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Van Der Werf et al.

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(54) **STRAIN OF SARS-ASSOCIATED
CORONAVIRUS AND APPLICATIONS
THEREOF**

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Related U.S. Application Data

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Dec. 2, 2003 (FR) 03 14152

(51) **Int. Cl.**

C12Q 1/70 (2006.01)
G01N 33/53 (2006.01)
G01N 33/542 (2006.01)
G01N 33/00 (2006.01)

(52) **U.S. Cl.** **435/5; 435/7.1; 435/7.9; 435/7.92; 435/7.94; 435/7.95**

(58) **Field of Classification Search** None
See application file for complete search history.

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Primary Examiner — Louise Humphrey

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(57) **ABSTRACT**

The invention relates to a novel strain of severe acute respiratory syndrome (SARS)-associated coronavirus, resulting from a sample collected in Hanoi (Vietnam), reference number 031589, nucleic acid molecules originating from the genome of same, proteins and peptides coded by said nucleic acid molecules and, more specifically, protein N and the applications thereof, for example, as diagnostic reagents and/or as a vaccine.

8 Claims, 116 Drawing Sheets

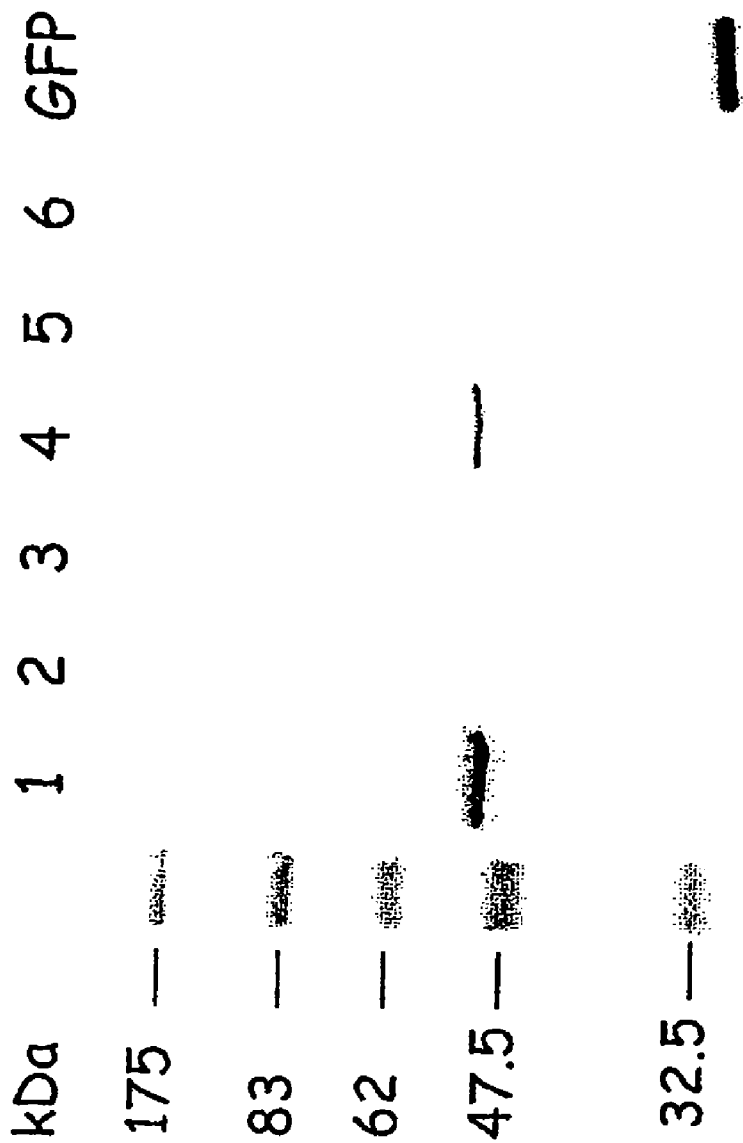


FIGURE 1

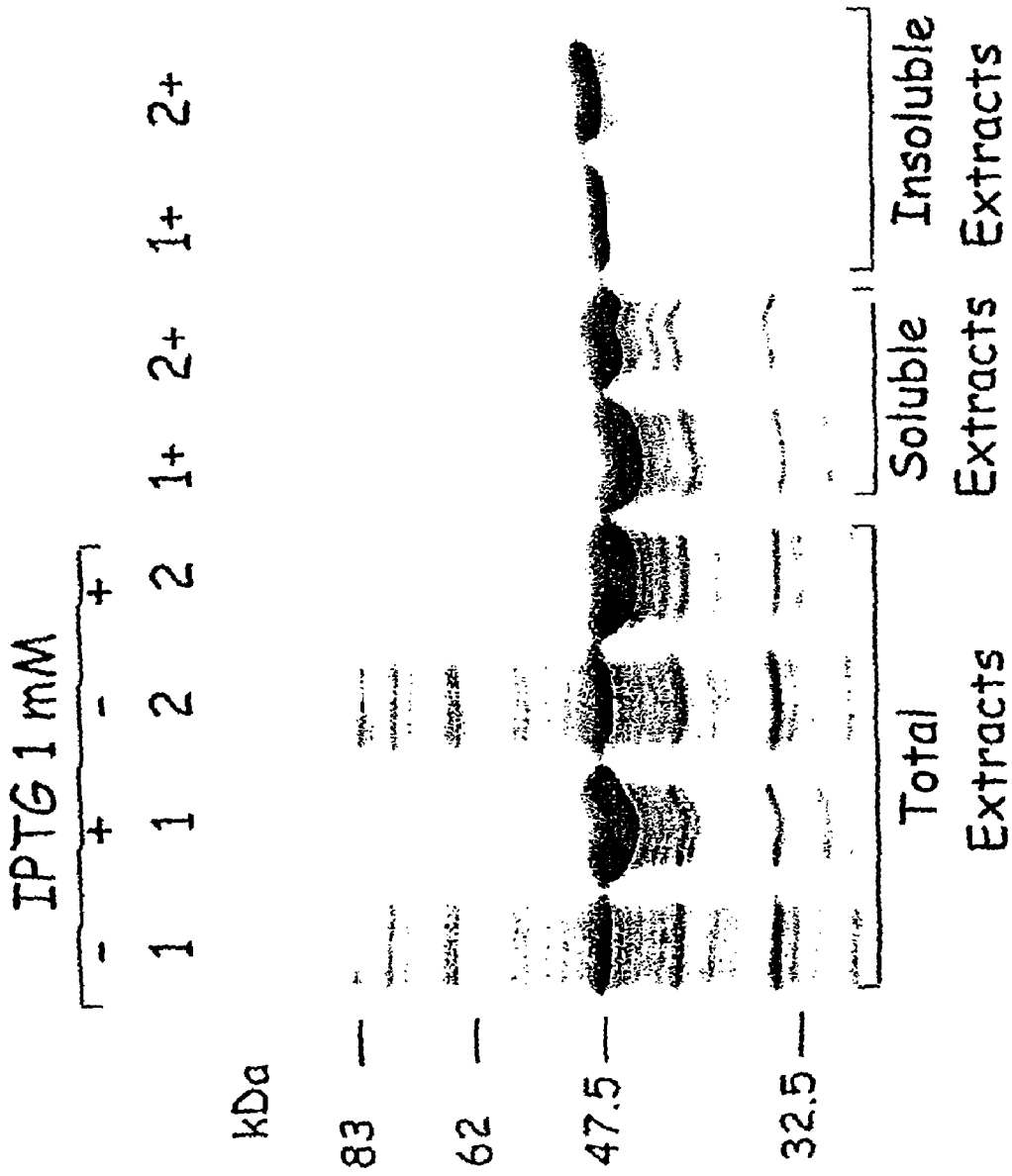


FIGURE 2

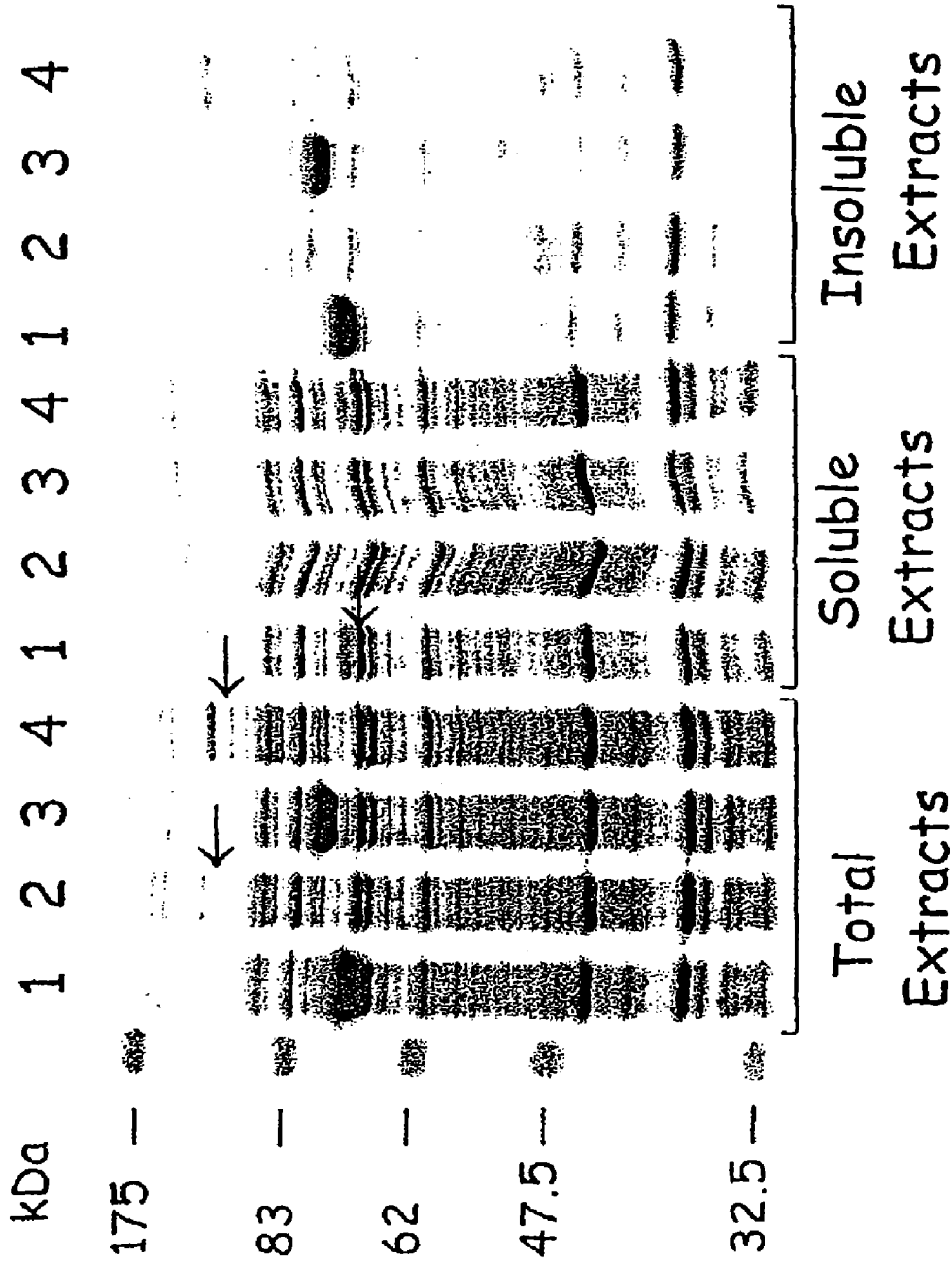


FIGURE 3

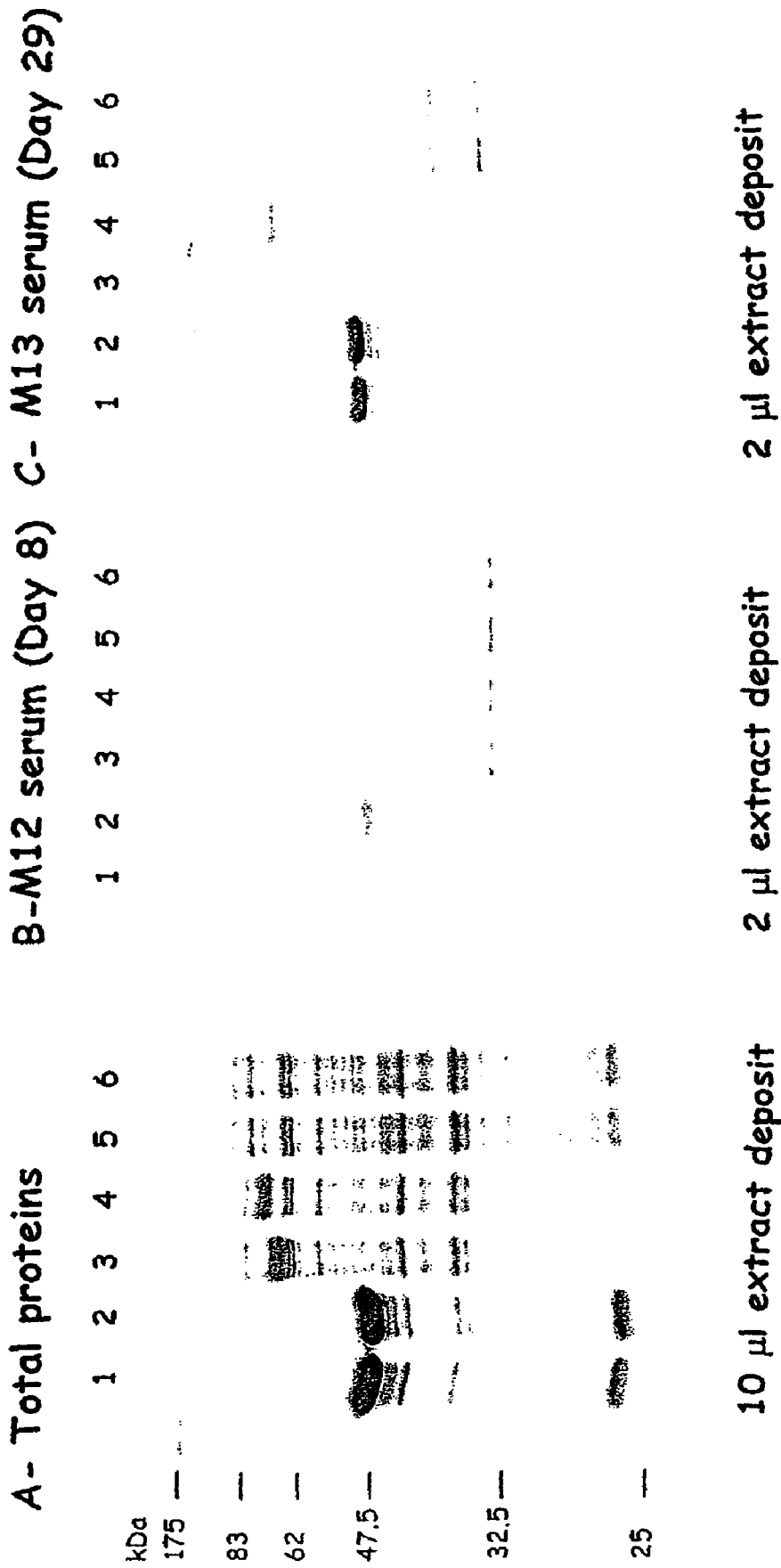


FIGURE 4

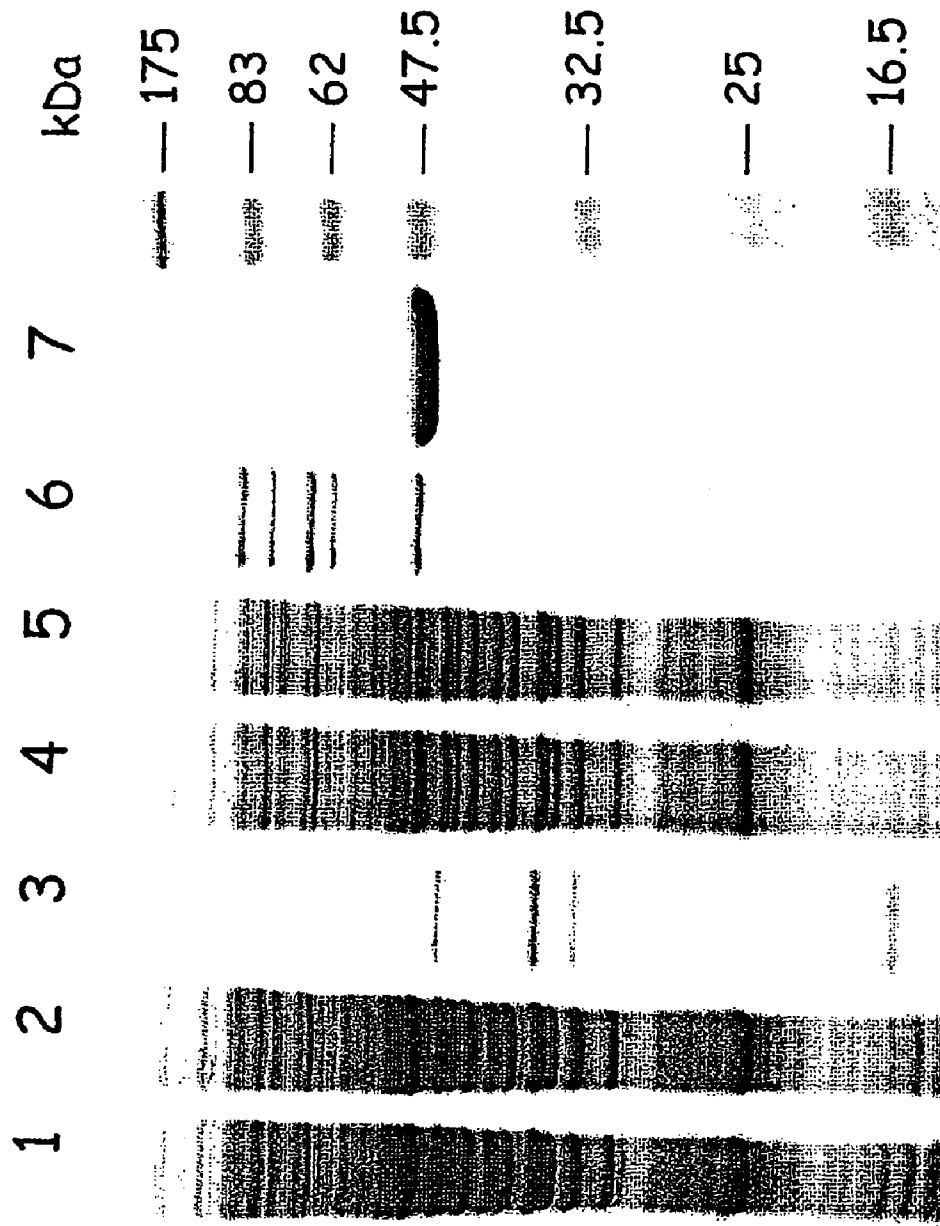


FIGURE 5

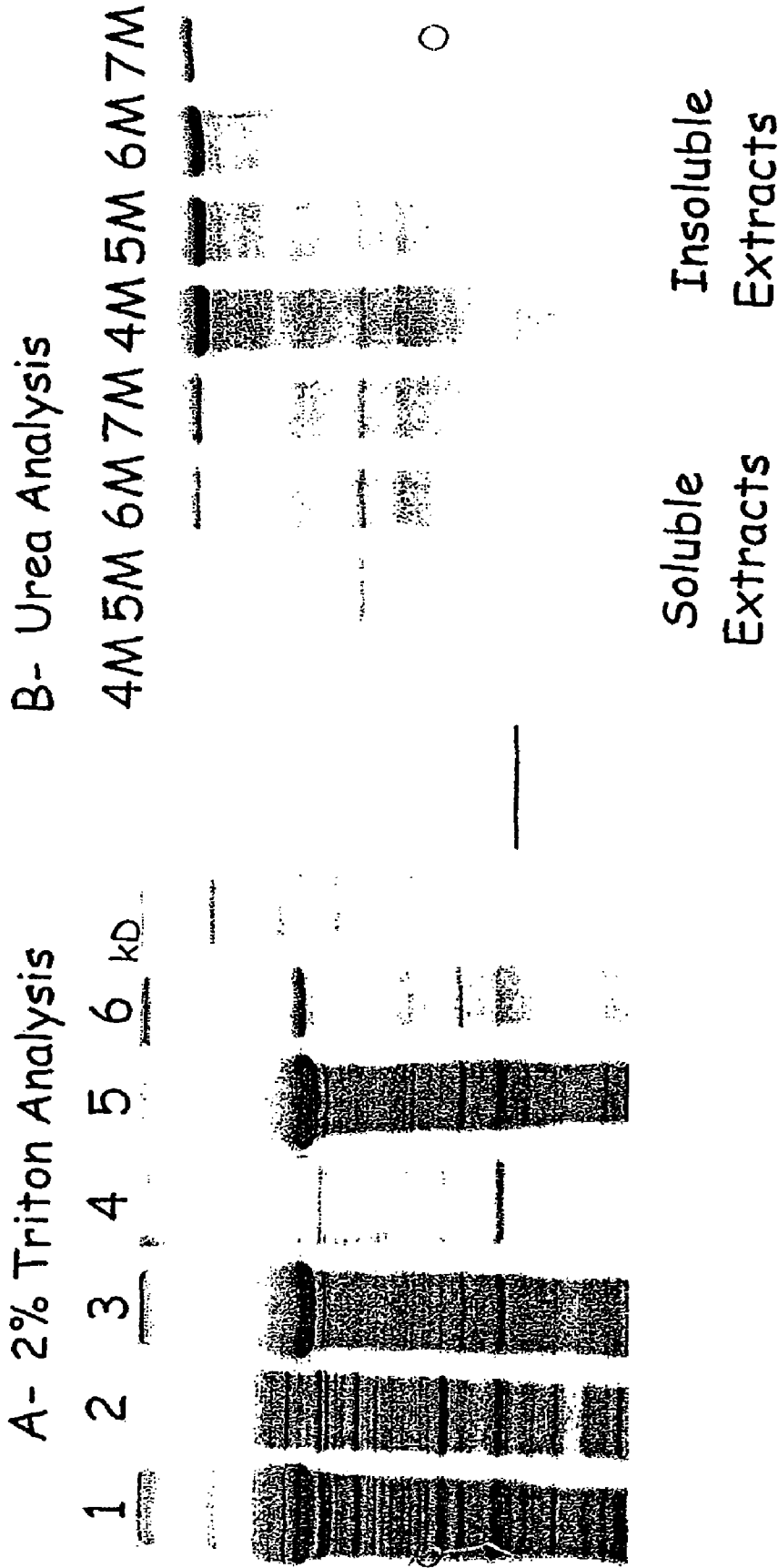


FIGURE 6

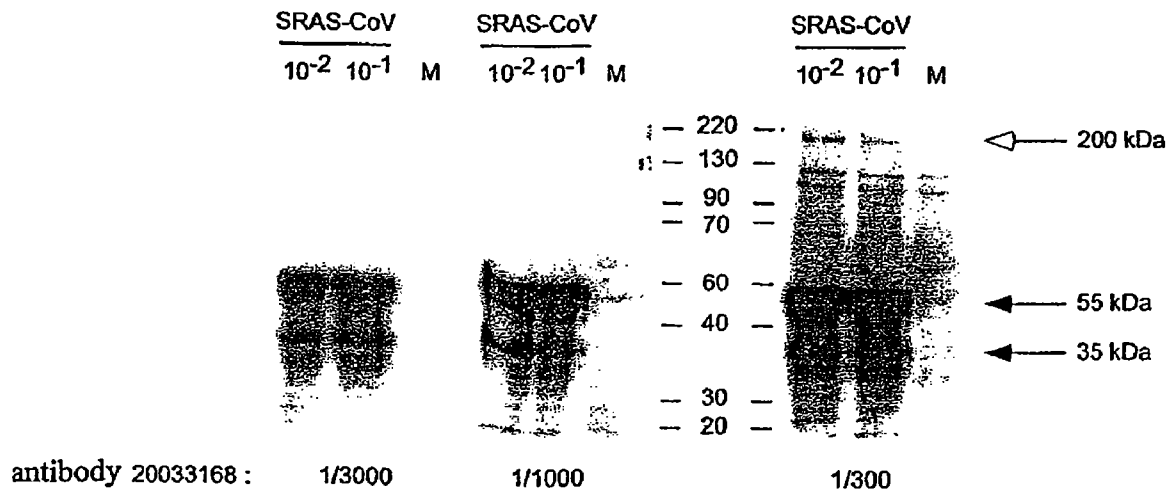


FIGURE 7

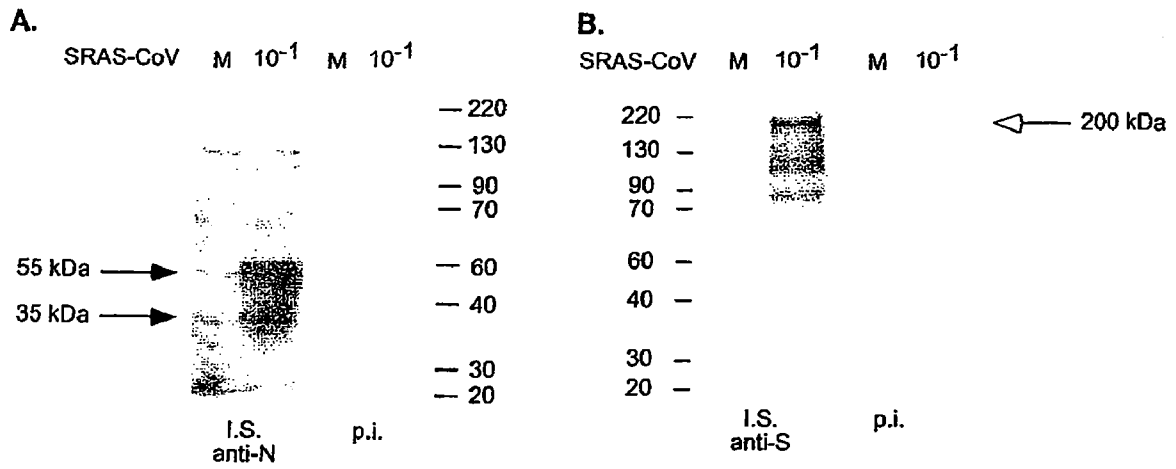
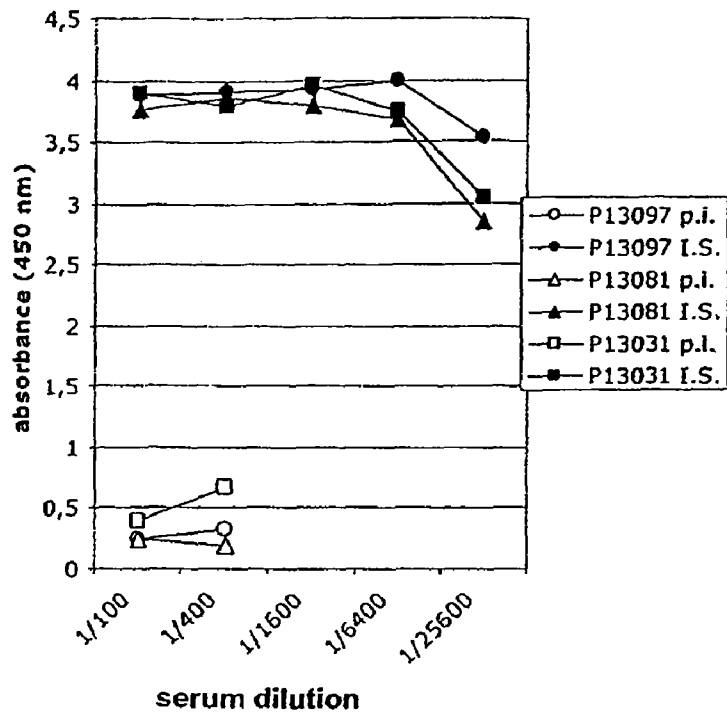


FIGURE 8

A



B

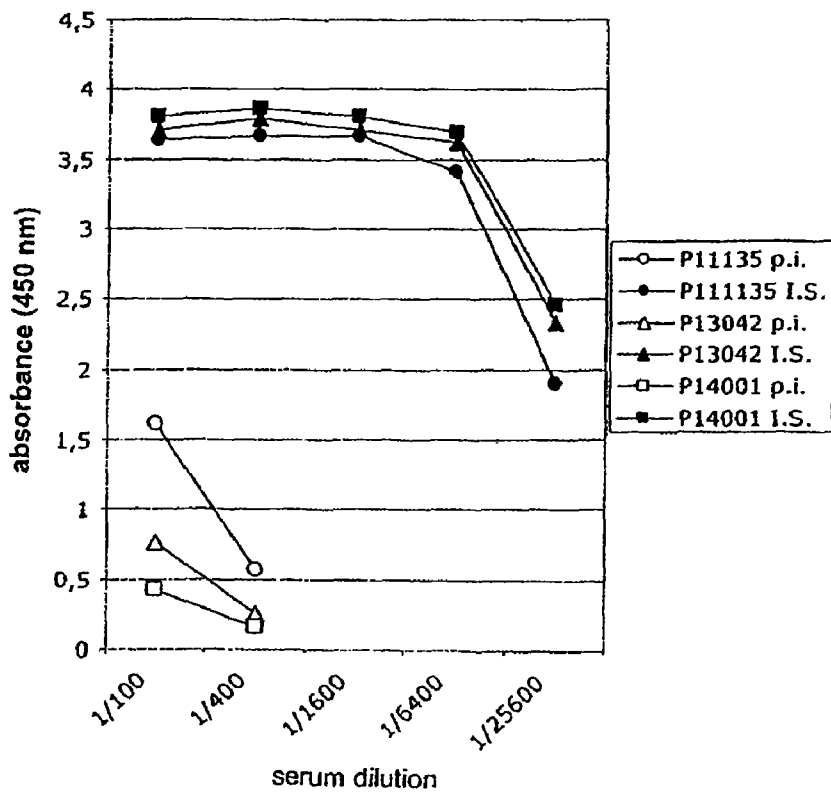


FIGURE 9

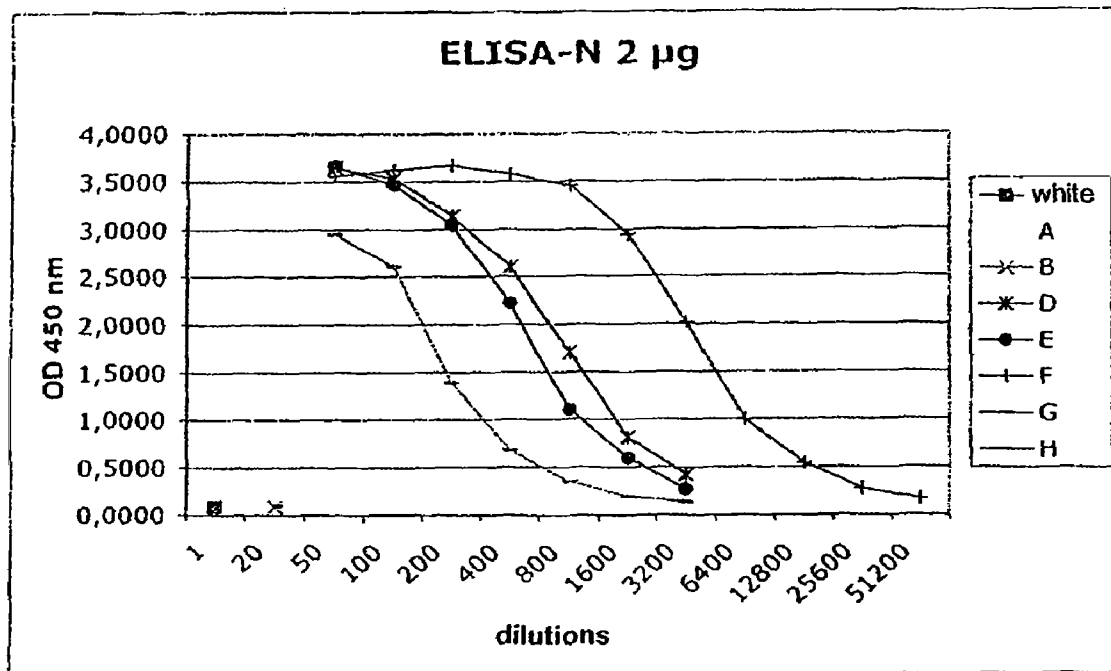
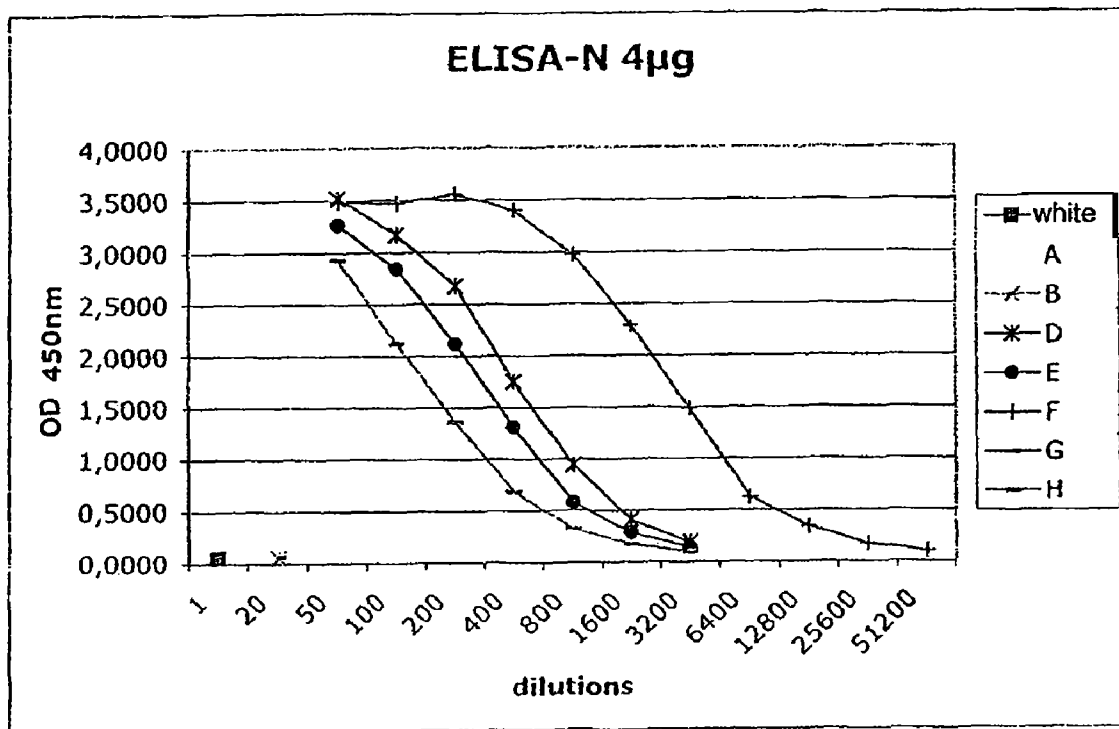


FIGURE 10a

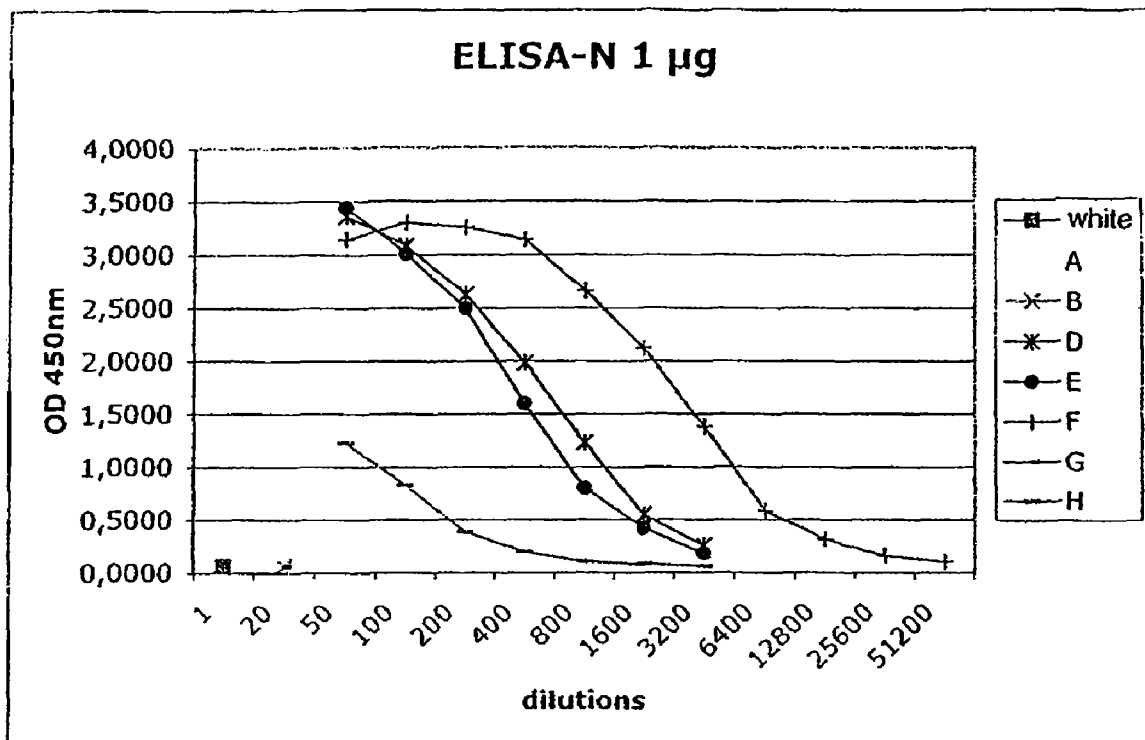


FIGURE 10b

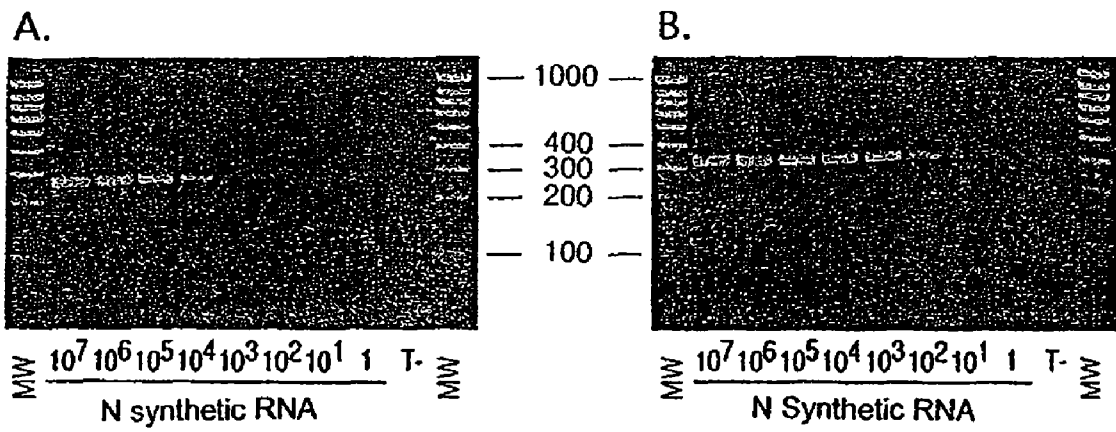


FIGURE 11

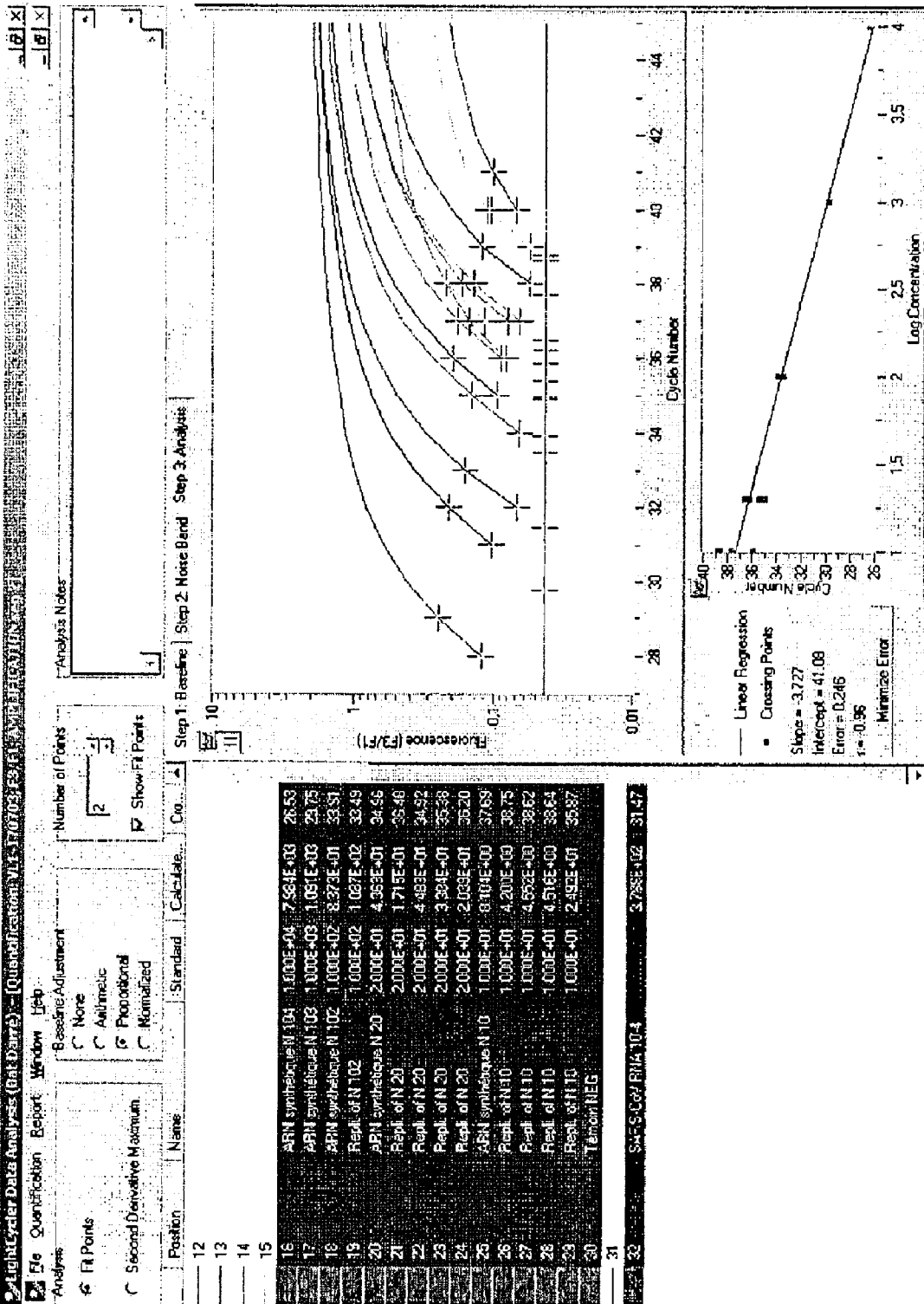


FIGURE 12

```

                                >< XhoII
                                >< Sau3AI
                                >< NdeII
                                >< MflI
                                >< MboI
                                >< DpnII
                                >< MnlI>< DpnI
                                >< BstYI
                                >< BspAI
                                >< Bsp143I
                                >< BglII
                                >< VneI
                                >< SphI
                                >< SnoI
                                >< RmaI
                                >< PaeI >< SduI
                                >< NspI >< NspII
                                >< NspHI >< HgiAI
                                >< NlaIII >< Bsp1286I
                                >< MaeI >< BmyI
                                >< ApaLI
                                >< Alw44I
                                >< Alw21I
                                >< SfcI
                                >< PstI
                                >< MnlI
                                >< Ksp632I
                                >< EarI
                                >< Eam1104I
                                >< HindII >< MboII >< EarI
                                >< HincII >< MaeIII >< Eam1104I
                                >< TthHB8I >< StyI
                                >< TaqI >< RmaI >< ScrFI
                                >< Sau3AI >< MaeI >< NciI
                                >< NdeII >< EcoT14I >< MspI
                                >< MboI >< Ecol30I >< MaeIII
                                >< DpnII >< BssT1I >< HpaII
                                >< DpnI >< BsaJI >< HapII
                                >< BspAI >< BlnI >< DsaV
                                >< Bsp143I >< AvrII >< BcnI
                                >< RmaI
                                >< Esp3I >< MaeII
                                >< BsmAI >< MaeI
                                >< HincII >< MaeII> < Eco57I >< Alw26I >< BsmBI
                                >< HincII > < AflIII > < DdeI
                                >< Alw26I >> < BsmBI
                                >< RmaI
                                >< Esp3I >< MaeII
                                >< BsmAI >< MaeI
                                >< HincII >< MaeII> < Eco57I >< Alw26I >< BsmBI
                                >< HincII > < AflIII > < DdeI
                                >< Alw26I >> < BsmBI

```

ATATTAGGTT TTTACCTACC CAGGAAAAGC CAACCAACCT CGATCTCTTG TAGATCTGTT CTCTAAACGA
 10 20 30 40 50 60 70

ACTTTAAAAT CTGTGTAGCT GTCGCTCGGC TGCATGCCTA GTGCACCTAC GCAGTATAAA CAATAATAAA
 80 90 100 110 120 130 140

TTTTACTGTC GTTGACAAGA AACGAGTAAC TCGTCCCTCT TCTGCAGACT GCTTACGGTT TCGTCCGTGT
 150 160 170 180 190 200 210

TGCAGTCGAT CATCAGCATA CCTAGGTTTC GTCCGGGTGT GACCGAAAGG TAAGATGGAG AGCCTTGTTT
 220 230 240 250 260 270 280

TTGGTGTCAA CGAGAAAACA CACGTCCAAC TCAGTTTGCC TGTCCTTCAG GTTAGAGACG TGCTAGTGCC
 290 300 310 320 330 340 350

FIGURE 13.1

```

                >< Sau96I
                >< PssI
                >< Pali
                >< NspIV
                >< MnlI
                >< HaeIII
                >< Eco0109I
                >< DraII>< MboII >< PmlI
                >< MnlI >< Cfr13I >< PmaCI
                >< Ksp632I >< BsuRI > < MaeII
                >< HinfI >< BsiZI>< EcoNI >< Eco72I
                >< EarI >< BshI >< BslI >< BsaAI
                >< PleI >< Eam1104I>< AsuI >< BsiYI>< BbrPI >< MnlI
TGGCTTCGGG GACTCTGTGG AAGAGGCCCT ATCGGAGGCA CGTGAACACC TCAAAAATGG CACTTGTGGT
    360          370          380          390          400          410          420

                >< Tru9I
                >< SfaNI
                >< MseI
                >< MaeII
                >< RmaI >< RsaI >< Csp6I >< BspWI >< MseI >< MaeII
                >< MaeI >< AluI >< AfaI >< AluI >> < MaeII
CTAGTAGAGC TGGAAAAGG CGTACTGCC CAGCTTGAAC AGCCCTATGT GTTCATTAAG CGTTCTGATG
    430          440          450          460          470          480          490

                >< Pali
                >< HaeIII
                >< Tru9I >< GdiII >< RsaI
                >< MseI >< EaeI >> McrI ><
                >< Esp4I >< BsuRI >> BsmI BsiEI ><
                >< AflII >< BshI >> AluI >< BscCI >< AfaI
CCTTAAGCAC CAATCACGGC CACAAGGTCG TTGAGCTGGT TGCAGAAATG GACGGCATTG AGTACGGTCG
    500          510          520          530          540          550          560

                >< NspI
                >< ScaI >< NspHI
                >< RsaI >< NlaIII
                > < Csp6I >< BslI
                >< BsrI >> BsiYI >> MboII
                >< AciI >> AfaI >> AflIII >> MunI >> AciI
TAGCGGTATA ACGTGGGAG TACTCGTGCC ACATGTGGGC GAAACCCCAA TTGCATACCG CAATGTTCTT
    570          580          590          600          610          620          630

                >< TthHB8I
                >< TaqI
                >< Sau3AI
                >< NdeII
                >< MboI
                >< DpnII
                > < DpnI
                >< ClaI
                >< Bsu15I
                >< BspDI
                >< BspAI
                > < Bsp143I
                >< Bsp106I
                >< BsiXI >> MaeIII >
                >< Cfr10I >> BscI>> SfaNI DdeI ><
                >< BscBI >> AluI >> BanIII >> BfrI ><
CTTCGTAAGA ACGGTAATAA GGGAGCCGGT GGTACATAGCT ATGGCATCGA TCTAAAGTCT TATGACTTAG
    640          650          660          670          680          690          700

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FIGURE 13.2


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>> MvaI      >> Hin6I      >> SduI      >> Csp45I
>> Ecl136I   >> HhaI      >> NspII     >> BstBI
>> BstOI     >> HaeII     >> HgiAI     >> Bsp119I
>> BstNI     >> Eco47III   >> Bsp1286I >> BsiCI
>> BsiLI     >> CfoI      >> BmyI      >> Bpu14I
>> ApyI >> DdeI >> Bsp143II >> AluI >> Alw21I >> AsuII
CTGGTTCAC T GAGCGCTCTG ATAAGAGCTA CGAGCACCAG ACACCCTTCG AAATTAAGAG TGCCAAGAAA
    990      1000      1010      1020      1030      1040      1050

                                >> Tru9I
                                >> MseI
                                >> BsmI
                                >> BscCI
                                >< MnlI
TTTGACACTT TCAAAGGGGA ATGCCCAAAG TTTGTGTTTC CTCTTAACTC AAAAGTCAAA GTCATTCAAC
    1060      1070      1080      1090      1100      1110      1120

>> PmlI
>> PmaCI
>> MaeII
>> Eco72I
>> BsaAI
>> BbrPI
>> AflIII   >> MnlI >> DdeI
CACGTGTTGA AAAGAAAAAG ACTGAGGGTT TCATGGGGCG TATACGCTCT GTGTACCCTG TTGCATCTCC
    1130      1140      1150      1160      1170      1180      1190

>> SfaNI
>> MaeIII   >> AccI
ACAGGAGTGT AACAAATATGC ACTTGTCTAC CTTGATGAAA TGTAATCATT GCGATGAAGT TTCATGGCAG
    1200      1210      1220      1230      1240      1250      1260

                                >> SinI
                                >> Sau96I
                                PssI ><
                                >> Psp5II
                                >> PpuMI
                                >> NspIV
                                >> NspHII
                                >> Eco47I
                                >> DraII
                                >> Cfr13I
                                >> BsiZI
                                >> Bme18I
                                >> AvaII
                                >> AsuI

>> MaeII
ACGTGCGACT TTCTGAAAAG CACTTGTGAA CATTGTGGCA CTGAAAATT AGTTATTGAA GGACCTACTA
    1270      1280      1290      1300      1310      1320      1330

                                EcoO109I >> AflIII >

                                Van9I ><
                                SinI ><
                                Sau96I ><
                                PflMI ><
                                NspIV ><
                                NspHII >
                                Eco47I ><
                                Cfr13I ><
                                BsiI ><
                                BsiZI ><
                                BsiYI ><
                                Bme18I ><
                                AvaII ><
                                AsuI ><

>> RsaI
>> NspI
>> NlaIV
>> NlaIII
>> NspHI >< KpnI
>> Eco64I
>> Csp6I
>> BscBI
>> BanI
>> Asp718
>> AfaI
>> AccBII
    
```

