonal antibodies mAb86 and mAb87 are used for the capture of the N protein.
- 19.** The immunocapture test as claimed in any one of claims 14 to 18, characterized in that the antibody used for the visualization of the N protein is the monoclonal antibody mAb57, produced by the hybridoma deposited at the CNCM on Dec. 1, 2004 under the number I-3330, said antibody being conjugated with a visualizing molecule or particle.
- 20.** The immunocapture test as claimed in any one of claims 14 to 18, characterized in that a combination of the mAb57 and mAb87 antibodies, conjugated with a visualizing molecule or particle, is used for the visualization of the N protein.
- 21.** A reagent for the detection of a SARS-associated coronavirus, characterized in that it is selected from the group consisting of:
- (a) a pair of primers as claimed in claim 5, or a probe as claimed in claim 6,
 - (b) a recombinant vector as claimed in any one of claims 7 to 9 or a modified cell as claimed in claim 10,
 - (c) an isolated coronavirus strain as claimed in claim 1 or claim 2 or a polynucleotide as claimed in either of claims 3 and 4,
 - (d) an antibody or an antibody fragment as claimed in claim 12,
 - (e) a combination of antibodies comprising the monoclonal antibodies mAb86 and/or mAb87, and the monoclonal antibody mAb57;
 - (f) a chip or a filter as claimed in claim 13.
- 22.** The use of a product selected from the group consisting of: a pair of primers as claimed in claim 5, a probe as claimed in claim 6, a recombinant vector as claimed in any one of claims 7 to 9, a modified cell as claimed in claim 10, an isolated coronavirus strain as claimed in claim 1 or claim 2, a polynucleotide as claimed in claim 3 or claim 4, for the preparation of a reagent for the detection and optionally genotyping of a SARS-associated coronavirus.
- 23.** A method for the detection of a SARS-associated coronavirus, from a biological sample, which method is characterized in that it comprises at least:
- (a) the extraction of nucleic acids present in said biological sample,
 - (b) the amplification of a fragment of ORF-N by RT-PCR with the aid of a pair of primers as claimed in claim 5, and
 - (c) the detection, by any appropriate means, of the amplification products obtained in (b).
- 24.** The method as claimed in claim 23, characterized in that step (b) of detection is carried out with the aid of at least one probe corresponding to positions 28561 to 28586, 28588 to 28608, 28541 to 28563 and 28565 to 28589 of the sequence of the polynucleotide as claimed in claim 3 or claim 4.
- 25.** A method for the detection of a SARS-associated coronavirus infection, from a biological sample, by indirect IgG ELISA using the N protein, which method is characterized in that the plates are sensitized with an N protein solution at a concentration of between 0.5 and 4 µg/ml, preferably 2 µg/ml, in a 10 mM PBS buffer, pH 7.2, phenol red at 0.25 ml/l.
- 26.** A method for the detection of a SARS-associated coronavirus infection, from a biological sample, by double epitope ELISA, characterized in that the serum to be tested is mixed with the visualizing antigen, said mixture then being brought into contact with the antigen attached to a solid support.
- 27.** An immune complex formed of a polyclonal or monoclonal antibody or antibody fragment as claimed in claim 11, and of a SARS-associated coronavirus protein or peptide.
- 28.** A SARS-associated coronavirus detection kit or box, characterized in that it comprises at least one reagent selected from the group consisting of: a pair of primers as claimed in claim 5, a probe as claimed in claim 6, a recombinant vector as claimed in any one of claims 7 to 9, a modified cell as claimed in claim 10, an isolated coronavirus strain as claimed in claim 1 or claim 2 and a polynucleotide as claimed in claim 3 or claim 4.
- 29.** A fragment of the polynucleotide as claimed in claim 3, characterized in that it includes at least one pair of bases or pairs of bases corresponding to the following positions: 7919 and 23220, 7919 and 25298, 16622 and 23220, 19064 and 23220, 16622 and 25298, 19064 and 25298, 23220 and 24872, 23220 and 26857, 24872 and 25298, 25298 and 26857.