FIGURE 13. 4

```

    >> Acc65I          >> SfcI          >> NlaIII         AccB7I >>
CATGTGGGTA CCTACCTACT AATGCTGTAG TGAAAATGCC ATGTCCTGCC TGTCAAGACC CAGAGATTGG
    1340          1350          1360          1370          1380          1390          1400

                                >> TthHB8I
                                >> TaqI>> MnlI
                                >> HinfI

    >> DdeI
ACCTGAGCAT AGTGTTCGAG ATTATCACAA CCACTCAAAC ATTGAAACTC GACTCCGCAA GGGAGGTAGG
    1410          1420          1430          1440          1450          1460          1470

                                >> PleI          >> AciI

    >> RmaI
    >> MnlI
    >> MaeI
ACTAGATGTT TTGGAGGCTG TGTGTTTGCC TATGTTGGCT GCTATAATAA GCGTGCCTAC TGGGTTCCCTC
    1480          1490          1500          1510          1520          1530          1540

                                NlaIV >>
                                >> BsrI
                                BscBI >>

    >> MaeI          >> BbvI          >> Fnu4HI          BscBI >>
ACTAGATGTT TTGGAGGCTG TGTGTTTGCC TATGTTGGCT GCTATAATAA GCGTGCCTAC TGGGTTCCCTC
    1480          1490          1500          1510          1520          1530          1540

                                XhoII >>
                                Sau3AI >>
                                NdeII >>
                                MflI >>
                                MboI >>

                                >> MaeIII
                                >> Eco31I          DpnII >>
                                >> HaeIII          >> BsrI          >> MnlI DpnI >
    >> RmaI          >> BsuRI          >> BsrI          >> BsmAI          BstYI >>
    >> MnlI          > < DdeI          >> BspWI          >> BsaI>< HphI          BspAI >>
    >> MaeI          >> BshI>> BglI          >> Alw26I          BspI43I >
GTGCTAGTGC TGATATTGGC TCAGGCCATA CTGGCATTAC TGGTGACAAAT GTGGAGACCT TGAATGAGGA
    1550          1560          1570          1580          1590          1600          1610

                                > < Tru9I
                                > < MseI
                                >> MaeII          >> Tru9I
                                >> HpaI
                                >> HindII
                                >> HincII
    >> AlwI          >> DdeI          >> AflIII          >> MseI
TCTCCTTGAG ATACTGAGTC GTGAACGTGT TAACATTAAC ATTGTTGGCG ATTTTCATTT GAATGAAGAG
    1620          1630          1640          1650          1660          1670          1680

                                >> MboII
                                >> BstXI          >> SfaNI          > < HinfI
GTTGCCATCA TTTTGGCATC TTTCTCTGCT TCTACAAGTG CCTTTATTGA CACTATAAAG AGTCTTGATT
    1690          1700          1710          1720          1730          1740          1750

                                >> StyI
                                >> MaeIII
                                >> EcoT14I
                                >> Eco130I
                                >> PleI
                                >> MaeIII
                                >> BssT1I          BslI >>
    >> HinfI>> AciI          >> BsaJI          BsiYI >>
ACAAGTCTTT CAAACCATT GTTGAGTCCT GCGGTAAC TAAGTTACC AAGGGAAAGC CCGTAAAAGC
    1760          1770          1780          1790          1800          1810          1820

                                >> Sau3AI
                                >> NdeII
                                >> MboI
                                >> DpnII
                                >> DpnI >> Tru9I
                                >> BspAI >> MseI
                                >> Bsp143I
                                >> Van91I
                                >> PflMI
                                >> DraIII
                                >> BslI
                                >> BsiYI
                                >> BbvI          >> MnlI
                                >> AccB7I          Fnu4HI >>

```

FIGURE 135

```

TGCTTGAAC ATTGGACAAC AGAGATCAGT TTTAACACCA CTGTGTGGTT TTCCCTCACA GGCTGCTGGT
 1830      1840      1850      1860      1870      1880      1890

      << ThaI
      << SfaNI
      << MvnI
      << HinPII
<< HinPII
      << Hin6I
<< Hin6I
      << HhaI
<< Sau3AI      << HhaI
<< NdeII      << CfoI
<< MboI      << CfoI
<< DpnII      << BstUI
      << DpnI      << BssHII
<< BspAI      << Bsp50I
      << Bsp143I      << AccII
GTTATCAGAT CAATTTTTCG GCGCACACTT GATCAGCAA ACCACTCAAT TCCTGATTTC CAAAGAGCAG
 1900      1910      1920      1930      1940      1950      1960

      << TthHB8I
      << StyI
      << NcoI
      << HindII
      << HincII
      << HinII
      << EcoT14I
      << Eco57I
      << TaqI>> Eco130I
<< SalI >> DsaI
<< RtrI >> BssT1I
      << BsaHI
      << BbiII>> NlaIII
      << AcyI >> HgaI
<< MaeIII
      << BbvI
      << MaeII >> AccI>> BsaJI HphI >>
CTGTCACCAT ACTTGATGGT ATTTCTGAAC AGTCATTACG TCTTGTGCGAC GCCATGGTTT ATACTTCAGA
 1970      1980      1990      2000      2010      2020      2030

      << RsaI
      << NdeI
      << MaeIII >> BsrI >< Csp6I
      << BspMI
      << MaeIII >> BsrI >< AfaI >> DdeI
CCTGCTCACC AACAGTGTC AATTATGGC ATATGTAAC TGTGGTCTTG TACAACAGAC TTCTCAGTGG
 2040      2050      2060      2070      2080      2090      2100

      << StuI
      << Pali
      << HaeIII
      << Eco147I
      << SduI
      << NspII
      << Bsp1286I
      << BmyI
      << DdeI
      << BsuRI
      << BshI
      << AatI
      << MnlI
      << DdeI >>
      << BfrI >>
TTGTCTAATC TTTTGGGCAC TACTGTTGAA AACTCAGGC CTATCTTTGA ATGGATTGAG GCGAAACTTA
 2110      2120      2130      2140      2150      2160      2170

      << TfiI
      << HinfI
      << SfaNI >> BsgI
      << FokI
      << Tth111I >>
      << AspI >>
GTGCAGGAGT TGAATTTCTC AAGGATGCTT GGGAGATTCT CAAATTTCTC ATTACAGGTG TTTTGCAT
 2180      2190      2200      2210      2220      2230      2240

```

FIGURE 13.6


```

Tru9I ><
MseI ><
HpaI >
HindII >
HincII >
>< Eco57I
CGTCAAGGGT CAAATACAGG TTGCTTCAGA TAACATCAAG GATTGTGTAA AATGCTTCAT TGATGTTGTT
2250          2260          2270          2280          2290          2300          2310

>< Sau3AI
>< NdeII
>< MboI
      > < MaeIII
      >< FbaI
>< DpnII
  >< DpnI
  >< BspAI
  >< Bsp143I
    >< TthHB8I
    >< TaqI
AACAAAGGCAC TCGAAATGTG CATTGATCAA GTCACTATCG CTGGCGCAAA GTTGCGATCA CTCAACTTAG
2320          2330          2340          2350          2360          2370          2380

      >< PvuII
      >< MaeII
      >< Bst1107I
      >< BsaAI
      >< BbvI
    >< HphI
    >< DrdI
    >< AccI
GTGAAGTCTT CATCGCTCAA AGCAAGGGAC TTTACCGTCA GTGTATACGT GGCAAGGAGC AGCTGCAACT
2390          2400          2410          2420          2430          2440          2450

    >< Tru9I
    >< NlaIV
    >< MseI
    >< MnlI
    >< Esp4I
    >< Eco64I
    >< BscBI
  >< NlaIII >< BanI
  >< AflIII
  >< BbvI
  >< AccB1I
  >< MaeIII
  >< TfiI
  >< HinfI
  >< HphI
  >< AfaI
ACTCATGCCT CTTAAGGCAC CAAAAGAAGT AACCTTTCTT GAAGGTGATT CACATGACAC AGTACTTAC
2460          2470          2480          2490          2500          2510          2520

      > < XhoI
      >< TthHB8I
    >< TthHB8I>< TaqI
      > < SlaI
      > < PaeR7I
      > < NspIII
    >< HphI >< HinfI
      > < Eco88I
      > < CcrI
    >< Esp3I >< BsaHI
      > < BcoI
    >< BsmAI >< BbiII
      > < AvaI
      >< HgaI
    >< TaqI > < Ama87I>< BsmBI
    >< DdeI>< MnlI
    >< Alw26I >< AcyI
TCTGAGGAGG TTGTTCTCAA GAACGGTGAA CTCGAAGCAC TCGAGACGCC CGTTGATAGC TTCACAAATG
2530          2540          2550          2560          2570          2580          2590

```

FIGURE 13.7

```

                >> Pali >> NlaIII
                >> HaeIII >> MnlI
                >> BsuRI >> DdeI >> Tru9I
                >> BshI >> BfrI >> MseI
    >> AluI          >> BsrI
GAGCTATCGT TGGCACACCA GTCTGTGTAA ATGGCCTCAT GCTCTTAGAG ATTAAGGACA AAGAACAATA
    2600          2610          2620          2630          2640          2650          2660

                >> VneI
                Tru9I ><
                >< SnoI
                >> SduI
                >> NspII
                MseI ><
                >> HgiAI
                Bsp1286I >< BslI ><
                BsiYI ><
                >< BmyI
                >< ApaLI
                >< Alw44I
                >> Tru9I >> Alw21I
                >> MseI
CTGCGCATTG TCTCCTGGTT TACTGGCTAC AAACAATGTC TTTCGCTTAA AAGGGGGTGC ACCAATTAAG
    2670          2680          2690          2700          2710          2720          2730

                >> TfiI
    >> MaeIII          >> MboII > < MaeIII >> HinfI AluI ><
GGTGTAACCT TTGGAGAAGA TACTGTTTGG GAAGTTCAAG GTTACAAGAA TGTGAGAATC ACATTTGAGC
    2740          2750          2760          2770          2780          2790          2800

                >> RsaI
                >> NlaIV
                MaeIII ><
                >< MspI >< KpnI
                >< HpaII
                >< HapII
                > < Eco64I
                >> SduI
                >> NspII >> TfiI >> BscBI
                >> HgiAI >> BanI
                >> Bsp1286I >> Asp718
                >> BmyI >> HinfI >> AfaI
                >> Alw21I >> AccB1I
                >> AflIII >> MseI >> AccI >> Acc65I
TTGATGAACG TGTGACAAA GTGCTTAATG AAAAGTGCTC TGTCTACACT GTGGAATCCG GTACCGAAGT
    2810          2820          2830          2840          2850          2860          2870

                >> Sau3AI
                >> NdeII
                >> MboI
                >> DpnII
                > < DpnI
                >> NspI
                >> NspHI >> MboII >> BspAI
                >> NlaIII >> BsrI >> Bsp143I
    >> DdeI          >> MnlI >> AlwNI >> BbsI >> AlwNI
TACTGAGTTT GCATGTGTTG TAGCAGAGGC TGTGTGAAG ACTTTACAAC CAGTTTCTGA TCTCCTTACC
    2880          2890          2900          2910          2920          2930          2940

                >> Sau3AI
                >> NdeII
                >> MboI
                >> DpnII
                >> DpnI
                >> BspAI

```

FIGURE 13.8

```

    >> NlaIII>> Bsp143I          >> AluI          >> SfaNI
AACATGGGTA TTGATCTTGA TGAGTGGAGT GTAGCTACAT TCTACTTATT TGATGATGCT GGTGAAGAAA
    2950          2960          2970          2980          2990          3000          3010

                                >> SfaNI
                                >> MnlI
    >> MboII          >> GsuI          >> Ksp632I          >> MnlI
                                >> BsaAI          >> EarI          >< MboII
    >> HphI >> MaeII>> BpmI          >> MnlI >> Eam1104I >< MboII
ACTTTTCATC ACGTATGTAT TGTTCCCTTT ACCCTCCAGA TGAGGAAGAA GAGGACGATG CAGAGTGTGA
    3020          3030          3040          3050          3060          3070          3080

                                >< RsaI
                                >> RsaI
    >> NlaIII
                                >> MnlI          >< FokI
                                >> Csp6I          Eco31I >>
                                >> Csp6I          >< MamI BsmAI ><
                                >< MboII          >< AfaI          >< BsiBI BsaI ><
    >> MboII          >> AfaI          >< BsaBIALw26I ><
GGAAGAAGAA ATTGATGAAA CCTGTGAACA TGAGTACGGT ACAGAGGATG ATTATCAAGG TCTCCCTCTG
    3090          3100          3110          3120          3130          3140          3150

    >> NlaIV>> PvuII>> XmnI
    >> Eco64I >> Psp5I >> TthHB8I
    >> MnlI >> DdeI          >> TaqI          >> MnlI          >> MboII
    >> BscBI>> NspBII >< MnlI          >> Ksp632I          >> MboII >< MboII
    >> BanI          >< MnlI          >> EarI          >> BsrI
    >> AccBII >> AluI >> Asp700I          >> Eam1104I >< MboII>< BbsI
GAATTTGGTG CCTCAGCTGA AACAGTTCGA GTTGAGGAAG AAGAAGAGGA AGACTGGCTG GATGATACTA
    3160          3170          3180          3190          3200          3210          3220

                                >> Tru9I
                                >> MseI          >< Eco57I
    >> FokI          >> BsrI>< MboII BsrI ><
    >> DdeI          CTGAGCAATC AGAGATTGAG CCAGAACCAG AACCTACACC TGAAGAACCA GTTAATCAGT TTAATGGTTA
    3230          3240          3250          3260          3270          3280          3290

    >> Tru9I          >> MnlI
    >> MseI          >> Tru9I >> HindII>< Tru9I          >> DraIII
    >> DraI          >> MseI >> HincII>< MseI          >> BspWI
TTTAAAACCTT ACTGACAATG TTGCCATTAA ATGTGTTGAC ATCGTTAAGG AGGCACAAAG TGCTAATCCT
    3300          3310          3320          3330          3340          3350          3360

                                >> VneI
                                >> SnoI
                                >< SduI
                                >< NspII
                                >< HgiAI
                                >< Bsp1286I
                                >< BmyI
                                >< ApaLI
    >> HphI          >< NlaIII          >< Alw44I
    >> BbvI          >< Fnu4HI          >> BspMI          >< Alw21I
ATGGTGATTG TAAATGCTGC TAACATACAC CTGAAACATG GTGGTGGTGT AGCAGGTGCA CTCAACAAGG
    3370          3380          3390          3400          3410          3420          3430

                                >> Sau96I
                                >> Pali
                                >> NspIV
                                >> HaeIII
    >> NlaIV          >> Cfr13I

```

FIGURE 13.9

```

    >< Eco64I
      >< BscBI
    >< BanI
      >< AccB1I>< NlaIII
CAACCAATGG TGCCATGCAA AAGGAGAGTG ATGATTACAT TAAGCTAAAT GGCCCTCTTA CAGTAGGAGG
    3440      3450      3460      3470      3480      3490      3500

    >< BsuRI
    >< Tru9I
    >< BsiZI
    >< MseI
    >< BshI
    >< MnlI
    >< AluI
    >< AsuI
    >< MnlI
    >< SinI
    >< Sau96I
    >< NspIV
    >< NspHI>< NspHII
    >< Eco47I
    >< Cfr13I
    >< NlaIII
    >< BspMI
    >< BsiZI
    >< Bme18I
    >< AvaII MnlI ><
    >< DdeI
    >< NspI>< AsuI FokI ><
GTCTTGTTTG CTTTCTGGAC ATAATCTTGC TAAGAAGTGT CTGCATGTTG TTGGACCTAA CCTAAATGCA
    3510      3520      3530      3540      3550      3560      3570

    >< Tru9I
    >< HphI> < MseI
    >< Esp4I
    >< AluI
    >< NdeI
    >< AflIII>< Fnu4HI
    >< BbvI
GGTGAGGACA TCCAGCTTCT TAAGGCAGCA TATGAAAATT TCAATTCACA GGACATCTTA CTGCACCAT
    3580      3590      3600      3610      3620      3630      3640

    RsaI ><
    Csp6I ><
    AfaI ><
    >< Eco57I
    >< BcgI
TGTTGTCAGC AGGCATATTT GGTGCTAAAC CACTTCAGTC TTTACAAGTG TGCGTGCAGA CGGTTTCGTAC
    3650      3660      3670      3680      3690      3700      3710

    >< BsqI
    >< BspMI
    >< BcgI/a
    >< AluI
    >< NlaIII
ACAGGTTTAT ATTGCAGTCA ATGACAAAGC TCTTTATGAG CAGGTTGTCA TGGATTATCT TGATAACCTG
    3720      3730      3740      3750      3760      3770      3780

    >< MnlI
    >< RmaI
    >< MaeI
    >< MnlI
    >< Eco57I
    >< NlaIV
    >< BscBI
    >< TfiI
    >< MboII
    >< HinfI
    >< DdeI
AAGCCTAGAG TGAAGCACC TAAACAAGAG GAGCCACCAA ACACAGAAGA TTCCAAAAC TCCAGAGAAAT
    3790      3800      3810      3820      3830      3840      3850

    >< Tru9I
    >< StuI
    >< Pali
    >< MseI
    >< MnlI
    >< MaeIII
    >< HaeIII
    >< Eco065I
    >< Eco147I
    >< Eco91I
    >< BsuRI
    >< BstXI ><
    >< RsaI
    >< Csp6I
    >< AfaI
    >< TthHB8I
    >< TaqI
    >< BshI
    >< BstPI
    >< AatI
    >< BstEII
CTGTCGTACA GAAGCCTGTC GATGTGAAGC CAAAATTTAA GGCCTGCATT GATGAGGTTA CCACAACACT
    3860      3870      3880      3890      3900      3910      3920

    TfiI ><
    NlaIII ><
    HinfI ><
    >< DdeI
    >< EcoRV
    >< HindIII

```

FIGURE 13.10

```

    >> BsrI      >> MboII      >> MaeIII      >> Eco32I      >> AluI
GGAAGAACT AAGTTTCTTA CCAATAAGTT ACTCTTGTTT GCTGATATCA ATGGTAAGCT TTACCATGAT
    3930      3940      3950      3960      3970      3980      3990

    >> NspI
    >> NspHI
    >> NlaIII
    >> MnlI      >> SfaNI
    >> DdeI      >> EcoNI
    >> BfrI      >> MboII >> BslI      >> NlaIII
    >> HphI      >> BsiYI      >> FokI
TCTCAGAACA TGCTTAGAGG TGAAGATATG TCTTTCCTTG AGAAGGATGC ACCTTACATG GTAGGTGATG
    4000      4010      4020      4030      4040      4050      4060

    >> SpeI
    >> RmaI
    >> MaeI      >> EcoRV >> HphI      >> SfaNI
    >> HphI      >> Eco32I      >> MnlI      >> DdeI
TTATCACTAG TGGTGATATC ACTTGTTGTTG TAATACCCTC CAAAAAGGCT GGTGGCACTA CTGAGATGCT
    4070      4080      4090      4100      4110      4120      4130

    >> ScrFI
    >> RsaI
    >> MvaI
    >> EcoRII
    >> Ecl136I
    >> DsaV
    >> Csp6I >> EcoNI
    >> BstOI
    >> BstNI
    >> BsiLI
    >> BsaJI
    >> BsaAI      >> BslI
    >> MaeII >> ApyI
    >> AluI      >> BsrI      >> AfaI      >> BsiYI
CTCAAGAGCT TTGAAGAAAG TGCCAGTTGA TGAGTATATA ACCACGTACC CTGGACAAGG ATGTGCTGGT
    4140      4150      4160      4170      4180      4190      4200

    >> Tru9I
    >> MseI
    >> DdeI      >> Esp4I      >> RsaI
    >> MnlI      >> BspWI      >> Csp6I
    >> FokI      >> AluI      >> AflIII      >> Eco57I >> AfaI
TATACACTTG AGGAAGCTAA GACTGCTCTT AAGAAATGCA AATCTGCATT TTATGTACTA CCTCAGAAG
    4210      4220      4230      4240      4250      4260      4270

    >> ScrFI
    >> MvaI
    >> EcoRII
    >> XmnI      >> EcoRII      >> NlaIII >>
    >> Ksp632I      >> RmaI      >> DsaV      >> Ksp632I >>
    >> EarI      >> TfiI >> MboII      >> BstOI      >> EarI
    >> Eam1104I      >> MaeI      >> BstNI      >> Eam1104I >>
    >> DdeI      >> HinfI      >> BsiLI      >> BsmAI >>
    >> BspWI      >> Asp700I      >> ApyI      >> Alw26I >>
CACCTAATGC TAAGGAAGAG ATTCTAGGAA CTGTATCCTG GAATTTGAGA GAAATGCTTG CTCATGCTGA
    4280      4290      4300      4310      4320      4330      4340

    >> VspI      >> Zsp2I
    >> Tru9I      >> Ppu10I
    >> MseI      >> NsiI
    >> MboII      >> NlaIII      >> FokI
    >> Eco57I      >> Mph1103I      >> FokI

```

FIGURE 13. 11

```

                >> AsnI           >> EcoT22I           >> BspWI
                >> AseI           >> AvaIII           >> BglI           >> MaeII
AGAGACAAGA AAATTAATGC CTATATGCAT GGATGTTAGA GCCATAATGG CAACCATCCA ACGTAAAGTAT
    4350         4360         4370         4380         4390         4400         4410

                >> SfaNI
                >> Tru9I           > < HindII           >> TfiI           >> SpeI
                >> MseI           > < HincII>< MboII           >> RmaI
                >> MnlI           >> DrdI >> HinfI           >> MaeI
AAAGGAATTA AAATTCAAGA GGCATCGTT GACTATGGTG TCCGATTCTT CTTTATACT AGTAAAGAGC
    4420         4430         4440         4450         4460         4470         4480

                >> MaeIII
                >> SfcI           >> Fnu4HI           >> MnlI
                >> AluI           >> AluI           >> AciI           >> MaeIII ><
CTGTAGCTTC TATTATTACG AAGCTGAACT CTCTAAATGA GCCGCTTGTC ACAATGCCAA TTGGTTATGT
    4490         4500         4510         4520         4530         4540         4550

                >> ThaI
                >> MvnI
                >> MboII
                >> HinPII
                >< HinPII
                >< Hin6I
                >< Hin6I
                >< HhaI
                >< HhaI
                >> Tru9I           >> HhaI
                >> NlaIII           >> Fnu4HI
                >> MseI           >> CfoI
                >> MnlI           >> CfoI
                >> Ksp632I           >> BstUI
                >> EarI           >< BssHII>< BspWI           >< Tru9I
                >> Eam1104I           >< Bsp50I           >< MseI
                >> BbvI           >< AccII           >> AluI           HphI ><
GACACATGGT TTTAATCTTG AAGAGGCTGC GCGCTGTATG CGTTCCTTA AAGCTCCTGC CGTAGTGTCA
    4560         4570         4580         4590         4600         4610         4620

                >> MaeIII
                >< SfaNI           >> AlwNI           >> MnlI >> MnlI>< DdeI
GTATCATCAC CAGATGCTGT TACTACATAT AATGGATACC TCACTTCGTC ATCAAAGACA TCTGAGGAGC
    4630         4640         4650         4660         4670         4680         4690

                >< SinI
                >< Sau96I
                >< NspIV
                >< NspHII
                >< SduI           >< Eco47I
                >< NspII           >< Cfr13I
                >< HgiAI           >< BsiZI
                >< Bsp1286I           >< Bme18I           >< RsaI
                >< BmyI           >< AvaII           >< Csp6I
                >< Alw21I           >< AsuI           >< AfaI
ACTTTGTAGA AACAGTTTCT TTGGCTGGCT CTTACAGAGA TTGGTCTTAT TCAGGACAGC GTACAGAGTT
    4700         4710         4720         4730         4740         4750         4760

                > < TthHB8I
                > < TaqI
                >< SduI
                >> Van91I           >> NspII
                >< Tru9I           >< RsaI >< PflMI           >< Eco24I
                >< MseI           >< HphI           >< BslI           >< Bsp1286I
                >< Esp4I           >< Csp6I           >< BsiYI           >< BmyI GsuI ><

```

FIGURE 13.12

```

      << AflIII << MaeIII      << AfaI << AccB7I << BanIIBpmI <<
AGGTGTTGAA TTTCTTAAGC GTGGTGACAA AATTGTGTAC CACACTCTGG AGAGCCCCGT CGAGTTTCAT
      4770      4780      4790      4800      4810      4820      4830

      << Tru9I
      << PleI << EcoNI
      << MnlI << BslI
      << BsmAI << BsiYI
<< MnlI      << HphI      << HinfI<< Alw26I<< AciI << MseI
CTTGACGGTG AGGTTCTTTC ACTTGACAAA CTAAAGAGTC TCTTATCCCT GCGGGAGGTT AAGACTATAA
      4840      4850      4860      4870      4880      4890      4900

      << AluI      << NdeI
AAGTGTTAC AACTGTGGAC AACACTAATC TCCACACACA GCTTGTGGAT ATGTCTATGA CATATGGACA
      4910      4920      4930      4940      4950      4960      4970

      << SinI
      << Sau96I
      << NspIV
      << NspHII
      << Eco47I
      << Cfr13I
      << BsiZI
      << Bme18I
      << AvaII
      << AsuI
      << MaeIII << Tru9I << MnlI
      << FokI << MseI << BspHI
GCAGTTTGGT CCAACATACT TGGATGGTGC TGATGTTACA AAAATTAAAC CTCATGTAAA TCATGAGGGT
      4980      4990      5000      5010      5020      5030      5040

      << RsaI
      << RmaI
      << MaeI
      << Csp6I
      << AfaI
      << RsaI
      << MaeI
      << MaeII << HindIII << ScaI
      << Eco105I << RsaI
      << BsaAI << AluI << Csp6I
      << AfaI
AAGACTTTCT TTGTACTACC TAGTGATGAC ACACTACGTA GTGAAGCTTT CGAGTACTAC CATACTCTTG
      5050      5060      5070      5080      5090      5100      5110

      << RsaI
      << NspI
      << NspHI
      << NlaIII
      << Csp6I << Tru9I
      << AflIII << MseI
      << AfaI << DraI
      << MnlI >
      << BslI <<
      << BsiYI <<
ATGAGAGTTT TCTTGTTAGG TACATGTCTG CTTTAAACCA CACAAAGAAA TGGAAATTTC CTCAGTTGG
      5120      5130      5140      5150      5160      5170      5180

      << Tru9I << Tru9I
      << MseI << MseI
      << MunI
      << RmaI
      << MaeI
      << AluI >
TGGTTTAACT TCAATPAAAT GGGCTGATAA CAATTGTTAT TTGTCTAGTG TTTTATTAGC ACTTCAACAG
      5190      5200      5210      5220      5230      5240      5250

      << SfaNI
      << SduI
      << NspII
      << Eco24I
      << Bsp1286I
      << BmyI
      << BbvI Fnu4HI <<
      << HphI >
      << MnlI
      << BanII << BspWI

```

FIGURE 13.13

```

CTTGAAGTCA AATTCAATGC ACCAGCACTT CAAGAGGCTT ATTATAGAGC CCGTGCTGGT GATGCTGCTA
5260          5270          5280          5290          5300          5310          5320

>< VneI
>< SnoI
    >< SduI
    >< NspII
    >< HgiAI
    >< Bsp1286I
    >< BmyI
>< ApaLI
>< Alw44I
    >< Alw21I
    >< AluI
    MboII ><
    >< HphI
ACTTTTGTGC ACTCATACTC GCTTACAGTA ATAAAACTGT TGGCGAGCTT GGTGATGTCA GAGAAACTAT
5330          5340          5350          5360          5370          5380          5390

    > < SphI
    > < PaeI
    > < NspI
    > < NspHI >< TfiI
    >< Tru9I
    >< SfcI > < NlaIII>< HinfI
    >< MseI
GACCCATCTT CTACAGCATG CTAATTTGGA ATCTGCAAAG CGAGTTCTTA ATGTGGTGTG TAAACATTGT
5400          5410          5420          5430          5440          5450          5460

    >< RsaI
    >< Tru9I
    > < Csp6I
    Esp4I >
    >< MseI
    >< AluI
    >< AfaI
    AflIII >
GGTCAGAAAA CTACTACCTT AACGGGTGTA GAAGCTGTGA TGTATATGGG TACTCTATCT TATGATAATC
5470          5480          5490          5500          5510          5520          5530

    >< RsaI
    >< MboII
    >< RmaI HinfI ><
    >< Csp6I
>< Tru9I
    >< SfaNI
    >< MaeI >< BbsI
>< MseI
    >< NlaIII
    >< AfaI
TTAAGACAGG TGTTTCCATT CCATGTGTGT GTGGTCGTGA TGCTACACAA TATCTAGTAC AACAAGAGTC
5540          5550          5560          5570          5580          5590          5600

    >< RsaI
    >< PleI
    > < DdeI
    >< Csp6I
    >< BsgI
    >< BspWI >< BspMI
    >< AfaI
TTCTTTTGTG ATGATGTCTG CACCACCTGC TGAGTATAAA TTACAGCAAG GTACATTCTT ATGTGCGAAT
5610          5620          5630          5640          5650          5660          5670

    >< RsaI
    >< Eco31I
    >< DdeI
    >< BsmAI
    >< Csp6I
    >< BsaI
    MnII ><
    >< AfaI >< BsrI
    >< Alw26I
    >< HphI >
GAGTACACTG GTAACATCA GTGTGGTCAT TACACTCATA TAACTGCTAA GGAGACCCTC TATCGTATTG
5680          5690          5700          5710          5720          5730          5740

    >< SstI
    >< SduI
    >< SacI
    >< NspII
    >< HgiAI
    >< Eco24I
    >< Ecl136II
    >< Bsp1286I
    >< BmyI

    >< SinI
    >< Sau96I
    >< NspIV
    >< NspHII
    > < RsaI
    >< MaeIII
    >< Eco47I
    >< Cfr13I
    >< BsiZI
    >< Bme18I

```

FIGURE 13. 14


```

    >< BanII
    >< Alw21I
    >< AluI
ACGGAGCTCA CCTTACAAAG ATGTCAGAGT ACAAAGGACC AGTGACTGAT GTTTTCTACA AGGAAACATC
    5750      5760      5770      5780      5790      5800      5810

    >< AvaII
    >< Csp6I>< AsuI
    > < AfaI >< BsrI>< AlwNI
    >< TthHB8I
    >< TaqI >< MaeIII
TTACTACTACA ACCATCAAGC CTGTGTCGTA TAAACTCGAT GGAGTTACTT ACACAGAGAT TGAACCAAAA
    5820      5830      5840      5850      5860      5870      5880

    >< RsaI
    >< Csp6I
    >< SfcI >< BbvI
    >< FokI
    >< Fnu4HI
    >< AfaI
TTGGATGGGT ATTATAAAAA GGATAATGCT TACTATACAG AGCAGCCTAT AGACCTTGTA CCAACTCAAC
    5890      5900      5910      5920      5930      5940      5950

    >< Tru9I ><
    >< SwaI ><
    >< MseI ><
    > < NspI
    > < NspHI
    > < NlaIII
    >< AflIII
    >< BsaBI ><
CATTACCAAA TGCGAGTTTT GATAATTTCA AACTCACATG TTCTAACACA AAATTTGCTG ATGATTTAAA
    5960      5970      5980      5990      6000      6010      6020

    >< MboII
    >< AluI >< AluI>< MaeIII
TCAAATGACA GGCTTCACAA AGCCAGCTTC ACGAGAGCTA TCTGTCACAT TCTTCCCAGA CTTGAATGGC
    6030      6040      6050      6060      6070      6080      6090

    >< SfcI
GATGTAGTGG CTATTGACTA TAGACACTAT TCAGCGAGTT TCAAGAAAGG TGCTAAATTA CTGCATAAGC
    6100      6110      6120      6130      6140      6150      6160

    >< Tru9I
    >< ScrFI
    >< MvaI
    >< MseI
    >< EcoRII
    >< Ecl136I
    >< DsaV
    >< BstOI
    >< BstNI
    >< BsiLI
    >< MunI
    >< BstXI
    >< ApyI
    >< MaeII
    >< DraIII
    >< BstXI
CAATTGTTG GCACATTAAC CAGGCTACAA CCAAGACAAC GTTCAAACCA AACACTTGGT GTTTACGTTG
    6170      6180      6190      6200      6210      6220      6230

    > < RsaI
    >< Csp6I
    > < AfaI>< BsrI
    >< MboII ><
    >< BbsI
TCTTTGGAGT ACAAAGCCAG TAGATACTTC AAATTCATTT GAAGTTCTGG CAGTAGAAGA CACACAAGGA
    6240      6250      6260      6270      6280      6290      6300

    >< HindII
    >< HincII
    >< MboII
    >< MnlI
    >< Eco57I
ATGGACAATC TTGCTTGTA AAGTCAACAA CCCACCTCTG AAGAAGTAGT GGAAAATCCT ACCATACAGA
    6310      6320      6330      6340      6350      6360      6370

```

FIGURE 13.15

```

                >< MaeIII
                >< MaeII
AGGAAGTCAT AGAGTGTGAC GTGAAAAC TA CCGAAGTTGT AGGCAATGTC ATACTTAAAC CATCAGATGA
    6380         6390         6400         6410         6420         6430         6440

                >< XhoII
                >< Sau3AI
                >< NlaIII
                >< NdeII
                >< MflI
                >< MboI
                >< DpnII
                >< DpnI
                >< BstYI
                >< BspAI
                >< BspHI >< Bsp143I>< Fnu4HI
                >< MnlI >< BbvI >< AlwI
<< Tru9I
<< MseI
    > < MaeIII
AGGTGTTAAA GTAACACAAG AGTTAGGTCA TGAGGATCTT ATGGCTGCTT ATGTGGAAAA CACAAGCATT
    6450         6460         6470         6480         6490         6500         6510

                >< SauI
                >< RmaI
                >< MstII
                >< MaeI
                >< Eco81I
                >< DdeI
                >< CvnI
                >< Bsu36I
                >< Bse21I
                >< BfrI> < Tru9I
                >< AxyI> < MseI>< MunI
                >< NlaIII
<< Tru9I
<< MseI
    >< AluI
    >< AocI >< DraI
    >< BbvI Fnu4HI ><
ACCATTAAGA AACCTAATGA GCTTTCAC TA GCCTTAGGTT TAAAAACAAT TGCCACTCAT GGTATTGCTG
    6520         6530         6540         6550         6560         6570         6580

                >< VspI
                >< StyI
                >< Tru9I
                >< EcoT14I
                >< MseI
                >< Eco130I
                >< AsnI
                >< Bst1I
                >< AseI
                >< BsaJI
                > < DdeI
                >< BslI
                >< BsiYI
                > < BfrI
                >< Fnu4HI
CAATTAATAG TGTTCCTTGG AGTAAAATTT TGGCTTATGT CAAACCATTC TTAGGACAAG CAGCAATTAC
    6590         6600         6610         6620         6630         6640         6650

                >< HinP1I
                >< Hin6I
                >< HhaI
                >< DdeI
                >< BbvI
                >< CfoI
                >< AflIII
                >< Tru9I
                >< MaeII>< MseI
                >< DraIII
                >< AcaI>> MseI
AACATCAAAT TGC GCTAAGA GATTAGCACA ACGTGTGTTT AACAAATTATA TGCCTTATGT GTTTACATTA
    6660         6670         6680         6690         6700         6710         6720

                >< RsaI
                >< Csp6I
                >< MunI >< AfaI
                > < RsaI>< XbaI
                >< Csp6I >< RmaI
                > < AfaI >< MaeI
                >< AluI
TTGTTCCAAT TGTGTACTTT TACTAAAAGT ACCAATTCTA GAATTAGAGC TTCACTACCT ACAACTATTG
    6730         6740         6750         6760         6770         6780         6790

                >< VspI
                >< Tru9I
                >< NaeI
                >< MspI
                >< MseI

```

FIGURE 13. 16

```

                >> HpaII
                >> HapII
                >> Cfr10I >> FokI
                >> AsnI
                >> Tru9I
                >> MseI
                >> SfaNI
                >> AseI>> HphI>> MaeIII
CTAAAAATAG TGTTAAGAGT GTTGCTAAAT TATGTTTGGA TGCCGGCATT AATTATGTGA AGTCACCCAA
    6800         6810         6820         6830         6840         6850         6860

                >> Tru9I    >> DdeI    MaeIII >
                >> MseI    >> BfrI    >> BbvI
ATTTTCTAAA TTGTTCAAAA TCGCTATGTG GCTATTGTTG TTAAGTATTT GCTTAGGTTT TCTAATCTGT
    6870         6880         6890         6900         6910         6920         6930

                >> SduI
                >> NspII
                >> HgiAI
                >> Bsp1286I
                > < RsaI
                >> Csp6I
                >> Fnu4HI > < AfaI
                >> Alw21I
GTAAGTCTG CTCTTGGTGT ACTCTTATCT AATTTTGGTG CTCCTTCTTA TTGTAATGGC GTTAGAGAAT
    6940         6950         6960         6970         6980         6990         7000

                Tru9I >>
                MseI >>
                >> Tru9I    > < MaeIII
                >> MseI    >> MaeII
                >> Fnu4HI
                >> BbvI >
TGTATCTTAA TTCGTCTAAC GTTACTACTA TGGATTCTCG TGAAGTTCTT TTCCTTGCA GCATTTGTTT
    7010         7020         7030         7040         7050         7060         7070

                > < TfiI
                >> MamI
                > < HinfI
                >> BsiBI
                >> XmnI>> MaeIII
                >> Asp700I
                >> AluI
                >> AfaI >>
                >> PleI>> HinfI
                >> BsaBI >> AluI
                >> XmnI>> MaeIII
                >> Asp700I
                >> AfaI >>
AAGTGGATTA GACTCCCTTG ATTCTTATCC AGCTCTTGAA ACCATTGAGG TGACGATTTT ATCGTACAAG
    7080         7090         7100         7110         7120         7130         7140

                >> Pali
                >> NspBII
                >> HaeIII
                >> GdiII
                >> Fnu4HI
                >> EaeI
                >> DdeI
                >> BsuRI
                >> BshI >> BslI
                >> MaeI
                >> AciI>> BsiYI
CTAGACTTGA CAATTTTAGG TCTGGCCGCT GAGTGGGTTT TGGCATATAT GTTGTTTACA AAATTCTTTT
    7150         7160         7170         7180         7190         7200         7210

                >> BspMI
                >> AluI
                >> RmaI
                >> MaeI
ATTTATTAGG TCTTTCAGCT ATAATGCAGG TGTTCTTTGG CTATTTTGCT AGTCATTTCA TCAGCAATTC
    7220         7230         7240         7250         7260         7270         7280

                RsaI >>
                >> MboII
                >> NlaIV
                >> Eco64I
                > < RsaI >> BscBI
                >> Csp6I >> BanI
                > < AfaI>> AccBI
                > < NlaIII
                >> RmaI
                >> MaeI
                >> BsiBI >>
                >> BsaBI >>
                >> AfaI >>

```

FIGURE 13.17


```

                >> FokI
                    >> BsmAI
                >> MnlI                    >> Alw26I    >> AciI
CCTCTACTTT GACAAGGCTG GTCAAAAGAC CTATGAGAGA CATCCGCTCT CCCATTTTGT CAATTTAGAC
    7710         7720         7730         7740         7750         7760         7770

                    >> VspI
                    >> Tru9I
                    >> MseI
                    >> AsnI
                > < AluI                    >> AseI                    >> BcgI/a
AATTTGAGAG CTAACAACAC TAAAGGTTCA CTGCCTATTA ATGTCATAGT TTTTGATGGC AAGTCCAAAT
    7780         7790         7800         7810         7820         7830         7840

                    >> SfcI    >> PvuII
                    >> RsaI    >> Psp5I
                >> PleI                    >> Csp6I    >> NspBII
                >> HinfI  >> DdeI    >> BcgI    >> AfaI    >> AluI
GCGACGAGTC TGCTTCTAAG TCTGCTTCTG TGTACTACAG TCAGCTGATG TGCCAACCTA TTCTGTTGCT
    7850         7860         7870         7880         7890         7900         7910

                                                    TthHB8I ><
                                                    TaqI ><
                                                    SalI ><
                                                    RtrI ><
                    >> ScaI                    HindII >
                    >> RsaI                    >> Tru9I    HincII >
                >> Csp6I                    >> SfaNI >< Eco57I
                >> AluI                    >> MaeII    >> AfaI    >> MseI    AccI ><
TGACCAAGCT CTTGTATCAG ACGTTGGAGA TAGTACTGAA GTTTCCGTTA AGATGTTTGA TGCTTATGTC
    7920         7930         7940         7950         7960         7970         7980

                    >> Tru9I
                    >> MseI
                > < Esp4I                    >> SfcI
                > < AflII                    >> BspWI  >> AluI
GACACCTTTT CAGCAACTTT TAGTGTTTCT ATGGAAAAAC TTAAGGCACT TGTGCTACA GCTCACAGCG
    7990         8000         8010         8020         8030         8040         8050

                                                    >> PvuII
                                                    >> Psp5I
                                                    >> NspBII
                                                    >> Fnu4HI
                >> AluI                    >> BbvI    >> AluI
AGTTAGCAA GGGTGTAGCT TTAGATGGTG TCCTTTCTAC ATTCGTGTCA GCTGCCCGAC AAGGTGTTGT
    8060         8070         8080         8090         8100         8110         8120

                                                    MaeIII ><
                >> HindII                    >> BsmAI    >> DdeI
                >> HincII                    >> FokI >< Alw26I >> BfrI
TGATACCGAT GTTGACACAA AGGATGTTAT TGAATGTCTC AAAC TTTCAC ATCACTCTGA CTTAGAAGTG
    8130         8140         8150         8160         8170         8180         8190

                                                    >> XhoII
                                                    Sau3AI ><
                                                    >> NdeII
                                                    >> MflI
                                                    >> MboI
                >> NlaIII >< HgaI
                >> HinfI >< DpnII
                                                    DpnI ><
    
```

FIGURE 13.19

```

Bsp143I ><
>< BsaHI >< BstYI
>< BbiII >< BspAI
>< MaeIII >< HphI >< NlaIII >< AcyI >< BglII
ACAGGTGACA GTTGTAACAA TTTCATGCTC ACCTATAATA AGGTTGAAAA CATGACGCCC AGAGATCTTG
8200 8210 8220 8230 8240 8250 8260

>< NspI
>< NspHI
>< NlaIII
>< HinPII
>< Hin6I
>< HhaI
>< CfoI >< BspWI >< MaeIII
GCGCATGTAT TGACTGTAAT GCAAGGCATA TCAATGCCCA AGTAGCAAAA AGTCACAATG TTTCACTCAT
8270 8280 8290 8300 8310 8320 8330

>< NspI
>< NspHI >< PvuII
>< NlaIII >< Psp5I
>< Eam1105I >< NspBII
>< BbvI >< Fnu4HI
>< AflIII >< AluI >< BbvI >< Fnu4HI
CTGGAATGTA AAAGACTACA TGTCTTTATC TGAACAGCTG CGTAAACAAA TTCGTAGTGC TGCCAAGAAG
8340 8350 8360 8370 8380 8390 8400

>< MboII >< RmaI
>< MaeI >< Eam1105I
AACAAACATAC CTTTtagact AACTTGTGCT ACAACTAGAC AGGTTGTCAA TGTCATAACT ACTAAAATCT
8410 8420 8430 8440 8450 8460 8470

>< Tru9I
>< Pali
>< MseI
>< HaeIII
>< ScaI >< Esp4I
>< RsaI >< Tru9I >< BsuRI
>< Csp6I >< MseI >< BshI
>< AfaI >< DraI >< AflII >< BbvI
CACTCAAGGG TGGTAAGATT GTTAGTACTT GTTTTAAACT TATGCTTAAG GCCACATTAT TGTGCGTTCT
8480 8490 8500 8510 8520 8530 8540

>< RsaI
>< Csp6I
>< BsrI >< NlaIII
>< Fnu4HI >< AfaI >< MaeIII
TGCTGCATTG GTTTGTATA TCGTTATGCC AGTACATACA TTGTCAATCC ATGATGGTTA CACAAATGAA
8550 8560 8570 8580 8590 8600 8610

>< MaeIII
>< MaeIII
>< MaeIII >< FokI
ATCATTGGTT ACAAAGCCAT TCAGGATGGT GTCACCTCGTG ACATCATTC TACTGATGAT TGTTTTGCAA
8620 8630 8640 8650 8660 8670 8680

>< NspI SfcI >
>< NspHI >< NlaIII Fnu4HI >>
>< NlaIII >< HgaI >< BstXI BbvI >>
ATAAACATGC TGGTTTTGAC GCATGGTTTA GCCAGCGTGG TGGTTCATAC AAAAATGACA AAAGCTGCC
8690 8700 8710 8720 8730 8740 8750

```

FIGURE 13. 20

```

                                >< ScrFI
                                >< ScrFI    >< RsaI
                                >< MvaI    >< MspI
                                >< EcoRII  >< HpaII
                                >< Ecl136I>< NciI
                                >< DsaV    >< HapII
                                >< BstOI>< DsaV
                                >< BstNI    >< Csp6I
                                >< BsiLI   >< BcnIDdeI ><
                                >< ApyI    >< AfaI
                                >< Fnu4HI
                                >< AluI
TGTAGTAGCT GCTATCATT CAAGAGAGAT TGGTTTCATA GTGCCCTGGCT TACCGGGTAC TGTGCTGAGA
8760          8770          8780          8790          8800          8810          8820

                                > < MaeIII   >< HphI
                                >< MnlI          >< BspWI
GCAATCAATG GTGACTTCTT GCATTTTCTA CCTCGTGTTC TTAGTGCTGT TGGCAACATT TGCTACACAC
8830          8840          8850          8860          8870          8880          8890

                                Tru9I >
                                SfaNI ><
                                >< RsaI
                                MseI >
                                >< BspWI          >< Fnu4HI >< Csp6I
                                >< BbvI>< MnlI          >< DdeI >< AfaI
CTTCCAAACT CATTGAGTAT AGTGATTTTG CTACCTCTGC TTGCGTTCTT GCTGCTGAGT GTACAATTTT
8900          8910          8920          8930          8940          8950          8960

                                > < RmaI
                                >< MnlI
                                > < MaeI
TAAGGATGCT ATGGGCAAAC CTGTGCCATA TTGTATGAC ACTAATTTGC TAGAGGGTTC TATTTCTTAT
8970          8980          8990          9000          9010          9020          9030

                                ScrFI >
                                MvaI >
                                MnlI ><
                                EcoRII ><
                                Ecl136I >
                                DsaV ><
                                BstOI >
                                >< NlaIV          BstNI >
                                >< FokI          BsiLI >
                                >< BscBI          ApyI >
                                >< AluI
AGTGAGCTTC GTCCAGACAC TCGTTATGTG CTTATGGATG GTTCCATCAT ACAGTTTCTT AACACTTACC
9040          9050          9060          9070          9080          9090          9100

                                >< RsaI
                                >< SfcI          >< NspI
                                >< ScaI          >< NspHI
                                >< SfaNI          >< RsaI          >< NlaIII
                                > < MaeIII   >< Csp6I          >< NlaIII
                                >< GsuI          >< AfaI          >< Csp6I
                                >< BpmI          >< DdeI          >< AccI   >< AfaI
TGGAGGGTTC TGTTAGAGTA GTAACAACCTT TTGATGCTGA GTACTGTAGA CATGGTACAT GCGAAAGGTC
9110          9120          9130          9140          9150          9160          9170

                                >< SstI
                                >< SduI
                                >< SacI
                                NspII ><
                                HgiAI ><
                                Eco24I ><
                                Bsp1286I ><

```

FIGURE 13.21

```

Ecl136II ><>< BmyI
BanII ><
>< Tru9I Alw21I ><
>< BsrI >< MseI >< AluI
AGAAGTAGGT ATTTGCCTAT CTACCACTGG TAGATGGGTT CTTAATAATG AGCATTACAG AGCTCTATCA
9180 9190 9200 9210 9220 9230 9240

>< TfiI
>< SfaNI >< HinfI >< AluI >< MnlI
GGAGTTTTCT GTGGTGTGA TGGATGAAT CTCATAGCTA ACATCTTTAC TCCTCTTG TG CAACCTGTGG
9250 9260 9270 9280 9290 9300 9310

>< MaeIII
HphI ><
>< Eco57I >< BbvI Fnu4HI ><
GTGCTTTAGA TGTGTCTGCT TCAGTAGTGG CTGGTGGTAT TATTGCCATA TTGGTACTT GTGCTGCCTA
9320 9330 9340 9350 9360 9370 9380

>< RsaI
>< Csp6I >< NlaIII
>< MaeII >< BbvI >< Fnu4HI
>< AflIII >< AfaI>< HphI >< BspWI
CTACTTTATG AAATTCAGAC GTGTTTTTGG TGAGTACAAC CATGTTGTTG CTGCTAATGC ACTTTTGT TT
9390 9400 9410 9420 9430 9440 9450

>< RsaI
>< NlaIV
>< KpnI
>< Eco64I >< ScrFI
>< Csp6I >< NciI
>< BscBI >< MspI
>< Asp718 >< HpaII
>< BanI >< AluI >< HinfI
>< AfaI >< HapII >< PleI
>< AccB1I >< BcnI >< DdeI
>< Acc65I >< AluI>< DsaV >< AccI
TTGATGTCTT TCACTATACT CTGTCTGGTA CCAGCTTACA GCTTTCTGCC GGGAGTCTAC TCAGTCTTTT
9460 9470 9480 9490 9500 9510 9520

>< RsaI
>< Csp6I
>< AfaI >< HphI >< HphI NlaIII ><
ACTTGACTT GACATCTAT TTCACCAATG ATGTTTCATT CTTGGCTCAC CTTCAATGGT TTGCCATGTT
9530 9540 9550 9560 9570 9580 9590

TTCTCCTATT GTGCCTTTTT GGATAACAGC AATCTATGTA TTCTGTATTT CTCTGAAGCA CTGCCATTGG
9600 9610 9620 9630 9640 9650 9660

>< TthHB8I
>< RsaI
>< MnlI
>< MnlI
>< Tru9I >< Csp6I
>< Tru9I >< PleI >< BcgI/a >< TaqI
>< MseI >< DdeI >< NlaIII >< BbvI
>< Eco57I >< BfrI >< HinfI >< MseI >< MaeIII >< AfaI Fnu4HI ><
TTCTTTAACA ACTATCTTAG GAAAAGAGTC ATGTTTAATG GAGTTACATT TAGTACCTTC GAGGAGGCTG
9670 9680 9690 9700 9710 9720 9730

>< RsaI
>< Csp6I
>< BcgI >< RsaI >< Csp6I >< BsmAI

```

FIGURE 13.22


```

    >> AfaI
CTTTGTGTAC CTTTTTGCTC AACAAGGAAA TGTACCTAAA ATTGCGTAGC GAGACACTGT TGCCACTTAC
  9740          9750          9760          9770          9780          9790          9800

    >> AfaI    >> Alw26I
    >> NlaIV
    >> RsaI
    >> DdeI
    >> Csp6I
    >> BscBI
    >> AfaI    >> BfrI    AluI >>
ACAGTATAAC AGGTATCTTG CTCTATATAA CAAGTACAAG TATTTTCAGTG GAGCCTTAGA TACTACCAGC
  9810          9820          9830          9840          9850          9860          9870

    >> Fnu4HI
    >> DdeI
    >> Fnu4HI    >> BfrI
    >> BbvI    >> AluI    >> BbvI
    >> DdeI    >> AlwNI
TATCGTGAAG CAGCTTGCTG CCACTTAGCA AAGGCTCTAA ATGACTTTAG CAACTCAGGT GCTGATGTTT
  9880          9890          9900          9910          9920          9930          9940

    >> SfcI
    >> PstI
    >> BsmI
    >> BscCI
TCTACCAACC ACCACAGACA TCAATCACTT CTGCTGTTCT GCAGAGTGGT TTTAGGAAAA TGGCATTCCC
  9950          9960          9970          9980          9990          10000        10010

    >> RsaI
    >> NlaIII
    >> MaeIII
    >> Csp6I
    >> AfaI
    >> Tru9I
    >> MseI
GTCAGGCAAA GTTGAAGGGT GCATGGTACA AGTAACCTGT GGAACTACAA CTCTTAATGG ATTGTGGTTG
  10020        10030        10040        10050        10060        10070        10080

    XhoII >>
    Sau3AI >>
    >> Tru9I    NdeII >>
    >> NspI    MflI >>
    >> NspHI   MboI >>
    >> NlaIII  DpnII >>
    >> MseI    BstYI >>
    >> MboII  BspAI >>
    >> AccI    >> AflIII
    >> BbsI    BglII >>
GATGACACAG TATACTGTCC AAGACATGTC ATTTGCACAG CAGAAGACAT GCTTAATCCT AACTATGAAG
  10090        10100        10110        10120        10130        10140        10150

    Pali >
    MscI >
    HaeIII >
    EaeI >>
    BsuRI >
    BshI >
    Bali >
    >> DpnI >> MboII
    >> Bsp143I
    >> AluI
ATCTGCTCAT TCGCAAATCC AACCATAGCT TTCTTGTTCA GGCTGGCAAT GTTCAACTTC GTGTTATTGG
  10160        10170        10180        10190        10200        10210        10220

    >> DdeI > < Tru9I
    >> BfrI > < MseI
    >> DdeI
CCATTCTATG CAAAATTGTC TGCTTAGGCT TAAAGTTGAT ACTTCTAACC CTAAGACACC CAAGTATAAA
  10230        10240        10250        10260        10270        10280        10290

    >> ScrFI
    >> MvaI
    >> EcoRII
    >> Ecl136I
    >> SphI

```

FIGURE 13.23


```

                >< EcoO109I
                >< Eco47I
    >< Sau3AI      >< DraII
    >< NdeII       >< Cfr13I
    >< MboI        >< BsiZI
    >< DpnII>< NlaIII >< BscBI
    >< DpnI >< HindII >< Bmel8I
    >< BspAI >< HincII >< AvaII
    >< Bsp143I    >< AsuI      >< MnlI
ACACAAGATC ATGTTGACAT ATTGGGACCT CTTTCTGCTC AAACAGGAAT TGCCGTCTTA GATATGTGTG
10720      10730      10740      10750      10760      10770      10780

                >< StyI
                >< RsaI
                >< EcoT14I
                >< Eco130I
    >< Fnu4HI      >< SfcI
    >< BbvI        >< Fnu4HI
    >< BbvI        >< AluI >< PstI
    >< BbvI        >< AluI >< PstI
CTGCTTTGAA AGAGCTGCTG CAGAATGGTA TGAATGGTGC TACTATCCTT GGTAGCACTA TTTTAGAAGA
10790      10800      10810      10820      10830      10840      10850

                >< StyI
                >< EcoT14I
                >< Eco130I
                >< BssT1I
    >< MboII      >< MaeIII>< BsaJI
TGAGTTTACA CCATTGATG TTGTTAGACA ATGCTCTGGT GTTACCTTCC AAGGTAAGTT CAAGAAAATT
10860      10870      10880      10890      10900      10910      10920

    >< SfaNI
    >< SduI
    >< NspII
    >< Tru9I>< Bsp1286I
    >< MseI >< BmyI
    >< MseI >< BmyI
GTTAAGGGCA CTCATCATTG GATGCTTTTA ACTTTCTTGA CATCACTATT GATTCTTGTT CAAAGTACAC
10930      10940      10950      10960      10970      10980      10990

                >< XmnI
                >< BsmI
                >< BscCI
    >< MaeIII    >< Asp700I
    >< MaeIII    >< Asp700I
AGTGGTCACT GTTTTCTTTT GTTTACGAGA ATGCTTTCTT GCCATTTACT CTTGGTATTA TGGCAATTGC
11000      11010      11020      11030      11040      11050      11060

    >< NspI
    >< NspHI
    >< NlaIII
    >< BspWI >< Fnu4HI>< BspWI >< BscCI
    >< BspWI >< Fnu4HI>< BspWI >< BscCI
TGCATGTGCT ATGCTGCTTG TTAAGCATAA GCACGCATTC TTGTGCTTGT TTCTGTACC TTCTCTTGCA
11070      11080      11090      11100      11110      11120      11130

                >< SfaNI
                >< RmaI
    >< NspI
    >< NlaIII
    >< NheI
    >< Tru9I
    >< BspWI >< MseI >< AccI>< NspHI>< AluI
    >< BspWI >< MseI >< AccI>< NspHI>< AluI
ACAGTTGCTT ACTTTAATAT GGTCTACATG CCTGCTAGCT GGGTGATGCG TATCATGACA TGGCTTGAAT
11140      11150      11160      11170      11180      11190      11200

                >< MamI
                >< HphI
                >< BspHI
                >< BsiBI
                >< NlaIII
                >< BsaBI >< NlaIII

```

FIGURE 13.25

```

                >< Tru9I
                >< MseI
    > < RmaI          > < Esp4I
    > < MaeI          >< Eco57I
                >< AluI          > < AflIII          >< AluI
TGGCTGACAC TAGCTTGTCT GGTATAGGC TTAAGGATTG TGTTATGTAT GCTTCAGCTT TAGTTTTGCT
    11210      11220      11230      11240      11250      11260      11270
                >< RmaI
                >< MaeII
                >< MaeI
    > < NlaIII      >< SfaNI      >< Fnu4HI
    >< BspHI >< AluI >< BbvI          >< AflIII
TATCTCATG ACAGCTCGCA CTGTTTATGA TGATGCTGCT AGACGTGTTT GGACACTGAT GAATGTCATT
    11280      11290      11300      11310      11320      11330      11340
                >< Sau96I
                >< Pali
                >< NspIV
                >< NlaIII
                >< HaeIII
                >< Sau3AI
                >< NdeII
                >< Cfr13I
                >< MboI
                >< BsuRI
                >< DpnII
                >< BsiZI
                >< DpnI
                >< BshI
                >< Bsp143I
                > < BfrI
                >< AccI
                >< BspAI>< AluI
                >< AsuI
ACACTTGTTT ACAAAGTCTA CTATGGTAAT GCTTTAGATC AAGCTATFTC CATGTGGGCC TTAGTTATTT
    11350      11360      11370      11380      11390      11400      11410
                >< RmaI
                >< NlaIII
                >< MaeI>< SfcI
    >< MaeIII      >< MnlI      >< MaeIII      >< AluI>< AluI
CTGTAACCTC TAACTATTCT GGTGTCGTTA CGACTATCAT GTTTTAGCT AGAGCTATAG TGTTTGTGTG
    11420      11430      11440      11450      11460      11470      11480
                >< BsrI
                >< NlaIII BfrI >
                DdeI >
TGTTGAGTAT TACCCATTGT TATTTATTAC TGGCAACACC TTACAGTGTA TCATGCTTGT TTATTGTTTC
    11490      11500      11510      11520      11530      11540      11550
                >< Pali
                >< HaeIII
                >< Fnu4HI
                >< BsuRI
    >< BbvI      >< Fnu4HI      >< BspWI
    >< BbvI      >< BspWI      >< BshI      >< Eco57I >< MaeIII
TTAGGCTATT GTTGCTGCTG CTTACTTGGC CTTTTCTGTT TACTCAACCG TTTACTTCAGG CTTACTCTTG
    11560      11570      11580      11590      11600      11610      11620
                >< ScrFI
                >< MvaI
                >< EcoRII
                >< Ecl136I
                >< DsaV
                >< BstOI
                >< BstNI
                >< BsiLI
                >< Eco31I
                >< BsmAI
                > < BsaJI
                >< BsaI
                >< BsaJI

```

FIGURE 13.26

```

                >< DrdI >< Alw26I
GTGTTTATGA C TACTTGGTC TCTACACAAG AATTTAGGTA TATGAACTCC >< ApyI DdeI ><
11630 11640 11650 11660 11670 11680 11690

                >< Tru9I
                >< MseI
>< SfaNI >< HindIII> < Tru9I
>< MnlI >< AluI > < MseI > < MnlI >< NlaIII
GAGTAGTATT GATGCTTTCA AGCTTAACAT TAAGTTGTTG GGTATTGGAG GTAAACCATG TATCAAGGTT
11700 11710 11720 11730 11740 11750 11760

                >< VneI
                >< SnoI
                >< SduI
                >< NspII
                >< HgiAI
                >< Bsp1286I
                >< BmyI >< RsaI
                >< RsaI >< ApaLI >< MboII
                >< Csp6I >< Alw44I >< Csp6I DdeI >
                >< AfaI >< MaeII >< Alw21I >< AfaI BfrI >
GCTACTGTAC AGTCTAAAAT GTCTGACGTA AAGTGACAT CTGTGGTACT GCTCTCGGTT CTTCAACAAC
11770 11780 11790 11800 11810 11820 11830

                >< NspII> < RsaI
                >< DraIII
                >< SduI>< Csp6I
                >< Bsp1286I
                >< BmyI > < AfaI >< MboII
TTAGAGTAGA GTCATCTTCT AAATTGTGGG CACAATGTGT ACAACTCCAC AATGATATTC TTCTTGCAAA
11840 11850 11860 11870 11880 11890 11900

                >< TthHB8I
                >< TaqI
                >< HindIII >< MboII SfcI ><
                >< AluI > < Eco57I >< NlaIII
AGACACAAC T GAAGCTTTTCG AGAAGATGGT TTCTCTTTTG TCTGTTTTGC TATCCATGCA GGGTGCTGTA
11910 11920 11930 11940 11950 11960 11970

                >< VspI
                >< Tru9I >< Ksp632I
                >< MseI >< TthHB8I >< EarI
                >< AsnI >< TaqI >< MboII >< Eam1104I
                >< AseI>< MnlI >< BcgI/a >< Eco57I >< Eco57I >< BcgI
GACATTAATA GGTGTGCGA GGAAATGCTC GATAACCGTG C TACTCTTCA GGCTATTGCT TCAGAATTTA
11980 11990 12000 12010 12020 12030 12040

                >< StuI
                >< ScrFI
                >< Pali
                >< MvaI>< HaeIII
                >< EcoRII>< Eco147I
                >< Ecl136I
                >< DsaV >< BsuRI
                >< BstOI
                >< BstNI
                >< BspWI
                >< BsiLI
                >< Fnu4HI >< BsaJI >< BshI TfiI ><
                >< NdeI >< BspWI>< MnlI >< BglI >< SfcI HinfI ><
                >< AciI >< ApyI>< AatI >< AluI
    
```

FIGURE 13. 27

```

GTTCTTTACC ATCATATGCC GCTTATGCCA CTGCCCAGGA GGCCTATGAG CAGGCTGTAG CTAATGGTGA
12050      12060      12070      12080      12090      12100      12110

    >> XmnI          >> Tru9I          >> SfaNI
    >> HphI          >> MseI          >> DdeI
    >> Asp700I      >> Eco57I
TTCTGAAGTC GTTCTCAAAA AGTTAAAGAA ATCTTTGAAT GTGGCTAAAT CTGAGTTTGA CCGTGATGCT
12120      12130      12140      12150      12160      12170      12180

                                XhoII >>
                                Sau3AI >>
                                NdeII >>
                                MnlI >
                                >> MnlI
                                >> MflI
                                >> MboI
                                DpnII >>
                                DpnI >>
                                DdeI >>
                                BstYI >>
                                >> BspWI          >> RsaIBspAI >>
                                >> BspAI          >> Csp6IBsp143I >>
                                >> Bsp143I        >> AfaIBglIII >>
>> NlaIII
GCCATGCAAC GCAAGTTGGA AAAGATGGCA GATCAGGCTA TGACCCAAAT GTACAAACAG GCAAGATCTG
12190      12200      12210      12220      12230      12240      12250

                                >> SpeI          >> Ksp632I > < HindIII
                                >> RmaI          >> DdeI >> SfaNI
                                >> MaeIII        >> MboII        >> Eam1104I >> BspWI
                                >> MaeI          >> BspWI        >> EarI >> BfrI >> AluI
AGGACAAGAG GGCAAAAGTA ACTAGTGCTA TGCAAACAAT GCTCTTCACT ATGCTTAGGA AGCTTGATAA
12260      12270      12280      12290      12300      12310      12320

                                >> ThaI
                                >> MvnI
                                >> HinPII
                                >> Hin6I
                                >> HhaI
                                >> CfoI
                                >> BstUI
                                >> Bsp50I
                                >> AccII
                                SfcI >>
>> Tru9I
>> MseI
TGATGCACTT AACACATTA TCAACAATGC GCGTGATGGT TGTGTTCCAC TCAACATCAT ACCATTGACT
12330      12340      12350      12360      12370      12380      12390

                                >> RsaI
                                >> NlaIV
                                >> Eco64I
                                >> Csp6I
                                >> BslI
                                >> BsiYI >> KpnI
                                >> BscBI
                                >> BanI
                                >> Asp718
                                >> AfaI
                                >> AccB1I          >> MaeIII
                                >> Acc65I          >> BsqI >>
ACAGCAGCCA AACTCATGGT TGTGTGCCCT GATTATGGTA CCTACAAGAA CACTTGTGAT GGTAACACCT
12400      12410      12420      12430      12440      12450      12460

                                >> Zsp2I
                                >> Ppu10I

```

FIGURE 13. 28

```

    >< NsiI
    >< Mph1103I
    >< NdeI>< EcoT22I
    >< AvaIII >< SfaNI
    >< SfaNI
    >< AciI
    DdeI ><
    BfrI ><
    TTACATATGC ATCTGCACTC TGGGAAATCC AGCAAGTTGT TGATGCGGAT AGCAAGATTG TTCAACTTAG
    12470 12480 12490 12500 12510 12520 12530

    >< PalI
    >< HaeIII >< MnlI >< DdeIDdeI ><
    >< BsuRI >< MaeIII >< BspWI
    >< Tru9I>< NlaIII
    >< MseI>< HphI > < XcmI>< BshI >< AluI BspWI ><
    TGAAATTAAC ATGGACAATT CACCAAATTT GGCTTGGCCT CTTATGTGTA CAGCTCTAAG AGCCAACTCA
    12540 12550 12560 12570 12580 12590 12600

    RsaI ><
    NlaIV ><
    KpnI ><
    >< Fnu4HI
    Eco64I ><
    Csp6I ><
    BscBI ><
    Asp718 ><
    AfaI ><
    >< AciI>< BanI
    AccB1I ><
    >< Tru9I
    >< PvuII
    >< Psp5I
    >< NspBII
    >< MseI >< HinfI >< PleI
    >< AluI > < SfcI >< DdeI>< BsrI >< PshAI Acc65I ><
    GCTGTAAAC TACAGAATAA TGAAGT GAGT CCAGTAGCAC TACGACAGAT GTCCTGTGCG GCTGGTACCA
    12610 12620 12630 12640 12650 12660 12670

    >< TthHB8I
    >< TaqI
    >< SfuI
    >< NspV
    >< MnlI
    >< LspI
    >< Csp45I
    >< BstBI
    >< Bsp119I
    >< BsiCI
    >< Bpu14I
    >< AsuII
    CACAACAGC TTGTACTGAT GACAATGCAC TTGCCTACTA TAACAATTCG AAGGGAGGTA GTTTGTGCT
    12680 12690 12700 12710 12720 12730 12740

    >< XhoII
    >< Sau3AI
    >< NdeII
    >< MflI
    >< MboI
    >< DpnII
    >< DpnI
    >< BstYI >< TfiI >< RsaI
    >< BspAI >< RmaI >< Csp6I
    >< Bsp143I >< HinfI >< Csp6I>< RsaI
    >< BglII >< MaeI >< DdeI >< AfaI>< AfaI
    GGCATTACTA TCAGACCACC AAGATCTCAA ATGGGCTAGA TTCCCTAAGA GTGATGGTAC AGGTACAATT
    12750 12760 12770 12780 12790 12800 12810

    >< Sau96I
    >< PssI
    >< Pali
    >< NspIV

```

FIGURE 13.29

```

                >< HaeIII
                >< Eco0109I
                >< DraII
                >< Cfr13I
                >< BsuRI
                >< NlaIV
                >< BsiZI
                >< BsrI
                >< BshI
                >< BscBI
                > < MaeIII
                >< AsuI
                >< RsaI
                >< Csp6I
                >< AfaI
TACACAGAAC TGGAACCACC TTGTAGGTT GTTACAGACA CACCAAAGG GCCTAAAGTG AAATACTTGT
12820      12830      12840      12850      12860      12870      12880

                >< SfcI
                > < MboII
                MaeII ><
                >< Fnu4HI >< RsaI
                >< Eco57I >< Csp6I
                > < BbsI
                >< Tru9I
                >< MseI >< MnlI
                >< BbvI
                >< AluI
                >< AfaI
ACTTCATCAA AGGCTTAAAC AACCTAAATA GAGGTATGGT GCTGGGCAGT TTAGCTGCTA CAGTACGTCT
12890      12900      12910      12920      12930      12940      12950

                >< RsaI
                >< SfcI >< Csp6I
                >< BspWI
                >< AfaI
                >< BspMI
                >< AccI ><
TCAGGCTGGA AATGCTACAG AAGTACCTGC CAATTCAACT GTGCTTTCCT TCTGTGCTTT TGCAGTAGAC
12960      12970      12980      12990      13000      13010      13020

                >< RmaI
                >< MnlI
                >< MaeI
                >< HphI
CCTGCTAAAG CATATAAGGA TTACCTAGCA AGTGGAGGAC AACCAATCAC CAACTGTGTG AAGATGTTGT
13030      13040      13050      13060      13070      13080      13090

                >< SinI
                >< Sau96I
                >< NspIV
                >< NspHII
                >< NlaIII
                >< Eco47I
                >< Eam1105I
                >< Cfr13I
                >< BsiZI
                >< Bme18I >< XcmI
                >< AvaI
                >< PleI ><
                >< AfaI
                >< AfaI
                >< MaeIII
                >< AluI
                >< AsuI > < HinfI
GTACACACAC TGGTACAGGA CAGGCAATTA CTGTAACACC AGAAGCTAAC ATGGACCAAG AGTCCTTGG
13100      13110      13120      13130      13140      13150      13160

                >< TfiI
                >< SfaNI
                >< NlaIII
                >< FokI
                >< HinfI
                >< MaeIII
TGGTGCTTCA TGTTGTCTGT ATTGTAGATG CCACATTGAC CATCCAAATC CTAAGGATT CTGTGACTTG
13170      13180      13190      13200      13210      13220      13230

                > < RsaI
                >< MaeII
                >< Csp6I
                > < AfaI
                >< DdeI
                >< BfrI
                >< BsrI
                >< BfrI
AAAGGTAAGT ACGTCAAAT ACCTACCACT TGTGCTAATG ACCCAGTGGG TTTTACACTT AGAAACACAG
13240      13250      13260      13270      13280      13290      13300

                >< ThaI

```

FIGURE 13.30


```

>< SfaNI
>< MvnI
>< BstUI
>< Bsp50I
>< AciI
>< RsaI
>< Csp6I
>< AfaI >< AciI
>< SfcI >< MaeIII
>< AccIISfaNI ><
TCTGTACCGT CTGCGGAATG TGGAAAGGTT ATGGCTGTAG TTGTGACCAA CTCCGCGAAC CCTTGATGCA
13310 13320 13330 13340 13350 13360 13370

>< Zsp2I
>< SfaNI
>< Mph1103I>< Tru9I
>< Ppu10I>< MaeII
>< NsiI>< FokI
>< EcoT22I >< MseI
Fnu4HI ><
BsgI ><
>< BbvI
>< AciI>< AvaIII >< DraI >< AciI >< Fnu4HI >< AciI ><
GTCTGCGGAT GCATCAACGT TTTTAAACGG GTTTGGCGTG TAAGTGCAGC CCGTCTTACA CCGTGC GGCA
13380 13390 13400 13410 13420 13430 13440

>< SpeI
>< ScaI
>< RsaI
>< RmaI
>< MaeI
>< Csp6I >< SfcI >< BspWI
>< BspWI >< AfaI >< AccI >< BcgI/a >< BcgI >
CAGGCACTAG TACTGATGTC GTCTACAGGG CTTTGTATAT TTACAACGAA AAAGTTGCTG GTTTTGCAAA
13450 13460 13470 13480 13490 13500 13510

>< ScrFI
>< MvaI
>< MnlI
>< EcoRII
>< Ecl136I
>< BstOI
>< BstNI
>< BslI
>< DsaV >< BsiYI
>< BsiLI
>< ApyI >< PleI
>< FokI >< HinfI
GTCCTAAAA ACTAATTGCT GTCGCTTCCA GGAGAAGGAT GAGGAAGGCA ATTTATTAGA CTCTTACTTT
13520 13530 13540 13550 13560 13570 13580

>< NlaIII
>< Ksp632I
>< EarI
>< Eam1104I
>< BsmAI
>< Tru9I
>< MnlI >< Alw26I >< MboII >< MseI
GTAGTTAAGA GGCATACTAT GTCTAACTAC CAACATGAAG AGACTATTTA TAACTTGGTT AAAGATTGTC
13590 13600 13610 13620 13630 13640 13650

>< RsaI
>< NlaIV
>< NlaIII
>< KpnI
>< HphI
>< Eco64I
>< Csp6I
>< BscBI
>< BanI
>< Asp718

```

FIGURE 13.31

```

                >< MaeIII >< AfaI
                > < AccB1I MaeII ><
    >< NspBII
    >< AciI >< NlaIII > < Acc65I > < HgaI
CAGCGGTTGC TGTCCATGAC TTTTCAAGT TTAGAGTAGA TGGTGACATG GTACCACATA TATCACGTCA
    13660      13670      13680      13690      13700      13710      13720

                >< MnlI
                >< MaeII
CGGTCTAACT AAATACACAA TGGCTGATTT AGTCTATGCT CTACGTCATT TTGATGAGGG TAATTGTGAT
    13730      13740      13750      13760      13770      13780      13790

    >< Tru9I
    >< MseI >< MaeIII >< MunI
ACATTAAGAAG AAATACTCGT CACATACAAT TGCTGTGATG ATGATTATTT CAATAAGAAG GATTGGTATG
    13800      13810      13820      13830      13840      13850      13860

                >< ThaI
                >< MvnI
                >< MluI
                >< BstUI
                >< Bsp50I
                >< RsaI
                >< HphI
    >< TfiI >< AflIII >< DdeI >< Csp6I Tru9I ><
    >< HinfI >< AccII >< BfrI >< AfaI MseI ><
ACTTCGTAGA GAATCCTGAC ATCTTAGCGG TATATGCTAA CTTAGGTGAG CGTGTACGCC AATCATTATT
    13870      13880      13890      13900      13910      13920      13930

                XhoII >
                Sau3AI >
                NdeII >
                MflI >
                MboI >
    > < SfaNI >< RsaI >< RsaI MboI >
    >< RsaI >< Csp6I >< Csp6I DpnII >
    >< Csp6I >< BspWI >< BspWI BstYI >
    >< AfaI >< SfaNI >< AfaI >< AfaI BspAI >
AAAGACTGTA CAATTCTGCG ATGCTATGCG TGATGCAGGC ATTGTAGGCG TACTGACATT AGATAATCAG
    13940      13950      13960      13970      13980      13990      14000

                > < ScrFI
                > < MvaI
                >< Fnu4HI
                >< EcoRII
                > < Ecl136I
                > < BstOI
                > < BstNI
    >< Tru9I >< RsaI >< BslI
    >< MseI >< RsaI >< HphI >< BsiYI
    >< DpnI >< Csp6I >< Csp6I >< BsiLI
    >< Bsp143I >< BsrI >< BbvI >< ApyI
    >< AlwI >< AfaI >< AfaI >< DsaV >< AciI
GATCTTAATG GGAAGTGGTA CGATTTCGGT GATTTTCGTAC AAGTAGCACC AGGCTGCGGA GTTCCATTG
    14010      14020      14030      14040      14050      14060      14070

                >< SfaNI
                >< RmaI >< HinfI
    >< TfiI >< SfaNI >< MamI >< MnlI >< Fnu4HIpleI ><
    >< HinfI >< FokI >< BsiBI >< MaeI >< DdeI
    >< HinfI >< FokI >< BsaBI >< BbvI >< BspWI NdeI ><
TGGATTGATA TTACTCATTG CTGATGCCCA TCCTCACTTT GACTAGGGCA TTGGCTGCTG AGTCCCATAT
    14080      14090      14100      14110      14120      14130      14140

    >< Sau3AI
    >< NdeII

```

FIGURE 13.32

```

    << MboI
    << MamI
    << DpnII
      << DpnI
        << BspWI
    << BspAI
      << Bsp143I
    << BsiBI
    << BsaBI << FokI
GGATGCTGAT CTCGCAAAAC CACTTATTAA GTGGGATTTG CTGAAATATG ATTTTACGGA AGAGAGACTT
14150      14160      14170      14180      14190      14200      14210

    << TthHB8I
    << TaqI
      << McrI
        << Ksp632I
        << EarI
        << Eam1104I
    << BsmAI
    << MboII
    << Alw26I
TGTCCTCTCG ACCGTTATTT TAAATATTGG GACCAGACAT ACCATCCCAA TTGTATTAAC TGTTTGGATG
14220      14230      14240      14250      14260      14270      14280

    << FokI
ATAGGTGTAT CCTTCATTGT GCAAACITTA ATGTGTTATT TTCTACTGTG TTCCACCTA CAAGTTTTGG
14290      14300      14310      14320      14330      14340      14350

    << SpeI
    << RmaI
    << MaeI
    << SspI
    << BsrI
ACCACTAGTA AGAAAAATAT TTGTAGATGG TGTTCCTTTT GTTGTTCCTCAA CTGGATACCA TTTTCGTGAG
14360      14370      14380      14390      14400      14410      14420

    << RsaI
    << HinfI << PfuI
      << Csp6I
    << AfaI
TTAGGAGTCG TACATAATCA GGATGTAAAC TTACATAGCT CGCGTCTCAG TTTCAAGGAA CTTTTAGTGT
14430      14440      14450      14460      14470      14480      14490

    << Zsp2I
    << SphI
    << Ppu10I
    << PaeI
    << NspI
  
```

FIGURE 13.33

```

    >> Sau3AI          >> NspHI
    >> NdeII           >> NsiI
    >> MboI            >> NlaIII
    >> DpnII           >> Mph1103I
    > < DpnI          >> Fnu4HI
    >> Fnu4HI>< BspWI >> EcoT22I
    >> BspAI           >> BspWI
    > < Bsp143I> < AvaIII > < AlwNI          >> RmaI
    >> AlwI            >> AluI           >> AluI   >> BbvI   >> MaeI
    ATGCTGCTGA TCCAGCTATG CATGCAGCTT CTGGCAATTT ATTGCTAGAT AAACGCACTA CATGCTTTTC
    14500      14510      14520      14530      14540      14550      14560

    >> ScrFI
    >> NciI
    >> MspI
    >> HpaII
    >> HapII
    >> Fnu4HI
    >> AlwNI           >> DsaV           >> Tru9I
    >> AluI            >> BcnI           >> MseI
    AGTAGCTGCA CTAACAAACA ATGTTGCTTT TCAAAGTGC AAACCCGGTA ATTTTAATAA AGACTTTTAT
    14570      14580      14590      14600      14610      14620      14630

    >> Tru9I
    >> MseI            >> MboII          DdeI >>
    GACTTTGCTG TGTCTAAAGG TTTCTTTAAG GAAGGAAGTT CTGTTGAACT AAAACACTTC TTCTTTGCTC
    14640      14650      14660      14670      14680      14690      14700

    >> FokI           EcoRV >>
    >> Fnu4HI        Eco32I >>
    AGGATGGCAA CGCTGCTATC AGTGATTATG ACTATTATCG TTATAATCTG CCAACAATGT GTGATATCAG
    14710      14720      14730      14740      14750      14760      14770

    >> VspI
    >> Tru9I
    >> MseI
    >> AsnI
    >> AseI
    >> MaeIII
    ACAACTCCTA TTCGTAAGTT AAGTTGTTGA TAAACTTTT GATTGTTACG ATGGTGGCTG TATTAATGCC
    14780      14790      14800      14810      14820      14830      14840

    >> Tru9I
    >> MseI           >> PvuII
    >> HpaI           >> Psp5I          > < XcmI
    >> HindII        >> NspBII        >> Tru9I          RmaI >>
    >> HincII        >> AluI           >> MseI          MaeI >>
    AACCAAGTAA TCGTTAACAA TCTGGATAAA TCAGCTGGTT TCCCATTAA TAAATGGGGT AAGGCTAGAC
    14850      14860      14870      14880      14890      14900      14910

    >> SfaNI          >> ThaI
    >> Sau3AI         >> MvnI
    >> NdeII         >> BstUI
    >> MboI          >> Bst1107I
    >> DpnII         >> BspWI >> FokI
    >> DpnI          >> Bsp50I
    >> PleI          >> Bsp143I      >> AccII>< DdeI
    >> HinfI>< MnlI >> BspAI >< AlwI          >> AccI
    TTTATTATGA CTCAATGAGT TATGAGGATC AAGATGCACT TTTCGCGTAT ACTAAGCGTA ATGTCATCCC
    14920      14930      14940      14950      14960      14970      14980

    >> SstI
    >> SduI
    >> SacI

```

FIGURE 13.34

```

                                >> NspII
                                >> HgiAI
                                >> Eco24I
                                >< Ecl136II
                                >> Bsp1286I
                                >> BmyI
                                >> BanII
                                >> Alw21I
                                >< Tru9I
                                >< TfiI
                                >< MseI
                                >< HinfI
                                >< Esp4I
                                >< AflIII
                                >< BspWI
                                >< AluI
                                >< AluI
TACTATAACT CAAATGAATC TTAAGTATGC CATTAGTGCA AAGAATAGAG CTCGCACCGT AGCTGGTGTGTC
14990      15000      15010      15020      15030      15040      15050

                                RmaI ><
                                >< MnlI
                                MaeI ><
                                >< Fnu4HI
                                >< AciI
TCTATCTGTA GTACTATGAC AAATAGACAG TTTCATCAGA AATTATTGAA GTCAATAGCC GCCACTAGAG
15060      15070      15080      15090      15100      15110      15120

                                >< Tru9I
                                >< MseI
>< AluI
GAGCTACTGT GGTAATTGGA ACAAGCAAGT TTTACGGTGG CTGGCATAAT ATGTTAAAAA CTGTTTACAG
15130      15140      15150      15160      15170      15180      15190

                                NspI ><
                                NspHI ><
                                NlaIII ><
                                >< NlaIII
                                DdeI ><
                                BspWI ><
                                >< MaeIII
                                BfrI ><
TGATGTAGAA ACTCCACACC TTATGGGTTG GGATTATCCA AAATGTGACA GAGCCATGCC TAACATGCTT
15200      15210      15220      15230      15240      15250      15260

                                >< Pali
                                >< HaeIII
                                >< BsuRI
                                >< BshI
                                >< MnlI
                                >< MaeIII
                                SfcI ><
AGGATAATGG CCTCTCTTGT TCTTGCTCGC AAACATAACA CTTGCTGTAA CTTATCACAC CGTTTCTACA
15270      15280      15290      15300      15310      15320      15330

                                Tru9I ><
                                ScrFI >
                                MvaI >
                                >< MseI
                                FokI ><
                                EcoRII ><
                                Ecl136I >
                                DsaV ><
                                BstOI >
                                BstNI >
                                >< NlaIII
                                >< CfoI>< Tru9I
                                >< Fnu4HI
                                BsiLI >
                                >< AluI
                                >< AviII >< MseI
                                >< AciI
                                ApyI >
GGTTAGCTAA CGAGTGTGCG CAAGTATTAA GTGAGATGGT CATGTGTGGC GGCTCACTAT ATGTTAAACC
15340      15350      15360      15370      15380      15390      15400

                                >< SfaNI
                                >< MspI
                                >< HpaII
                                >< HapII
                                >< HphI
                                >< BspWI
                                >< Tru9I
                                MaeIII ><
                                >< MseI
                                AluI ><

```

FIGURE 13.35

```

AGGTGGAACA TCATCCGGTG ATGCTACAAC TGCTTATGCT AATAGTGTCT TTAACATTTG TCAAGCTGTT
15410      15420      15430      15440      15450      15460      15470

                                << DrdI
<< BspWI                                << AluI                                > < AciI
ACAGCCAATG TAAATGCACT TCTTTCAACT GATGGTAATA AGATAGCTGA CAAGTATGTC CGCAATCTAC
15480      15490      15500      15510      15520      15530      15540

                                << Sau3AI
                                << NdeII
                                << MboI
                                > < MamI
                                << FbaI
                                << DpnII
                                << DpnI
                                << BspHI
                                << BspAI
                                << Bsp143I
                                << BsiQI
                                << SfcI                                > < BsiBI>< NlaIII
                                << BsmAI                                > < BsaBI>< FokI
                                << Alw26I                                << BclI>< EcoRI                                FokI ><
AACACAGGCT CTATGAGTGT CTCTATAGAA ATAGGGATGT TGATCATGAA TTCGTGGATG AGTTTACGC
15550      15560      15570      15580      15590      15600      15610

                                << TfiI
                                << SfaNI
                                << NlaIII
                                << BspMI                                << HinfI                                << MaeIII
TTACCTGCGT AAACATTTCT CCATGATGAT TCTTTCTGAT GATGCCGTTG TGTGCTATAA CAGTAACTAT
15620      15630      15640      15650      15660      15670      15680

                                > < RmaI
                                << NheI << Tru9I
                                > < MaeI                                << Tru9I
<< Fnu4HI                                << AluI << MseI                                << MseI                                MnlI ><
GCGGCTCAAG GTTTAGTAGC TAGCATTAAAG AACTTTAAGG CAGTCTTTA TTATCAAAAT AATGTGTTCA
15690      15700      15710      15720      15730      15740      15750

                                << SinI
                                << Sau96I
                                << PssI
                                << Psp5II
                                << PpuMI
                                << NspIV
                                << NspHII
                                << Eco0109I
                                << Eco47I
                                << DraII
                                << Cfr13I
                                << Bsi2I
                                << Bme18I
                                << DdeI
                                << NlaIII                                << BsmAI
                                << DdeI                                << Alw26I                                << AsuI                                << MnlI
TGTCTGAGGC AAAATGTTGG ACTGAGACTG ACCTTACTAA AGGACCTCAC GAATTTTGCT CACAGCATA
15760      15770      15780      15790      15800      15810      15820

                                << XhoII
                                << Sau3AI
                                << NdeII
                                << MflI
                                << MboI

```

FIGURE 13. 36

```

                >< RsaI          >< DpnII
                >< MaeII        >< DpnI          > < SspI
                >< Csp6I        >< BstYI        HinPII ><
    >< Tru9I
    >< RmaI          >< BsaAI          >< BspMI        Hin6I ><
    >< MaeI          >< AflIII       >< BspAI        HhaI ><
    >< BspWI>< MseI          >< AfaI      >< AlwI>< Bsp143I   CfoI ><
AATGCTAGTT AAACAAGGAG ATGATTACGT GTACCTGCCT TACCCAGATC CATCAAGAAT ATTAGGCGCA
    15830      15840      15850      15860      15870      15880      15890

                >< RsaI          >< SfaNI
                >< TthHB8I      >< Csp6I        >< MaeIII
                >< TaqI          >< AfaI          BsrI ><
GGCTGTTTTG TCGATGATAT TGTCAAAACA GATGGTACAC TTATGATTGA AAGGTTCGTG TCACTGGCTA
    15900      15910      15920      15930      15940      15950      15960

    > < FokI
    >< BspWI
    TTGATGCTTA CCCACTTACA AAACATCCTA ATCAGGAGTA TGCTGATGTC TTCACTTGT ATTACAATA
    15970      15980      15990      16000      16010      16020      16030

                >< Van9II
                >< PflMI
                >< NspI
    > < Pali>< NspHI
    > < MscI>< NlaIII
    > < HaeIII
    > < BsuRI
    >< BsrI
    >< EaeI >< BslI >< NspI
    > < BshI>< BsiYI >< NspHI
                >< NlaIII
                >< AflIII >< AflIII
    >< MaeIII >< AluI > < BalI>< AccB7I >< NlaIII
CATTAGAAAG TTACATGATG AGCTTACTGG CCACATGTTG GACATGTATT CCGTAATGCT AACTAATGAT
    16040      16050      16060      16070      16080      16090      16100

                >< RsaI> < NlaIV
                >< MnlI
    >< Csp6I >< DdeI          >< RsaI
                >< BsrI >< MnlI          >< Csp6I
    >< AfaI> < BscBI          >< AfaI          SfcI ><
AACACCTCAC GGTACTGGGA ACCTGAGTTT TATGAGGCTA TGTACACACC ACATACAGTC TTGCAGGCTG
    16110      16120      16130      16140      16150      16160      16170

                >< NlaIV
                >< EcoNI
                >< Eco31I
    >< Eco64I>< BsmAI
                >< BscBI >< BslI
    >< BanI >< BsiYI
    >< AciI >< BsaI
    >< BspWI          >< AccB1I>< Alw26I BbvI ><
TAGGTGCTTG TGTATTGTGC AATTCACAGA CTTCACTTCG TTGCGGTGCC TGTATTAGGA GACCATTCTT
    16180      16190      16200      16210      16220      16230      16240

                >< Tth111I
                >< Fnu4HI >< NlaIII          > < Tru9I
    >< BspWI >< AspI          > < MseI
ATGTTGCAAG TGCTGCTATG ACCATGTCAT TTCAACATCA CACAAATTAG TGTTGTCTGT TAATCCCTAT
    16250      16260      16270      16280      16290      16300      16310

    >< ScrFI
    >< MvaI

```

FIGURE 13. 37

```

>< EcoRII
  >< Ecl136I
>< DsaV
  >< BstOI
  >< BstNI
  >< BsiLI
  >< BsaJI
  >< ApyI
  >< MaeIII >< MaeIII
  >< MnlI
  >< MaeI
  >< RmaI
  >< MnlI
  >< BspWI ><
  >< AluI
GTTTGCAATG CCCAGGTTG TGATGTCAC TATGTGACAC AACTGTATCT AGGAGGTATG AGCTATTATT
  16320      16330      16340      16350      16360      16370      16380

  >< MaeIII
  >< MnlI
GCAAGTCACA TAAGCCTCCC ATTAGTTTTT CATTATGTGC TAATGGTCAG GTTTTTGGTT TATACAAAAA
  16390      16400      16410      16420      16430      16440      16450

  >< NspI
  >< NspHI
  >< NlaIII>< MaeIII>< MaeIII
  >< AflIII
  >< AspI
  >< NspI
  >< NspHI
  >< NlaIII
  >< AflIII
  >< AspI
  >< AflIII
CACATGTGTA GGCAGTGACA ATGTCAC TCTCAATGCG ATAGCAACAT GTGATTGGAC TAATGCTGGC
  16460      16470      16480      16490      16500      16510      16520

  >< RsaI
  >< P1eI
  >< DdeI
  >< Csp6I
  >< BsmAI >< HinfI
  >< Alw26I
  >< AfaI
  >< HindIII
  >< MnlI
  >< DdeI ><
  >< AluI >< Fnu4HI >< BbvI
GATTACATAC TTGCCAACAC TTGTACTGAG AGACTCAAGC TTTTCGCAGC AGAAACGCTC AAAGCCACTG
  16530      16540      16550      16560      16570      16580      16590

  >< ThaI
  >< ScaI
  >< RsaI >< RsaI
  >< MvnI
  >< Csp6I >< Csp6I
  >< BstUI
  >< Bsp50I
  >< Tru9I
  >< MseI >< NdeI
  >< AfaI >< AfaI
  >< AluI
  >< AccII
  >< MnlI >
AGGAAACATT TAAGCTGTCA TATGGTATTG CCACTGTACG CGAAGTACTC TCTGACAGAG AATTGCATCT
  16600      16610      16620      16630      16640      16650      16660

  >< MaeIII ><
  >< MaeIII
  >< EcoO65I
  >< Eco91I
  >< BstPI
  >< BstEII
  >< BsrI
  >< SfaNI
  >< NlaIII
  >< RmaI
  >< MaeI
  >< RmaI
  >< MaeI
  >< BsrI
TTCATGGGAG GTTGGAAAAC CTAGACCACC ATTGAACAGA AACTATGTCT TFACTGGTTA CCGTGTAACT
  16670      16680      16690      16700      16710      16720      16730

  >< RsaI ><
  >< MnlI
  >< HphI
  >< RsaI
  >< RsaI
  >< Csp6I
  >< Csp6I
  >< SfaNI
  >< Csp6I ><
  >< AfaI
  >< AfaI
  >< MaeIII
  >< HphI AfaI ><
AAAAATAGTA AAGTACAGAT TGGAGAGTAC ACCTTTGAAA AAGGTGACTA TGGTGATGCT GTTGTGTACA
  16740      16750      16760      16770      16780      16790      16800

```

FIGURE 13. 38


```

    >< RsaI
    >< Csp6I
    >< AfaI
GAGGTACTAC GACATACAAG TTGAATGTTG GTGATTACTT TGTGTGACA TCTCACACTG TAATGCCACT
    16810      16820      16830      16840      16850      16860      16870

    >< HphI
    >< HindII
    >< HincII
    DdeI ><
    BfrI ><

    >< VneI
    >< SnoI
    >< SduI
    >< NspII
    >< HgiAI
    > < SduI
    > < NspII
    >< DraIII
    >< Bsp1286I
    >< BmyI
    >< BspWI >< DraIII
    >< RsaI
    >< ApaLI >< RmaI
    >< Bsp1286I
    >< Csp6I
    >< Alw44I >< MaeI
    >< BmyI
    >< BsrI
    >< Alw21I
    >< AfaI
    DdeI >
TAGTGCACCT ACTCTAGTGC CACAAGAGCA CTATGTGAGA ATTACTGGCT TGTACCCAAC ACTCAACATC
    16880      16890      16900      16910      16920      16930      16940

    StyI ><
    SinI >
    Sau96I >
    NspIV >
    EcoT14I ><
    Eco47I >
    Eco130I ><
    >< ScaI Cfr13I >
    BssT1I ><
    >< SphI >< RsaI BsiZI >
    >< PaeI BsaJI ><
    >< NlaIII Bme18I >
    >< NspI>< Csp6I AvaII >
    >< NspHI>< AfaI AsuI >
    >< RmaI
    >< MaeI
TCAGATGAGT TTTCTAGCAA TGTTGCAAAT TATCAAAGG TCGGCATGCA AAAGTACTCT ACACTCCAAG
    16950      16960      16970      16980      16990      17000      17010

    >< ScrFI
    >< RsaI
    >< MvaI
    >< EcoRII
    >< Ecl136I
    > < Csp6I
    >< BstOI
    >< BstNI
    >< XcmI >< BslI
    >< NspHII >< BsiYI
    >< BsiLI
    >< ApyI >< BsrI
    >< DsaV>< AfaI > < HinfI>< PleI
GACCACCTGG TACTGGTAAG AGTCATTTTG CCATCGGACT TGCTCTCTAT TACCCATCTG CTCGCATAGT
    17020      17030      17040      17050      17060      17070      17080

    >< SfaNI
    >< SphI >< PvuII
    >< PaeI >< Psp5I
    >< NspI >< NspBII
    >< NspHI >< Fnu4HI
    >< Bst1107I > < NlaIII>< BspWI
    >< SspI
    >< AccI >< NlaIII >< AluI >< BbvI
    > < MseI
GTATACGGCA TGCTCTCATG CAGCTGTTGA TGCCCTATGT GAAAAGGCAT TAAAATATTT GCCCATAGAT
    17090      17100      17110      17120      17130      17140      17150

```

FIGURE 13.39

```

> < ThaI
>< ThaI
> < MvnI
>< MvnI >< ThaI
> < HinP1I
>< HinP1I
>< HinP1I >< MvnI
> < Hin6I
>< Hin6I
> < HhaI
>< HhaI >< HhaI
> < CfoI
>< CfoI >< CfoI
> < BstUI
>< BstUI >< BstUI
>< BssHII
>< BspMI
> < Bsp50I
>< Bsp50I>< Bsp50I
>< TfiI >< Hin6I> < AccII
>< HinFI >< AccII >< AccII
>< EcoRI
AAATGTAGTA GAATCATACC TGCGCGTGCG CGCGTAGAGT GTTTTGATAA ATTCAAAGTG AATTC AACAC
17160 17170 17180 17190 17200 17210 17220

>< Zsp2I
>< Ppul0I
>< NsiI
>< Mph1103I
>< EcoT22I
>< BsgI > < AvaIII >< DrdI
TAGAACAGTA TGTTTTCTGC ACTGTAAATG CATTGCCAGA AACAACTGCT GACATTGTAG TCTTTGATGA
17230 17240 17250 17260 17270 17280 17290

>< RmaI
>< MaeI >< MaeII
AATCTCTATG GCTACTAATT ATGACTTGAG TGTTGTCAAT GCTAGACTTC GTGCAAAACA CTACGTCTAT
17300 17310 17320 17330 17340 17350 17360

>< Sau3AI
>< NdeII
>< MboI
>< DpnII
>< DpnI
>< BspAI >< RmaI
>< AlwI>< Bsp143I > < AciI >< MaeI SspI ><
ATTGGCGATC CTGCTCAATT ACCAGCCCCC CGCACATTGC TGAATAAAGG CACTAGAA CCAGAATATT
17370 17380 17390 17400 17410 17420 17430

>< SinI
>< Sau96I
>< NspIV >< StyI
>< NspHII >< NspI
>< Eco47I >< NspHI
>< Cfr13I >< NlaIII
>< Bsi2I >< EcoT14I
>< BsgI >< Eco130I
>< Bme18I >< BssTII
>< AvaII >< BsaJI
>< Tru9I
>< MseI
>< AsuI> < AflIII
TTAATTCAGT GTGCAGACTT ATGAAAACAA TAGGTCCAGA CATGTTCTT GGAACCTGTC GCCGTTGTCC
17440 17450 17460 17470 17480 17490 17500

```

FIGURE 13. 40

```

                >< HindII
                >< HincII
                >< AluI
TGCTGAAATT GTTGACACTG TGAGTGCTTT AGTTTATGAC AATAAGCTAA AAGCACACAA GGATAAGTCA
 17510      17520      17530      17540      17550      17560      17570

>< AluI                >< NlaIII
GCTCAATGCT TCAAAATGTT CTACAAAGGT GTTATTACAC ATGATGTTTC ATCTGCAATC AACAGACCTC
 17580      17590      17600      17610      17620      17630      17640

                >< MnlI
>< EcoNI
  >< BslI
  >< BsiYI
AAATAGGCGT TGTAAGAGAA TTTCTTACAC GCAATCCTGC TTGGAGAAAA GCTGTTTTTA TCTCACCTTA
 17650      17660      17670      17680      17690      17700      17710

                >< HphI
                >< AluI
                >< SfcI                >< DdeI                >< TfiI
                > < AluI                >< BfrI                >< HinfI
TAATTCACAG AACGCTGTAG CTTCAAAAAT CTTAGGATTG CCTACGCAGA CTGTTGATTC ATCACAGGGT
 17720      17730      17740      17750      17760      17770      17780

                >< Tth111I                > < HindII
                >< AspI                > < HincII
                >< AspI                >< AciI
TCTGAATATG ACTATGTCAT ATTCACACAA ACTACTGAAA CAGCACACTC TTGTAATGTC AACCGCTTCA
 17790      17800      17810      17820      17830      17840      17850

                >< XhoII
                >< Sau3AI
                >< NdeII
                >< MflI
                >< MboI
                >< MamI
                >< DpnII
                >< DpnI
                >< BstYI
                >< BspAI
                >< Bsp143I
                >< BsiBI
                >< BsaBI
                >< BglII
                >< BspWI
ATGTGGCTAT CACAAGGGCA AAAATTGGCA TTTTGTGCAT AATGTCTGAT AGAGATCTTT ATGACAACT
 17860      17870      17880      17890      17900      17910      17920

                >< XbaI
                >< RmaI                >< MaeIII
                >< MaeI                >< MaeII                BsrI ><
GCAATTTACA AGTCTAGAAA TACCACGTCG CAATGTGGCT ACATTACAAG CAGAAAATGT AACTGGACTT
 17930      17940      17950      17960      17970      17980      17990

                >< Sau3AI
                >< NdeII
                >< MboII
                >< MboI
                > < FokI
                >< DpnII                >< NlaIV
                >< DpnI                >< Eco64I
                >< BspAI                >< BscBI
                >< Bsp143I
                >< BanI                MnlI ><
>< Tru9I                >< BbsI > < BsrI                >< AccBI                >< DdeI
>< MseI>< SfcI

```

FIGURE 13. 41


```

TGTTGACACT GAAAATAACA CAGAATTCAC CAGAGTTAAT GCAAAACCTC CACCAGGTGA CCAGTTTAAA
18350      18360      18370      18380      18390      18400      18410

                >< ScrFI
                >< MvaI
                >< EcoRII
                >< Ecl136I
                >< DsaV
                >< BstOI
                >< BstNI
                >< BsiLI
                >< BsaJI
                >< ApyI
                >< NlaIII
                >< NlaIII
                >< NlaIII
                >< HinPII
                >< Hin6I
                >< Tth111I
                >< HinFI
                >< AspI
                >< PleI
                >< AspI
                >< CfoI
                >< AluI
                >< SinI
                >< Sau96I
                >< NspIV
                >< NspHII
                >< Eco47I
                >< Cfr13I
                >< ScaI
                >< RsaI
                >< Csp6I
                >< AfaI
                >< BsiZI
                >< Bme18I
                >< AvaII
                >< AsuI
                >< MaeII
                >< AflIII
                >< MaeIII>< MaeII
AATGAAGTAC TTTGTCAAGA TTGGACCTGA AAGAACGTGT TGTCTGTGTG ACAAACGTGC AACTTGCTTT
18560      18570      18580      18590      18600      18610      18620

                >< TfiI
                >< HinFI
                >< Tth111I
                >< AspI
TCTACTTCAT CAGATACTTA TGCCTGCTGG AATCATTCTG TGGGTTTTGA CTATGTCTAT AACCCATTTA
18630      18640      18650      18660      18670      18680      18690

                >< ScrFI
                RsaI ><
                >< MvaI
                >< EcoRII
                Ecl136I ><
                >< DsaV
                Csp6I ><
                BstXI ><
                >< MaeIII
                >< Eco065I
                >< Eco91I
                >< BstPI
                >< Eco57I> < BstEII
                >< MaeIII >< NlaIII
                >< AfaI ><
TGATTGATGT TCAGCAGTGG GGCTTTACGG GTAACCTTCA GAGTAACCAT GACCAACATT GCCAGGTACA
18700      18710      18720      18730      18740      18750      18760

                >< SfaNI
                >< RnaI
                >< NspI
                >< NspHI

```

FIGURE 13.43

```

                >> NlaIII                >> RmaI
                >> MaeI                >> NlaIII                Tru9I >>
>> NlaIII    >> BspWI                >> MaeI                >> NlaIII
    > < AflIII                >> BspHI                MseI >>
TGGAAATGCA CATGTGGCTA GTTGTGATGC TATCATGACT AGATGTTTAG CAGTCCATGA GTGCTTTGTT
    18770      18780      18790      18800      18810      18820      18830

    >> ThaI
    >> MvnI
>> HinPII
>> Hin6I
    >> HbaI
    >> CfoI
    >> BstUI
    >> Bsp50I
    >> AccII
                >> EcoNI > < MnlI
                >> BslI                >> Tru9I
                >> BsiYI                >> DdeI >> MseI
AAGCGCGTTG ATGGTCTGT TGAATACCCT ATTATAGGAG ATGAACTGAG GGTAAATTCT GCTTGCAGAA
    18840      18850      18860      18870      18880      18890      18900

    >> RsaI
    >> Csp6I
    >> AfaI    >> NlaIII                >> BspWI                >> MboII                > < NlaIII
AAGTACAACA CATGGTTGTG AAGTCTGCAT TGCTTGCTGA TAAGTTTCCA GTTCTTCATG ACATTGGAAA
    18910      18920      18930      18940      18950      18960      18970

                >> SauI
                >> MstII
                >> Eco81I
                >> DdeI
                >> CvnI
                >> Bsu36I
                >> Bse21I
                >> AxyI
                >> AocI    >> MnlI    >> SfaNI
                >> Bpu1102I
TCCAAGGCT ATCAAGTGTG TGCCTCAGGC TGAAGTAGAA TGGAAGTTCT ACGATGCTCA GCCATGTAGT
    18980      18990      19000      19010      19020      19030      19040

                >> MnlI
    >> HindIII
    >> AluI    >> MboII                >> Eam1104I
                >> Ksp632I
                >> EarI
GACAAAGCTT ACAAATAGA GGAACCTTC TATTCTTATG CTACACATCA CGATAAATTC ACTGATGGTG
    19050      19060      19070      19080      19090      19100      19110

                >> Sau3AI
                >> NdeII
                >> MboI
                >> MaeII > < MaeIII
                >> DpnII
                >> DpnI
                >> BspAI
                >> MaeIII >> Bsp143I                >> MunI                HinfI >
                >> DrdI >>
TTTGTTTGTT TTGGAATTGT AACGTGATC GTTACCCAGC CAATGCAATT GTGTGTAGGT TTGACACAAG
    19120      19130      19140      19150      19160      19170      19180

                Zsp2I >>
                >> SphI
                > < Ppu10I
                >> PaeI
                >> NspI
                >> NspHI
                >> NlaIII
    >> ScrFI
    >> MvaI
    >> EcoRII                Mph1103I >>

```

FIGURE 1344

```

                << Ecl136I
                << DsaV
                << BstOI
                << BstNI
                << BsiLI
                << ApyI
                << PleI
AGTCTTGTC AACTTGAAC TACCAGGCTG TGATGGTGGT AGTTTGTATG TGAATAAGCA TGCATTCCAC
19190      19200      19210      19220      19230      19240      19250

                << Tru9I
                > < MunI
                << TthHB8I
                << BcgI/a << TaqI
                << AluI
                << MseI
                << DraI
                << BcgI
ACTCCAGCTT TCGATAAAAG TGCATTTACT AATTTAAAGC AATTGCCTTT CTTTTACTAT TCTGATAGTC
19260      19270      19280      19290      19300      19310      19320

                << PleI
                << NlaIII
                << BsmAI
                << HinfI>< Alw26I
                SfaNI <<
                << MaeII
                BsaAI <<
                AflIII <<
CTTGTGAGTC TCATGGCAA CAAGTAGTGT CGGATATTGA TTATGTTCCA CTCAAATCTG CTACGTGTAT
19330      19340      19350      19360      19370      19380      19390

                Zsp2I >
                << ScaI
                Ppu10I ><
                << RsaINsiI >
                Mph1103I >
                << SfaNIEcoT22I >
                > < RsaI >< Csp6I
                << Csp6I
                << NlaIII> < AfaI >< AfaI
TACACGATGC AATTTAGGTG GTGCTGTTTG CAGACACCAT GCAAATGAGT ACCGACAGTA CTTGGATGCA
19400      19410      19420      19430      19440      19450      19460

                << FokI
TATAATATGA TGATTTCTGC TGGATTTAGC CTATGGATTT ACAAACAATT TGATACTTAT AACCTGTGGA
19470      19480      19490      19500      19510      19520      19530

                << ScrFI
                << MvaI
                << MaeIII
                << EcoRII
                << Ecl136I
                << DsaV
                << BstOI
                << BstNI
                << BsiLI
                << ApyI
                << Tru9I
                << MseI
ATACATTTAC CAGGTTACAG AGTTTAGAAA ATGTGGCTTA TAATGTTGTT AATAAAGGAC ACTTTGATGG
19540      19550      19560      19570      19580      19590      19600

                << SgrAI
                << NaeI
                << MspI
                << HpaII
                << HapII
                << Cfr10I
                << BspWI
                > < VspI
                > < Tru9I
                > < MseI
                > < AsnI
                > < AseI
ACACGCCGGC GAAGCACCTG TTTCATCAT TAATAATGCT GTTTACACAA AGGTAGATGG TATTGATGTG
19610      19620      19630      19640      19650      19660      19670

```

FIGURE 13. 45

```

>< XhoII
>< Sau3AI
>< NdeII
>< MflI
>< MboI
>< DpnII
  >< DpnI
>< BstYI
>< BspAI
  >< BspI43I
>< BglII
GAGATCTTTG AAAATAAGAC AACACTTCCT GTTAATGTTG CATTGAGCT TTGGGCTAAG CGTAACATTA
19680      19690      19700      19710      19720      19730      19740
                                     >< MaeIII
                                     >< EspI
                                     >< DdeITru9I ><
                                     >< CeliIMseI ><
                                     >< AluI >< BpuII02I
>< Tru9I
>< MseI
>< BsrI
AACCAGTGCC AGAGATTAAG ATACTCAATA ATTTGGGTGT TGATATCGCT GCTAATACTG TAATCTGGGA
19750      19760      19770      19780      19790      19800      19810
                                     >< Fnu4HI
                                     >< EcoRV
                                     >< Eco32I
>< MseI
>< BbvI
>< NspI
>< NspHI
>< NlaIII
  >< BsgI
    >< AflIII
CTACAAAAGA GAAGCCCCAG CACATGTATC TACAATAGGT GTCTGCACAA TGA CTGACAT TGCCAAGAAA
19820      19830      19840      19850      19860      19870      19880
>< DdeI>< MboII
CCTACTGAGA GTGCTTGTTT CTTCACTTACT GTCTTGTTT ATGGTAGAGT GGAAGGACAG GTAGACCTTT
19890      19900      19910      19920      19930      19940      19950
                                     SinI ><
                                     Sau96I ><
                                     NspIV ><
                                     NspHII ><
                                     NlaIV ><
                                     Eco47I ><
                                     CfrI3I ><
                                     >< BslI
                                     BsiZI ><
                                     >< BsiYI
                                     BscBI ><
                                     BmeI8I ><
                                     AvaII ><
                                     AsuI ><
>< Tru9I
>< MseI
TTAGAAACGC CCGTAATGGT GTTTAAATAA CAGAAGGTT AGTCAAAGGT CTAACACCTT CAAAGGGACC
19960      19970      19980      19990      20000      20010      20020
                                     >< VspI
                                     >< Tru9I
                                     >< PleI
                                     >< MseI
>< RmaI
>< NheI
>< MaeI
>< HgaI>< AluI
AGCACAAAGCT AGCGTCAATG GAGTCACATT AATTGGAGAA TCAGTAAAAA CACAGTTTAA CTACTTTAAG
20030      20040      20050      20060      20070      20080      20090
                                     >< Tru9I ><
                                     >< Tru9I
                                     >< MseI ><
                                     >< MseI
>< AsnI >< TfiI
>< HinfI>< AseI >< HinfI
>< DdeI >< MnlI Tru9I ><
>< BsmAI >< DdeI

```

FIGURE 1346


```

>< AccI
AAAGTAGACG GCATTATTCA ACAGTTGCCT GAAACCTACT TTACTCAGAG CAGAGACTTA GAGGATTTTA
20100      20110      20120      20130      20140      20150      20160

>< Alw26I >< BfrIMseI ><
>< TthHB8I
>< TaqI
    >< SstI
    >< SduI
    >< SacI
    >< PaeR7I
    >< NspIII
    >< NspII
    >< HgiAI
    >< Eco88I
    >< XhoI>< Eco24I
    >< XcmI
    >< XhoI>< Eco24I
    >< Ecl136II
    >< SlaI>< Bsp1286I
    >< CcrI>< BmyI
    >< BcoI>< BanII
    >< Ama87I
    >< AvaI>< Alw21I
    >< BspAI
    >< Bsp143I
    >< AluI
    >< EcoRI
    >< FokI>< AluI ><
AGCCCAGATC ACAAATGGAA ACTGACTTTC TCGAGCTCGC TATGGATGAA TTCATACAGC GATATAAGCT
20170      20180      20190      20200      20210      20220      20230

>< TthHB8I
>< TaqI
>< SfuI
>< NspV
>< LspI
>< Csp45I
>< BstBI
>< Bsp119I
>< BsiCI
>< Bpu14I
>< AsuII >< BcgI
>< MboII
>< BbsI Tru9I ><
>< NlaIII >< AciIMseI ><
CGAGGGCTAT GCCTTCGAAC ACATCGTTTA TGGAGATTTT AGTCATGGAC AACTGGCGG TCTTCATTTA
20240      20250      20260      20270      20280      20290      20300

>< HphI
>< HinPII
>< Hin6I
>< EspI >< HhaI >< TfiI
>< DdeI >< HaeII
>< CelII >< Eco47III >< Tru9I
>< Bpu1102I >< CfoI >< HinfI >< MseI
>< BfrI >< Bsp143II >< MnlI
ATGATAGGCT TAGCCAAGCG CTCACAAGAT TCACCACTTA AATTAGAGGA TTTTATCCCT ATGGACAGCA
20310      20320      20330      20340      20350      20360      20370

>< MstI
>< HinPII
>< Hin6I
>< HhaI
>< FspI
>< FdiII
>< CfoI
>< SfaNI >< AviII
>< Sau3AI ><
>< NdeII ><
>< MboI ><
>< DpnII ><
>< DpnI ><
>< BspAI ><
>< Bsp143I ><
CAGTGAAAAA TTACTTCATA ACAGATGCGC AAACAGGTTT ATCAAAATGT GTGTGTTCTG TGATTGATCT
20380      20390      20400      20410      20420      20430      20440

>< TthHB8I

```

FIGURE 13. 47

```

                >< Tth111I
                >< TaqI
                >< AspI                > < MaeIII                MaeIII ><
TTTACTTGAT GACTTTGTCG AGATAATAAA GTCACAAGAT TTGTCAGTGA TTTCAAAAGT GGTCAAGGTT
    20450         20460         20470         20480         20490         20500         20510

                >< NspI
                >< NspHI
                >< NlaIII
                >< FokI

    >< MunI                > < NlaIII                >< AflIII
ACAATTGACT ATGCTGAAAT TTCATTCATG CTTGGTGTA AGGATGGACA TGTTGAAACC TTCTACCCAA
    20520         20530         20540         20550         20560         20570         20580

                >< SfaNI
                >< ScrFI
                >< MvaI
    >< EcoRII
                >< Ecl136I
    >< DsaV
                >< BstOI                >< SfaNI
                >< BstNI                >< RsaI    BspWI ><
                >< BsiLI                > < Csp6I                BsmI >
                >< BspWI                >< ApyI                >< AfaI    BscCI ><
AACTACAAGC AAGTCAAGCG TGGCAACCAG GTGTTGCGAT GCCTAACTTG TACAAGATGC AAAGAATGCT
    20590         20600         20610         20620         20630         20640         20650

    >< Eco57I >< MaeIII                >< HphI
TCTTGAAAAG TGTGACCTTC AGAATTATGG TGAAAATGCT GTTATACCAA AAGGAATAAT GATGAATGTC
    20660         20670         20680         20690         20700         20710         20720

                > < RsaI
                >< Csp6I
                >< Bst1107I                >< Tru9I                >< AluI
                >< AccI                >< MseI                > < AfaINlaIII ><
GCAAAGTATA CTCAACTGTG TCAATACTTA AATACACTTA CTTTAGCTGT ACCCTACAAC ATGAGAGTTA
    20730         20740         20750         20760         20770         20780         20790

                >< ScrFI
                >< RsaI
                >< MvaI
    >< EcoRII >< NspBII
                >< Ecl136I                >< SduI
                > < Csp6I                >< NspII
                >< BstOI >< PvuII>< HgiAI
                >< BstNI                >< DdeI
                >< BsiLI >< Psp5I>< Bsp1286I
                >< ApyI >< AluI >< BmyI
                >< DsaV>< AfaI                >< Alw21I
TTCAC TTGG TGCTGGCTCT GATAAAGGAG TTGCACCAGG TACAGCTGTG CTCAGACAAT GGTGCCAAC
    20800         20810         20820         20830         20840         20850         20860

                >< XhoII
                >< Tru9I
                >< Sau3AI
                >< NdeII
    >< TthHB8I >< MseI
                >< MflI
                >< MboI
                >< MamI
                >< DpnII
                >< TfiI >< DpnI

```

FIGURE 13. 48

```

                >< BstYI                > < TfiI
                >< BspAI                > < HinfI
                >< HinfI>< BspI43I        >< Esp3I    >< Tru9I
                >< BsiBI    >< Tth111I    >< BsmBI    >< MseI
                >< BsaBI    >< BsmAI    > < BsmAI
    >< BsrI    >< TaqI >< BglII    >< AspI    >< Alw26I >< HgaI> < Alw26I
TGGCACACTA CTTGTCGATT CAGATCTTAA TGA CTTCGTC TCCGACGCAG ATTCTACTTT AATTGGAGAC
    20870      20880      20890      20900      20910      20920      20930

                >< StyI
                >< SinI
                >< Sau96I
                > < SinI
                > < Sau96I
                >< PssI
                >< Psp5II
                > < PpuMI
                > < NspIV
                >< NspHII
                >< NlaIV
                > < Eco0109I
                > < Eco47I
                > < DraII
                > < Cfr13I
                > < BsiZI
                >< BscBI
                >< Bme18I
                > < AvaII
                >< AfaI
                >< RsaI
                > < Csp6I
                >< AfaI
                >< RmaI
                >< NspIV
NspHII ><
                >< MaeI
                >< EcoT14I
                >< Eco47I
                >< Eco130I
                >< Cfr13I
                >< BstT1I
                >< BsiZI
                >< BsaJI
                >< Bme18I
                >< BlnI
                >< AvrII
                >< AvaII
                >< AsuI
                >< AflIII ><
TGTGCAACAG TACATACGGC TAATAAATGG GACCTTATTA TTAGCGATAT GTATGACCCT AGGACCAAAC
    20940      20950      20960      20970      20980      20990      21000

    >< NspI
    >< NspHI
    >< NlaIII >< PleI
    >< MaeIII    >< HinfI
ATGTGACAAA AGAGAAATGAC TCTAAAGAAG GGT TTTTCAC TTATCTGTGT GGATTTATAA AGCAAAAAC
    21010      21020      21030      21040      21050      21060      21070

    >< ScrFI
    >< MvaI
    >< EcoRII
    >< Ecl136I
    >< DsaV
    >< BstOI
    >< BstNI
    >< BsiLI
    >< BsaJI
    >< BsaJI    >< SfcI
    >< ApyI
    >< AluI
    >< BscCI
    >< BsmI
    >< BsmI
    >< BscCIHindIII ><> < AluI
AGCCCTGGGT GGT TCTATAG CTGTAAAGAT AACAGAGCAT TCTTGGAAATG CTGACCTTTA CAAGCTTATG
    21080      21090      21100      21110      21120      21130      21140

                >< Zsp2I
                >< Ppu10I
                >< NsiI
                >< Mph1103I    Tru9I ><
                >< EcoT22I
                >< MseI
    >< BshI
    >< NlaIII>< AluI
    >< BcgI
    >< AvaIII >< SfaNIBcgI/a ><
GGCCATTCT CATGGTGGAC AGCTTTTGTT ACAAATGTAA ATGCATCATC ATCGGAAGCA TTTTAAATG
    21150      21160      21170      21180      21190      21200      21210

```

FIGURE 13.49

```

                >< Zsp2I
                >< SphI
        >< Ppu10I
                >< PaeI
                >< NspI
                >< NspHI
                >< NsiI
                >< NlaIII
        > < NlaIII
                >< Mph1103I
                >< EcoT22I
        > < AvaIII   >< MnlI
GGGCTAACTA TCTTGGCAAG CCGAAGGAAC AAATTGATGG CTATACCATG CATGCTAACT ACATTTCTG
  21220      21230      21240      21250      21260      21270      21280

                >< MboII
                >< GsuI
                >< BsrI
                >< BpmI
                >< BbsI
                >< NlaIII
        >< MnlI ><
        >< Tru9I ><
        >< Tru9I
        >< MseI ><
        >< MseI ><
        >< MseI
        >< MnlI ><
GAGGAACACA AATCCTATCC AGTTGTCTTC CTATTCACCTC TTTGACATGA GCAAATTTCC TCTTAAATTA
  21290      21300      21310      21320      21330      21340      21350

                >< Tru9I
                >< MseI
                >< Esp4I> < TfiI
                >< BsmAI
                >< Alw26I
                >< AflIII> < HinfI
                >< MboII
                >< EarI
                >< Eam1104I ><
AGAGGAACTG CTGTAATGTC TCTTAAGGAG AATCAAATCA ATGATATGAT TTATTCTCTT CTGGAAAAAG
  21360      21370      21380      21390      21400      21410      21420

                >< Tru9I
                >< MseI
                >< HindII
                >< HincII
                >< HpaI AflIII >
GTAGGCTTAT CATTAGAGAA AACAACAGAG TTGTGGTTTC AAGTGATATT CTTGTTAACA ACTAAACGAA
  21430      21440      21450      21460      21470      21480      21490

                >< VneI
                >< SnoI
                >< SduI
                >< NspII
        >< HpaII
                >< HgiAI
        >< HapII
        >< Cfr10I
                >< Bsp1286I
                >< MspI>< BmyI
                >< ApaLI
                >< Alw44I
                >< MaeI >< MaeIII >< AgeI >< Alw21I
CATGTTTATT TTCTTATTAT TTCTTACTCT CACTAGTGGT AGTGACCTTG ACCGGTGCAC CACTTTTGAT
  21500      21510      21520      21530      21540      21550      21560

                > < AluI
                >< MnlI
GATGTTCAAG CTCCTAATTA CACTCAACAT ACTTCATCTA TGAGGGGGGT TTAATATCCT GATGAAATTT
  21570      21580      21590      21600      21610      21620      21630

>< Sau3AI

```

FIGURE 13. 50

```

>< NdeII
>< MboI
>< DpnII
  >< DpnI          >< Tru9I
>< BspAI          >< MseI > < MboII
  >< Bsp143I      >< DdeI          >< MaeIII
TTAGATCAGA CACTCTTTAT TTAACTCAGG ATTTATTTCT TCCATTTTAT TCTAATGTTA CAGGGTTTCA
  21640      21650      21660      21670      21680      21690      21700

  >< VspI
  >< Tru9I
  >< MseI
  >< AsnI          >< Tru9I          >< FokI
  >< AseI >< MaeII >< MseI >< BbvI > < Fnu4HI
TACTATTAAT CATACGTTTG GCAACCCTGT CATACCTTTT AAGGATGGTA TTTATTTTGC TGCCACAGAG
  21710      21720      21730      21740      21750      21760      21770

          >< BslI
          >< DsaI>< BsiYI          >< NlaIII
          >< BsaJI          > < MaeIII
AAATCAAATG TTGTCCTGG TTGGGTTTTT GGTCTACCA TGAACAACAA GTCACAGTCG GTGATTATTA
  21780      21790      21800      21810      21820      21830      21840

          >< NspI
  >< Tru9I          >< NspHI
  >< MseI          >< NlaIII
  >< HphI          >< MaeIII          >< MaeIII
TTAACAATTC TACTAATGTT GTTATACGAG CATGTAACCT TGAATTGTGT GACAACCCTT TCTTTGCTGT
  21850      21860      21870      21880      21890      21900      21910

  >< StyI          >< Zsp2I
  >< NlaIII          >< Tru9I
  >< NcoI >< RsaI          >< Ppu10I TthHB8I ><
  >< EcoT14I          >< NsiI          >< TaqI
  >< Eco130I          >< MseI          SfaNI ><
  >< DsaI>< Csp6I          >< Mph1103I RsaI ><
  >< BssT1I          >< TthHB8I >< EcoT22I Csp6I ><
  >< BsaJI>< AfaI          >< TaqI >< AvaIII AfaI ><
TTCTAAACCC ATGGGTACAC AGACACATAC TATGATATTC GATAATGCAT TTAATTGCAC TTTCGAGTAC
  21920      21930      21940      21950      21960      21970      21980

          >< Tru9I
          >< MseI
          >< DraI
ATATCTGATG CCTTTTCGCT TGATGTTTCA GAAAAGTCAG GTAATTTTAA ACACTTACGA GAGTTTGTGT
  21990      22000      22010      22020      22030      22040      22050

          >< Sau3AI
          >< NdeII
          >< MboI
  >< Tru9I          >< DpnII
  >< MseI          >< DpnI
  >< DraI          >< BspAI
          >< SfcI Bsp143I ><
TTAAAAATAA AGATGGGTTT CTCTATGTTT ATAAGGGCTA TCAACCTATA GATGTAGTTC GTGATCTACC
  22060      22070      22080      22090      22100      22110      22120

          >< Tru9I
  >< Tru9I          > < Tru9I          >< MseI
  >< MseI          > < MseI          >< MnlI
TTCTGGTTTT AACACTTTGA AACCTATTTT TAAGTTGCCT CTTGGTATTA ACATTACAAA TTTTAGAGCC
  22130      22140      22150      22160      22170      22180      22190

```

FIGURE 13.51

```

> < SduI>< SfcI
    >< PvuII
    >< Psp5I
> < NspII
    >< NspBII
> < MaeII > < Fnu4HI
> < Bsp1286I >< PstI           Tru9I >
    >< BmyI>< Fnu4HI           MseI >
    >< BspMI
    >< BbvI           >< AluI           >< BbvI
    >< HphI
ATTCTTACAG CCTTTTCACC TGCTCAAGAC ATTTGGGGCA CGTCAGCTGC AGCCTATTTT GTTGGCTATT
    22200      22210      22220      22230      22240      22250      22260

    >< SfaNI
    >< RsaI
    > < Csp6I
    >< AfaI           >< AlwNI
>< DraI
TAAAGCCAAC TACATTTATG CTCAAGTATG ATGAAAATGG TACAATCACA GATGCTGTTG ATTGTTCTCA
    22270      22280      22290      22300      22310      22320      22330

    > < Tru9I
    > < MseI
    >< AluI
AAATCCACTT GCTGAACTCA AATGCTCTGT TAAGAGCTTT GAGATTGACA AAGGAATTTA CCAGACCTCT
    22340      22350      22360      22370      22380      22390      22400

    >< SauI
    >< MstII
    >< Eco81I
    >< DdeI
    >< CvnI
    >< Bsu36I
    >< Bse21I
    >< AxyI           >< TfiI
    >< MnlI           >< AocI   >< MnlI   >< HinfI   >< SspI           >< MnlI
AATTCAGGG TTGTTCCCTC AGGAGATGTT GTGAGATTCC CTAATATTAC AAACCTGTGT CCTTTTGGAG
    22410      22420      22430      22440      22450      22460      22470

    >< Zsp2I
    >< Ppu10I
    >< NsiI
    > < NlaIII
    >< Mph1103I
    >< EcoT22I
    >< AvaIII
    >< Tru9I
    >< MseI
AGGTTTTTAA TGCTACTAAA TTCCCTTCTG TCTATGCATG GGAGAGAAAA AAAATTTCTA ATTGTGTTGC
    22480      22490      22500      22510      22520      22530      22540

    >< SduI
    >< NspII
    >< HgiAI
    >< Bsp1286I
    >< BmyI           >< Tru9I
    >< Alw21I       >< MseI           DdeI ><
TGATTACTCT GTGCTCTACA ACTCAACATT TTTTCAACC TTTAAGTGCT ATGGCGTTTC TGCCACTAAG
    22550      22560      22570      22580      22590      22600      22610

    >< Sau3AI
    >< NdeII
    >< MboI
    >< DpnII
    >< DpnI

```

FIGURE 13.52

```

    >> BspAI                >> TfiI
      >> Bsp143I           >> HinfI
TTGAATGATC TTTGCTTCTC CAATGTCTAT GCAGATTCTT TTGTAGTCAA GGGAGATGAT GTAAGACAAA
22620      22630      22640      22650      22660      22670      22680

    >> ScrFI
      >> MvaI
>> HinPII
>> Hin6I
  >> HhaI
    >> HaeII
  >> EcoRII
    >> Ecl136I
  >> DsaV
  >> CfoI
    >> BstOI
    >> BstNI
  >> Bsp143II
  >> BsiLI
  >> ApyI          > < BsrI                                >< NlaIII
TAGCGCCAGG ACAAACGGT GTTATTGCTG ATTATAATTA TAAATTGCCA GATGATTCA TGGGTTGTGT
22690      22700      22710      22720      22730      22740      22750

          >> SfaNI
          >> RmaI
          >> MaeI
CCTTGCTTGG AATACTAGGA ACATTGATGC TACTTCAACT GGTAAATTATA ATTATAAATA TAGGTATCTT
22760      22770      22780      22790      22800      22810      22820

          >> BsrI                                >> OdeI ><
          >> BfrI ><

    >> Sau96I
      >> Pali
      >> NspIV
  > < HindIII
    >> HaeIII
    >> Eco0109I
    >> DraII
  >> DdeI
    >> Cfr13I
    >> BsuRI
    >> BsiZI
    >> BshI
  >> BfrI >> Pssi
  >> NlaIII >> AsuI >< BsmAI
    >> AluI          >> Alw26I                                BspWI ><
AGACATGGCA AGCTTAGGCC CTTGAGAGA GACATATCTA ATGTGCCTTT CTCCCCTGAT GGCAAACCTT
22830      22840      22850      22860      22870      22880      22890

          >> Tru9I
          >> Pali
          >> MscI
          >> HaeIII
        >> EaeI >< MseI
    >> Tru9I          >> BsuRI
    >> MseI          >> BshI
    >> BspMI        >> Bali                                BsrI ><
GCACCCACC TGCTCTTAAT TGTTATTGGC CATTAAATGA TTATGGTTTT TACACCACTA CTGGCATTGG
22900      22910      22920      22930      22940      22950      22960

          >> Sau96I ><
          >> PalINspIV ><
  > < MspI  NspHII ><
    >> HaeIII

```

FIGURE 13.53

```

> < HpaII Eco47I ><
  >< DsaI
> < HapII Cfr13I ><
  >< BsuRISinI ><
  >< GdiII BsiZI ><
    >< BsaJI
    >< ScaI
    >< RsaI
  >< Csp6I
    >< AfaI
CTACCAACCT TACAGAGTTG TAGTACTTTC TTTTGAACCT TTAAATGCAC CGGCCACGGT TTGTGGACCA
  22970      22980      22990      23000      23010      23020      23030
    >< Tru9I
    >< RsaI
  >< Tru9I >< EaeI Bme18I ><
  >< MseI >< Cfr10I AvaII ><
    >< DraI >< BshI AsuI ><
AAATTATCCA CTGACCTTAT TAAGAACCAG TGTGTCAATT TTAATTTTAA TGGACTCACT GGTACTGGTG
  23040      23050      23060      23070      23080      23090      23100
    >< Tru9I
    >< MseI >< BsrI
  >< MseI >< BsrI
  >< Tru9I
  >< MseI >< BsrI
  >< MseI >< HinfI >< AfaI
AAATTATCCA CTGACCTTAT TAAGAACCAG TGTGTCAATT TTAATTTTAA TGGACTCACT GGTACTGGTG
  23040      23050      23060      23070      23080      23090      23100
  >< Tru9I
  >< MseI
  >< MboII
  >< HpaI
  >< HindII
  >< HincII
  >< Pali
  >< HaeIII
  >< GdiII
  >< EaeI
  >< BsuRI
  >< BshI
  TfiI ><
  HinfI ><
TGTTAACTCC TTCTTCAAAG AGATTTCAAC CATTTCACA APTTGGCCGT GATGTTTCTG ATTTCACTGA
  23110      23120      23130      23140      23150      23160      23170
  >< XhoII
  >< TthHB8I
  >< TaqI
  >< Sau3AI
  >< NdeII
  >< MflI
  >< MboI
  >< DpnII
  >< DpnI
  >< BstYI
  >< BspAI
  >< SspI
  >< AlwI >< Bsp143I
  >< HphI
TTCCGTTCGA GATCCTAAAA CATCTGAAAT ATTAGACATT TCACCTTGCT CTTTTGGGGG TGTAAGTGTA
  23180      23190      23200      23210      23220      23230      23240
  >< ScrFI
  >< MvaI
  >< EcoRII
  >< Ecl136I
  >< DsaV
  >< BstOI
  >< BstNI
  >< BsiLI
  >< ApyI
  >< Tru9I
  >< MseI
  >< HpaI
  >< HindII
  >< Eco57I
  >< BsgI >< HincII
ATTACACCTG GAACAAATGC TTCATCTGAA GTTGCTGTTC TATATCAAGA TGTTAACTGC ACTGATGTTT
  23250      23260      23270      23280      23290      23300      23310
  >< Sau3AI
  >< NlaIII
  >< NdeII
  >< MboI
  >< DpnII
  >< DpnI
  >< HinPII

```

FIGURE 13. 54


```

                >> BspWI
                >< BspAI
                >< SfcI
                >> Bsp143I
                >< AluI> < CfoI
                >< PleI ><
                >< BsrI
CTACAGCAAT TCATGCAGAT CAACTCACAC CAGCTTGGCG CATATATTCT ACTGGAAACA ATGTATTCCA
    23320      23330      23340      23350      23360      23370      23380

                >< TthHB8I
                >< TaqI
                >< SalI
                >< RtrI
                >< NspI
                >< EspI >< NspHI
                >< DdeI >< NlaIII
                >< CelII >< HindII
                >< Bpu1102I>< HincII
>< HinfI
GACTCAAGCA GGCTGTCTTA TAGGAGCTGA GCATGTCGAC ACTTCTTATG AGTGGCACAT TCCTATTGGA
    23390      23400      23410      23420      23430      23440      23450

                > < SnaBI
                >< ScaI
                >< RsaI
                >< RmaI
                >< MaeII >< MaeI
                > < Eco105I
                >< Csp6I
                >< BsaAI
                >< AfaI
>< AluI
GCTGGCATTT GTGCTAGTTA CCATACAGTT TCTTTATTAC GTAGTACTAG CCAAAAATCT ATTGTGGCTT
    23460      23470      23480      23490      23500      23510      23520

                >< RmaI
                >< MaeIII
                >< MaeI
                >< AfaI
                >< AfaI
>< AluI
GCTGGCATTT GTGCTAGTTA CCATACAGTT TCTTTATTAC GTAGTACTAG CCAAAAATCT ATTGTGGCTT
    23460      23470      23480      23490      23500      23510      23520

                >< MunI
ATACTATGTC TTTAGGTGCT GATAGTTCAA TTGCTTACTC TAATAACACC ATTGCTATAC CTAATAACTT
    23530      23540      23550      23560      23570      23580      23590

                >< RsaI ><
                >< MnlI
                >< Csp6I ><
                >< AfaI ><
                >< SfcI
TTCAATTAGC ATTACTACAG AAGTAATGCC TGTTTCTATG GCTAAAACCT CCGTAGATTG TAATATGTAC
    23600      23610      23620      23630      23640      23650      23660

                > < TfiI
                > < HinfI
                >< AciI
                > < AluI
ATCTGCGGAG ATTCTACTGA ATGTGCTAAT TTGCTTCTCC AATATGGTAG CTTTTCGACA CAACTAAATC
    23670      23680      23690      23700      23710      23720      23730

>< VneI
    >< SduI
    >< NspII
    >< HgiAI
>< SnoI>< DdeI
    >< Bsp1286I
    >< BmyI
    >< BbvI
>< ApaLI
>< Alw44I
    >< Alw21I
    >< Fnu4HI
    >< BspAI
    >< AflIII
    >< PmlI
    >< PmaCI
    >< MaeII
    >< Eco72I
    >< BsaAI
    >< BbrPI
    >< DpnI
    >< DpnII >< AlwI
    >< Bsp143I >< BbrPI
    >< BspAI >< AflIII
GTGCACTCTC AGGTATTGCT GCTGAACAGG ATCGCAACAC ACGTGAAGTG TTCGCTCAAG TCAAACAAAT
    23740      23750      23760      23770      23780      23790      23800

```

FIGURE 13.55

```

>< RsaI
>< Csp6I
>< AfaI
GTACAAAACC CCAACTTTGA AATATTTTGG TGGTTTTAAT TTTTCACAAA TATTACCTGA CCCTCTAAAG
23810 23820 23830 23840 23850 23860 23870

>< MnlI
>< MnlI
>< DdeI >< MnlI
CCAAC TAAGA GGTCTTTTAT TGAGGACTTG CTCTTTAATA AGGTGACACT CGCTGATGCT GGCTTCATGA
23880 23890 23900 23910 23920 23930 23940

>< XhoII
>< Sau3AI
>< StyI
>< RmaI
>< MaeI
>< EcoT14I
>< Eco130I
>< BssT1I >< VspI
>< BsmI
>< BscCI
>< BsaJI
>< BlnI
>< AvrII
>< XhoII
>< Sau3AI
>< RmaI
>< NdeII
>< MflI
>< MboI
>< MaeI
>< DpnII
>< HphI> < DpnI
>< BstYI
>< BspAI
> < Bsp143I
>< BglII
>< MstI
>< HinPII
>< Hin6I
>< HhaI
>< FspI
>< FdiII
>< CfoI
>< AviII
AGCAATATGG CGAATGCCTA GGTGATATTA ATGCTAGAGA TCTCATTTGT GCGCAGAAGT TCAATGGACT
23950 23960 23970 23980 23990 24000 24010

>< RmaIRsaI ><
>< MnlI
>< Fnu4HI
>< Fnu4HI Csp6I ><
>< BspWI >< BbvI
>< BbvI >< BspWI >< MaeIAfaI ><
TACAGTGTG CCACCTCTGC TCACTGATGA TATGATTGCT GCCTACACTG CTGCTCTAGT TAGTGGTACT
24020 24030 24040 24050 24060 24070 24080

>< MboII
>< HinPII
>< Hin6I
>< HhaI
>< HaeII
>< Fnu4HI >< Ksp632I
>< CfoI >< EarI
>< FokI >< BspWI >< Eam1104I
>< BbvI
>< Bsp143II
GCCACTGCTG GATGGACAT TGGTGCTGGC GCTGCTCTTC AAATACCTTT TGCTATGCAA ATGGCATATA
24090 24100 24110 24120 24130 24140 24150

Tru9I ><
MseI ><
>< MaeIII
GGTTCAATGG CATTGGAGTT ACCCAAATG TTCTCTATGA GAACCAAAAA CAAATCGCCA ACCAATTTAA
24160 24170 24180 24190 24200 24210 24220

MaeII ><
>< Fnu4HI
>< TfiI
>< HinfI
>< BbvI
>< AluI
CAAGGCGATT AGTCAAATC AAGAATCACT TACAACAACA TCAACTGCAT TGGGCAAGCT GCAAGACGTT
24230 24240 24250 24260 24270 24280 24290

>< Tru9I
>< MseI
>< HpaI
>< HindII >< BsmI >< Tru9I
>< HincII>< BscCI >< MseI
>< DdeI
>< Tru9I >< BfrI
>< MseI >< AluI

```

FIGURE 13. 56

```

GTTAACCAGA ATGCTCAAGC ATTAACACA CTTGTAAAC AACTTAGCTC TAATTTGGT GCAATTTCAA
 24300      24310      24320      24330      24340      24350      24360

      >< ThaI
      >< SpoI
      >< NruI
      >< MvnI
      >< BstUI          >< TthHB8I
      >< Bsp68I       >< TaqI          >< RsaI
    >< EcoRV >< Bsp50I   >< MnlI          >< Csp6I          >< Tru9I
    >< Eco32I >< AccII >< MnlI      >< AciI>< AfaI      >< MseI
GTGTGCTAAA TGATATCCTT TCGCGACTTG ATAAAGTCGA GGCGGAGGTA CAAATTGACA GGTAAATTAC
 24370      24380      24390      24400      24410      24420      24430

      >< MaeIII >< BbvI          >< Fnu4HI   BbvI ><
AGGCAGACTT CAAAGCCTTC AACCTATGT AACACAACAA CTAATCAGGG CTGCTGAAAT CAGGGCTTCT
 24440      24450      24460      24470      24480      24490      24500

      >< Fnu4HI          >< HindII
    >< BspWI          >< DdeI          >< HincII
GCTAATCTTG CTGCTACTAA AATGTCTGAG TGTGTTCTTG GACAATCAAA AAGAGTTGAC TTTGTGGAA
 24510      24520      24530      24540      24550      24560      24570

      > < NspI
      > < NspHI
      > < NlaIII
      >< MaeIII
      >< NlaIII
      >< MboII          >< FokI
    >< Fnu4HI >< BbsI          BsaAI ><
      >< AciI>< BbvI          >< AflIII
AGGGCTACCA CTTTATGTCC TTCCCACAAG CAGCCCCGCA TGGTGTGTC TTCCTACATG TCACGTATGT
 24580      24590      24600      24610      24620      24630      24640

      >< ScrFI
      >< MvaI
    >< EcoRII
      >< Ecl136I
      >< BstOI
      >< BstNI          >< HinPI
    >< MnlI >< BslI          >< Hin6I
    >< DsaV>< BsiYI          >< HhaI
      >< BsiLI          >< HaeII
    >< BsaJI>< HphI          >< CfoI          >< NlaIII
      >< ApyI          >< Bsp143II >< BspHI          EcoNI ><
GCCATCCCAG GAGAGGAACT TCACCACAGC GCCAGCAATT TGTCATGAAG GCAAAGCATA CTTCCCTCGT
 24650      24660      24670      24680      24690      24700      24710

      >< MnlI
    >< BslI          >< Tru9I
    >< BsiYI          >< MseI          >< MnlI
GAAGGTGTTT TTGTGTTTAA TGGCACTTCT TGGTTTATTA CACAGAGGAA CTTCTTTTCT CCACAAATAA
 24720      24730      24740      24750      24760      24770      24780

      >< DdeI          >< Tru9I
      >< BsmAI          >< SfaNI
    >< SfcI          >< Alw26I          >< MseIAlwI ><
TTACTACAGA CAATACATT GTCTCAGGAA ATTGTGATGT CGTTATGGC ATCATTAAACA ACACAGTTTA
 24790      24800      24810      24820      24830      24840      24850

>< Sau3AI
>< NdeII

```

FIGURE 13.57

```

>< MboI           >< P1eI           > < ScaI
>< DpnII          >< MnlI           > < Ksp632I       > < RsaI
  >< DpnI          >< DdeI >< HinfI       >< MboII
>< BspAI          >< BspWI          > < Eam1104I      >< Csp6I
  >< Bsp143I       >< AluI           > < EarI > < AluI > < AfaI > < HphI
TGATCCTCTG CAACCTGAGC TTGACTCATT CAAAGAAGAG CTGGACAAGT ACTTCAAAAA TCATACATCA
  24860      24870      24880      24890      24900      24910      24920

  >< Sau3AI
  >< NdeII
  >< MboI
>< MamI
  >< DpnII
  >> DpnI
  >< BspAI
  >< Bsp143I
  >< BsiBI           >< Tru9I           >< HindII
  >< BsaBI           >< MseI           >< HincII       AciI ><
CCAGATGTTG ATCTTGCGCA CATTTCAGGC ATTAACGCTT CTGTCGTCAA CATTCAAAAA GAAATTGACC
  24930      24940      24950      24960      24970      24980      24990

  >< Tru9I
  > < TfiI
  >< MnlI           >< SwaI
  >< EcoNI          >< MseI
  >< BslI           > < HinfI
>< MnlI>< BsiYI       >< DraI
GCCTCAATGA GGTGCTAAA AATTAAATG AATCACTCAT TGACCTCAA GAATTGGGAA AATATGAGCA
  25000      25010      25020      25030      25040      25050      25060

  >< StyI
  >< Pali
  >< HaeIII
  >< EcoT14I
  >< Eco130I
  >< BsuRI
  >< BssTII
  >< Tru9I>< BshI           NlaIII ><
  >< MseI >< BsaJI           MaeIII ><
  >> BstXI
ATATATATAA TGGCCTTGGT ATGTTTGGCT CGGCTTCATT GCTGGACTAA TTGCCATCGT CATGGTTACA
  25070      25080      25090      25100      25110      25120      25130

  > < SphI
  > < PaeI
  >< SpeI           > < NspI
  > < RmaI         > < NspHI
  >< NlaIII       > < NlaIII
  > < MaeI         >< MnlI>< BbvI Fnu4HI ><
ATCTTGCTTT GTTGCATGAC TAGTTGTTGC AGTTGCCTCA AGGTCATG CTCTTGTTGGT TCTTGCTGCA
  25140      25150      25160      25170      25180      25190      25200

  >< FokI
  >> DdeI
>< MnlI >< P1eI>< HinfI >< BsrI
AGTTTGATGA GGATGACTCT GAGCCAGTTC TCAAGGGTGT CAAATTACAT TACACATAAA CGAACTTATG
  25210      25220      25230      25240      25250      25260      25270

  >< Sau3AI
  >< NdeII
  >< MboI
  >< DpnII
  > < DpnI

```

FIGURE 13.58

```

                << BspAI
                > < BspI43I
                << BsgI                << AlwI                << BsrI                BspWI >
GATTTGTTTA TGAGATTTT TACTCTTGGG TCAATTACTG CACAGCCAGT AAAAATTGAC AATGCTTCTC
25280      25290      25300      25310      25320      25330      25340

                << ScaI
                << RsaI
                << Csp6I                << SfcI
                << AfaI                << NlaIII                << AciI                << MnlI                FokI >
CTGCAAGTAC TGTTTCATGCT ACAGCAACGA TACCGCTACA AGCCTCACTC CCTTTCGGAT GGCTTGTTAT
25350      25360      25370      25380      25390      25400      25410

                > < HinPII
                > < Hin6I
                << HhaI                RmaI >>
                << HaeII                << HinPII                NheI >>
                << Eco47III                << Hin6I                MaeI >>
                << CfoI                << HhaI                Fnu4HI >>
                << BspWI                << BspI43II                << CfoI                AluI >>
TGGCGTTGCA TTCTTGCTG TTTTCAGAG CGCTACCAA ATAATTGCGC TCAATAAAG ATGGCAGCTA
25420      25430      25440      25450      25460      25470      25480

                << EcoNI
                << BslI
                << BsiYI                << MaeIII
                << BbvI                << BsrI                << BbvI                > < Fnu4HI                BbvI >>
GCCCTTTATA AGGGCTTCCA GTTCATTTGC AATTTACTGC TGCTATTTGT TACCATCTAT TCACATCTTT
25490      25500      25510      25520      25530      25540      25550

                << SfcI                << HinPII
                << PstI                << Hin6I                << RsaI                Zsp2I >>
                > < Fnu4HI                << HhaI                << Csp6I                Ppu10I >>
                << BspMI                << MnlI                << CfoI                << AfaI                << MnlI                NsiI >>
                << BspMI                << MnlI                << CfoI                << AfaI                << MnlI                Mph1103I >>
TGCTTGTCGC TGCAGGTATG GAGGCGCAAT TTTGTACCT CTATGCCTTG ATATATTTTC TACAATGCAT
25560      25570      25580      25590      25600      25610      25620
                << MnlI                AvaIII >>

                << SfaNI
                << NspI
                << NspHI
                << NlaIII                << SfaNI
CAACGCATGT AGAATTATTA TGAGATGTTG GCTTTGTTGG AAGTGCAAAT CCAAGAACCC ATTACTTTAT
25630      25640      25650      25660      25670      25680      25690

                << Bst1107I
                << AccI                MaeIII >>
GATGCCAACT ACTTTGTTTG CTGGCACACA CATAACTATG ACTACTGTAT ACCATATAAC AGTGTCACAG
25700      25710      25720      25730      25740      25750      25760

                << MboII
                << HphI                BstXI >>
                << MunI >> MaeIII << MaeIII                << Eco57I                << BbsI MnlI >
ATACAATTGT CGTTACTGAA GGTGACGGCA TTTCAACACC AAAACTCAAA GAAGACTACC AAATTGGTGG
25770      25780      25790      25800      25810      25820      25830

                << RsaI
                > < NlaIII
                << HphI
                << Tru9I >> Tth111I << Csp6I
                << DdeI                << DdeI                << MseI >> AspI                << AfaI

```

FIGURE 13.59

```

TTATTCTGAG GATAGGCACT CAGGTGTTAA AGACTATGTC GTTGTACATG GCTATTTTCAC CGAAGTTTAC
25840      25850      25860      25870      25880      25890      25900

                > < HinFI >< PleI
                >> BsrI
                Tru9I ><
                MseI ><
>< AluI >< AccI >> SfcI >< AlwNI >> MboII HindIII >
TACCAGCTTG AGTCTACACA AATTACTACA GACTACTGGTA TTGAAAATGC TACATTCTTC ATCTTTAACA
25910      25920      25930      25940      25950      25960      25970

                > < TthHB8I
                >< Tru9I >> TaqI >< Ksp632I
                >< MseI >> MboII >< EarI BspWI ><
>< AluI >> Eco57I >< Eam1104I AlwI ><
AGCTTGTTAA AGACCCACCG AATGTGCAAAA TACACACAAT CGACGGCTCT TCAGGAGTTG CTAATCCAGC
25980      25990      26000      26010      26020      26030      26040

>< XhoII
>< Sau3AI
>< NlaIV
>< NdeII
>< MflI
>< MboI
>< DpnII
>> DpnI
>< BstYI
>< BstI
>< BspAI
>< Bsp143I
>< BscBI
>< BamHI >< AlwI
AATGGATCCA ATTTATGATG AGCCGACGAC GACTACTAGC GTGCCTTTGT AAGCACAAGA AAGTGAGTAC
26050      26060      26070      26080      26090      26100      26110

                > < Tru9I
                >< RsaI
                > < MseI
                >< MboII
                >< MaeII
                >< RsaI
>< Csp6I >> Csp6I >< Tru9I >< Csp6I
> < AfaI >> AfaI >< MseI >< AfaI
GAACTTATGT ACTCATTCGT TTCGGAAGAA ACAGGTACGT TAATAGTTAA TAGCGTACTT CTTTTTCTTG
26120      26130      26140      26150      26160      26170      26180

                >< TthHB8I
                >< TaqI
                >< RmaI
                >< HinPII > < RsaI
                > < MaeIII >> Hin6I Fnu4HI ><
                >< MaeI >> RmaI >< HhaI >< Csp6I
                >< FokI >> MaeI >< CfoI >< BbvI > < AfaI
CTTTCGTGGT ATTCTTGCTA GTCACACTAG CCATCCTTAC TGCCTTCGA TTGTGTGCGT ACTGCTGCAA
26190      26200      26210      26220      26230      26240      26250

                >< Tru9I
                >< Tru9I
                >< MseI
                >< MvnI
>< SspI >< MaeII
                >< HpaI >> BstUI Ksp632I >
                >< HindII >> MaeII >< Bsp50I >< MboII EarI >
                >< HincII >> AccI >< AccII Eam1104I >
TATTGTAAAC GTGAGTTTAG TAAAACCAAC GGTTTACGTC TACTCGCGTG TAAAAATCT GAACTCTTCT
26260      26270      26280      26290      26300      26310      26320

```

FIGURE 13.60

```

    >> Sau3AI
    >> NdeII
    >> MboI
    >> DpnII
    >> MboII>> DpnI
    >> XmnI >> BspAI> < Eco57I
    >> Asp700I>> Bsp143I
GAAGGAGTTC CTGATCTTCT GGTCTAAACG AACTAACTAT TATTATTATT CTGTTTGGAA CTTTAACATT
26330      26340      26350      26360      26370      26380      26390

    >> ScrFI
    >> MvaI
    >> EcoRII
    >> Ecl136I
    >> DsaV NlaIV >>
    >> BstOI
    >> RsaI
    >> MnlI
    >> Tru9I
    >> BstNI RmaI >>
    >> Csp6I
    >> MseI
    >> BsiLI MaeI >>
    > < NlaIII
    >> AfaI
    > < AluI
    >> ApyIBscBI >>
GCTTATCATG GCAGACAACG GTACTATTAC CGTTGAGGAG CTTAAACAAC TCCTGGAACA ATGGAACCTA
26400      26410      26420      26430      26440      26450      26460

    >> ScrFI
    >> RmaI
    >> MvaI
    >> MaeI
    >> EcoRII
    >> Ecl136I
    >> DsaV
    >> BstOI
    >> BstNI
    >> BsiLI
    >> ApyI >> MaeIII
GTAATAGGTT TCCTATTCCT AGCCTGGATT ATGTTACTAC AATTTGCCTA TTCTAATCGG AACAGGTTTT
26470      26480      26490      26500      26510      26520      26530

    >> Pali
    >> Msci
    >> MnlI >> MaeIII
    >> HaeIII
    >> EaeI
    >> BsuRI
    >> BsrI
    >> RsaI
    >> Csp6I >> HindIII
    >> AfaI >> AluI
    >> BspWI
    >> BshI
    >> Ball
    >> BbvI Fnu4HI >>
TGACATAAT AAAGCTTGTT TTCCTCTGGC TCTTGTGGCC AGTAACACTT GCTTGTTTTG TGCTTGCTGC
26540      26550      26560      26570      26580      26590      26600

    >> VspI
    >> Tru9I
    >> MseI
    >> SfcI >> AsnI
    >> HphI
    >> BsrI
    >> AccI >> AseI>> MaeIII>> AciI
TGCTACAGA ATTAATTGGG TGACTGGCGG GATTGCGATT GCAATGGCTT GTATTGTAGG CTTGATGTGG
26610      26620      26630      26640      26650      26660      26670

    >> EspI
    >> Eco57I
    >> DdeI
    >> CelII
    >> Epu1102I
    >> RsaI
    >> Csp6I

```

FIGURE 13.61


```

CCACGCCGGT AGCAACGACA ATATTGCTTT GCTAGTACAG TAAGTGACAA CAGATGTTTC ATCTTGTTGA
 27030      27040      27050      27060      27070      27080      27090

    << ScrFI
    << MvaI
      << MaeIII
    << EcoRII
    << Ecl136I
    << DsaV
    << BstOI
    << BstNI
    << BsiLI
    << ApyI
    << MnlI
    << TfiI
    << HinFI >>
CTTCCAGGTT ACAATAGCAG AGATATTGAT TATCATTATG AGGACTTTC A GGATTGCTAT TTGGAATCTT
 27100      27110      27120      27130      27140      27150      27160

    << MaeII
    << BsmAI
    << Tru9I
    << MnlI
    << Alw26I
    << MseI
    << DdeI
    << MboII
GACGTTATAA TAAGTTCAAT AGTGAGACAA TTATTTAAGC CTCTAACTAA GAAGAATTAT TCGGAGTTAG
 27170      27180      27190      27200      27210      27220      27230

    << Ksp632I
    << MboII
    << EarI
    << MboII
    << NlaIII Eam1104I >>
ATGATGAAGA ACCTATGGAG TTAGATTATC CATAAAACGA ACATGAAAAT TATTCTCTTC CTGACATTGA
 27240      27250      27260      27270      27280      27290      27300

    << RsaI >> RsaI
    << Csp6I >> Csp6I
    << AluI
    << MnlI
    << AfaI >> AfaI
TTGTATTTAC ATCTTGCGAG CTATATCACT ATCAGGAGTG TGTTAGAGGT ACGACTGTAC TACTAAAAGA
 27310      27320      27330      27340      27350      27360      27370

    << MnlI
    << HphI
    << HphI
    << MnlI
ACCTTGCCCA TCAGGAACAT ACGAGGGCAA TTCACCATTT CACCCTCTTG CTGACAATAA ATTTGCACTA
 27380      27390      27400      27410      27420      27430      27440

    << RmaI
    << MaeI
    << TthHB8I
    << TaqI
    << RsaI
    << Csp6I
    << BbvI
    << AfaI
    << AluI
    << Sau3AI >
    << PvuII
    << Psp5I
    << NspBII
    << NdeII >
    << MboI >
    << Fnu4HI
    << DpnII >
    << BspAI >
    << RmaI
    << MaeI
    << AfaI
    << AluI
ACTTGCACTA GCACACACTT TGCTTTTGCT TGTGCTGACG GTACTCGACA TACCTATCAG CTGCGTGCAA
 27450      27460      27470      27480      27490      27500      27510

    << SstI
    << SduI
    << SacI
    << NspII
    << HgiAI
    << Eco24I
    << Ecl136II
    << BspWI
    << Bsp1286I
    << BmyI
    << BanII
    << Alw21I
    << HphI
    << DpnI
    << MnlI

```

FIGURE 13. 63

```

>< Bsp143I          >< MnlI          > < AluI      BbvI ><
GATCAGTTTC ACCAAAACCTT TTCATCAGAC AAGAGGAGGT TCAACAAGAG CTCTACTCGC CACTTTTTCT
27520          27530          27540          27550          27560          27570          27580

SstI ><
SduI ><
SacI ><
NspII ><
HgiAI ><
Eco24I ><
Ecl136II ><
Bsp1286I ><
BmyI ><
BanII ><
Alw21I ><
AluI ><
>< Fnu4HI          >< RmaI          >< Tru9I
GCTCTAGTAT TTTAATACT TTTAATACT TTGCTTCACC ATTAAGAGAA AGACAGAATG AATGAGCTCA
27590          27600          27610          27620          27630          27640          27650

>< Tru9I          >< Tru9I
>< MseI          >< MseI
CTTTAATTGA CTTCTATTTG TGCTTTTTAG CCTTTCGCT ATTCCTTGTT TTAATAATGC TTATTATATT
27660          27670          27680          27690          27700          27710          27720

>< XhoII
>< XbaI
> < ScrFI
>< Sau3AI
>< RmaI
>< NdeII
> < MvaI
>< MflI
>< MboI
>< EcoRII>< MaeI
> < Ecl136I
>< DpnII
>< DpnI
>< BstYI
> < BstOI
> < BstNI
>< TthHB8I >< BspAI          > < RsaI
>< DsaV>< Bsp143I          >< MboII
> < BsiLI          >< Csp6I
>< TaqI > < ApyI > < AlwI > < AfaI          >< NlaIII
TTGGTTTTCA CTCGAAATCC AGGATCTAGA AGAACCTTGT ACCAAAGTCT AAACGAACAT GAAACTTCTC
27730          27740          27750          27760          27770          27780          27790

>< HinP1I
>< Hin6I
>< HhaI
>< RsaI >< HaeII
>< SfcI          >< Eco47III
>< Csp6I>< CfoI SfaNI ><
>< AfaI >< Bsp143II
ATTGTTTTGA CTTGTATTTT TCTATGCAGT TGCATATGCA CTGTAGTACA GCGCTGTGCA TCTAATAAAC
27800          27810          27820          27830          27840          27850          27860

>< XhoII
>< Sau3AI
>< NdeII
> < MnlI
>< MflI

```

FIGURE 13.64

```

    >> MboI
    >> DpnII
      >> DpnI      >> RsaI
    >> BstYI  >> MboII
  >> NlaIII>> BspAI      >> Csp6I >> RmaI
    >> AlwI >> Bsp143I  >> AfaI >> MaeI
CTCATGTGCT TGAAGATCCT TGTAAGGTAC AACACTAGGG GTAATACTTA TAGCACTGCT TGGCTTTGTG
27870      27880      27890      27900      27910      27920      27930

  >> SduI
  >> RmaI
  >> NspII
  >> MaeI
  >> HgiAI
  >> Bsp1286I
  >> BmyI
  >> Alw21I
CTCTAGGAAA GGTTTTACCT TTTCATAGAT GGCACACTAT GGTTCAAACA TGCACACCTA ATGTTACTAT
27940      27950      27960      27970      27980      27990      28000

    > < XhoII
    > < Sau3AI > < Van9I
      >> PvuII
      >> Psp5I
    > < NdeII > < PflMI
    > < MflI>< NspBII
    > < DpnII      >> HinP1I
      >> Bsp143I      >> Hin6I
    > < BstYI > < BslI >> HhaI >> RmaI >> Asp718 >> Eco91I
    > < BspAI > < BsiYI>< CfoI >> MaeI >> AfaI >> BstPI
    > < MboI>< AluI>< BspWI >> BspWI >> AccB1I >> BstEII
  >> AlwI >> DpnI > < AccB7I >> AluI >> Acc65I >> BbvI
CAACTGTCAA GATCCAGCTG GTGGTGCCT TATAGCTAGG TGTTGGTACC TTCATGAAGG TCACCAAAC
28010      28020      28030      28040      28050      28060      28070

    >> SinI
    >> Sau96I
    >> NspIV
  NspHII ><
  NlaIV ><
    >> Eco47I
    >> Cfr13I
    >> BsiZI
  BscBI ><
  >> Esp3I >> Csp6I >> Tru9I >> Bme18I
  >> BsmAI >> BsmBI >> MseI >> Tru9I >> AvaII
  >> Alw26I >> AfaI >> DraI >> MseI >> AsuI
GCTGCATTTA GAGACGTA CTGTTGTTTT AATAAACGAA CAAATTAAAA TGTCTGATAA TGGACCCCAA
28080      28090      28100      28110      28120      28130      28140

    >> SinI
    >> Sau96I
    >> NspIV
      >> NspHII
      >> NlaIV
    >> Eco47I
    >> Cfr13I
    >> BsiZI
      >> BscBI
    >> SduI
    >> NspII
    >> Bsp1286I
    >> BmyI
  >> MaeII >> AciI
    >> SinI
    >> Sau96I
    >> NspIV
      >> NspHII
      >> NlaIV
    >> Eco47I
    >> Cfr13I
    >> BsiZI
      >> BscBI
    >> AvaII >> TfiI
    >> AsuI >> HinfI >> MnlI

```

FIGURE 13. 65

```

TCAAACCAAC GTAGTGCCCC CCGCATTACA TTTGGTGGAC CCACAGATTC AACTGACAAT AACCGAATG
28150      28160      28170      28180      28190      28200      28210

                >> HinP1I >> StyI
                >> HaeII
                > < PalI >> Hin6I >> EcoT14I
                > < HaeIII >> HhaI>> Eco130I
                >> BspWI >> BssT1I
                > < BsuRI >> Bsp143II
                >> HgaI> < BshI >> CfoI>> BsaJI >> HgaI
GAGGACGCAA TGGGGCAAGG CAAAACAGC GCCGACCCCA AGGTTTACCC AATAATACTG CGTCTTGTT
28220      28230      28240      28250      28260      28270      28280

                >> TthHB8I
                > < ScrFI
                >> PalI
                >> PaeR7I
                >> NspIII
                > < MvaI
                >> HaeIII
                >> EcoRII
                >> Eco88I
                >> XhoI > < Ecl136I
                >> DsaV
                >> BsuRI
                >> SlaI > < BstOI
                >> MnlI>> TaqI> < BstNI
                >> CcrI > < BsiLI
                >> HinfI >> BshI
                >> TfiI>> BcoI>> BsaJI
                >> MnlI >> DdeI >> AvaI > < ApyI
                >> AluI >> DdeI > < NlaIII >> BfrI >> Ama87I >> MnlI
CACAGCTCTC ACTCAGCATG GCAAGGAGGA ACTTAGATTC CCTCGAGGCC AGGGCGTTCC AATCAACACC
28290      28300      28310      28320      28330      28340      28350

                >> SinI
                >> Sau96I
                >> NspIV
                >> NspHII
                >> Eco47I
                >> Cfr13I
                >> BsiZI
                >> Bme18I > < Ksp632I
                >> AvaII > < Eam1104I
                >> AsuI > < EarI > < AluI>> MboII >> MaeIII
AATAGTGGTC CAGATGACCA AATTGGCTAC TACCGAAGAG CTACCCGACG AGTTCGTGGT GGTGACGGCA
28360      28370      28380      28390      28400      28410      28420

                >> SstI
                >> SduI
                >> SacI
                >> NspII
                >> HgiAI
                >> EspI
                >> Eco24I >> Sau96I
                >> Ecl136II >> StyI >> PalI
                >> DdeI >> RmaI >> NspIV
                >> CelII >> MaeI >> HaeIII
                >> Bsp1286I >> EcoT14I >> Cfr13I
                >> Bpu1102I >> Eco130I >> BsuRI
                >> BmyI >> BssT1I > < BsrI
                >> BanII >> RsaI >> BsaJI >> BsiZI

```

FIGURE 13.66


```

> < BshI   > < BbvI   >< MnlI >< BspWI   >< SfaNI   AfaI ><
AACAAACAAGG CCAAAGTGC ACTAAGAAAT CTGCTGCTGA GGCATCTAAA AAGCCTCGCC AAAAAAGTAC
28850      28860      28870      28880      28890      28900      28910

                                >< Tth111I
                                >< SinI
                                >< Sau96I
                                >< NspIV
                                >< NspHII
> < MaeII
                                >< Eco47I
                                >< Cfr13I
                                >< BsmBI
                                >< BsiZI   >< StyI
                                >< Bme18I  >< EcoT14I
                                >< AvaII   >< Eco130I
                                >< AsuI    >< BssTII
                                >< Alw26I> < AspI   >< BsaJI
                                >< RsaI
                                >< MaeIII
                                >< MaeII   >< Esp3I
                                >< Csp6I   >< BsmAI
                                >< AfaI    >< Alw26I> < AspI   >< BsaJI
TGCCACAAAA CAGTACAACG TCACTCAAGC ATTTGGGAGA CGTGGTCCAG AACAAACCCA AGGAAATTTT
28920      28930      28940      28950      28960      28970      28980

>< SinI
>< Sau96I
>< NspIV
>< NspHII
>< NlaIV
>< Eco47I
>< Cfr13I
>< BsiZI
>< BscBI
>< Bme18I
>< AvaII
>< AsuI
                                >< Pali
                                >< HaeIII
                                >< GdiI
                                >< Fnu4HI
                                >< EaeI
                                >< BsuRI
                                >< BshI
                                >< AciI
                                >< BspWI >
GGGGACCAAG ACCTAATCAG ACAAGGAACT GATTACAAAC ATTGGCCGCA AATTGCACAA TTTGCTCCAA
28990      29000      29010      29020      29030      29040      29050

                                >< BsmI
                                >< NlaIII
                                >< BscCI >< MnlI >< MaeIII
                                >< MaeIII
                                >< NlaIII
GTGCCTCTGC ATTCTTTGGA ATGTCACGCA TTGGCATGGA AGTCACACCT TCGGGAACAT GGCTGACTTA
29060      29070      29080      29090      29100      29110      29120

                                >< XhoII
                                >< Sau3AI
                                >< NdeII
                                >< MflI
                                >< MboI
                                >< FokI
                                >< Tru9I
                                >< DpnII
                                >< NlaIV
                                >< DpnI
                                >< NlaIII
                                >< BstYI
                                >< Tth111I
                                >< MseI
                                >< BspAI
                                >< MaeII
                                >< BscBI >< BstXI>< AlwI> < Bsp143I
                                >< AspI
                                >< BspWI ><
TCATGGAGCC ATTAAATTGG ATGACAAAGA TCCACAATTC AAAGACAACG TCATACTGCT GAACAAGCAC
29130      29140      29150      29160      29170      29180      29190

                                EspI ><
                                DdeI ><
                                CclII ><
                                Bpu1102I ><
                                AluI ><
                                >< HgaI
ATTGACGCAT AAAAAACATT CCCACCAACA GAGCCTAAAA AGGACAAAAA GAAAAAGACT GATGAAGCTC
29200      29210      29220      29230      29240      29250      29260

```

FIGURE 1368

```

                >> PleI
                >> MboII
    >> Fnu4HI
    >> BspWI
    >> BsmAI
    >> Alw26I
    >> AciI
    >> Fnu4HI
    >> BbvI
    >> AciI
    >> NlaIII
    >> MboII
    >> Ksp632I
    >> GsuI
    >> MaeIII
    >> EarI
    >> Fnu4HI
    >> HinfI
    >> Eam1104I
    >> BpmI
AGCCTTTGCC GCAGAGACAA AAGAAGCAGC CCACTGTGAC TCTTCTTCCT GCGGCTGACA TGGATGATTT
29270      29280      29290      29300      29310      29320      29330

                >> NlaIII
                >> HinfI
                NlaIII >>
    >> FokI
    >> AluI
    >> TfiI
    >> DdeI
    >> BspHI
CTCCAGACAA CTTCAAATTT CCATGAGTGG AGCTTCTGCT GATTCAACTC AGGCATAAAC ACTCATGATG
29340      29350      29360      29370      29380      29390      29400

                >> MaeII
                >> AccI
ACCACACAAG GCAGATGGGC TATGTAAACG TTTTCGCAAT TCCGTTTACG ATACATAGTC TACTCTTGTG
29410      29420      29430      29440      29450      29460      29470

                >> Tru9I
                >> Tru9I
                >> MseI
                >> MseI
    >> XmnI
    >> EcoRI
    >> MaeIII
    >> Asp700I
    >> BsgI
    >> HpaI
    >> HindII
    >> HincII
    >> Tru9I
    >> MseI
CAGAATGAAT TCTCGTAACT AAACAGCACA AGTAGGTTTA GTTAACTTTA ATCTCACATA GCAATCTTTA
29480      29490      29500      29510      29520      29530      29540

                XorII >
                TthHB8I >
                TaqI >
                Sau3AI >
                RsaI >>
                >> ThaIPvuI >
                NdeII >>
                >> MnlI
                >> MvnIMcrI >
                MboI >>
                DpnII >>
                DpnI >>
                Csp6I >>
                >> BstUI
                >> HaeIII
                >> BspCI >
                >> BspAI >>
                >> TthHB8I >>
                >> Bsp50I
                >> Pali
                >> Bsp143I >>
                >> BsuRI
                >> BsiEI >
                >> BshIAfaI >>

    >> MnlI
    >> MaeIII
    >> TaqI
    >> AciI
    >> MnlI
    >> AccII
ATCAATGTGT AACATTAGGG AGGACTTGAA AGAGCCACCA CATTTCATC GAGGCCACGC GGAGTACGAT
29550      29560      29570      29580      29590      29600      29610

                >> SduI
                >> NspII
                >> MboII
                >> VspI
                >> Ksp632I
                >> Eco24I
                >> Tru9I
    >> RsaI
    >> RmaI
    >> Fnu4HI
    >> Bsp1286I
    >> MseI
    >> Csp6I
    >> MaeI
    >> EarI
    >> BmyI
    >> AsnI
    >> AfaI
    >> BbvI
    >> AluI
    >> Eam1104I
    >> BanII
    >> AseI

```

FIGURE 13.69

```
CGAGGGTACA GTGAATAATG CTAGGGAGAG CTGCCTATAT GGAAGAGCCC TAATGTGTAA AATTAATTTT
 29620      29630      29640      29650      29660      29670      29680

                >< Tru9I   >< DdeI
                >< MseI   >< BfrI
                >< NlaIII  > < AluI
AGTAGTGCTA TCCCATGTG ATTTAATAG CTTCTTAGGA GAATGACAAA AAAAAAAAAA AAAAAA
 29690      29700      29710      29720      29730      29740
```


SRAS serology : Indirect N technique(first set)

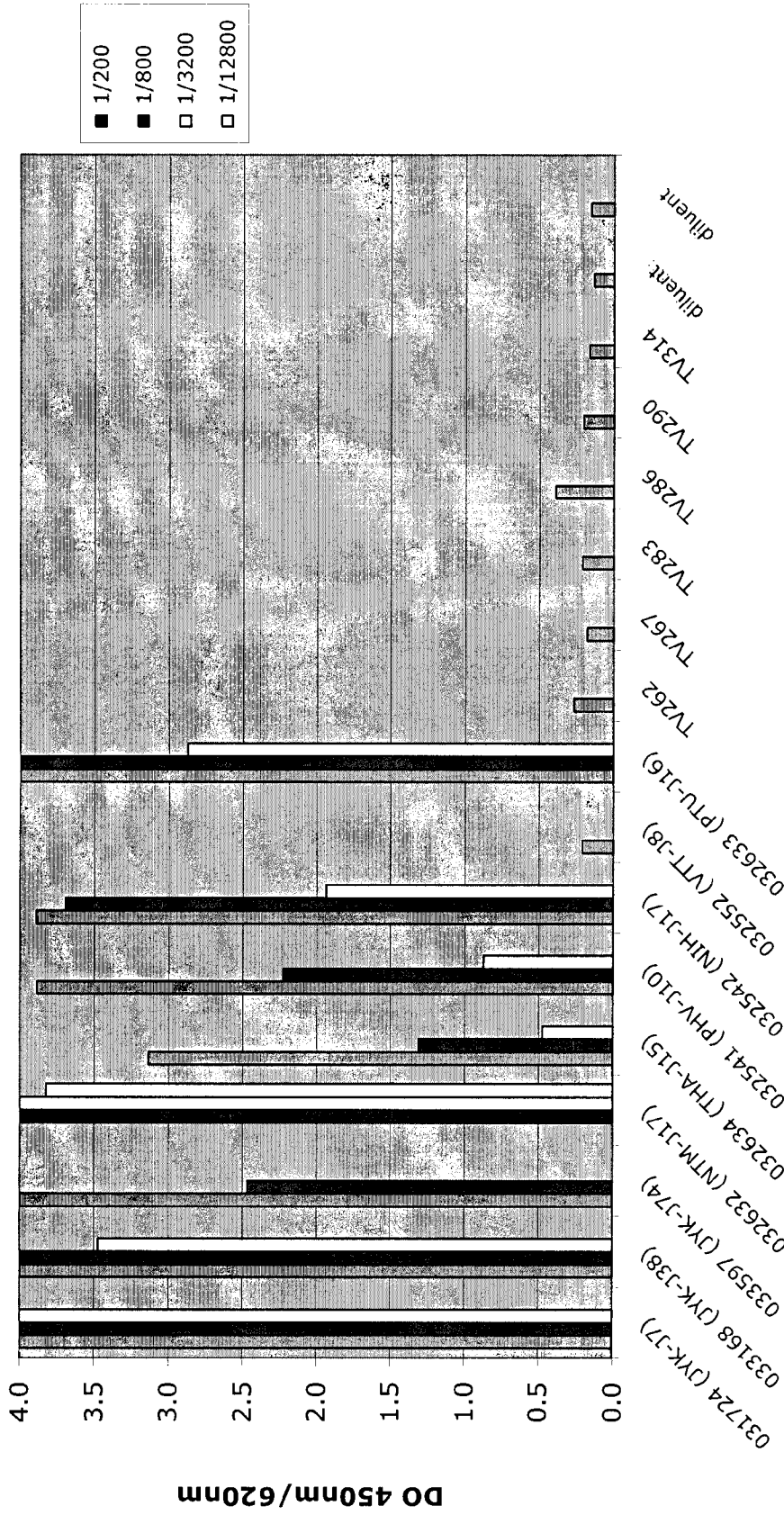


FIGURE 14

SRAS serology : Indirect N Technique (Second set)

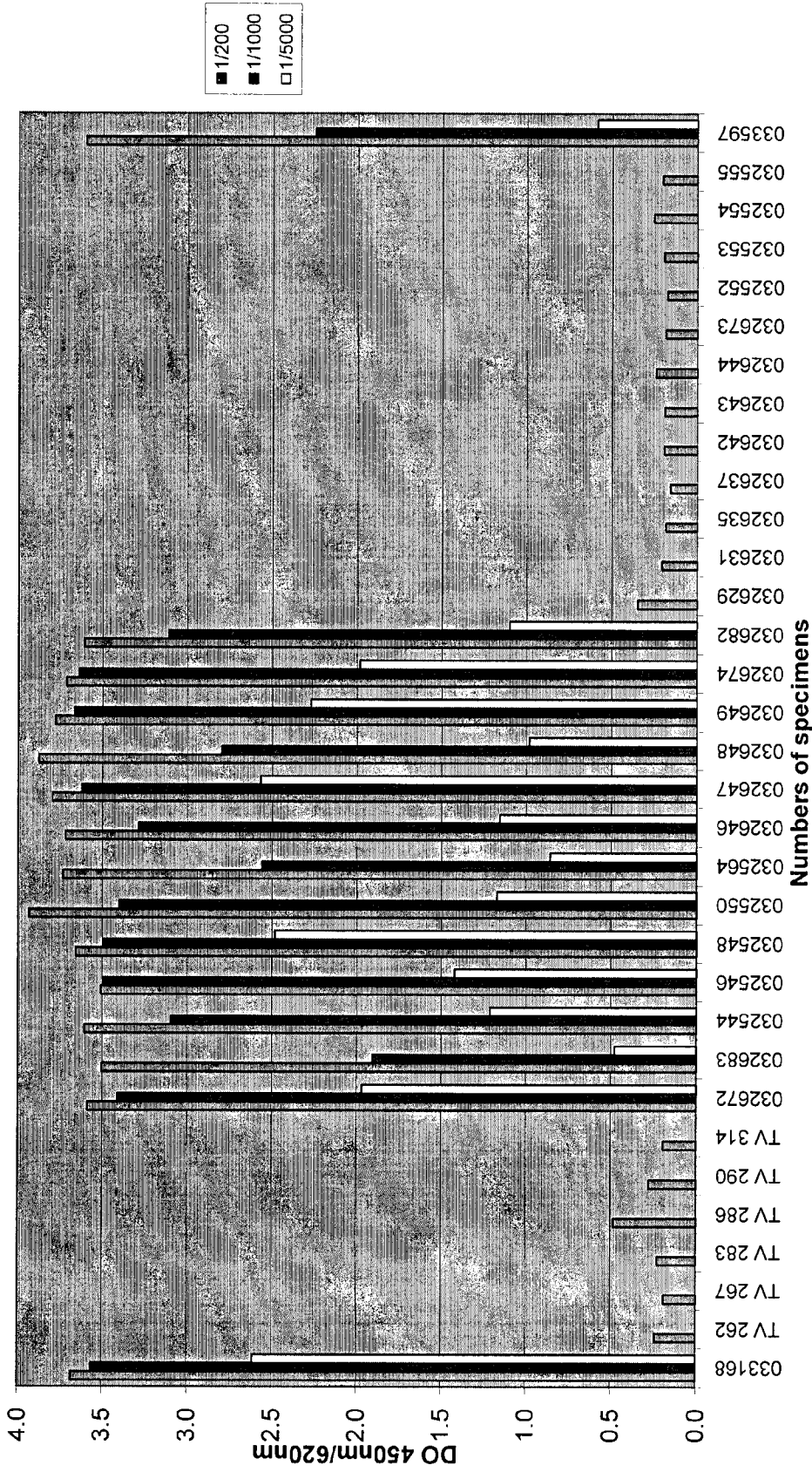


FIGURE 15

SRAS serology: Double Epitope Technique (First set)

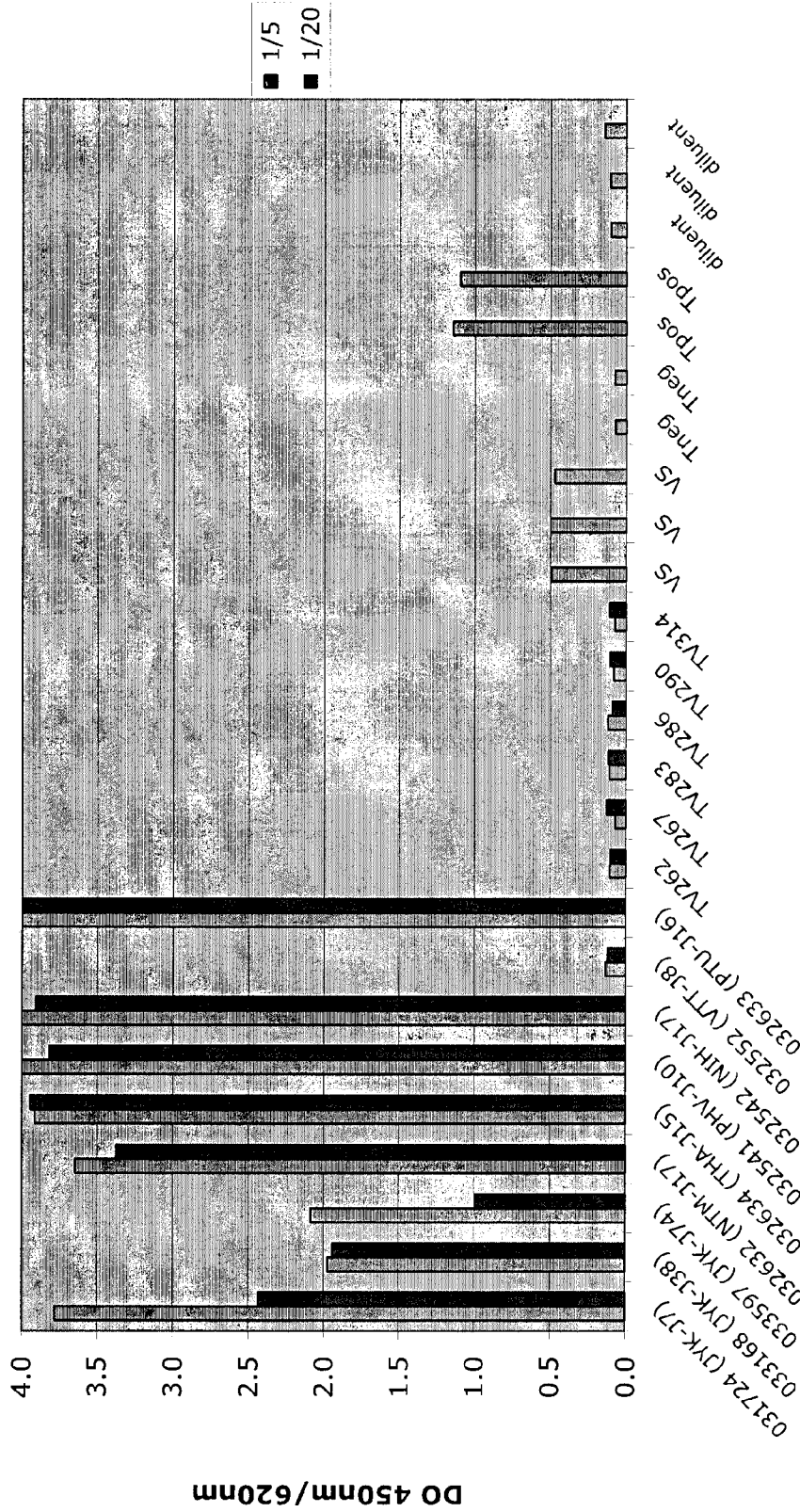


FIGURE 16

SRAS serology : Double Epitope Technique (Second set)

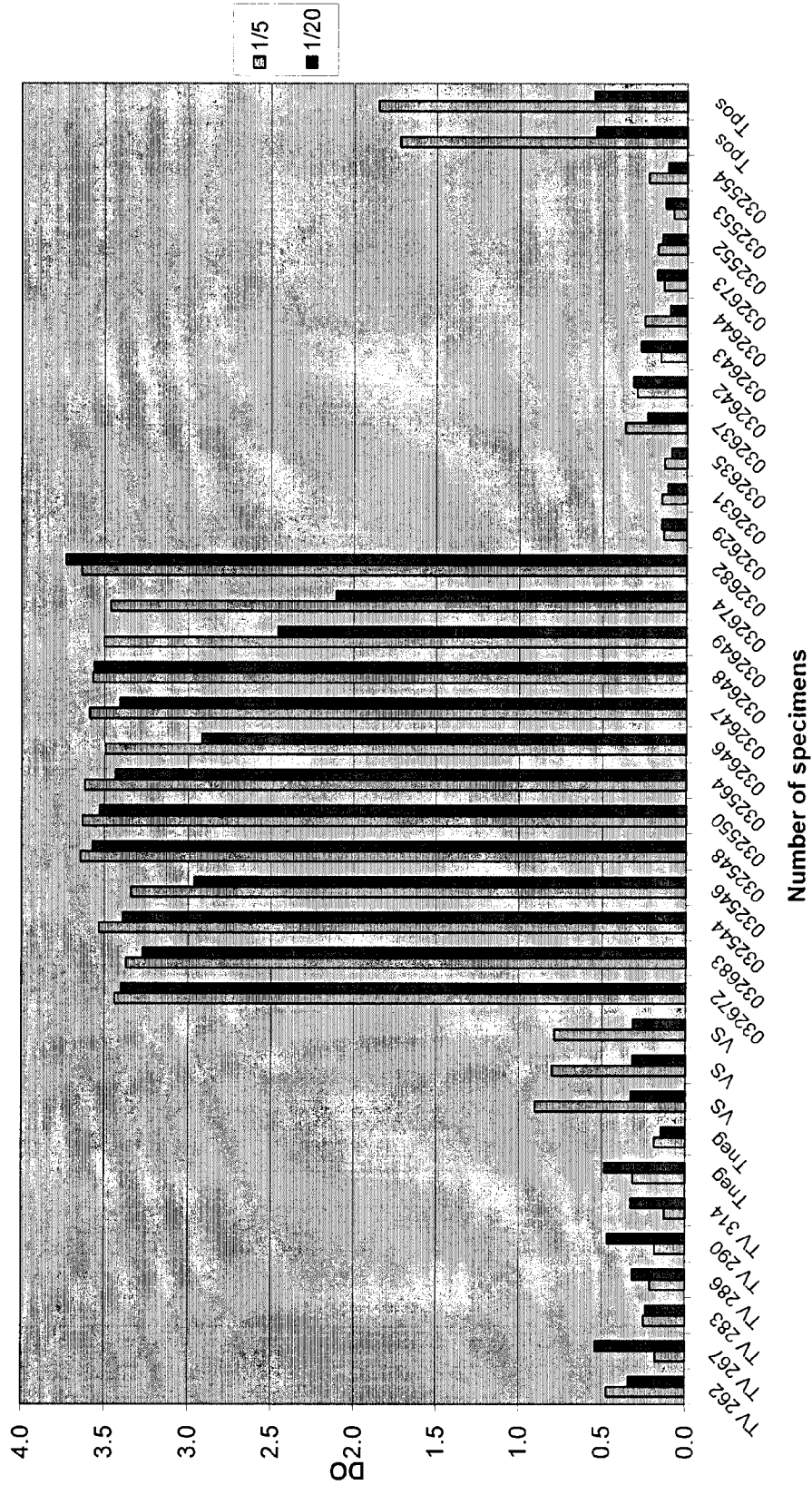


FIGURE 17

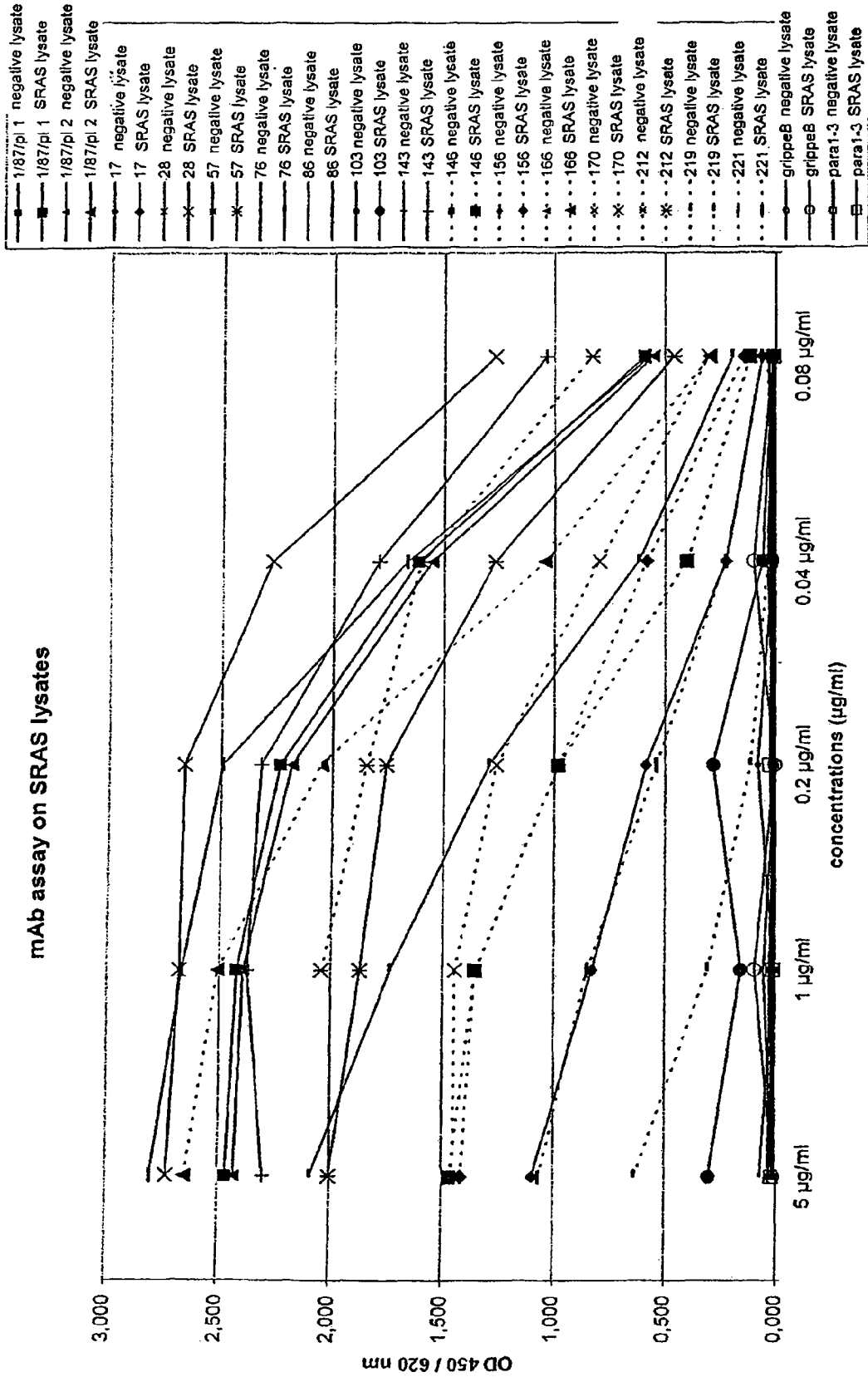
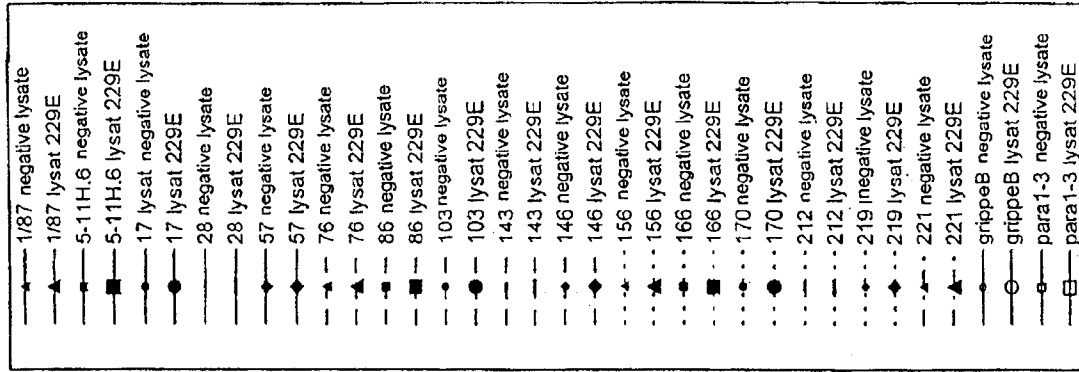


FIGURE 18



mAb assay on HCoV-229E lysates

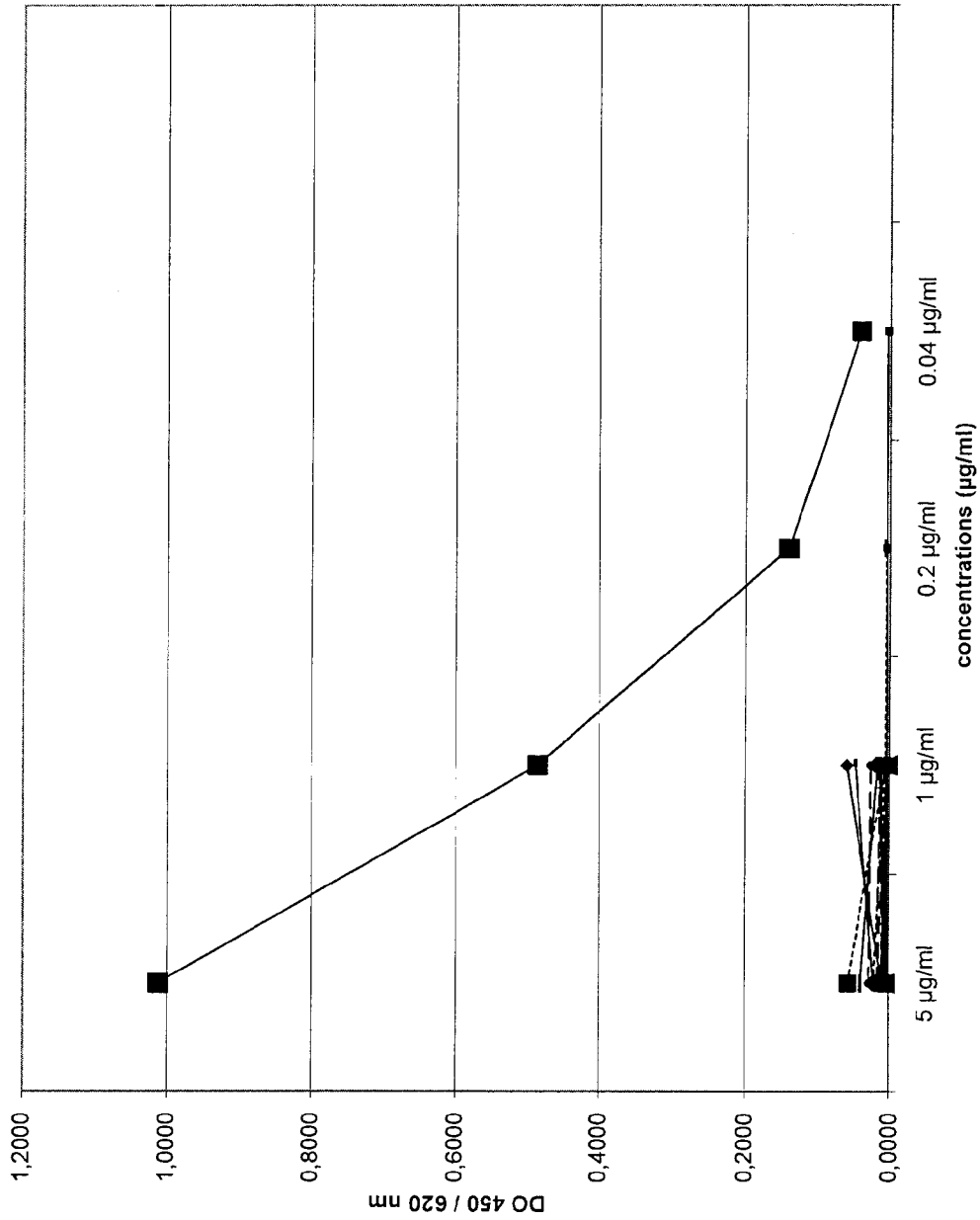


FIGURE 19

#para1-3
#grippeB
#221
#219
#212
#170
#166
#156
#146
#143
#103
#86
#76
#57
#28
#17
1/87

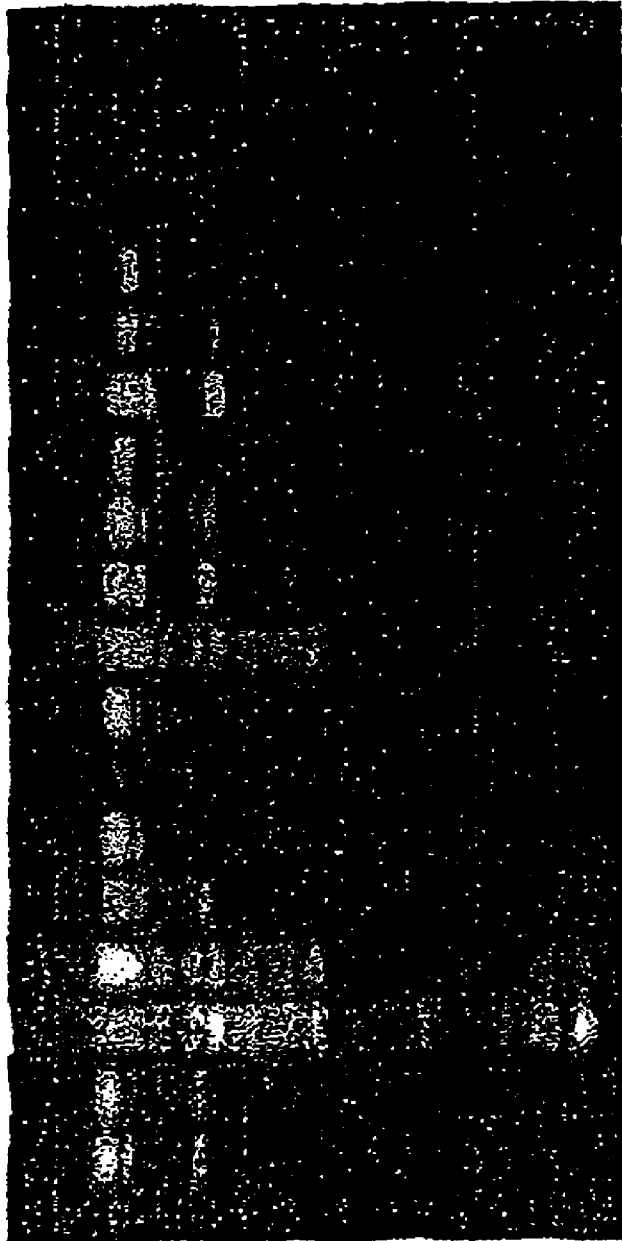


FIGURE 20

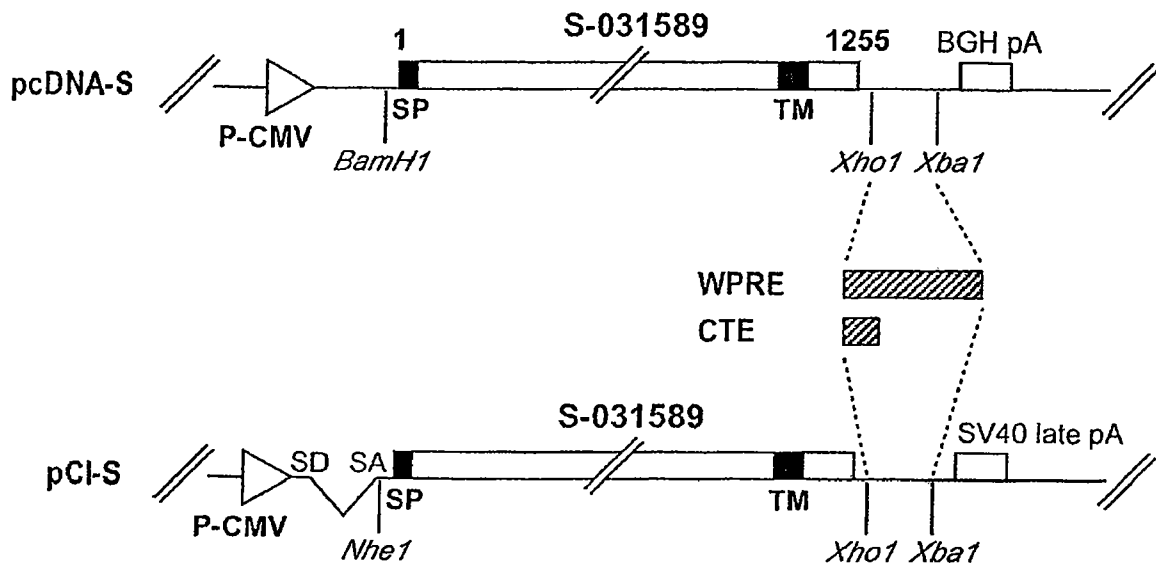


FIGURE 21

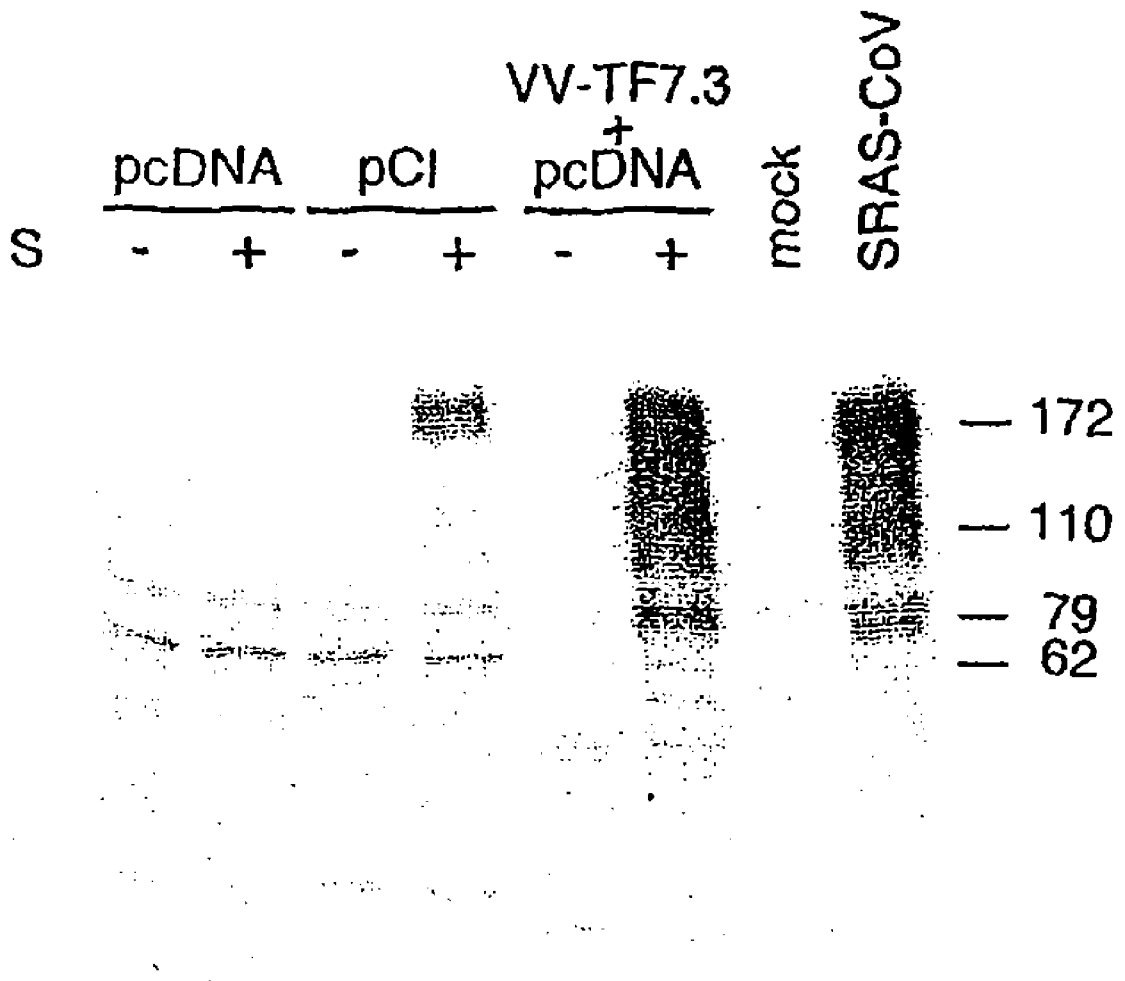
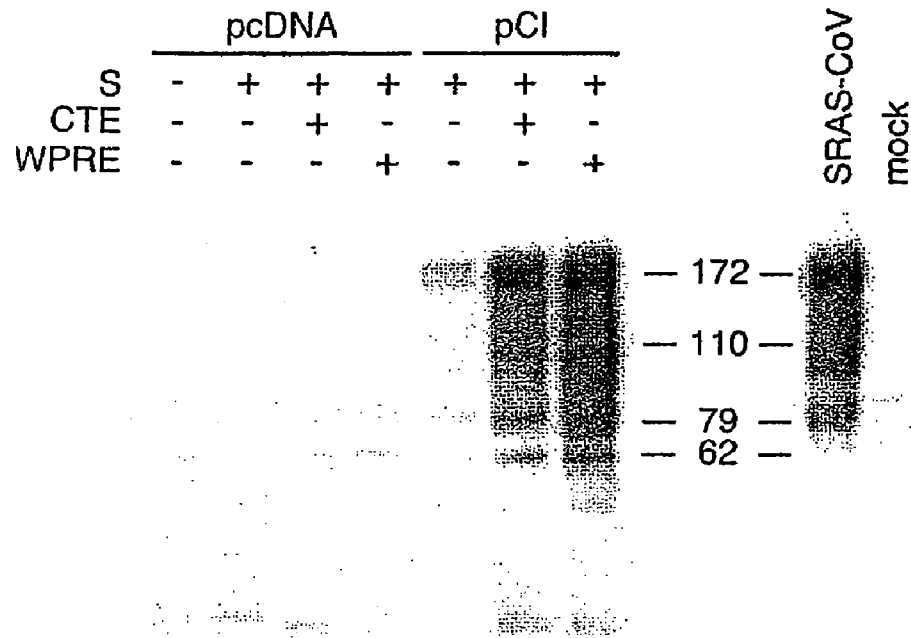


FIGURE 22

A.



B.

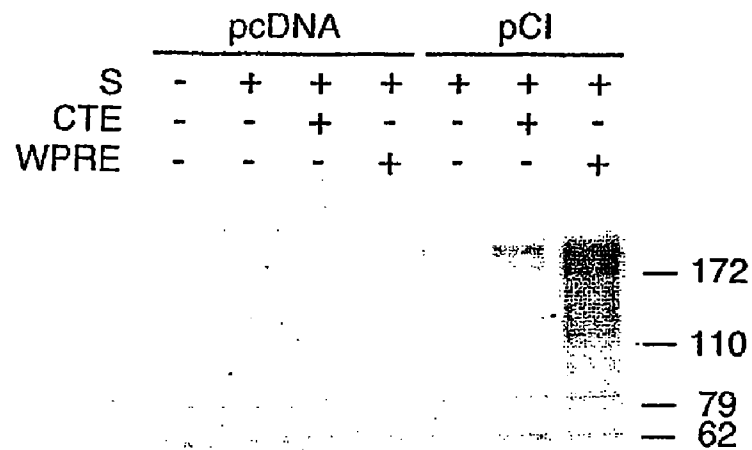


FIGURE 23

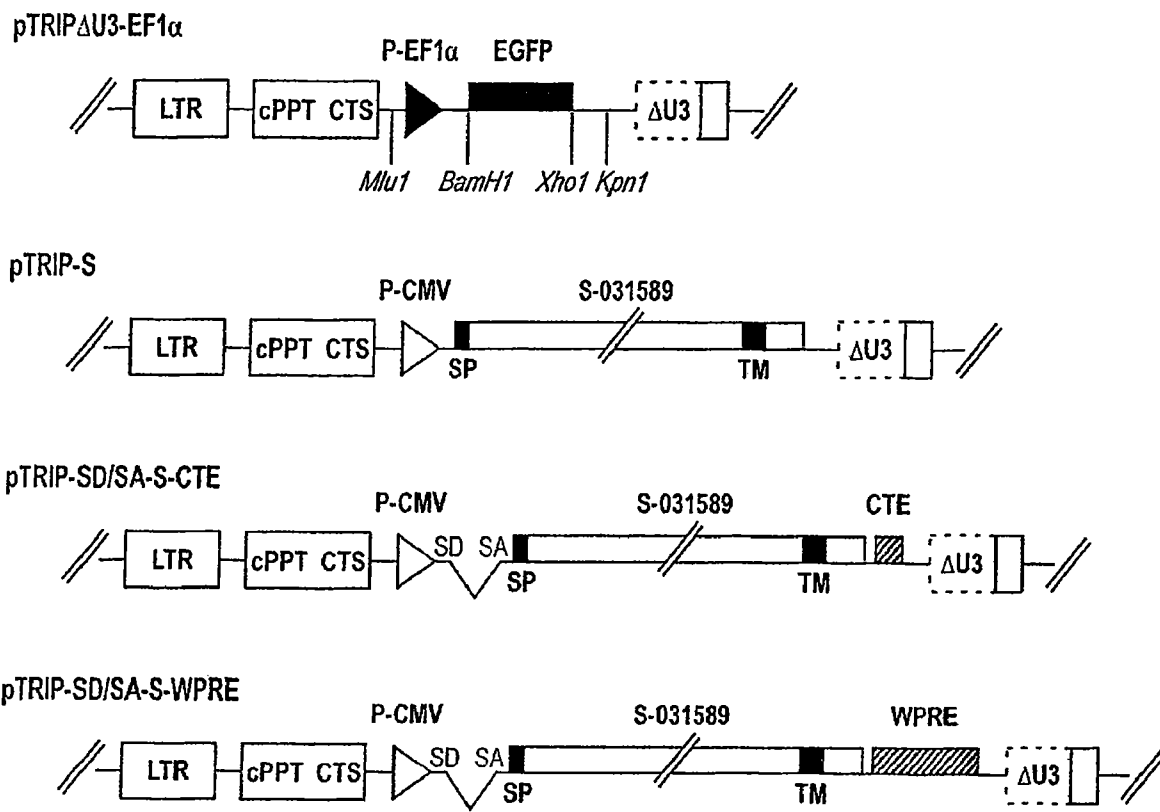


FIGURE 24

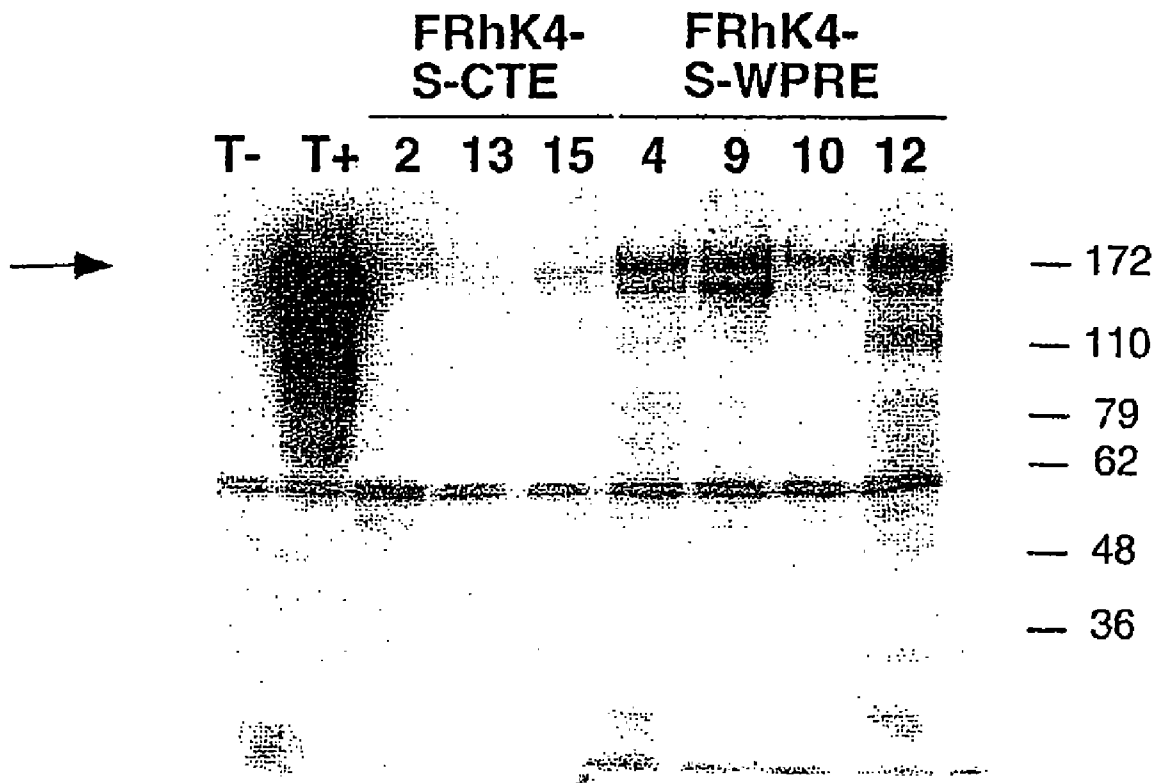


FIGURE 25

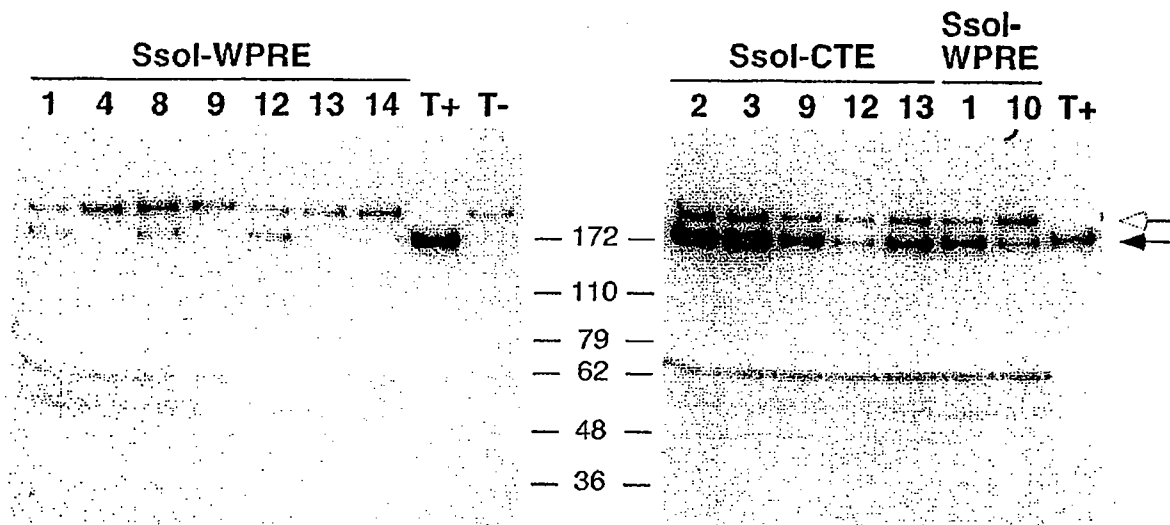
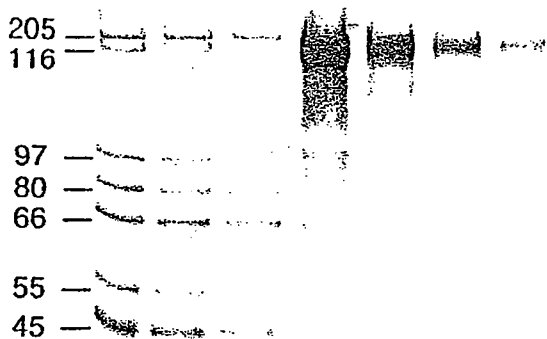


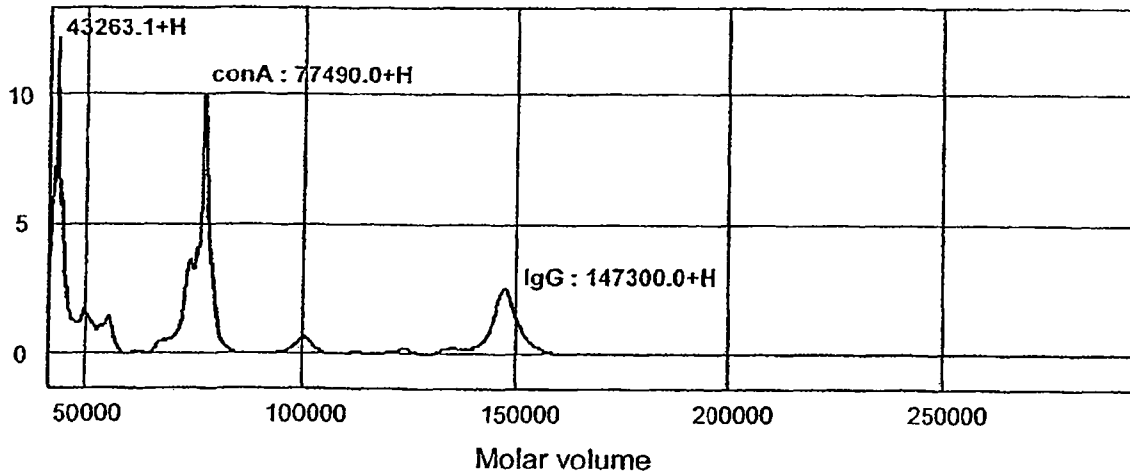
FIGURE 26

A.

MM (ng)			Ssol (μ g)		
120	60	30	2	.5	.13 .03



B.



C.

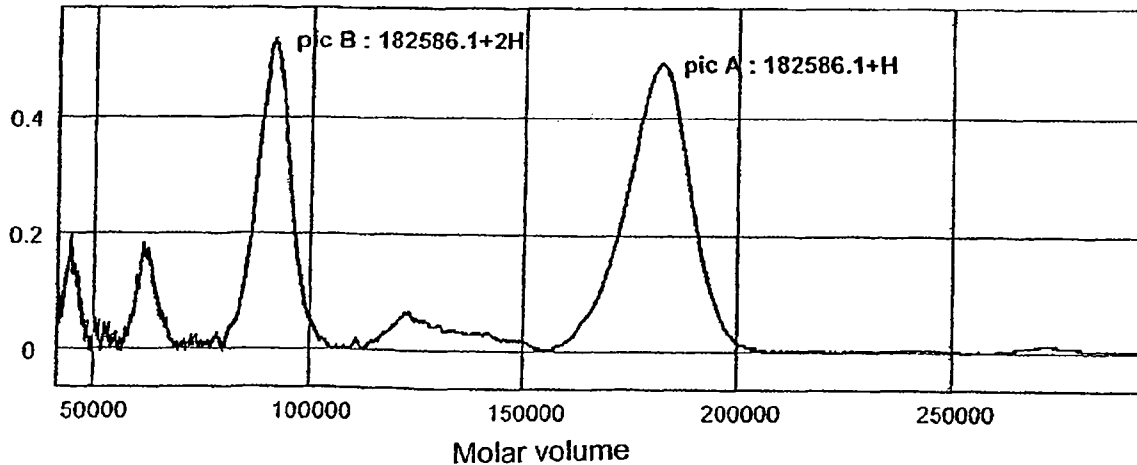


FIGURE 27 A-C

D.

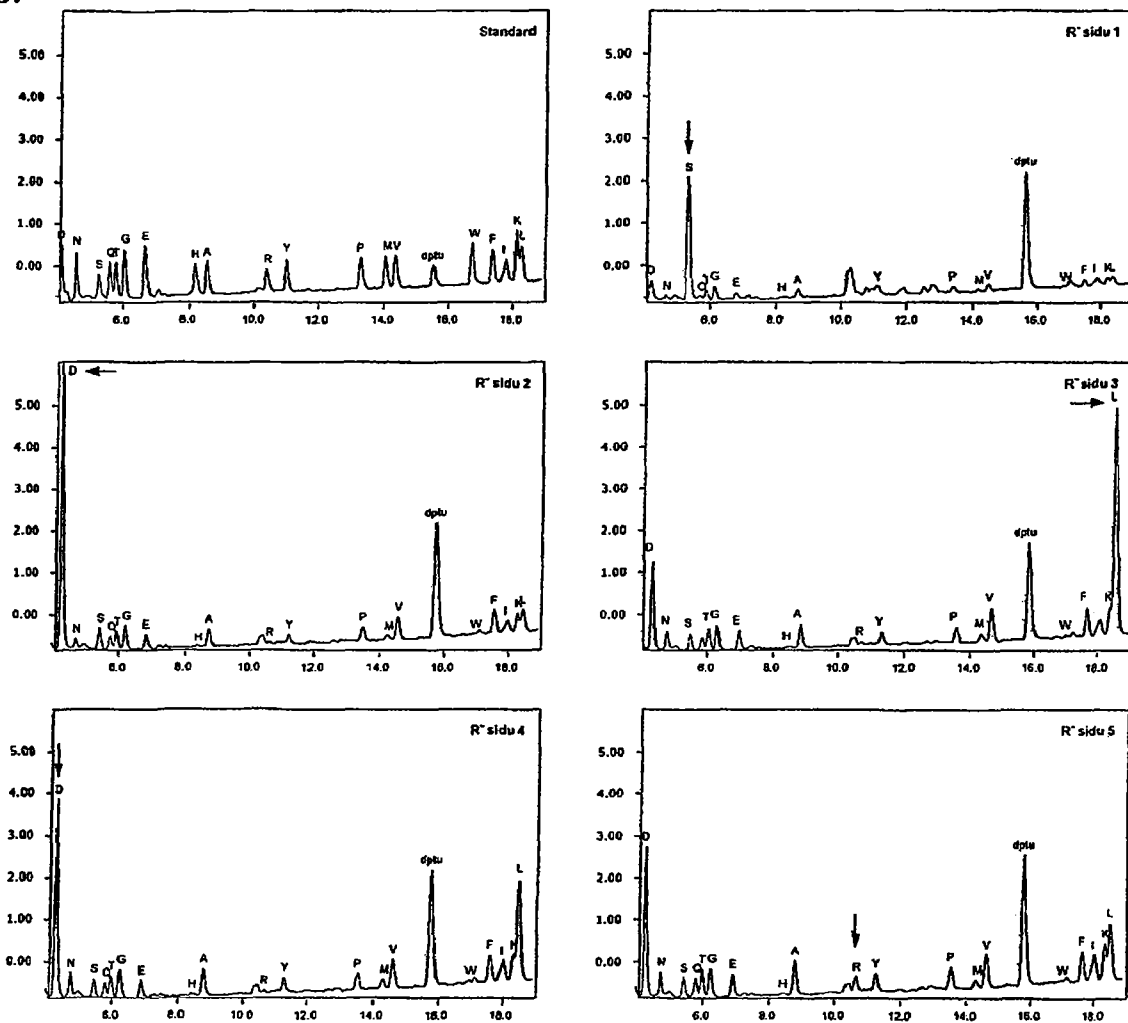


FIGURE 27 D

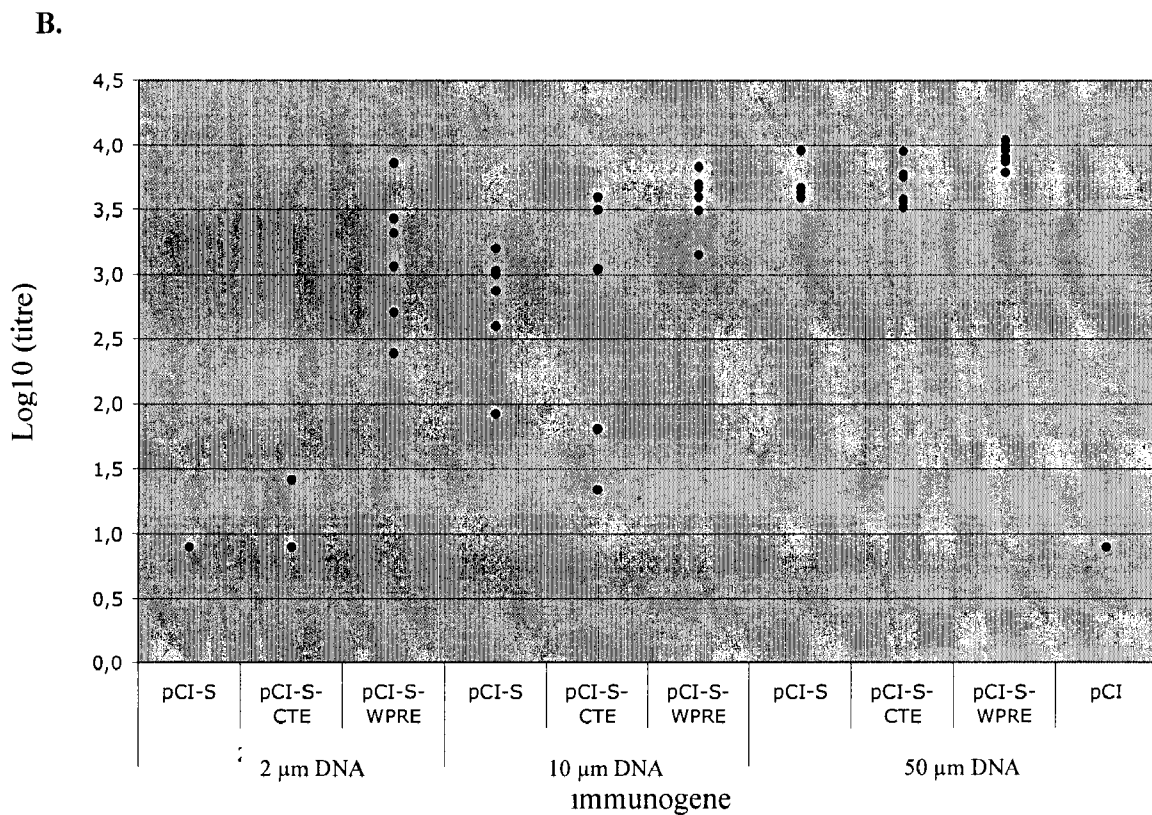
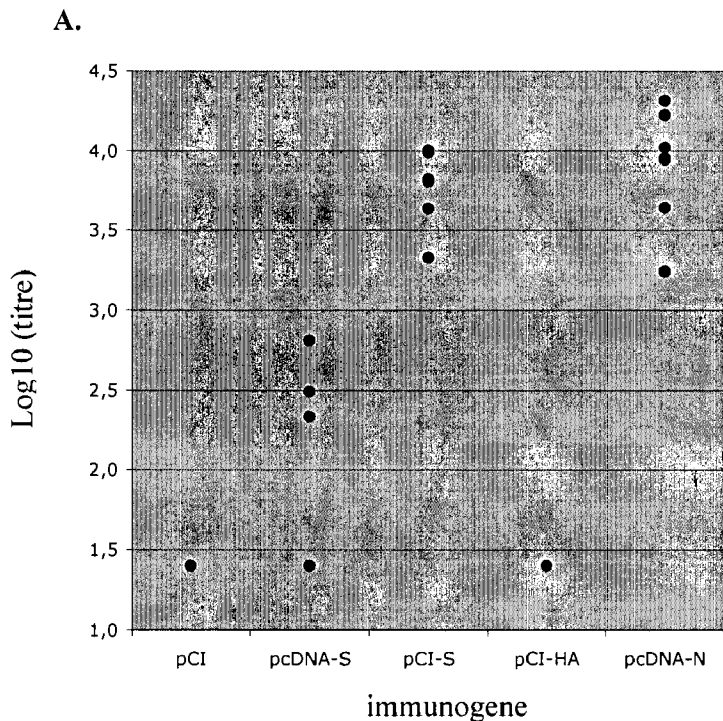


FIGURE 28

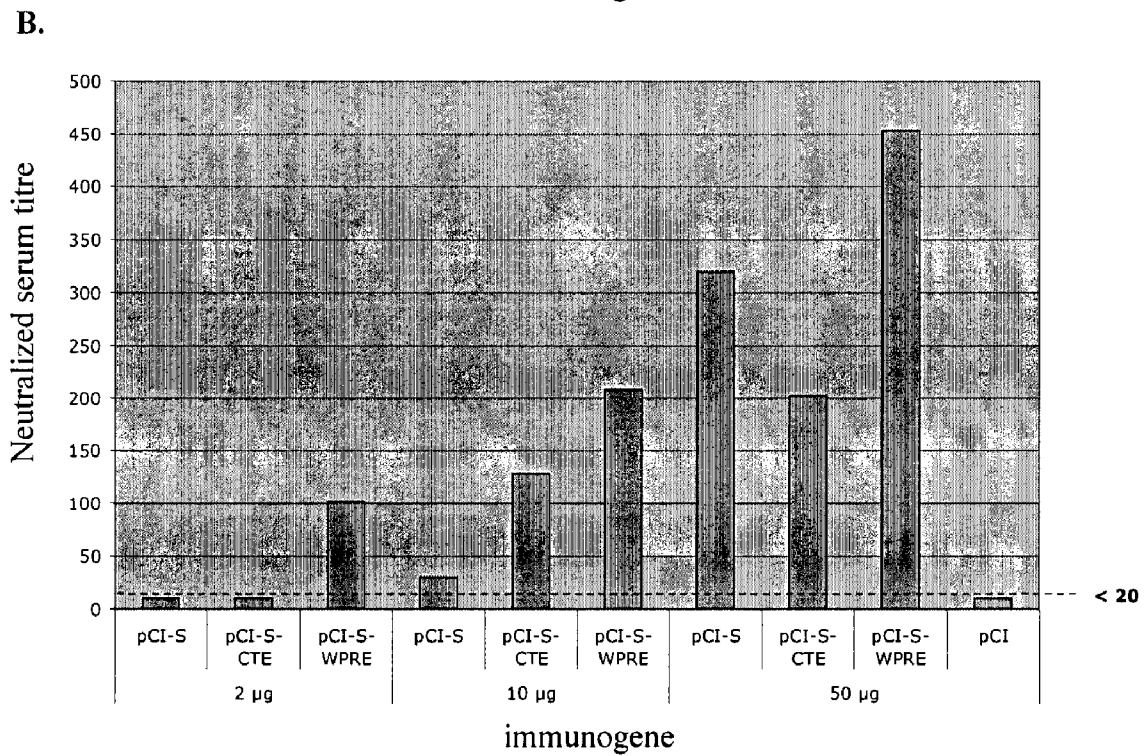
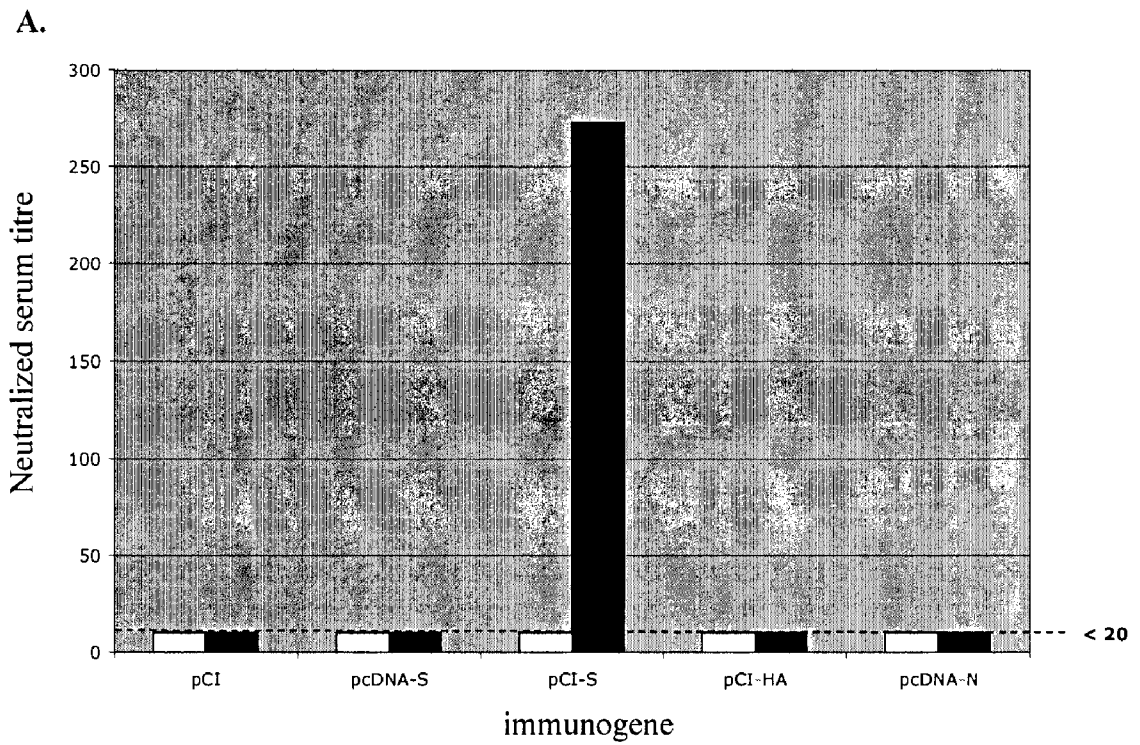
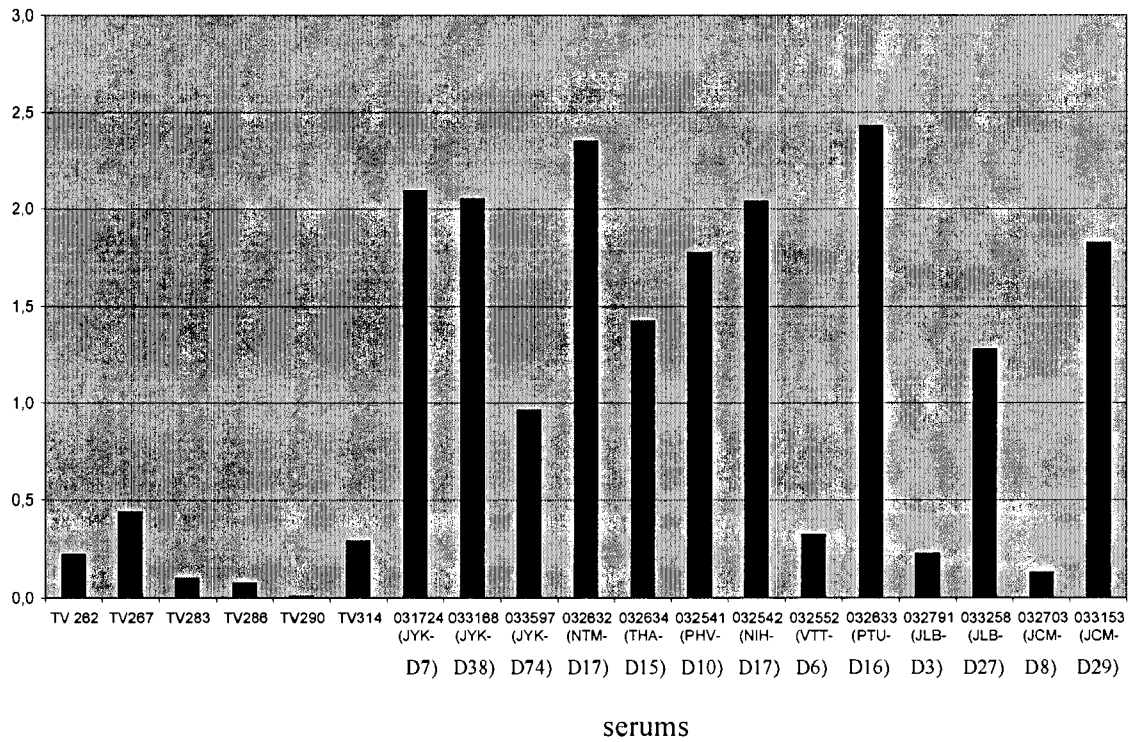


FIGURE 29



serums
FIGURE 30

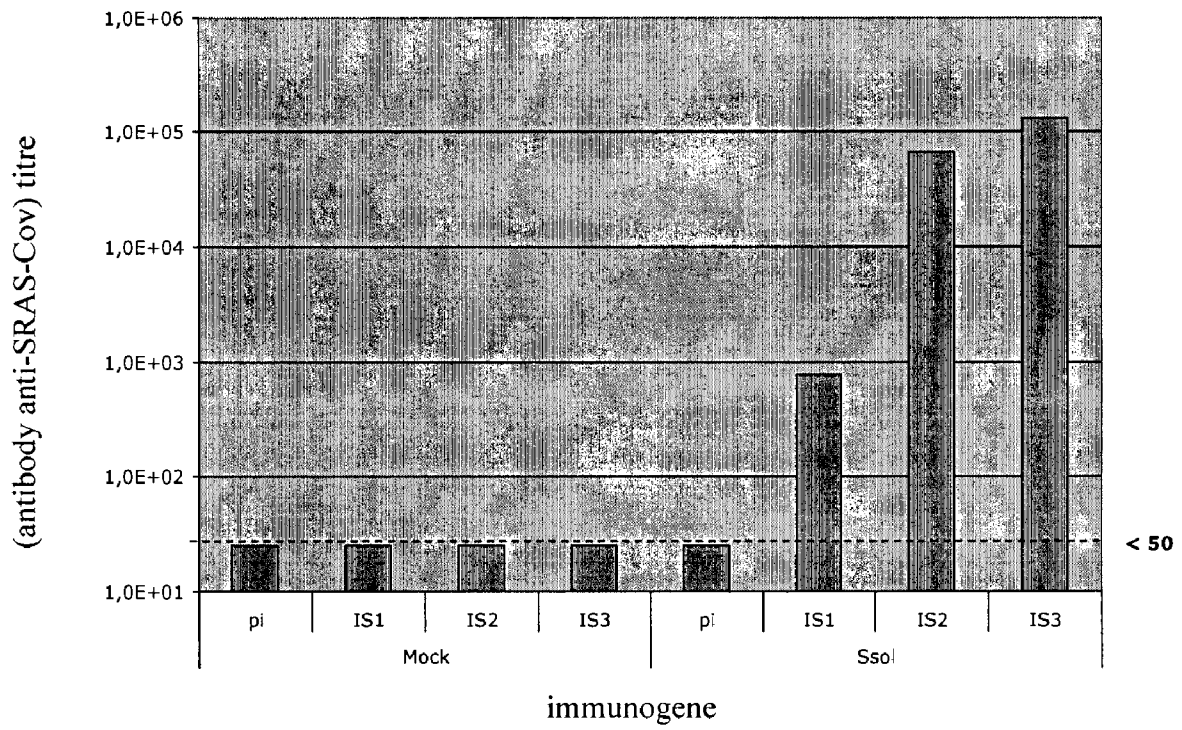


FIGURE 31

I-3059 1 CTCTTCTGGAAAAGGTAGGCTTATCATTAGAGAAAACAACAGAGTTGTGGTTTCAAGTG
S-040530

I-3059 61 ATATTCTTGTTAACAACTAAACGAACATGTTATTTTCTTATTATTCTTACTCTCACTA
S-040530 1 GG" T" C" " C" " " " " C" " " " C" " " " C" " G" C" " G" " C" " G" " C" "

I-3059 121 GTGGTAGTGACCTTGACCGGTGCACCACCTTTGATGATGTTCAAGCTCCTAATTACTCT
S-040530 44 " C" " C" " C" " " " " G" " " " " " " " " " " " " C" " " C" " " C" " G" " G" " C" " C" " C" " " " " " C" "

I-3059 181 AACATACTT CATCTATGAGGGGGTTACTATCCTGATGAAATTTTAGATCAGACACT
S-040530 104 " G" " C" " CAG" " G" " " " C" " " " " C" " " G" " " " " C" " " C" " G" " C" " " C" " GAGC" " " " " C

I-3059 240 CTTTATTTAACTCAGGATTTATTTCTTCCATTTTATCTAATGTTACAGGGTTTCATACT
S-040530 163 " G" " CC" " G" " C" " " " " " CC" " G" " C" " G" " C" " " C" " CAGC" " C" " G" " C" " C" " C" " C" " C" "

I-3059 300 ATTAATCATAAGTTTGGCAACCCGTGCATACCTTTTAAAGCATGGTATTTATTTTGCCTGCC
S-040530 223 " C" " C" " C" " C" " C" " " " " " " C" " G" " C" " C" " C" " " " C" " C" " C" " C" " C" " C" " C" " "

I-3059 360 ACAGAGAAATCAAATGTTGTCCSTGGTGGGTTTGGTTCTACCATGAACAACAAGTCA
S-040530 283 " C" " " " " GAGC" " C" " G" " G" " G" " C" " " " " G" " C" " CAGC" " " " " " " " " " " AGC

I-3059 420 CAGTCGGTGATTATTTAACAATCTACTAATGTTGTTATACGAGCATGTAACCTTGAA
S-040530 343 " " " AGC" " " " " C" " C" " C" " " " " CAGC" " C" " C" " G" " G" " C" " G" " C" " C" " " " " " C" " G

I-3059 480 TTGTGTGACAACCCCTTCTTTGCTGTTTCTAAACCCATGGGTACACAGACACATACTATG
S-040530 403 C" " " " C" " " " " " " C" " " " " C" " G" " C" " " " " " " " " " C" " C" " " " C" " C" " C" " "

I-3059 540 ATATTCGATAATGCATTTAATTGCACTTTTCGAGTACATATCTGATGCCTTTTCGCTTGAT
S-040530 463 " " C" " " " " C" " C" " C" " C" " C" " " " " " C" " " " " " " " CAGC" " C" " " " CAGC" " G" " C

I-3059 600 GTTTCAGAAAAGTCAGGTAATTTTAAACACTTACGAGAGTTTGTGTTAAAAATAAGAT
S-040530 523 " GAGC" " G" " " AGC" " C" " C" " C" " G" " " C" " G" " G" " " " C" " " " " C" " G" " C" " G" " C

I-3059 660 GGGTTTCTCTATGTTATAAGGGCTATCAACCTATAGATGTAGTTCGTGATCTACCTTCT
S-040530 583 " C" " C" " G" " C" " G" " C" " " " " " " C" " G" " C" " C" " C" " G" " GA" " A" " C" " G" " CAGC

I-3059 720 GGTTTTAACTTTGAAACCTATTTTAAAGTTCCTCTGGTATTAACATTACAAATTTT
S-040530 643 " C" " C" " " " " CC" " " " G" " C" " C" " C" " C" " " " C" " G" " C" " C" " " " " C" " C" " C" " C

I-3059 780 AGAGCCATTCTTACAGCCTTTTACCTGCTCAAGACATTTGGGGCACGTCAGCTGCAGCC
S-040530 703 C" G" " " " " C" " G" " C" " " " " AGC" " " " C" " G" " " " C" " " " " " CAGC" " C" " C" " "

I-3059 840 TATTTGTTGGCTATTTAAAGCCAACCTATTTATGCTCAAGTATGATGAAAATGGTACA
S-040530 763 " C" " C" " G" " " " " CC" " G" " " " T" " C" " C" " C" " " " " G" " " " C" " C" " G" " C" " C" " C

I-3059 900 ATCACAGATGCTGTGATTGTTCTCAAAATCCACTTGTGAACTCAAATGCTCTGTAAAG
S-040530 823 " " " " C" " C" " C" " G" " C" " CAGC" " G" " C" " C" " G" " C" " G" " G" " G" " " AGC" " G" " "

I-3059 960 AGCTTTGAGATTGACAAAGGAATTTACCAGACCTCTAATTTAGGGTTGTTCCCTCAGGA
S-040530 883 " " " " C" " " " " C" " " " G" " C" " C" " " " " " AGC" " C" " " " A" " G" " G" " TAGC" " C

I-3059 1020 GATGTTGTGAGATTCCCTAATATTACAACTTGTGTCCTTTTGGAGAGGTTTTTAAATGCT
S-040530 943 " " " " G" " C" " G" " " " " C" " " " C" " C" " C" " " " C" " C" " C" " C" " A" " G" " C" " C" " C

I-3059 1080 ACTAAATCCCTTCTGTCTATGCATGGGAGAGAAAAAATTTCTAATGTGTTGCTGAT
S-040530 1003 " C" " G" " " " CAGC" " G" " C" " C" " " " " C" " G" " G" " G" " CAGC" " C" " C" " G" " C" " C

I-3059 1140 TACTCTGTGCTCTACAACCTCAACATTTTTTCAACCTTTAAGTGCATGGCGTTTCTGCC
S-040530 1063 " " " AGC" " " " " G" " " " " " C" " C" " C" CAGC" " " " C" " " " " " C" " " " " " GAGC" " "

I-3059 1200 ACTAAGTTGAATGATCTTTGCTTCTCCAATGTCTATGCAGATTCTTTTGTAGTCAAGGGA
S-040530 1123 " C" " " C" " " C" " C" " G" " " " " AG" " C" " G" " C" " C" CAGC" " C" " G" " G" " " " C

I-3059 1260 GATGATGTAAGACAAATAGCGCCAGGACAAACTGGTGTATTGCTGATTATAATTATAAA
S-040530 1183 " C" " C" " G" " " " " G" " C" " C" " T" " C" " G" " C" " C" " G" " C" " C" " C" " C" " C" " C" " C" " G

FIGURE 32.1

I-3059 2697 GGTGCTGGCGCTGCTCTTCAAAATACCTTTTGCTATGCAAATGGCATATAGGTTCAATGGC
 S-040530 2620 ""A""C""A""C""C""G""G""C""C""C""C""G""G""C""C""CC""C""G""G""C""G""

I-3059 2757 ATTGGAGTTACCCAAAATGTTCTCTATGAGAACCAAAACAATCGCCAACCAATTTAAC
 S-040530 2680 ""C""C""G""G""G""C""G""G""C""G""G""G""G""G""G""G""G""G""G""G""G""G""G""G""

I-3059 2817 AAGGCGATTAGTCAAATTCAGAATCACTTACAACAACATCAACTGCATTGGGCAAGCTG
 S-040530 2740 ""C""C""C""C""G""C""G""GAGC""G""C""C""CAGC""C""CC""C""G""G""G""G""G""G""

I-3059 2877 CAAGACGTTGTTAACCAGAATGCTCAAGCATTAACACACTTGTTAAACAACCTTAGCTCT
 S-040530 2800 ""G""G""G""G""G""G""G""C""C""G""C""C""G""G""G""G""G""G""G""G""G""G""G""G""AGC

I-3059 2937 AATTTGGTGCAATTTCAAGTGCTAAATGATATCCTTTTCGCGACTTGATAAAGTCGAG
 S-040530 2860 ""C""C""C""C""C""CAGCTC""C""G""C""C""G""GAGCA""G""G""C""C""G""G""

I-3059 2997 GCGGAGGTACAAATGACAGGCTAATTACAGGCAGACTTCAAAGCCTTCAAACCTATGTA
 S-040530 2920 ""C""A""G""G""C""C""C""G""C""C""AC""C""G""GTC""G""G""G""C""G""G""

I-3059 3057 ACACAACAATAATCAGGGCTGCTGAAATCAGGGCTTCTGCTAATCTTGCTGCTACTAAA
 S-040530 2980 ""C""G""G""G""G""A""C""C""G""C""C""CAGC""C""C""G""C""C""C""C""G""

I-3059 3117 ATGCTGAGTGTGTTCTTGGACAATCAAAAAGAGTTGACTTTTGTGGAAAGGGCTACCAC
 S-040530 3040 ""AGC""C""G""G""G""G""GAGC""G""G""G""G""G""G""G""C""C""C""C""C""C""T""

I-3059 3177 CTTATGTCCTTCCCACAAGCAGCCCCGCATGGTGTGTCTTCTACATGTCACGTATGTG
 S-040530 3100 ""G""AG""C""G""C""C""C""C""C""G""G""G""G""C""G""C""G""C""C""

I-3059 3237 CCATCCCAGGAGAGGAACTTCACCACAGCGCCAGCAATTTGTCATGAAGGCAAAGCATA
 S-040530 3160 ""TAG""C""C""C""C""C""C""C""C""C""C""C""C""C""G""G""G""C""C""

I-3059 3297 TTCCTCGTGAAGGTGTTTGTGTTAATGGCACTTCTTGGTATTATACACAGAGGAAC
 S-040530 3220 ""C""G""G""C""G""C""C""C""C""CAGC""C""C""C""C""C""C""

I-3059 3357 TTCTTTCTCCACAAAATAATTACTACAGACAATACATTTGTCTCAGGAAATGTGATGTC
 S-040530 3280 ""CAGC""C""G""C""C""C""C""C""C""C""C""G""C""C""C""C""G""G""G""

I-3059 3417 GTTATTGGCATCATTAAACAACACAGTTTATGATCCTCTGCAACCTGAGCTTGACTCATT
 S-040530 3340 ""G""C""C""C""C""C""C""C""C""G""C""C""C""C""G""C""G""AGC""

I-3059 3477 AAAGAAGAGCTGGACAAGTACTTCAAAAATCATAATCACCAGATGTTGATCTTGCCGAC
 S-040530 3400 ""G""G""A""G""C""A""G""T""A""G""C""C""C""C""C""C""C""C""G""C""G""G""T

I-3059 3537 ATTTCAGGCATTAACGCTTCTGTGCTCAACATTCAAAAAGAAATGACCGCCTCAATGAG
 S-040530 3460 ""CAGC""C""C""C""C""C""G""G""C""C""G""G""G""C""A""A""G""C""A

I-3059 3597 GTCGCTAAAATTTAAATGAATCACTCATTGACCTTCAAGAATGGGAAAATATGAGCAA
 S-040530 3520 ""G""C""G""CC""G""C""GAGC""G""C""C""G""G""G""C""C""G""G""C""G""G""

I-3059 3657 TATATTAATGGCCTTGGTATGTTTGGCTCGGCTTCAATGCTGGACTAATGGCCATCGTC
 S-040530 3580 ""C""C""G""C""C""C""G""G""G""C""C""C""C""C""G""C""G""G""G""G""

I-3059 3717 ATGGTTACAATCTTGCTTTGTTGCATGACTAGTTGTTGCAGTTGCCTCAAGGGTGCATGC
 S-040530 3640 ""G""C""C""C""G""C""C""C""C""C""C""C""C""G""G""A""C""C""C""

I-3059 3777 TCTGTGGTCTTCTGCTGCAAGTTTGATGAGGATGACTCTGAGCCAGTTCTCAAGGGTGTC
 S-040530 3700 AGC""CAGC""C""C""C""C""AGC""C""G""C""G""G""C""G""

I-3059 3837 AAATTACATTACACATAAACGAACCTATGGATTGTTTATGAGATTTTFACTCTTGGAT
 S-040530 3760 ""GC""G""C""C""G""T""CGA""

I-3059 3897 CAATTACTGCACAGCCAGTAAAATGACAATGCTTCTCCTGCAAGT

FIGURE 32.3

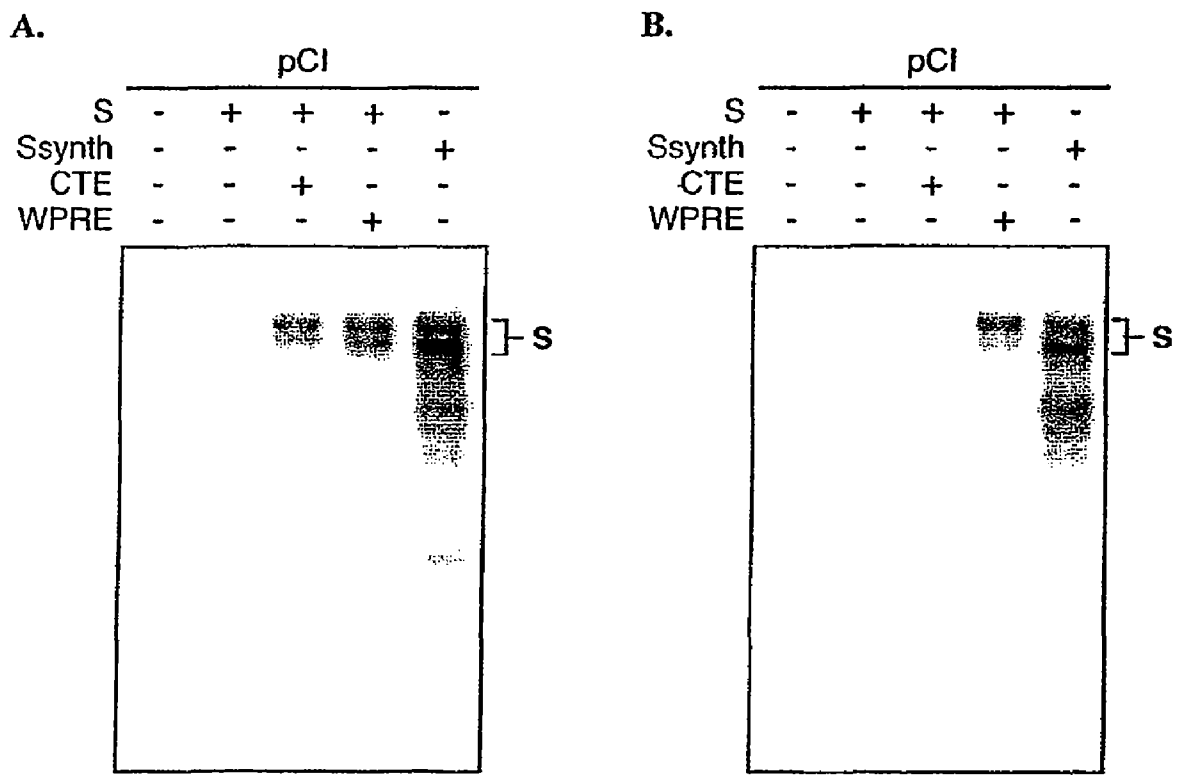
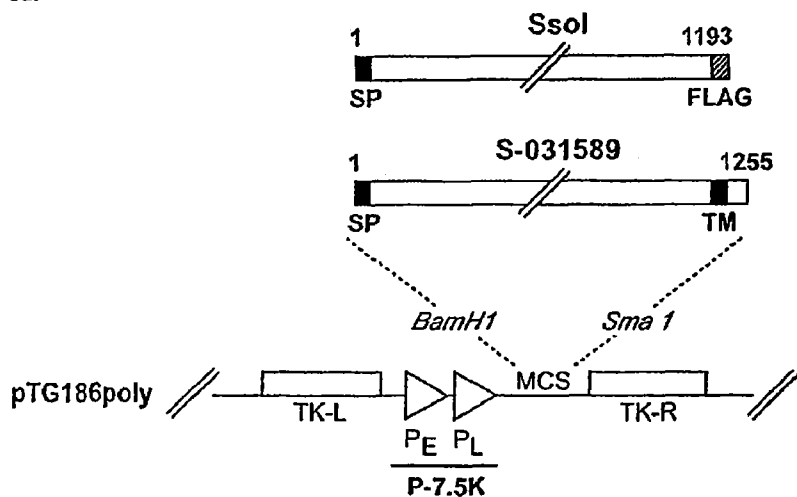
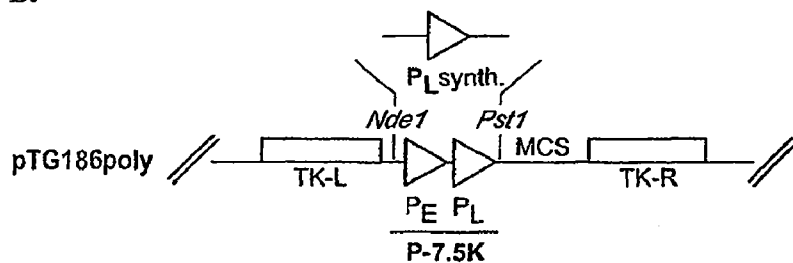


FIGURE 33

A.



B.



C.

CATATG AGC [T]₂₀GGCATATAAATA GACTC GGCGCGCC AT CTGCAG
Nde1 promoteur 480 *Asc1* *Pst1*

FIGURE 34 A-C

D.

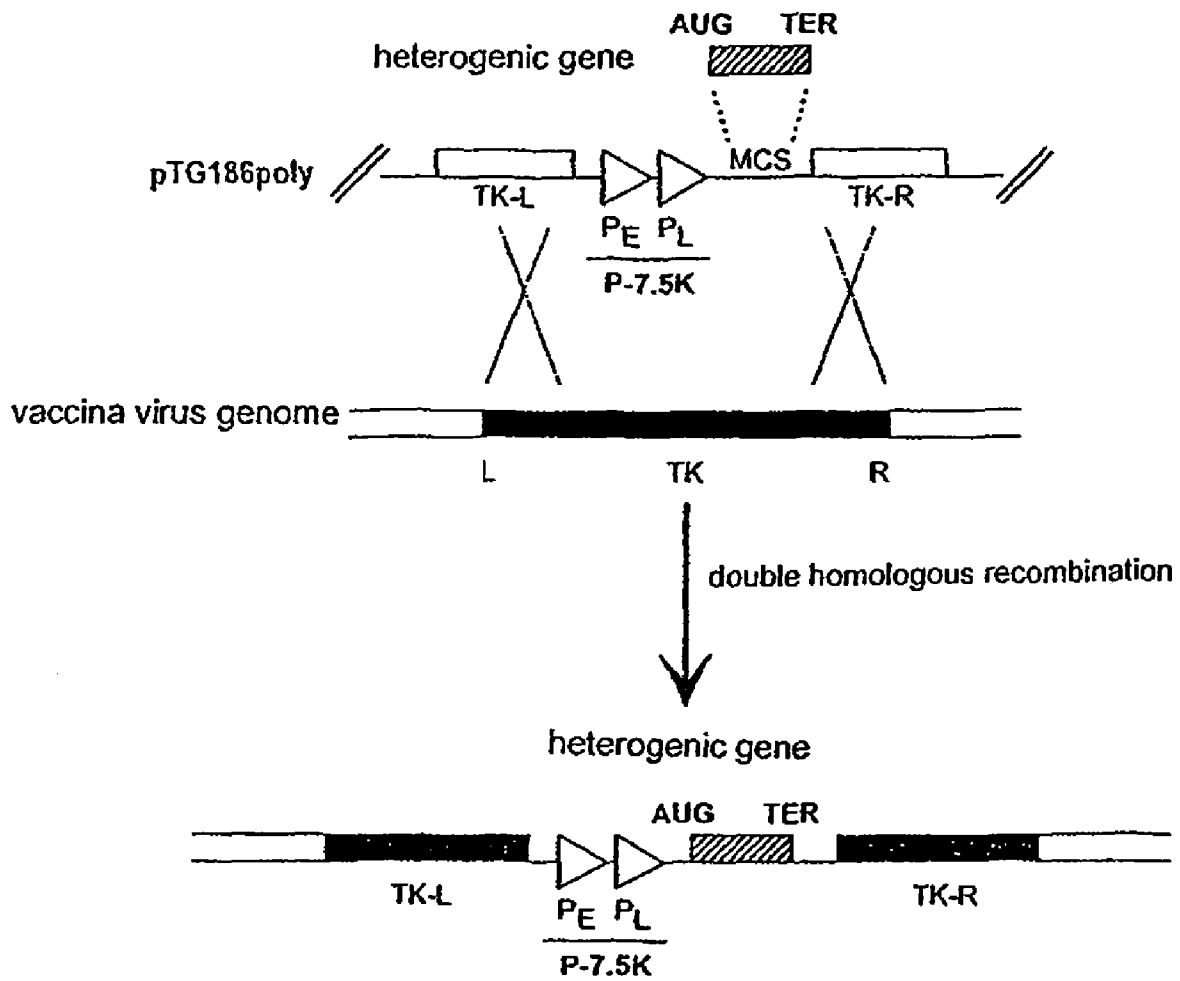
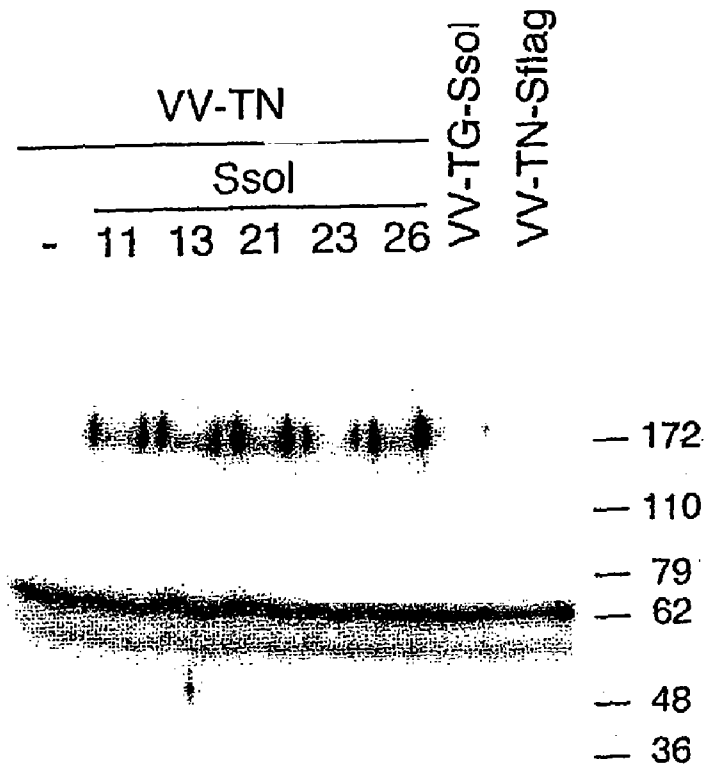


FIGURE 34 D

A.



B.

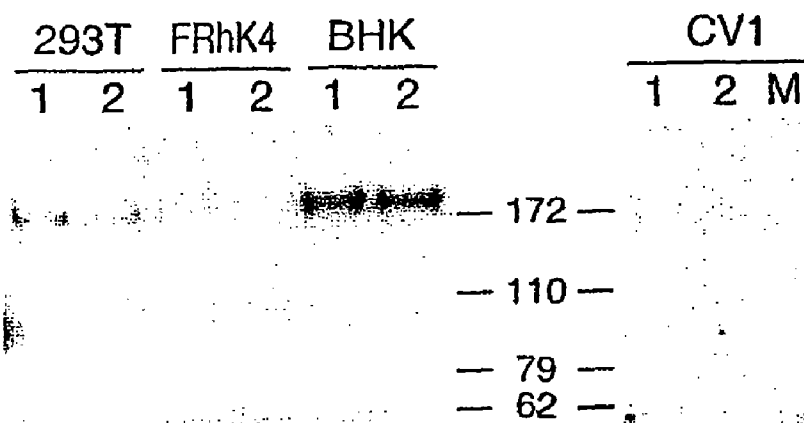


FIGURE 36



FIGURE 37

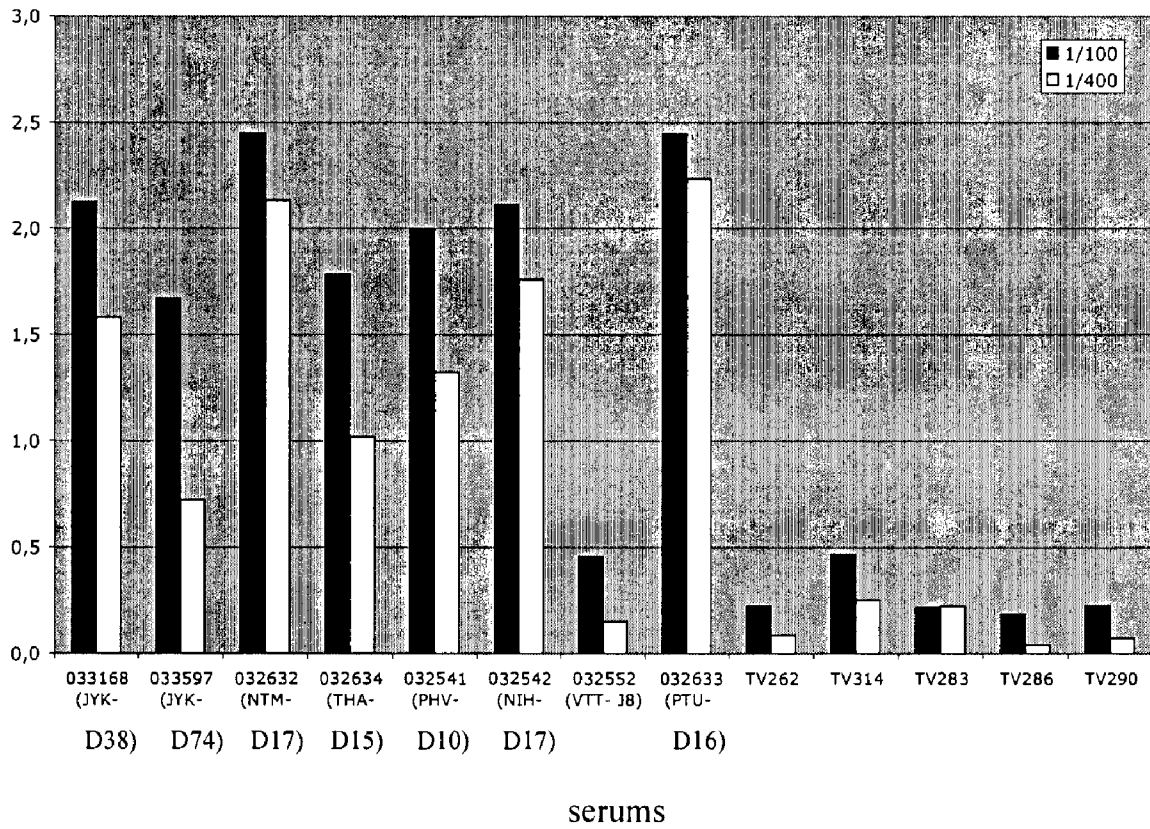
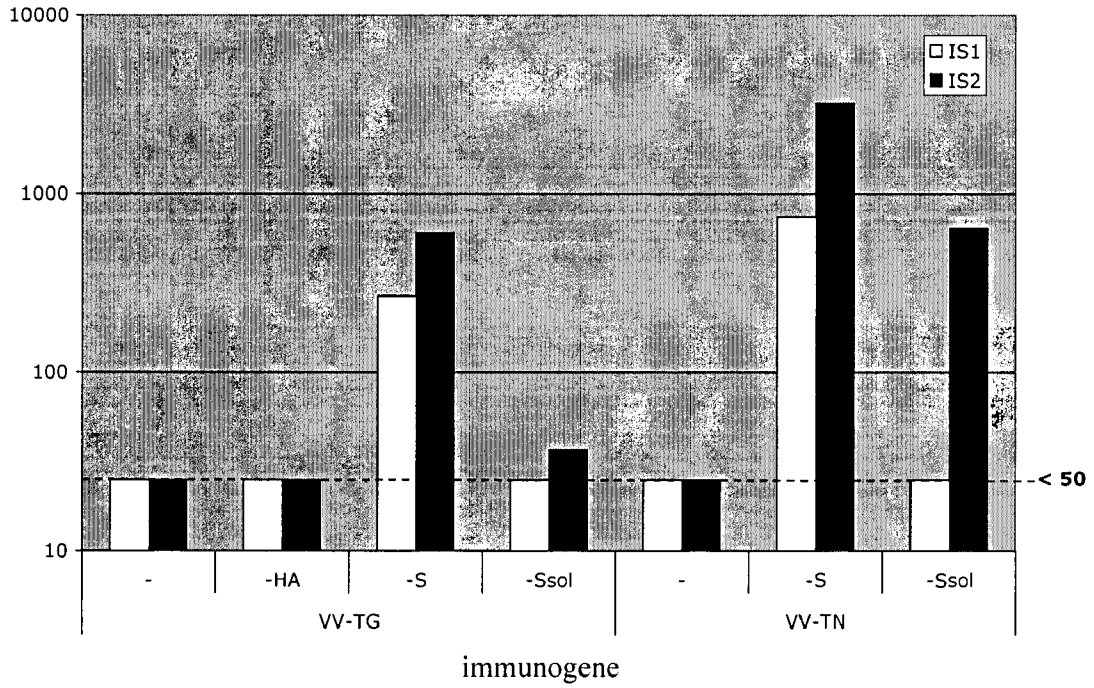


FIGURE 38

A.



B.

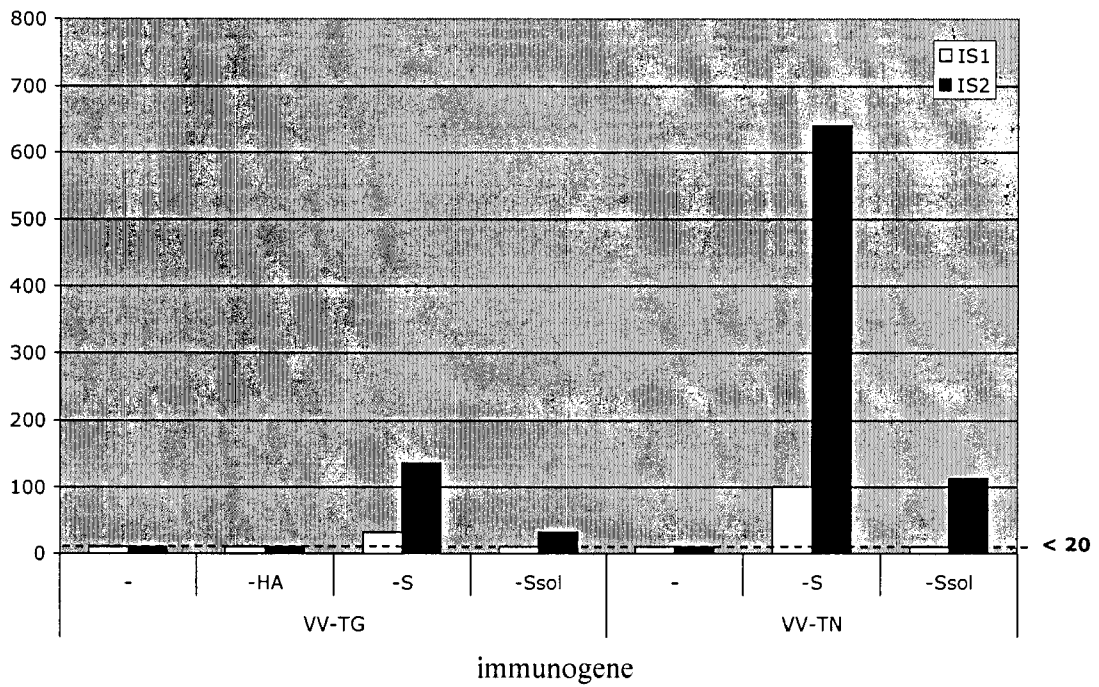


FIGURE 39

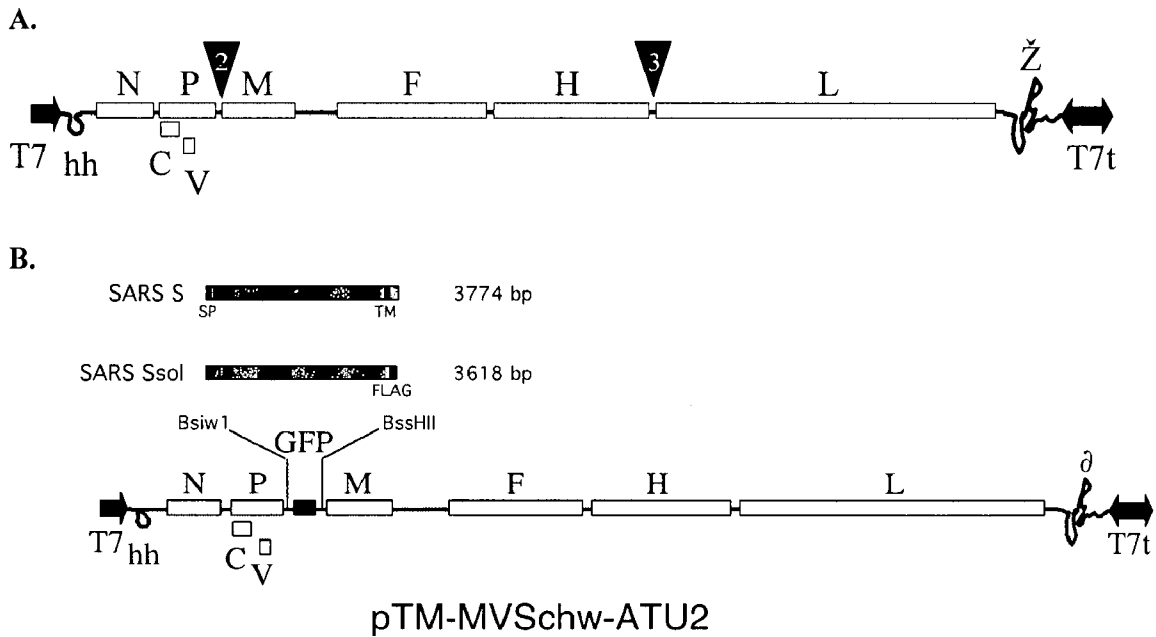


FIGURE 40

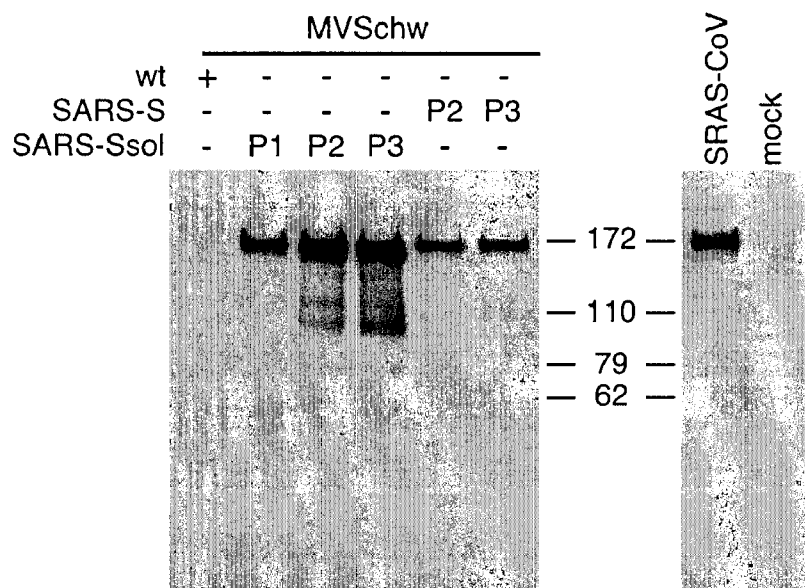


FIGURE 41

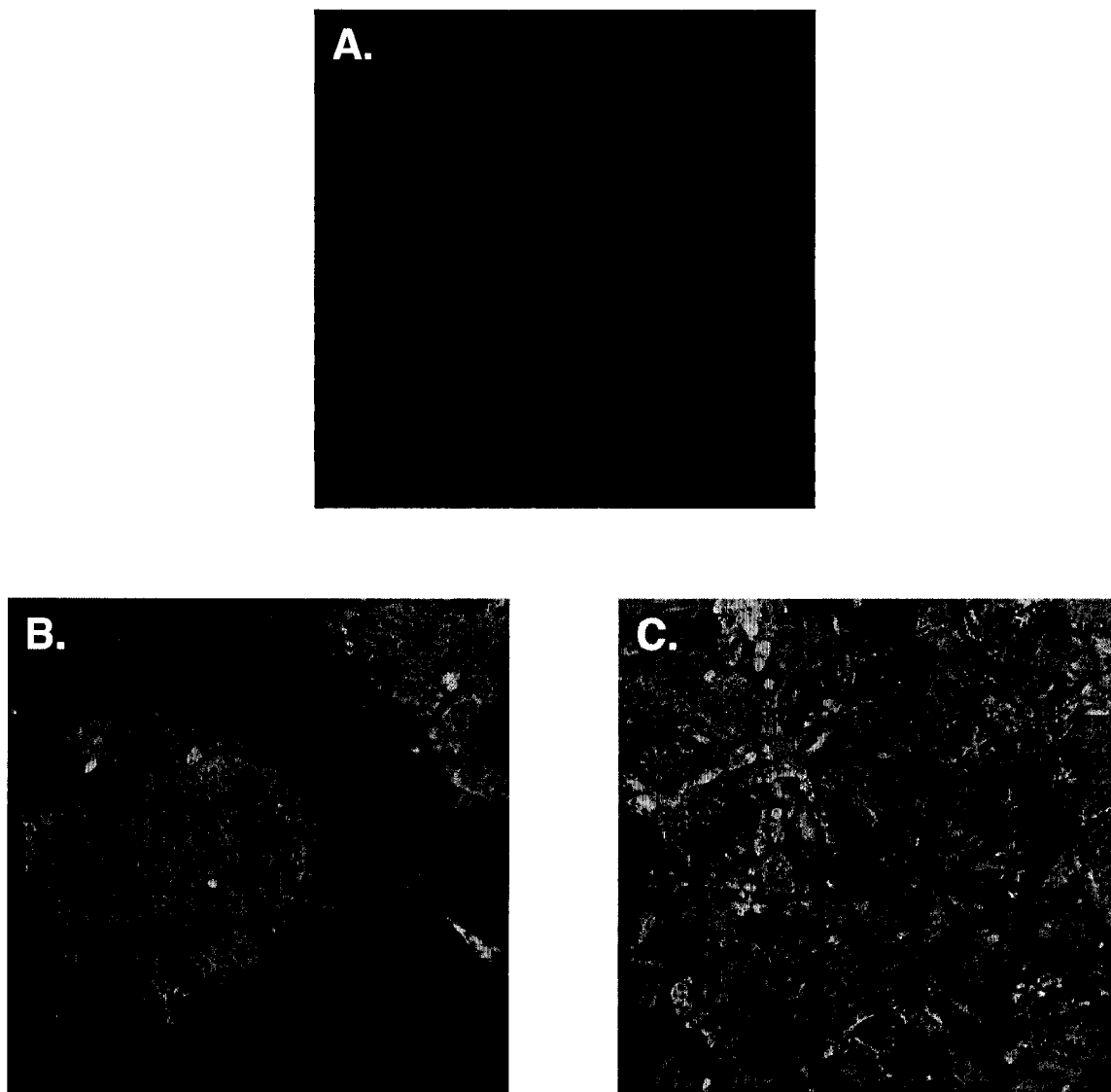


FIGURE 42

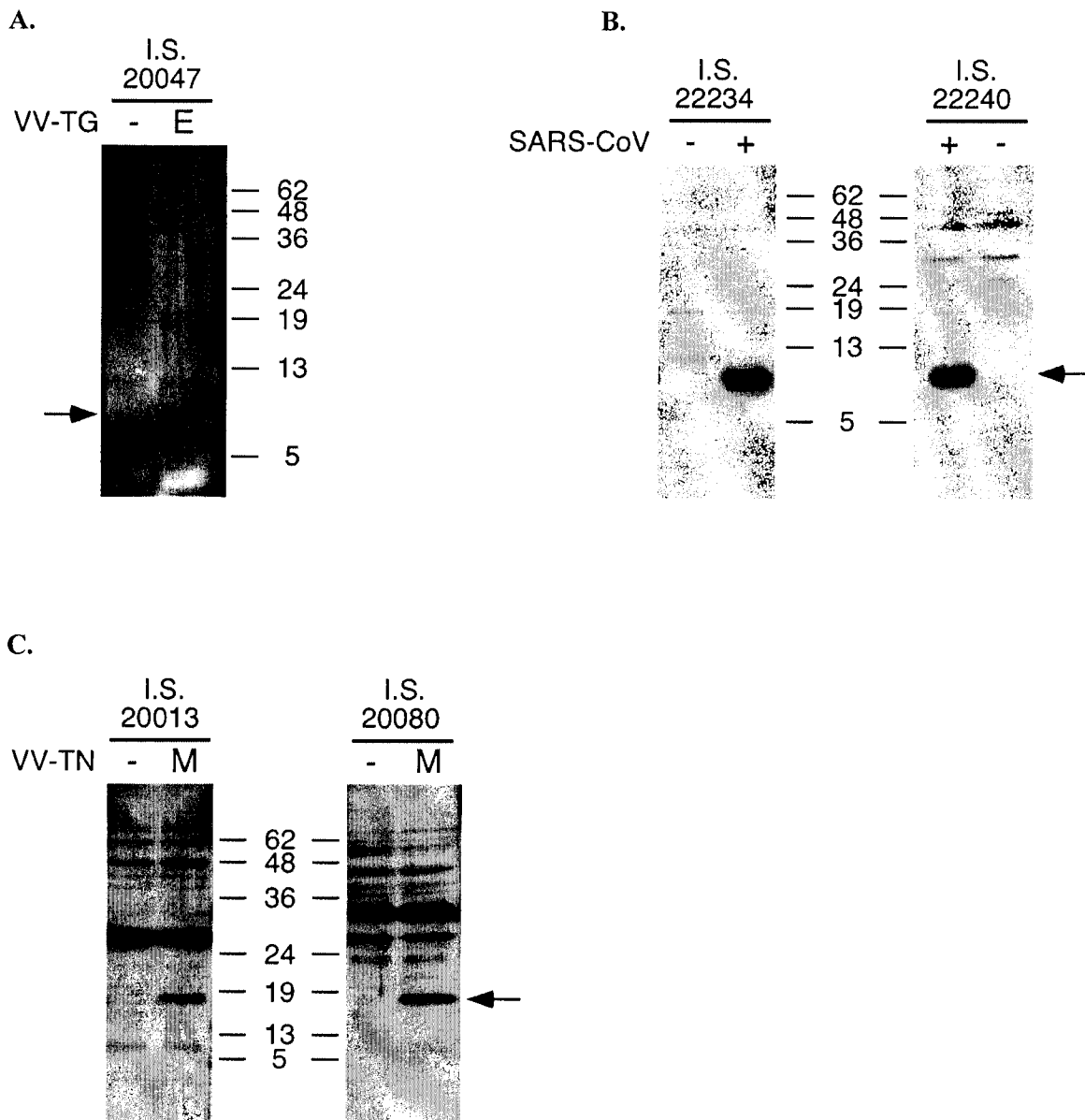


FIGURE 43

**STRAIN OF SARS-ASSOCIATED
CORONAVIRUS AND APPLICATIONS
THEREOF**

This is a division of application Ser. No. 10/581,356, filed Feb. 8, 2007, now U.S. Pat. No. 7,736,850, which is a continuation of International Application No. PCT/FR2004/003106, filed Dec. 2, 2004, both of which are incorporated herein by reference.

The present invention relates to a novel strain of severe acute respiratory syndrome (SARS)-associated coronavirus derived from a sample recorded under No. 031589 and collected in Hanoi (Vietnam), to nucleic acid molecules derived from its genome, to the proteins and peptides encoded by said nucleic acid molecules and to their applications, in particular as diagnostic reagents and/or as vaccine.

Coronavirus is a virus containing single-stranded RNA, of positive polarity, of approximately 30 kilobases which replicates in the cytoplasm of the host cells; the 5' end of the genome has a capped structure and the 3' end contains a polyA tail. This virus is enveloped and comprises, at its surface, peplomeric structures called spicules.

The genome comprises the following open reading frames or ORFs, from its 5' end to its 3' end: ORF1a and ORF1b corresponding to the proteins of the transcription-replication complex, and ORF-S, ORF-E, ORF-M and ORF-N corresponding to the structural proteins S, E, M and N. It also comprises ORFs corresponding to proteins of unknown function encoded by: the region situated between ORF-S and ORF-E and overlapping the latter, the region situated between ORF-M and ORF-N, and the region included in ORF-N.

The S protein is a membrane glycoprotein (200-220 kDa) which exists in the form of spicules or spikes emerging from the surface of the viral envelope. It is responsible for the attachment of the virus to the receptors of the host cell and for inducing the fusion of the viral envelope with the cell membrane.

The small envelope protein (E), also called sM (small membrane), which is a nonglycosylated transmembrane protein of about 10 kDa, is the protein present in the smallest quantity in the virion. It plays a powerful role in the coronavirus budding process which occurs at the level of the intermediate compartment in the endoplasmic reticulum and the Golgi apparatus.

The M protein or matrix protein (25-30 kDa) is a more abundant membrane glycoprotein which is integrated into the viral particle by an M/E interaction, whereas the incorporation of S into the particles is directed by an S/M interaction. It appears to be important for the viral maturation of coronaviruses and for the determination of the site where the viral particles are assembled.

The N protein or nucleocapsid protein (45-50 kDa) which is the most conserved among the coronavirus structural proteins is necessary for encapsidating the genomic RNA and then for directing its incorporation into the virion. This protein is probably also involved in the replication of the RNA.

When the host cell is infected, the reading frame (ORF) situated in 5' of the viral genome is translated into a polypeptide which is cleaved by the viral proteases and then releases several nonstructural proteins such as the RNA-dependent RNA polymerase (Rep) and the ATPase helicase (Hel). These two proteins are involved in the replication of the viral genome and in the generation of transcripts which are used in the synthesis of the viral proteins. The mechanisms by which these subgenomic mRNAs are produced are not completely understood; however, recent facts indicate that the sequences for regulation of transcription at the 5' end of each gene

represent signals which regulate the discontinuous transcription of the subgenomic mRNAs.

The proteins of the viral membrane (S, E and M proteins) are inserted into the intermediate compartment, whereas the replicated RNA (+ strand) is assembled with the N (nucleocapsid) protein. This protein-RNA complex then combines with the M protein contained in the membranes of the endoplasmic reticulum and the viral particles form when the nucleocapsid complex buds into the endoplasmic reticulum. The virus then migrates across the Golgi complex and eventually leaves the cell, for example by exocytosis. The site of attachment of the virus to the host cell is at the level of the S protein.

Coronaviruses are responsible for 15 to 30% of colds in humans and for respiratory and digestive infections in animals, especially cats (FIPV: Feline infectious peritonitis virus), poultry (IBV: Avian infectious bronchitis virus), mice (MHV: Mouse hepatitis virus), pigs (TGEV: Transmissible gastroenteritis virus, PEDV: Porcine Epidemic diarrhea virus, PRCoV: Porcine Respiratory Coronavirus, HEV: Hemagglutinating encephalomyelitis Virus) and bovines (BCoV: Bovine coronavirus).

In general, each coronavirus affects only one species; in immunocompetent individuals, the infection induces optionally neutralizing antibodies and cell immunity, capable of destroying the infected cells.

An epidemic of atypical pneumonia, called severe acute respiratory syndrome (SARS) has spread in various countries (Vietnam, Hong Kong, Singapore, Thailand and Canada) during the first quarter of 2003, from an initial focus which appeared in China in the last quarter of 2002. The severity of this disease is such that its mortality rate is about 3 to 6%. The determination of the causative agent of this disease is underway by numerous laboratories worldwide.

In March 2003, a new coronavirus (SARS-CoV or SARS virus) was isolated, in association with cases of severe acute respiratory syndrome (T. G. KSIAZEK et al., *The New England Journal of Medicine*, 2003, 348, 1319-1330; C. DROSTEN et al., *The New England Journal of Medicine*, 2003, 348, 1967-1976; Peiris et al., *Lancet*, 2003, 361, 1319).

Genomic sequences of this new coronavirus have thus been obtained, in particular those of the Urbani isolate (Genbank accession No. AY274119.3 and A. MARRA et al., *Science*, May 1, 2003, 300, 1399-1404) and the Toronto isolate (Tor2, Genbank accession No. AY278741 and A. ROTA et al., *Science*, 2003, 300, 1394-1399).

The organization of the genome is comparable with that of other known coronaviruses, thus making it possible to confirm that SARS-CoV belongs to the Coronaviridae family; open reading frames ORF1a and 1b and open reading frames corresponding to the S, E, M and N proteins, and to proteins encoded by: the region situated between ORF-S and ORF-E (ORF3), the region situated between ORF-S and ORF-E and overlapping. ORF-E (ORF4), the region situated between ORF-M and ORF-N (ORF7 to ORF11) and the region corresponding to ORF-N (ORF13 and ORF14), have in particular been identified.

Seven differences have been identified between the sequences of the Tor2 and Urbani isolates; 3 correspond to silent mutations (c/t at position 16622 and a/g at position 19064 of ORF1b, t/c at position 24872 of ORF-S) and 4 modify the amino acid sequence of respectively: the proteins encoded by ORF1a (c/t at position 7919 corresponding to the A/V mutation), the S protein (g/t at position 23220 corresponding to the A/S mutation), the protein encoded by ORF3

(a/g at position 25298 corresponding to the R/G mutation) and the M protein (t/c at position 26857 corresponding to the S/P mutation).

In addition, phylogenetic analysis shows that SARS-CoV is distant from other coronaviruses and that it did not appear by mutation of human respiratory coronaviruses nor by recombination between known coronaviruses (for a review, see Holmes, J. C. I., 2003, 111, 1605-1609).

The determination and the taking into account of new variants are important for the development of reagents for the detection and diagnosis of SARS which are sufficiently sensitive and specific, and immunogenic compositions capable of protecting populations against epidemics of SARS.

The inventors have now identified another strain of SARS-associated coronavirus which is distinguishable from the Tor2 and Urbani isolates.

The subject of the present invention is therefore an isolated or purified strain of severe acute respiratory syndrome-associated human coronavirus, characterized in that its genome has, in the form of complementary DNA, a serine codon at position 23220-23222 of the gene for the S protein or a glycine codon at position 25298-25300 of the gene for ORF3, and an alanine codon at position 7918-7920 of ORF1a or a serine codon at position 26857-26859 of the gene for the M protein, said positions being indicated in terms of reference to the Genbank sequence AY274119.3.

According to an advantageous embodiment of said strain, the DNA equivalent of its genome has a sequence corresponding to the sequence SEQ ID No: 1; this coronavirus strain is derived from the sample collected from the bronchoalveolar washings from a patient suffering from SARS, recorded under the No. 031589 and collected at the Hanoi (Vietnam) French hospital.

In accordance with the invention, said sequence SEQ ID No: 1 is that of the deoxyribonucleic acid corresponding to the ribonucleic acid molecule of the genome of the isolated coronavirus strain as defined above.

The sequence SEQ ID No: 1 is distinguishable from the Genbank sequence AY274119.3 (Tor2 isolate) in that it possesses the following mutations:

g/t at position 23220; the alanine codon (gct) at position 577 of the amino acid sequence of the Tor2 S protein is replaced by a serine codon (tct),

a/g at position 25298; the arginine codon (aga) at position 11 of the amino acid sequence of the protein encoded by the Tor2 ORF3 is replaced by a glycine codon (gga).

In addition, the sequence SEQ ID No: 1 is distinguishable from the Genbank sequence AY278741 (Urbani isolate) in that it possesses the following mutations:

t/c at position 7919; the valine codon (gtt) in position 2552 of the amino acid sequence of the protein encoded by ORF1a is replaced by an alanine codon (gct),

t/c at position 16622: this mutation does not modify the amino acid sequence of the proteins encoded by ORF1b (silent mutation),

g/a at position 19064: this mutation does not modify the amino acid sequence of the proteins encoded by ORF1b (silent mutation),

c/t at position 24872: this mutation does not modify the amino acid sequence of the S protein, and

c/t at position 26857: the proline codon (ccc) at position 154 of the amino acid sequence of the M protein is replaced by a serine codon (tcc).

Unless otherwise stated, the positions of the nucleotide and peptide sequences are indicated with reference to the Genbank sequence AY274119.3.

The subject of the present invention is also an isolated or purified polynucleotide, characterized in that its sequence is that of the genome of the isolated coronavirus strain as defined above.

According to an advantageous embodiment of said polynucleotide, it has the sequence SEQ ID No: 1.

The subject of the present invention is also an isolated or purified polynucleotide, characterized in that its sequence hybridizes under high stringency conditions with the sequence of the polynucleotide as defined above.

The terms "isolated or purified" mean modified "by the hand of humans" from the natural state; in other words if an object exists in nature, it is said to be isolated or purified if it is modified or extracted from its natural environment or both. For example, a polynucleotide or a protein/peptide naturally present in a living organism is neither isolated nor purified; on the other hand, the same polynucleotide or protein/peptide separated from coexisting molecules in its natural environment, obtained by cloning, amplification and/or chemical synthesis is isolated for the purposes of the present invention. Furthermore, a polynucleotide or a protein/peptide which is introduced into an organism by transformation, genetic manipulation or by any other method, is "isolated" even if it is present in said organism. The term purified as used in the present invention means that the proteins/peptides according to the invention are essentially free of association with the other proteins or polypeptides, as is for example the product purified from the culture of recombinant host cells or the product purified from a nonrecombinant source.

For the purposes of the present invention, high stringency hybridization conditions are understood to mean temperature and ionic strength conditions chosen such that they make it possible to maintain the specific and selective hybridization between complementary polynucleotides.

By way of illustration, high stringency conditions for the purposes of defining the above polynucleotides are advantageously the following: the DNA-DNA or DNA-RNA hybridization is performed in two steps: (1) prehybridization at 42° C. for 3 hours in phosphate buffer (20 mM, pH 7.5) containing 5×SSC (1×SSC corresponds to a 0.15 M NaCl+0.015 M sodium citrate solution), 50% formamide, 7% sodium dodecyl sulfate (SDS), 10×Denhardt's, 5% dextran sulfate and 1% salmon sperm DNA; (2) hybridization for 20 hours at 42° C. followed by 2 washings of 20 minutes at 20° C. in 2×SSC+2% SDS, 1 washing of 20 minutes at 20° C. in 0.1×SSC+0.1% SDS. The final washing is performed in 0.1×SSC+0.1% SDS for 30 minutes at 60° C.

The subject of the present invention is also a representative fragment of the polynucleotide as defined above, characterized in that it is capable of being obtained either by the use of restriction enzymes whose recognition and cleavage sites are present in said polynucleotide as defined above, or by amplification with the aid of oligonucleotide primers specific for said polynucleotide as defined above, or by transcription in vitro, or by chemical synthesis.

According to an advantageous embodiment of said fragment, it is selected from the group consisting of: the cDNA corresponding to at least one open reading frame (ORF) chosen from: ORF1a, ORF1b, ORF-S, ORF-E, ORF-M, ORF-N, ORF3, ORF4, ORF7 to ORF11, ORF13 and ORF14 and the cDNA corresponding to the noncoding 5' or 3' ends of said polynucleotide.

According to an advantageous feature of this embodiment, said fragment has a sequence selected from the group consisting of:

5

the sequences SEQ ID NO: 2 and 4 representing the cDNA corresponding to the ORF-S which encodes the S protein,

the sequences SEQ ID NO: 13 and 15 representing the cDNA corresponding to the ORF-E which encodes the E protein,

the sequences SEQ ID NO: 16 and 18 representing the cDNA corresponding to the ORF-M which encodes the M protein,

the sequences SEQ ID NO: 36 and 38 representing the cDNA corresponding to the ORF-N which encodes the N protein,

the sequences representing the cDNA corresponding respectively: to ORF1a and ORF1b (ORF1ab, SEQ ID NO: 31), to ORF3 and ORF4 (SEQ ID NO: 7, 8), to ORF7 to 11 (SEQ ID NO: 19, 20) to ORF13 (SEQ ID NO: 32) and to ORF14 (SEQ ID NO: 34), and

the sequences representing the cDNAs corresponding respectively to the noncoding 5' (SEQ ID NO: 39 and 72) and 3' (SEQ ID NO: 40, 73) ends of said polynucleotide.

The subject of the present invention is also a cDNA fragment encoding the S protein, as defined above, characterized in that it has a sequence selected from the group consisting of the sequences SEQ ID NO: 5 and 6 (Sa and Sb fragments).

The subject of the present invention is also a cDNA fragment corresponding to ORF1a and ORF1b as defined above, characterized in that it has a sequence selected from the group consisting of the sequences SEQ ID NO: 41 to 54 (L0 to L12 fragments).

The subject of the present invention is also a polynucleotide fragment as defined above, characterized in that it has at least 15 consecutive bases or base pairs of the sequence of the genome of said strain including at least one of those situated in position 7979, 16622, 19064, 23220, 24872, 25298 and 26857. Preferably this is a fragment of 20 to 2500 bases or base pairs, preferably from 20 to 400.

According to an advantageous embodiment of said fragment, it includes at least one pair of bases or base pairs corresponding to the following positions: 7919 and 23220, 7919 and 25298, 16622 and 23220, 19064 and 23220, 16622 and 25298, 19064 and 25298, 23220 and 24872, 23220 and 26857, 24872 and 25298, 25298 and 26857.

The subject of the present invention is also primers of at least 18 bases capable of amplifying a fragment of the genome of a SARS-associated coronavirus or of the DNA equivalent thereof.

According to an embodiment of said primers, they are selected from the group consisting of:

the pair of primers No. 1 corresponding respectively to positions 28507 to 28522 (sense primer, SEQ ID NO: 60) and 28774 to 28759 (antisense primer, SEQ ID NO: 61) of the sequence of the polynucleotide as defined above,

the pair of primers No. 2 corresponding respectively to positions 28375 to 28390 (sense primer, SEQ ID NO: 62) and 28702 to 28687 (antisense primer, SEQ ID NO: 63) of the sequence of the polynucleotide as defined above, and

the pair of primers consisting of the primers SEQ ID Nos: 55 and 56.

The subject of the present invention is also a probe capable of detecting the presence of the genome of a SARS-associated coronavirus or of a fragment thereof, characterized in that it is selected from the group consisting of: the fragments as defined above and the fragments corresponding to the following positions of the polynucleotide sequence as defined

6

above: 28561 to 28586, 28588 to 28608, 28541 to 28563 and 28565 to 28589 (SEQ ID NO: 64 to 67).

The probes and primers according to the invention may be labeled directly or indirectly with a radioactive or nonradioactive compound by methods well known to persons skilled in the art so as to obtain a detectable and/or quantifiable signal. Among the radioactive isotopes used, there may be mentioned ^{32}P , ^{33}P , ^{35}S , ^3H or ^{125}I . The nonradioactive entities are selected from ligands such as biotin, avidin, streptavidin, digoxigenin, haptens, dyes, luminescent agents such as radioluminescent, chemoluminescent, bioluminescent, fluorescent and phosphorescent agents.

The invention encompasses the labeled probes and primers derived from the preceding sequences.

Such probes and primers are useful for the diagnosis of infection by a SARS-associated coronavirus.

The subject of the present invention is also a method for the detection of a SARS-associated coronavirus, from a biological sample, which method is characterized in that it comprises at least:

(a) the extraction of nucleic acids present in said biological sample,

(b) the amplification of a fragment of ORF-N by RT-PCR with the aid of a pair of primers as defined above, and

(c) the detection, by any appropriate means, of the amplification products obtained in (b).

The amplification products (amplicons) in (b) are 268 bp for the pair of primers No. 1 and 328 bp for the pair of primers No. 2.

According to an advantageous embodiment of said method, the step (b) of detection is carried out with the aid of at least one probe corresponding to positions 28561 to 28586, 28588 to 28608, 28541 to 28563 and 28565 to 28589 of the sequence of the polynucleotide as defined above.

Preferably, the SARS-associated coronavirus genome is detected and optionally quantified by PCR in real time with the aid of the pair of primers No. 2 and probes corresponding to positions 28541 to 28563 and 28565 to 28589 labeled with different compounds, in particular different fluorescent agents.

The real time RT-PCR which uses this pair of primers and this probe is very sensitive since it makes it possible to detect 10^2 copies of RNA and up to 10 copies, of RNA; it is in addition reliable and reproducible.

The invention encompasses the single-stranded, double-stranded and triple-stranded polydeoxyribonucleotides and polyribonucleotides corresponding to the sequence of the genome of the isolated strain of coronavirus and its fragments as defined above, and to their sense or antisense complementary sequences, in particular the RNAs and cDNAs corresponding to the sequence of the genome and of its fragments as defined above.

The present invention also encompasses the amplification fragments obtained with the aid of primers specific for the genome of the purified or isolated strain as defined above, in particular with the aid of primers or pairs of primers as defined above, the restriction fragments formed by or comprising the sequence of fragments as defined above, the fragments obtained by transcription *in vitro* from a vector containing the sequence SEQ ID NO: 1 or a fragment as defined above, and fragments obtained by chemical synthesis. Examples of restriction fragments are deduced from the restriction map of the sequence SEQ ID NO: 1 illustrated by FIG. 13. In accordance with the invention, said fragments are either in the form of isolated fragments, or in the form of mixtures of fragments. The invention also encompasses fragments modified, in relation to the preceding ones, by removal

or addition of nucleotides in a proportion of about 15%, relative to the length of the above fragments and/or modified in terms of the nature of the nucleotides, as long as the modified nucleotide fragments retain a capacity for hybridization with the genomic or antigenomic RNA sequences of the isolate as defined above. 5

The nucleic acid molecules according to the invention are obtained by conventional methods, known per se, following standard protocols such as those described in *Current Protocols in Molecular Biology* (Frederick M. AUSUBEL, 2000, Wiley and son Inc., Library of Congress, USA). For example, they may be obtained by amplification of a nucleic sequence by PCR or RT-PCR or alternatively by total or partial chemical synthesis. 10

The subject of the present invention is also a DNA or RNA chip or filter, characterized in that it comprises at least one polynucleotide or one of its fragments as defined above. 15

The DNA or RNA chips or filters according to the invention are prepared by conventional methods, known per se, such as for example chemical or electrochemical grafting of oligonucleotides on a glass or nylon support. 20

The subject of the present invention is also a recombinant cloning and/or expression vector, in particular a plasmid, a virus, a viral vector or a phage comprising a nucleic acid fragment as defined above. Preferably, said recombinant vector is an expression vector in which said nucleic acid fragment is placed under the control of appropriate elements for regulating transcription and translation. In addition, said vector may comprise sequences (tags) fused in phase with the 5' and/or 3' end of said insert, which are useful for the immobilization and/or detection and/or purification of the protein expressed from said vector. 25 30

These vectors are constructed and introduced into host cells by conventional recombinant DNA and genetic engineering methods which are known per se. Numerous vectors into which a nucleic acid molecule of interest may be inserted in order to introduce it and to maintain it in a host cell are known per se; the choice of an appropriate vector depends on the use envisaged for this vector (for example replication of the sequence of interest, expression of this sequence, maintenance of the sequence in extrachromosomal form or alternatively integration into the chromosomal material of the host), and on the nature of the host cell. 35 40

In accordance with the invention, said plasmid is selected in particular from the following plasmids: 45

the plasmid, called SARS-S, contained in the bacterial strain deposited under the No. I-3059, on Jun. 20, 2003, at the Collection Nationale de Cultures de Microorganismes, 25 rue du Docteur Roux, 75724 Paris Cedex 15; it contains the cDNA sequence encoding the S protein of the SARS-CoV strain derived from the sample recorded under the No. 031589, said sequence corresponding to the nucleotides at positions 21406 to 25348 (SEQ ID NO: 4), with reference to the Genbank sequence AY274119.3, 50 55

the plasmid, called SARS-S1, contained in the bacterial strain deposited under the No. I-3020, on May 12, 2003, at the Collection Nationale de Cultures de Microorganismes, 25 rue du Docteur Roux, 75724 Paris Cedex 15; it contains a 5' fragment of the cDNA sequence encoding the S protein of the SARS-CoV strain derived from the sample recorded under the No. 031589, as defined above, said fragment corresponding to the nucleotides at positions 21406 to 23454 (SEQ ID NO: 5), with reference to the Genbank sequence AY274119.3 Tor2, 60 65

the plasmid, called SARS-S2, contained in the bacterial strain deposited under the No. I-3019, on May 12, 2003,

at the Collection Nationale de Cultures de Microorganismes, 25 rue du Docteur Roux, 75724 Paris Cedex 15; it contains a 3' fragment of the cDNA sequence encoding the S protein of the SARS-CoV strain derived from the sample recorded under the number No. 031589, as defined above, said fragment corresponding to the nucleotides at positions 23322 to 25348 (SEQ ID NO: 6), with reference to the Genbank sequence accession No. AY274119.3,

the plasmid, called SARS-SE, contained in the bacterial strain deposited under the No. I-3126, on Nov. 13, 2003, at the Collection Nationale de Cultures de Microorganismes, 25 rue du Docteur Roux, 75724 Paris Cedex 15; it contains the cDNA corresponding to the region situated between ORF-S and ORF-E and overlapping ORF-E of the SARS-CoV strain derived from the sample recorded under the No. 031589, as defined above, said region corresponding to the nucleotides at positions 25110 to 26244 (SEQ ID NO: 8), with reference to the Genbank sequence accession No. AY274119.3,

the plasmid, called SARS-E, contained in the bacterial strain deposited under the No. I-3046, on May 28, 2003, at the Collection Nationale de Cultures de Microorganismes, 25 rue du Docteur Roux, 75724 Paris Cedex 15; it contains the cDNA sequence encoding the E protein of the SARS-CoV strain derived from the sample recorded under the No. 031589, as defined above, said sequence corresponding to the nucleotides at positions 26082 to 26413 (SEQ ID NO: 15), with reference to the Genbank sequence accession No. AY274119.3,

the plasmid, called SARS-M, contained in the bacterial strain deposited under the No. I-3047, on May 28, 2003, at the Collection Nationale de Cultures de Microorganismes, 25 rue du Docteur Roux, 75724 Paris Cedex 15; it contains the cDNA sequence encoding the M protein of the SARS-CoV strain derived from the sample recorded under the No. 031589, as defined above; said sequence corresponding to the nucleotides at positions 26330 to 27098 (SEQ ID NO: 18), with reference to the Genbank sequence accession No. AY274119.3,

the plasmid, called SARS-MN, contained in the bacterial strain deposited under the No. I-3125, on Nov. 13, 2003, at the Collection Nationale de Cultures de Microorganismes, 25 rue du Docteur Roux, 75724 Paris Cedex 15; it contains the cDNA sequence corresponding to the region situated between ORF-M and ORF-N of the SARS-CoV strain derived from the sample recorded under the No. 031589 and collected in Hanoi, as defined above, said sequence corresponding to the nucleotides at positions 26977 to 28218 (SEQ ID NO: 20), with reference to the Genbank accession No. AY274119.3,

the plasmid, called SARS-N, contained in the bacterial strain deposited under the No. I-3048, on Jun. 5, 2003, at the Collection Nationale de Cultures de Microorganismes, 25 rue du Docteur Roux, 75724 Paris Cedex 15; it contains the cDNA encoding the N protein of the SARS-CoV strain derived from the sample recorded under the No. 031589, as defined above, said sequence corresponding to the nucleotides at positions 28054 to 29430 (SEQ ID NO: 38), with reference to the Genbank sequence accession No. AY274119.3; thus, this plasmid comprises an insert of sequence SEQ ID NO: 38 and is contained in a bacterial strain which was deposited under the No. I-3048, on Jun. 5, 2003, at the Collection Nationale de Cultures de Microorganismes, 25 rue du Docteur Roux, 75724 Paris Cedex 15,

the plasmid, called SARS-5'NC, contained in the bacterial strain deposited under the No. I-3124, on Nov. 7, 2003, at the Collection Nationale de Cultures de Microorganismes, 25 rue du Docteur Roux, 75724 Paris Cedex 15; it contains the cDNA corresponding to the noncoding 5' end of the genome of the SARS-CoV strain derived from the sample recorded under the No. 031589, as defined above, said sequence corresponding to the nucleotides at positions 1 to 204 (SEQ ID NO: 39), with reference to the Genbank sequence accession No. AY274119.3,

the plasmid called SARS-3'NC, contained in the bacterial strain deposited under the No. I-3123 on Nov. 7, 2003, at the Collection Nationale de Cultures de Microorganismes, 25 rue du Docteur Roux, 75724 Paris Cedex 15; it contains the cDNA sequence corresponding to the non-coding 3' end of the genome of the SARS-CoV strain derived from the sample recorded under the No. 031589, as defined above, said sequence corresponding to that situated between the nucleotide and position 28933 to 29727 (SEQ ID NO: 40), with reference to the Genbank sequence accession No. AY274119.3, ends with a series of nucleotides a,

the expression plasmid, called pIV2.3N, containing a cDNA fragment encoding a C-terminal fusion of the N protein (SEQ ID NO: 37) with a polyhistidine tag,

the expression plasmid, called pIV2.3S_C, containing a cDNA fragment encoding a C-terminal fusion of the fragment corresponding to positions 475 to 1193 of the amino acid sequence of the S protein (SEQ ID NO: 3) with a polyhistidine tag,

the expression plasmid, pIV2.3S_L, containing a cDNA fragment encoding a C-terminal fusion of the fragment corresponding to positions 14 to 1193 of the amino acid sequence of the S protein (SEQ ID NO: 3) with a polyhistidine tag,

the expression plasmid, called pIV2.4N, containing a cDNA fragment encoding a N-terminal fusion of the N protein (SEQ ID NO: 3) with a polyhistidine tag,

the expression plasmid, called pIV2.4S_C or pIV2.4S₁, containing an insert encoding a N-terminal fusion of the fragment corresponding to positions 475 to 1193 of the amino acid sequence of the S protein (SEQ ID NO: 3) with a polyhistidine tag, and

the expression plasmid, called pIV2.4S_L, containing a cDNA fragment encoding an N-terminal fusion of the fragment corresponding to positions 14 to 1193 of the amino acid sequence of the S protein (SEQ ID NO: 3) with a polyhistidine tag.

According to an advantageous feature of the expression plasmid as defined above, it is contained in a bacterial strain which was deposited under the No. I-3117, on Oct. 23, 2003, at the Collection Nationale de Cultures de Microorganismes, 25 rue du Docteur Roux, 75724 Paris Cedex 15.

According to another advantageous feature of the expression plasmid as defined above, it is contained in a bacterial strain which was deposited under the No. I-3118, on Oct. 23, 2003, at the Collection Nationale de Cultures de Microorganismes, 25 rue du Docteur Roux, 75724 Paris Cedex 15.

According to another feature of the expression plasmid as defined above, it is contained in a bacterial strain which was deposited at the CNCM, 25 rue du Docteur Roux, 75724 Paris Cedex 15 under the following numbers:

- a) strain No. I-3118, deposited on Oct. 23, 2003,
- b) strain No. I-3019, deposited on May 12, 2003,
- c) strain No. I-3020, deposited on May 12, 2003,
- d) strain No. I-3059, deposited on Jun. 20, 2003,
- e) strain No. I-3323, deposited on Nov. 22, 2004,

- f) strain No. I-3324, deposited on Nov. 22, 2004,
- g) strain No. I-332, deposited on Dec. 1, 2004,
- h) strain No. I-3327, deposited on Dec. 1, 2004,
- i) strain No. I-3332, deposited on Dec. 1, 2004,
- j) strain No. I-3333, deposited on Dec. 1, 2004,
- k) strain No. I-3334, deposited on Dec. 1, 2004,
- l) strain No. I-3335, deposited on Dec. 1, 2004,
- m) strain No. I-3336, deposited on Dec. 1, 2004,
- n) strain No. I-3337, deposited on Dec. 1, 2004,
- o) strain No. I-3338, deposited on Dec. 2, 2004,
- p) strain No. I-3339, deposited on Dec. 2, 2004,
- q) strain No. I-3340, deposited on Dec. 2, 2004,
- r) strain No. I-3341, deposited on Dec. 2, 2004.

The subject of the present invention is also a nucleic acid insert of viral origin, characterized in that it is contained in any of the strains as defined above in a)-r).

The subject of the present invention is also a nucleic acid containing a synthetic gene allowing optimized expression of the S protein in eukaryotic cells, characterized in that it possesses the sequence SEQ ID NO: 140.

The subject of the present invention is also an expression vector containing a nucleic acid containing a synthetic gene allowing optimized expression of the S protein, which vector is contained in the bacterial strain deposited at the CNCM, on Dec. 1, 2004, under the No. I-3333.

According to one embodiment of said expression vector, it is a viral vector, in the form of a viral particle or in the form of a recombinant genome.

According to an advantageous feature of this embodiment, this is a recombinant viral particle or a recombinant viral genome capable of being obtained by transfection of a plasmid according to paragraphs g), h) and k) to r) as defined above, in an appropriate cellular system, that is to say, for example, cells transfected with one or more other plasmids intended to transcomplement certain functions of the virus that are deleted in the vector and that are necessary for the formation of the viral particles.

The expression "S protein family" is understood here to mean the complete S protein, its ectodomain and fragments of this ectodomain which are preferably produced in a eukaryotic system.

The subject of the present invention is also a lentiviral vector encoding a polypeptide of the S protein family, as defined above.

The subject of the present invention is also a recombinant measles virus encoding a polypeptide of the S protein family, as defined above.

The subject of the present invention is also a recombinant vaccinia virus encoding a polypeptide of the S protein family, as defined above.

The subject of the present invention is also the use of a vector according to paragraphs e) to r) as defined above, or of a vector containing a synthetic gene for the S protein, as defined above, for the production, in a eukaryotic system, of the SARS-associated coronavirus S protein or of a fragment of this protein.

The subject of the present invention is also a method for producing the S protein in a eukaryotic system, comprising a step of transfecting eukaryotic cells in culture with a vector chosen from the vectors contained in the bacterial strains mentioned in paragraphs e) to r) above or a vector containing a synthetic gene allowing optimized expression of the S protein.

The subject of the present invention is also a cDNA library characterized in that it comprises fragments as defined above,

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in particular amplification fragments or restriction fragments, cloned into a recombinant vector, in particular an expression vector (expression library).

The subject of the present invention is also cells, in particular prokaryotic cells, modified by a recombinant vector as defined above.

The subject of the present invention is also a genetically modified eukaryotic cell expressing a protein or a polypeptide as defined above. Quite obviously, the terms "genetically modified eukaryotic cell" do not denote a cell modified with a wild-type virus.

According to an advantageous embodiment of said cell, it is capable of being obtained by transfection with any of the vectors mentioned in paragraphs i) to l) above.

According to an advantageous feature of this embodiment, this is the cell FRhK4-Ssol-30, deposited at the CNCM on Nov. 22, 2004, under the No. I-3325.

The recombinant vectors as defined above and the cells transformed with said expression vectors are advantageously used for the production of the corresponding proteins and peptides. The expression libraries derived from said vectors, and the cells transformed with said expression libraries are advantageously used to identify the immunogenic epitopes (B and T epitopes) of the SARS-associated coronavirus proteins.

The subject of the present invention is also the purified or isolated proteins and peptides, characterized in that they are encoded by the polynucleotide or one of its fragments as defined above.

According to an advantageous embodiment of the invention, said protein is selected from the group consisting of:

the S protein having the sequence SEQ ID NO: 3 or its ectodomaine

the E protein having the sequence SEQ ID NO: 14

the M protein having the sequence SEQ ID NO: 17

the N protein having the sequence SEQ ID NO: 37

the proteins encoded by the ORFs: ORF1a, ORF1b, ORF3, ORF4 and ORF7 to ORF11, ORF13 and ORF14 and having the respective sequence, SEQ ID NO: 74, 75, 10, 12, 22, 24, 26, 28, 30, 33 and 35.

The terms "ectodomaine of the S protein" and "soluble form of the S protein" will be used interchangeably below.

According to an advantageous embodiment of the invention, said polypeptide consists of the amino acids corresponding to positions 1 to 1193 of the amino acid sequence of the S protein.

According to another advantageous embodiment of the invention, said peptide is selected from the group consisting of:

a) the peptides corresponding to positions 14 to 1193 and 475 to 1193 of the amino acid sequence of the S protein,

b) the peptides corresponding to positions 2 to 14 (SEQ ID NO: 69) and 100 to 221 of the amino acid sequence of the M protein; these peptides correspond respectively to the ectodomaine and to the endodomaine of the M protein, and

c) the peptides corresponding to positions 1 to 12 (SEQ ID NO: 70) and 53 to 76 (SEQ ID NO: 71) of the amino acid sequence of the E protein; these peptides correspond respectively to the ectodomaine and to the C-terminal end of the E protein, and

d) the peptides of 5 to 50 consecutive amino acids, preferably of 10 to 30 amino acids, inclusive or partially or completely overlapping the sequence of the peptides as defined in a), b) or c).

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The subject of the present invention is also a peptide, characterized in that it has a sequence of 7 to 50 amino acids including an amino acid residue selected from the group consisting of:

the alanine situated at position 2552 of the amino acid sequence of the protein encoded by ORF1a,

the serine situated at position 577 of the amino acid sequence of the S protein of the SARS-CoV strain as defined above,

the glycine at position 11 of the amino acid sequence, of the protein encoded by ORF3 of the SARS-CoV strain as defined above,

the serine at position 154 of the amino acid sequence of the M protein of the SARS-CoV strain as defined above.

The subject of the present invention is also an antibody or a polyclonal or monoclonal antibody fragment which can be obtained by immunization of an animal with a recombinant vector as defined above, a cDNA library as defined above or alternatively a protein or a peptide as defined above, characterized in that it binds to at least one of the proteins encoded by SARS-CoV as defined above.

The invention encompasses the polyclonal antibodies, the monoclonal antibodies, the chimeric antibodies such as the humanized antibodies, and fragments thereof (Fab, Fv, scFv).

A subject of the present invention is also a hybridoma producing a monoclonal antibody against the N protein, characterized in that it is chosen from the following hybridomas: the hybridoma producing the monoclonal antibody 87, deposited at the CNCM on Dec. 1, 2004 under the number I-3328,

the hybridoma producing the monoclonal antibody 86, deposited at the CNCM on Dec. 1, 2004 under the number I-3329,

the hybridoma producing the monoclonal antibody 57, deposited at the CNCM on Dec. 1, 2004 under the number I-3330, and

the hybridoma producing the monoclonal antibody 156, deposited at the CNCM on Dec. 1, 2004 under the number I-3331.

The subject of the present invention is also a polyclonal or monoclonal antibody or antibody fragment directed against the N protein, characterized in that it is produced by a hybridoma as defined above.

For the purposes of the present invention, the expression chimeric antibody is understood to mean, in relation to an antibody of a particular animal species or of a particular class of antibody, an antibody comprising all or part of a heavy chain and/or of a light chain of an antibody of another animal species or of another class of antibody.

For the purposes of the present invention, the expression humanized antibody is understood to mean a human immunoglobulin in which the residues of the CDRs (Complementary Determining Regions) which form the antigen-binding site are replaced by those of a nonhuman monoclonal antibody possessing the desired specificity, affinity or activity. Compared with the nonhuman antibodies, the humanized antibodies are less immunogenic and possess a prolonged half-life in humans because they possess only a small proportion of nonhuman sequences given that practically all the residues of the FR (Framework) regions and of the constant (Fc) region of these antibodies are those of a consensus sequence of human immunoglobulins.

A subject of the present invention is also a protein chip or filter, characterized in that it comprises a protein, a peptide or alternatively an antibody as defined above.

The protein chips according to the invention are prepared by conventional methods known per se. Among the appropri-

ate supports on which proteins may be immobilized, there may be mentioned those made of plastic or glass, in particular in the form of microplates.

The subject of the present invention is also reagents derived from the isolated strain of SARS-associated coronavirus, derived from the sample recorded under the No. 031589, which are useful for the study and diagnosis of the infection caused by a SARS-associated coronavirus, said reagents are selected from the group consisting of:

- (a) a pair of primers, a probe or a DNA chip as defined above,
- (b) a recombinant vector or a modified cell as defined above,
- (c) an isolated coronavirus strain or a polynucleotide as defined above,
- (d) a protein or a peptide as defined above,
- (e) an antibody or an antibody fragment as defined above, and
- (f) a protein chip as defined above.

These various reagents are prepared and used according to conventional molecular biology and immunology techniques following standard protocols such as those described in *Current Protocols in Molecular Biology* (Frederick M. AUSUBEL, 2000, Wiley and Son Inc., Library of Congress, USA), in *Current Protocols in Immunology* (John E. Coligan, 2000, Wiley and Son Inc., Library of Congress, USA) and in *Antibodies: A Laboratory Manual* (E. Howell and D. Lane, Cold Spring Harbor Laboratory, 1988).

The nucleic acid fragments according to the invention are prepared and used according to conventional techniques as defined above. The peptides and proteins according to the invention are prepared by recombinant DNA techniques, known to persons skilled in the art, in particular with the aid of the recombinant vectors as defined above. Alternatively, the peptides according to the invention may be prepared by conventional techniques of solid or liquid phase synthesis, known to persons skilled in the art.

The polyclonal antibodies are prepared by immunizing an appropriate animal with a protein or a peptide as defined above, optionally coupled to KLH or to albumin and/or combined with an appropriate adjuvant such as (complete or incomplete) Freund's adjuvant or aluminum hydroxide; after obtaining a satisfactory antibody titer, the antibodies are harvested by collecting serum from the immunized animals and enriched with IgG by precipitation, according to conventional techniques, and then the IgGs specific for the SARS-CoV proteins are optionally purified by affinity chromatography on an appropriate column to which said peptide or said protein is attached, as defined above, so as to obtain a monospecific IgG preparation.

The monoclonal antibodies are produced from hybridomas obtained by fusion of B lymphocytes from an animal immunized with a protein or a peptide as defined above with myelomas, according to the Köhler and Milstein technique (Nature, 1975, 256, 495-497); the hybridomas are cultured in vitro, in particular in fermenters or produced in vivo, in the form of ascites; alternatively, said monoclonal antibodies are produced by genetic engineering as described in American patent U.S. Pat. No. 4,816,567.

The humanized antibodies are produced by general methods such as those described in International application WO 98/45332.

The antibody fragments are produced from the cloned V_H and V_L regions, from the mRNAs of hybridomas or splenic lymphocytes of an immunized mouse; for example, the Fv, scFv or Fab fragments are expressed at the surface of filamentous phages according to the Winter and Milstein tech-

nique (Nature, 1991, 349, 293-299); after several selection steps, the antibody fragments specific for the antigen are isolated and expressed in an appropriate expression system, by conventional techniques for cloning and expression of recombinant DNA.

The antibodies or fragments thereof as defined above are purified by conventional techniques known to persons skilled in the art, such as affinity chromatography.

The subject of the present invention is additionally the use of a product selected from the group consisting of: a pair of primers, a probe, a DNA chip, a recombinant vector, a modified cell, an isolated coronavirus strain, a polynucleotide, a protein or a peptide, an antibody or an antibody fragment and a protein chip as defined above, for the preparation of a reagent for the detection and optionally genotyping/serotyping of a SARS-associated coronavirus.

The proteins and peptides according to the invention, which are capable of being recognized and/or of inducing the production of antibodies specific for the SARS-associated coronavirus, are useful for the diagnosis of infection with such a coronavirus; the infection is detected, by an appropriate technique—in particular EIA, ELISA, RIA, immunofluorescence—in a biological sample collected from an individual capable of being infected.

According to an advantageous feature of said use, said proteins are selected from the group consisting of the S, E, M and/or N proteins and the peptides as defined above.

The S, E, M and/or N proteins and the peptides derived from these proteins as defined above, for example the N protein, are used for the indirect diagnosis of a SARS-associated coronavirus infection (serological diagnosis; detection of an antibody specific for SARS-CoV), in particular by an immunoenzymatic method (ELISA).

The antibodies and antibody fragments according to the invention, in particular those directed against the S, E, M and/or N proteins and the derived peptides as defined above, are useful for the direct diagnosis of a SARS-associated coronavirus infection; the detection of the protein(s) of SARS-CoV is carried out by an appropriate technique, in particular EIA, ELISA, RIA, immunofluorescence, in a biological sample collected from an individual capable of being infected.

The subject of the present invention is also a method for the detection of a SARS-associated coronavirus, from a biological sample, which method is characterized in that it comprises at least:

- (a) bringing said biological sample into contact with at least one antibody or one antibody fragment, one protein, one peptide or alternatively one protein or peptide chip or filter as defined above, and
- (b) visualizing by any appropriate means antigen-antibody complexes formed in (a), for example by EIA, ELISA, RIA, or by immunofluorescence.

According to one advantageous embodiment of said process, step (a) comprises:

- (a₁) bringing said biological sample into contact with at least a first antibody or an antibody fragment which is attached to an appropriate support, in particular a microplate,
- (a₂) washing the solid phase, and
- (a₃) adding at least a second antibody or an antibody fragment, different from the first, said antibody or antibody fragment being optionally appropriately labeled.

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This method, which makes it possible to capture the viral particles present in the biological sample, is also called immunocapture method.

For example:

step (a₁) is carried out with at least a first monoclonal or polyclonal antibody or a fragment thereof, directed against the S, M and/or E protein, and/or a peptide corresponding to the ectodomain of one of these proteins (M2-14 or E1-12 peptides)

step (a₃) is carried out with at least one antibody or an antibody fragment directed against another epitope of the same protein or preferably against another protein, preferably against an inner protein such as the N nucleoprotein or the endodomain of the E or M protein, more preferably still these are antibodies or antibody fragments directed against the N protein which is very abundant in the viral particle; when an antibody or an antibody fragment directed against an inner protein (N) or against the endodomain of the E or M proteins is used, said antibody is incubated in the presence of detergent, such as Tween 20 for example, at concentrations of the order of 0.1%.

step (b) for visualizing the antigen-antibody complexes formed is carried out, either directly with the aid of a second antibody labeled for example with biotin or an appropriate enzyme such as peroxidase or alkaline phosphatase, or indirectly with the aid of an anti-immunoglobulin serum labeled as above. The complexes thus formed are visualized with the aid of an appropriate substrate.

According to a preferred embodiment of this aspect of the invention, the biological sample is mixed with the visualizing monoclonal antibody prior to its being brought into contact with the capture monoclonal antibodies. Where appropriate, the serum-visualizing antibody mixture is incubated for at least 10 minutes at room temperature before being applied to the plate.

The subject of the present invention is also an immunocapture test intended to detect an infection by the SARS-associated coronavirus by detecting the native nucleoprotein (N protein), in particular characterized in that the antibody used for the capture of the native viral nucleoprotein is a monoclonal antibody specific for the central region and/or for a conformational epitope.

According to one embodiment of said test, the antibody used for the capture of the N protein is the monoclonal antibody mAb87, produced by the hybridoma deposited at the CNCM on Dec. 1, 2004 under the number I-3328.

According to another embodiment of said immunocapture test, the antibody used for the capture of the N protein is the monoclonal antibody mAb86, produced by the hybridoma deposited at the CNCM on Dec. 1, 2004 under the number I-3329.

According to another embodiment of said immunocapture test, the monoclonal antibodies mAb86 and mAb87 are used for the capture of the N protein.

In the immunocapture tests according to the invention, it is possible to use, for visualizing the N protein, the monoclonal antibody mAb57, produced by the hybridoma deposited at the CNCM on Dec. 1, 2004 under the number I-3330, said antibody being conjugated with a visualizing molecule or particle.

In accordance with said immunocapture test, a combination of the antibodies mAb57 and mAb87, conjugated with a visualizing molecule or particle, is used for the visualization of the N protein.

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A visualizing molecule may be a radioactive atom, a dye, a fluorescent molecule, a fluorophore, an enzyme; a visualizing particle may be for example: colloidal gold, a magnetic particle or a latex bead.

The subject of the present invention is also a reagent for detecting a SARS-associated coronavirus, characterized in that it is selected from the group consisting of:

- (a) a pair of primers or a probe as defined above,
- (b) a recombinant vector as defined above or a modified cell as defined above,
- (c) an isolated coronavirus strain as defined above or a polynucleotide as defined above,
- (d) an antibody or an antibody fragment as defined above,
- (e) a combination of antibodies comprising the monoclonal antibodies mAb86 and/or mAb87, and the monoclonal antibody mAb57, as defined above,
- (f) a chip or a filter as defined above.

The subject of the present invention is also a method for the detection of a SARS-associated coronavirus infection, from a biological sample, by indirect IgG ELISA using the N protein, which method is characterized in that the plates are sensitized with an N protein solution at a concentration of between 0.5 and 4 µg/ml, preferably to 2 µg/ml, in a 10 mM PBS buffer pH 7.2, phenol red at 0.25 ml/l.

The subject of the present invention is additionally a method for the detection of a SARS-associated coronavirus infection, from a biological sample, by double epitope ELSA, characterized in that the serum to be tested is mixed with the visualizing antigen, said mixture then being brought into contact with the antigen attached to a solid support.

According to one variant of the tests for detecting SARS-associated coronaviruses, these tests combine an ELSA using the N protein, and another ELSA using the S protein, as described below.

The subject of the present invention is also an immune complex formed of a polyclonal or monoclonal antibody or antibody fragment as defined above, and of a SARS-associated coronavirus protein or peptide.

The subject of the present invention is additionally a SARS-associated coronavirus detection kit, characterized in that it comprises at least one reagent selected from the group consisting of: a pair of primers, a probe, a DNA or RNA chip, a recombinant vector, a modified cell, an isolated coronavirus strain, a polynucleotide, a protein or a peptide, an antibody, and a protein chip as defined above.

The subject of the present invention is additionally an immunogenic composition, characterized in that it comprises at least one product selected from the group consisting of:

- a) a protein or a peptide as defined above,
- b) a polynucleotide of the DNA or RNA type or one of its representative fragments as defined above, having a sequence chosen from:
 - (i) the sequence SEQ ID NO: 1 or its RNA equivalent
 - (ii) the sequence hybridizing under high stringency conditions with the sequence SEQ ID NO: 1,
 - (iii) the sequence complementary to the sequence SEQ ID NO: 1 or to the sequence hybridizing under high stringency conditions with the sequence SEQ ID NO: 1,
 - (iv) the nucleotide sequence of a representative fragment of the polynucleotide as defined in (i), (ii) or (iii),
 - (v) the sequence as defined in (i), (ii), (iii) or (iv), modified, and
- c) a recombinant expression vector comprising a polynucleotide as defined in b), and
- d) a cDNA library as defined above, said immunogenic composition being capable of inducing protective humoral or cellular immunity specific for the

SARS-associated coronavirus, in particular the production of an antibody directed against a specific epitope of the SARS-associated coronavirus.

The proteins and peptides as defined above, in particular the S, M, E and/or N proteins and the derived peptides, and the nucleic acid (DNA or RNA) molecules encoding said proteins or said peptides are good candidate vaccines and may be used in immunogenic compositions for the production of a vaccine against the SARS-associated coronavirus.

According to an advantageous embodiment of the compositions according to the invention, they additionally contain at least one pharmaceutically acceptable vehicle and optionally carrier substances and/or adjuvants.

The pharmaceutically acceptable vehicles, the carrier substances and the adjuvants are those conventionally used.

The adjuvants are advantageously chosen from the group consisting of oily emulsions, saponin, mineral substances, bacterial extracts, aluminum hydroxide and squalene.

The carrier substances are advantageously selected from the group consisting of unilamellar liposomes, multilamellar liposomes, micelles of saponin or solid microspheres of a saccharide or auriferous nature.

The compositions according to the invention are administered by the general route, in particular by the intramuscular or subcutaneous route or alternatively by the local, in particular nasal (aerosol) route.

The subject of the present invention is also the use of an isolated or purified protein or peptide having a sequence selected from the group consisting of the sequences SEQ ID NO: 3, 10, 12, 14, 17, 22, 24, 26, 28, 30, 33, 35, 37, 69, 70, 71, 74 and 75 to form an immune complex with an antibody specifically directed against an epitope of the SARS-associated coronavirus.

The subject of the present invention is also an immune complex consisting of an isolated or purified protein or peptide having a sequence selected from the group consisting of the sequences SEQ ID NO: 3, 10, 12, 14, 17, 22, 24, 26, 28, 30, 33, 35, 37, 69, 70, 71, 74 and 75, and of an antibody specifically directed against an epitope of the SARS-associated coronavirus.

The subject of the present invention is also the use of an isolated or purified protein or peptide having a sequence selected from the group-consisting of the sequences SEQ ID NO: 3, 10, 12, 14, 17, 22, 24, 26, 28, 30, 33, 35, 37, 69, 70, 71, 74 and 75 to induce the production of an antibody capable of specifically recognizing an epitope of the SARS-associated coronavirus.

The subject of the present invention is also the use of an isolated or purified polynucleotide having a sequence selected from the group consisting of the sequences SEQ ID NO: 1, 2, 4, 7, 8, 13, 15, 16, 18, 19, 20, 31, 36 and 38 to induce the production of an antibody directed against the protein encoded by said polynucleotide and capable of specifically recognizing an epitope of the SARS-associated coronavirus.

The subject of the present invention is also monoclonal antibodies recognizing the native S protein of a SARS-associated coronavirus.

The subject of the present invention is also the use of a protein or a polypeptide of the S protein family, as defined above, or of an antibody recognizing the native S protein, as defined above, to detect an infection by a SARS-associated coronavirus, in a biological sample.

The subject of the present invention is also a method for detecting an infection by a SARS-associated coronavirus, in a biological sample, characterized in that the detection is carried out by ELISA using the recombinant S protein, expressed in a eukaryotic system.

According to an advantageous embodiment of said method, it is a double epitope ELISA method, and the serum to be tested is mixed with the visualizing antigen, said mixture then being brought into contact with the antigen attached to a solid support.

The subject of the present invention is also an immune complex consisting of a monoclonal antibody or antibody fragment recognizing the native S protein, and of a protein or a peptide of the SARS-associated coronavirus.

The subject of the present invention is also an immune complex consisting of a protein or a polypeptide of the S protein family, as defined above, and of an antibody specifically directed against an epitope of the SARS-associated coronavirus.

The subject of the present invention is additionally a SARS-associated coronavirus detection kit or box, characterized in that it comprises at least one reagent selected from the group consisting of: a protein or polypeptide of the S protein family, as defined above, a nucleic acid encoding a protein or peptide of the S protein family, as defined above, a cell expressing a protein or polypeptide of the S protein family, as defined above, or an antibody recognizing the native S protein of a SARS-associated coronavirus.

The subject of the present invention is an immunogenic and/or vaccine composition, characterized in that it comprises a polypeptide or a recombinant protein of the S protein family, as defined above, obtained in a eukaryotic expression system.

The subject of the present invention is also an immunogenic and/or vaccine composition, characterized in that it comprises a vector or recombinant virus, expressing a protein or a polypeptide of the S protein family, as defined above.

In addition to the preceding features, the invention further comprises other features, which will emerge from the description which follows, which refers to examples of use of the polynucleotide representing the genome of the SARS-CoV strain derived from the sample recorded under the number 031589, and derived cDNA fragments which are the subject of the present invention, and to Table I presenting the sequence listing:

TABLE I

Sequence listing			
Identification number	Sequence	Position of the cDNA with reference to Genbank AY274119.3	Deposit number at the CNCM of the corresponding plasmid
SEQ ID NO: 1	genome of the strain derived from the sample 031589	—	—
SEQ ID NO: 2	ORF-S*	21406-25348	—
SEQ ID NO: 3	S protein	—	—
SEQ ID NO: 4	ORF-S**	21406-25348	I-3059
SEQ ID NO: 5	Sa fragment	21406-23454	I-3020
SEQ ID NO: 6	Sb fragment	23322-25348	I-3019
SEQ ID NO: 7	ORF-3 + ORF-4*	25110-26244	—
SEQ ID NO: 8	ORF-3 + ORF-4**	25110-26244	I-3126
SEQ ID NO: 9	ORF3	—	—
SEQ ID NO: 10	ORF-3 protein	—	—
SEQ ID NO: 11	ORF4	—	—
SEQ ID NO: 12	ORF-4 protein	—	—
SEQ ID NO: 13	ORF-E*	26082-26413	—
SEQ ID NO: 14	E protein	—	—
SEQ ID NO: 15	ORF-E**	26082-26413	I-3046
SEQ ID NO: 16	ORF-M*	26330-27098	—
SEQ ID NO: 17	M protein	—	—

TABLE I-continued

Sequence listing			
Identification number	Sequence	Position of the cDNA with reference to Genbank AY274119.3	Deposit number at the CNCM of the corresponding plasmid
SEQ ID NO: 18	ORF-M**	26330-27098	I-3047
SEQ ID NO: 19	ORF7 to 11*	26977-28218	—
SEQ ID NO: 20	ORF7 to 11**	26977-28218	I-3125
SEQ ID NO: 21	ORF7	—	—
SEQ ID NO: 22	ORF7 protein	—	—
SEQ ID NO: 23	ORF8	—	—
SEQ ID NO: 24	ORF8 protein	—	—
SEQ ID NO: 25	ORF9	—	—
SEQ ID NO: 26	ORF9 protein	—	—
SEQ ID NO: 27	ORF10	—	—
SEQ ID NO: 28	ORF10 protein	—	—
SEQ ID NO: 29	ORF11	—	—
SEQ ID NO: 30	ORF11 protein	—	—
SEQ ID NO: 31	Orf1ab	265-21485	—
SEQ ID NO: 32	ORF13	28130-28426	—
SEQ ID NO: 33	ORF13 protein	—	—
SEQ ID NO: 34	ORF14	—	—
SEQ ID NO: 35	ORF14 protein	28583-28795	—
SEQ ID NO: 36	ORF-N*	28054-29430	—
SEQ ID NO: 37	N protein	—	—
SEQ ID NO: 38	ORF-N**	28054-29430	I-3048
SEQ ID NO: 39	noncoding 5***	1-204	I-3124
SEQ ID NO: 40	noncoding 3***	28933-29727	I-3123
SEQ ID NO: 41	ORF1ab	30-500	—
SEQ ID NO: 42	Fragment L0	—	—
SEQ ID NO: 43	Fragment L1	211-2260	—
SEQ ID NO: 44	Fragment L2	2136-4187	—
SEQ ID NO: 45	Fragment L3	3892-5344	—
SEQ ID NO: 46	Fragment L4b	4932-6043	—
SEQ ID NO: 47	Fragment L4	5305-7318	—
SEQ ID NO: 48	Fragment L5	7275-9176	—
SEQ ID NO: 49	Fragment L6	9032-11086	—
SEQ ID NO: 50	Fragment L7	10298-12982	—
SEQ ID NO: 51	Fragment L8	12815-14854	—
SEQ ID NO: 52	Fragment L9	14745-16646	—
SEQ ID NO: 53	Fragment L10	16514-18590	—
SEQ ID NO: 54	Fragment L11	18500-20602	—
SEQ ID NO: 55	Fragment L12	20319-22224	—
SEQ ID NO: 56	Sense N primer	—	—
SEQ ID NO: 57	Antisense	—	—
SEQ ID NO: 58	N primer	—	—
SEQ ID NO: 59	Sense S _C primer	—	—
SEQ ID NO: 60	Sense S _L primer	—	—
SEQ ID NO: 61	Antisense S _C and S _L primer	—	—
SEQ ID NO: 62	Sense primer series 1	28507-28522	—
SEQ ID NO: 63	Antisense primer series 1	28774-28759	—
SEQ ID NO: 64	Sense primer series 2	28375-28390	—
SEQ ID NO: 65	Antisense primer series 2	28702-28687	—
SEQ ID NO: 66	Probe 1/series 1	28561-28586	—
SEQ ID NO: 67	Probe 2/series 1	28588-28608	—
SEQ ID NO: 68	Probe 1/series 2	28541-28563	—
SEQ ID NO: 69	Probe 2/series 2	28565-28589	—
SEQ ID NO: 70	Anchor primer 14T	—	—
SEQ ID NO: 71	Peptide M2-14	—	—
SEQ ID NO: 72	Peptide E1-12	—	—
SEQ ID NO: 73	Peptide E53-76	—	—
SEQ ID NO: 74	Noncoding 5**	1-204	—
SEQ ID NO: 75	Noncoding 3**	28933-29727	—
SEQ ID NO: 76	ORF1a protein	—	—
SEQ ID NO: 77	ORF1b protein	—	—
SEQ ID NO: 78	Primers	—	—
SEQ ID NO: 79	Pseudogene of S	—	—
SEQ ID NO: 80	Primers	—	—
SEQ ID NO: 81	Aa1-13 of S	—	—

TABLE I-continued

Sequence listing			
Identification number	Sequence	Position of the cDNA with reference to Genbank AY274119.3	Deposit number at the CNCM of the corresponding plasmid
SEQ ID NO: 150	Polypeptide	—	—
SEQ ID NO: 151-158	Primers	—	—
*PCR amplification product (amplicon)			
**Insert cloned into the plasmid deposited at the CNCM and to the appended drawings in which:			
FIG. 1 illustrates Western-blot analysis of the expression in vitro of the recombinant proteins N, S _C and S _L from the expression vectors pIVEX. Lane 1: pIV2.3N. Lane 2: pIV2.3S _C . Lane 3: pIV2.3S _L . Lane 4: pIV2.4N. Lane 5: pIV2.4S ₁ or pIV2.4S _C . Lane 6: pIV2.4S _L . The expression of the GFP protein expressed from the same vector is used as a control.			
FIG. 2 illustrates the analysis, by polyacrylamide gel electrophoresis under denaturing conditions (SDS-PAGE) and staining with Coomassie blue, of the expression in vivo of the N protein from the expression vectors pIVEX. The <i>E. coli</i> BL21(DE3)pDIA17 strain transformed with the recombinant vectors pIVEX is cultured at 30° C. in LB medium, in the presence or in the absence of inducer (IPTG 1 mM). Lane 1: pIV2.3N. Lane 2: pIV2.4N.			
FIG. 3 illustrates the analysis, by polyacrylamide gel electrophoresis under denaturing conditions (SDS-PAGE) and staining with Coomassie blue, of the expression in vivo of the S _L and S _C polypeptides from the expression vectors pIVEX. The <i>E. coli</i> BL21(DE3)pDIA17 strain transformed with the recombinant vectors pIVEX is cultured at 30° C. in LB medium, in the presence or in the absence of inducer (IPTG 1 mM). Lane 1: pIV2.3S _C . Lane 2: pIV2.3S _L . Lane 3: pIV2.4S ₁ . Lane 4: pIV2.4S _L .			
FIG. 4 illustrates the antigenic activity of the recombinant N, S _L and S _C proteins produced in the <i>E. coli</i> BL21(DE3)pDIA17 strain transformed with the recombinant vectors pIVEX. A: electrophoresis (SDS-PAGE) of the bacterial lysates. B and C: Western-blot with the sera, obtained from the same patient infected with SARS-CoV, collected 8 days (B: serum M12) and 29 days (C: serum M13) respectively after the onset of the SARS symptoms. Lane 1: pIV2.3N. Lane 2: pIV2.4N. Lane 3: pIV2.3S _C . Lane 4: pIV2.4S ₁ . Lane 5: pIV2.3S _L . Lane 6: pIV2.4S _L .			
FIG. 5 illustrates the purification on an Ni-NTA agarose column of the recombinant N protein produced in the <i>E. coli</i> BL21(DE3)pDIA17 strain from the vector pIV2.3N. Lane 1: total bacterial extract. Lane 2: soluble extract. Lane 3: insoluble extract. Lane 4: extract deposited on the Ni-NTA column. Lane 5: unbound proteins. Lane 6: fractions of peak 1. Lane 7: fractions of peak 2.			
FIG. 6 illustrates the purification of the recombinant S _C protein from the inclusion bodies produced in the <i>E. coli</i> BL21(DE3)pDIA17 strain transformed with pIV2.4S ₁ . A: Treatment with Triton X-100 (2%): Lane 1: total bacterial extract. Lane 2: soluble extract. Lane 3: insoluble extract. Lane 4: supernatant after treatment with Triton X-100 (2%). Lanes 5 and 6: pellet after treatment with Triton X-100 (2%). B: Treatment with 4 M, 5 M, 6 M and 7 M urea of the soluble and insoluble extracts.			
FIG. 7 represents the immunoblot produced with the aid of a lysate of cells infected with SARS-CoV and a serum from a patient suffering from a typical pneumonia.			
FIG. 8 represents immunoblots produced with the aid of a lysate of cells infected with SARS-CoV and rabbit immunosera specific for the nucleoprotein N (A) and for the spicule protein S (B). I.S.: immune serum. p.i.: preimmune serum. The anti-N immune serum was used at 1/50 000 and the anti-S immune serum at 1/10 000.			
FIG. 9 illustrates the ELISA reactivity of the rabbit monospecific polyclonal sera directed against the N protein or the short fragment of the S protein (S _C), toward the corresponding recombinant proteins used for immunization. A: rabbits P13097, P13081 and P13031 immunized with the purified recombinant N protein. B: rabbits P11135, P13042 and P14001 immunized with a preparation of inclusion bodies corresponding to the short fragment of the S protein (S _C). I.S.: immune serum. p.i.: preimmune serum.			
FIG. 10 illustrates the ELISA reactivity of the purified recombinant N protein, toward sera from patients suffering from a typical pneumonia caused by SARS-CoV. FIG. 10A: ELISA plates prepared with the N protein at the concentration of 4 µg/ml and 2 µg/ml. FIG. 10B: ELISA plate prepared with the N protein at the concentration of 1 µg/ml. The sera designated A, B, D, E, F, G, H correspond to those of Table IV.			
FIG. 11 illustrates the amplification by RT-PCR of decreasing quantities of synthetic RNA of the SARS-CoV N gene (10 ¹ to 1 copy), with the aid of pairs of primers No. 1 (N+/28507, N-/28774) (A) and No. 2 (N+/28375, N-/28702) (B). T: amplification performed in the absence of RNA. MW: DNA marker.			
FIG. 12 illustrates the amplification by RT-PCR in real time of synthetic RNA for the SARS-CoV N gene: decreasing quantities of synthetic RNA as replica (repli.; lanes 16 to 29) and of viral RNA diluted 1/20 × 10 ⁻⁴ (lane 32) were amplified by RT-PCR in real time with the aid of the kit "Light Cycler RNA Amplification Kit Hybridization Probes" and pairs of primers and probes of the No. 2 series, under the conditions described in Example 8.			
FIG. 13 (FIG. 13.1 to 13.7) represents the restriction map of the sequence SEQ ID NO: 1 corresponding to the DNA equivalent of the genome of the SARS-CoV strain derived from the sample recorded under the number 031589.			
FIG. 14 shows the result of the SARS serology test by indirect N ELISA (1 st series of sera tested).			
FIG. 15 shows the result of the SARS serology test by indirect N ELISA (2 nd series of sera tested).			
FIG. 16 presents the result of the SARS serology test by double epitope N ELISA (1 st series of sera tested).			
FIG. 17 shows the result of the SARS serology test by double epitope N ELISA (2 nd series of sera tested).			
FIG. 18 illustrates the test of reactivity of the anti-N monoclonal antibodies by ELISA on the native nucleoprotein N of SARS-CoV. The antibodies were tested in the form of hybridoma culture supernatants by indirect ELISA using an irradiated lysate of VeroE6 cells infected with SARS-CoV as antigen (SARS lysate curves). A negative control for reactivity is performed for each antibody on a lysate of uninfected VeroE6 cells (negative lysate curves).			
Several monoclonal antibodies of known specificity were used as negative control antibodies: para 1-3 directed against the antigens of the parainfluenza virus type 1-3 (Bio-Rad) and influenza B directed against the antigens of the influenza virus type B (Bio-Rad).			

TABLE I-continued

Sequence listing		
Identification number	Sequence	Position of the cDNA with reference to Genbank AY274119.3

FIG. 19 illustrates the test of reactivity of the anti-N of SARS-CoV monoclonal antibodies by ELISA on the native antigens of the human coronavirus 229E (HCoV-229E). The antibodies were tested in the form of hybridoma culture supernatants by an indirect ELISA test using a lysate of MRC-5 cells infected with the human coronavirus 229E as antigen (229E lysate curves). A negative control for immunoreactivity was performed for each antibody on a lysate of noninfected MRC-5 cells (negative lysate curves). The monoclonal antibody 5-11H.6 directed against the S protein of the human coronavirus 229E (Sizun et al. 1998, J. Virol. Met. 72: 145-152) is used as positive control antibody. The antibodies para-1-3 directed against the antigens of the parainfluenza virus type 1-3 (Bio-Rad) and influenza B directed against the antigens of the influenza virus type B (Bio-Rad) were added to the panel of monoclonal antibodies tested.

FIG. 20 shows a test of reactivity of the anti-N of SARS-CoV monoclonal antibodies by Western blotting on the denatured native nucleoprotein N of SARS-CoV. A lysate of VeroE6 cells infected with SARS-CoV was prepared in the loading buffer according to Laemmli and caused to migrate in a 12% SDS polyacrylamide gel and then the proteins were transferred onto PVDF membrane. The anti-N monoclonal antibodies tested were used for the immunoblotting at the concentration of 0.05 µg/ml. The visualization is carried out with anti-mouse IgG(H + L) antibodies coupled to peroxidase (NA93IV, Amersham) and the ECL+ system. Two monoclonal antibodies were used as negative controls for reactivity: influenza B directed against the antigens of the influenza virus type B (Bio-Rad) and para-1-3 directed against the antigens of the parainfluenza virus type 1-3 (Bio-Rad).

FIG. 21 presents the plasmids for expression in mammalian cells of the SARS-CoV S protein. The cDNA for the SARS-CoV S was inserted between the BamHI and XhoI sites of the expression plasmid pcDNA3.1(+)(Clontech) in order to obtain the plasmid pcDNA-S and between the NheI and XhoI sites of the expression plasmid pCI (Promega) in order to obtain the plasmid pCI-S. The WPRE and CTE sequences were inserted between each of the two plasmids pcDNA-S and pCI-S between the XhoI and XbaI sites in order to obtain the plasmids pcDNA-S-CTE, pcDNA-S-WPRE, pCI-S-CTE and pCI-S-WPRE, respectively. SP: signal peptide predicted (aa 1-13) with the software signalP v2.0 (Nielsen et al., 1997, Protein Engineering, 10: 1-6)

TM: transmembrane region predicted (aa 1196-1218) with the software TMHMM v2.0 (Sonnhammer et al., 1998, Proc. of Sixth Int. Conf. on Intelligent Systems for Molecular Biology, pp. 175-182, AAAI Press). It should be noted that the amino acids W1194 and P1195 are possibly part of the transmembrane region with the respective probabilities of 0.13 and 0.42

P-CMV: cytomegalovirus immediate/early promoter.

BGH pA: polyadenylation signal of the bovine growth hormone gene

SV40 late pA: SV40 virus late polyadenylation signal

SD/SA: splice donor and acceptor sites

WPRE: sequences of the "Woodchuck Hepatitis Virus posttranscriptional regulatory element" of the woodchuck hepatitis virus

CTE: sequences of the "constitutive transport element" of the Mason-Pfizer simian retrovirus

FIG. 22 illustrates the expression of the S protein after transfection of VeroE6 cells. Cellular extracts were prepared 48 hours after transfection of VeroE6 cells with the plasmids pcDNA, pcDNA-S, pCI and pCI-S. Cellular extracts were also prepared 18 hours after infection with the recombinant vaccinia virus VV-TF7.3 and transfection with the plasmids pcDNA or pcDNA-S. As a control, extracts of VeroE6 cells were prepared 8 hours after infection with SARS-CoV at a multiplicity of infection of 3. They were separated on an 8% SDS acrylamide gel and analyzed by Western blotting with the aid of an anti-S rabbit polyclonal antibody and an anti-rabbit IgG(H + L) polyclonal antibody coupled to peroxidase (NA934V, Amersham). A molecular mass ladder (kDa) is presented in the figure. SARS-CoV: extract of VeroE6 cells infected with SARS-CoV

Mock: control extract of noninfected cells

FIG. 23 illustrates the effect of the CTE and WPRE sequences on the expression of the S protein after transfection of VeroE6 and 293T cells. Cellular extracts were prepared 48 hours after transfection of VeroE6 cells (A) or 293T cells (B) with the plasmids pcDNA, pcDNA-S, pcDNA-S-CTE, pcDNA-S-WPRE, pCI-S, pCI-S-CTE and pCI-S-WPRE separated on 8% SDS polyacrylamide gel and analyzed by Western blotting with the aid of an anti-S rabbit polyclonal antibody and an anti-rabbit IgG(H + L) polyclonal antibody coupled to peroxidase (NA934V, Amersham). A molecular mass ladder (kDa) is presented in the figure. SARS-CoV: extract of VeroE6 cells prepared 8 hours after infection with SARS-CoV at a multiplicity of infection of 3.

Mock: control extract of noninfected VeroE6 cells

FIG. 24 presents defective lentiviral vectors with central DNA flap for the expression of SARS-CoV S. The cDNA for the SARS-CoV S protein was cloned in the form of a BamHI-XhoI fragment into the plasmid pTRIPAU3-CMV containing a defective lentiviral vector TRIP with central DNA flap (Sirven et al., 2001, Mol. Ther., 3: 438-448) in order to obtain the plasmid pTRIP-S. The optimum expression cassettes consisting of the CMV virus immediate/early promoter, a splice signal, cDNA for S and either of the posttranscriptional signals CTE or WPRE were substituted for the cassette EF1α-EGFP of the defective lentiviral expression vector with central DNA flap TRIPAU3-EF1α (Sirven et al., 2001, Mol. Ther., 3: 438-448) in order to obtain the plasmids pTRIP-SD/SA-S-CTE and pTRIP-SD/SA-S-WPRE.

SP: signal peptide

TM: transmembrane region

P-CMV: cytomegalovirus immediate/early promoter

P-EF1α: EF1α gene promoter

SD/SA: splice donor and acceptor sites

WPRE: sequences of the "Woodchuck Hepatitis Virus posttranscriptional regulatory element" of the woodchuck hepatitis virus

CTE: sequences of the "constitutive transport element" of the Mason-Pfizer simian retrovirus

LTR: long terminal repeat AU3; LTR deleted for the "promoter/enhancer" sequences

cPPT: "polypurine tract cis-active sequence"

CTS: "central termination sequence"

TABLE I-continued

Sequence listing			
Identification number	Sequence	Position of the cDNA with reference to Genbank AY274119.3	Deposit number at the CNCM of the corresponding plasmid

FIG. 25 shows the Western-blot analysis of the expression of the SARS-CoV S by cell lines transduced with the lentiviral vectors TRIP-SD/SA-S-WPRE and TRIP-SD/SA-S-CTE. Cellular extracts were prepared from established lines FrhK4-S-CTE and FrhK4-S-WPRE after transduction with the lentiviral vectors TRIP-SD/SA-S-CTE and TRIP-SD/SA-S-WPRE respectively. They were separated on an 8% SDS acrylamide gel and analyzed by Western blotting with the aid of an anti-S rabbit polyclonal antibody and an anti-rabbit IgG(H + L) conjugate coupled to peroxidase. A molecular mass ladder (kDa) is presented in the figure.

T-: control extract of FrhK-4 cells

T+: extract of FrhK-4 cells prepared 24 hours after infection with SARS-CoV at a multiplicity of infection of 3.

FIG. 26 relates to the analysis of the expression of Ssol polypeptide by cell lines transduced with the lentiviral vectors TRIP-SD/SA-Ssol-WPRE and TRIP-SD/SA-Ssol-CTE. The secretion of the Ssol polypeptide was determined in the supernatant of a series of cell clones isolated after transduction of FrhK-4 cells with the lentiviral vectors TRIP-SD/SA-Ssol-WPRE and TRIP-SD/SA-Ssol-CTE. 5 µl of supernatant, diluted 1/2 in loading buffer according to Laemmli, were analyzed by Western blotting, visualized with an anti-FLAG monoclonal antibody (M2, Sigma) and an anti-mouse IgG(H + L) conjugate coupled to peroxidase.

T-: supernatant of the parental FrhK-4 line.

T+: supernatant of FBHK cells infected with a recombinant vaccinia virus expressing the Ssol polypeptide. The solid arrow indicates the Ssol polypeptide, while the empty arrow indicates a cross reaction with a protein of cellular origin.

FIG. 27 shows the results relating to the analysis of the purified Ssol polypeptide

A, 8, 2, 0.5 and 0.125 µg of recombinant Ssol polypeptide purified by anti-FLAG affinity chromatography and gel filtration (G75) were separated on 8% SDS polyacrylamide gel. The Ssol polypeptide and variable quantities of molecular mass markers (MM) were visualized by staining with silver nitrate (Gelcode SilverSNAP stain kit II, Pierce).

B. Standard markers for analysis by SELDI-TOF mass spectrometry

IgG: bovine IgG of MM 147300

ConA: conalbumin of MM 77490

HRP: horseradish peroxidase analyzed as a control and of MM 43240

C. Analysis by mass spectrometry (SELDI-TOF) of the recombinant Ssol polypeptide.

The peaks A and B correspond to the single and double charged Ssol polypeptide.

D. Sequencing of the N-terminal end of the recombinant Ssol polypeptide. 5 Edman degradation cycles in liquid phase were carried out on an ABI494 sequencer (Applied Biosystems).

FIG. 28 illustrates the influence of a splicing signal and of the CTE and WPRE sequences on the efficacy of the gene immunization with the aid of plasmid DNA encoding the SARS-CoV S

A. Groups of 7 BALB/c mice were immunized twice at 4 weeks' interval with the aid of 50 µg of plasmid DNA of pCI, pcDNA-S, pCI-S, pcDNA-N and pCI-HA.

B. Groups of 6 BALB/c mice were immunized twice at 4 weeks' interval with the aid of 2 µg, 10 µg or 50 µg of plasmid DNA of pCI, pCI-S, pCI-S-CTE and pCI-S-WPRE.

The immune sera collected 3 weeks after the second immunization were analyzed by indirect ELISA using a lysate of VeroE6 cells infected with SARS-CoV as antigen. The anti-SARS-CoV antibody titers are calculated as the reciprocal of the dilution producing a specific OD of 0.5 after visualization with an anti-mouse IgG polyclonal antibody coupled to peroxidase (NA931V, Amersham) and TMB (KPL).

FIG. 29 shows the seroneutralization of the

ineffectivity of SARS-CoV with the antibodies induced in

mice after gene immunization with the aid of plasmid

DNA encoding SARS-CoV S. Pools of immune sera collected

3 weeks after the second immunization were prepared for

each of the groups of experiments described in

FIG. 28 and evaluated for their capacity to seroneutralize the infectivity of 100 TCID50 of SARS-CoV on FRhK-4 cells. 4 points are produced for each of the 2-fold dilutions tested from 1/20. The seroneutralizing titer is calculated according to the Reed and Munch method as the reciprocal of the dilution neutralizing the infectivity of 2 wells out of 4.

A. Groups by BALB/c mice immunized twice at 4 weeks' interval with the aid of 50 µg of plasmid DNA of pCI, pcDNA-S, pCI-S, pcDNA-N and pCI-HA. □: preimmune serum. ■: immune serum.

B. Groups of BALB/c mice immunized twice at 4 weeks' interval with the aid of 2 µg, 10 µg or 50 µg of plasmid DNA of pCI, pCI-S, pCI-S-CTE and pCI-S-WPRE.

FIG. 30 illustrates the immunoreactivity of the recombinant Ssol polypeptide toward sera from patients suffering from SARS. The reactivity of sera from patients was analyzed by indirect ELISA test against solid phases prepared with the aid of the purified recombinant Ssol polypeptide. The antibodies from patients reacting with the solid phase at a dilution of 1/400 are visualized with a human anti-IgG(H + L) polyclonal antibody coupled to peroxidase (Amersham NA933V) and TMB plus H202 (KPL). The sera of probable SARS cases are identified by a National Reference Center for Influenza Viruses serial number and by the initials of the patient and the number of days elapsed since the onset of symptoms, where appropriate. The TV sera are control sera from subjects which were collected in France before the SARS epidemic which occurred in 2003.

FIG. 31 shows the induction of antibodies directed against SARS-CoV after immunization with the recombinant Ssol polypeptide. Two groups of 6 mice were immunized at 3 weeks' interval with 10 µg of recombinant Ssol polypeptide (Ssol group) adjuvanted with aluminum hydroxide or, as a control, of adjuvant alone (mock group). Three successive immunizations were performed and the immune sera were collected 3 weeks after each of the three immunizations (IS1, IS2, IS3). The immune sera were analyzed per pool for each of the 2 groups by indirect ELISA using a lysate of VeroE6 cells infected with SARS-CoV as antigen. The anti-SARS-CoV antibody titers are calculated as the reciprocal of the dilution producing a specific OD of 0.5 after visualization with an anti-mouse IgG polyclonal antibody coupled to peroxidase (Amersham) and TMB (KPL).

FIG. 32 presents the nucleotide alignment of the sequences of the synthetic gene 040530 with the sequence of the wild-type gene of the SARS-CoV isolate 031589. 1-3059 corresponds to nucleotides 21406-25348 of the SARS-CoV isolate 031589 deposited at the C.N.C.M. under the number 1-3059 (SEQ ID NO: 4, plasmid pSARS-S) S-040530 is the sequence of the synthetic gene 040530.

TABLE I-continued

Sequence listing		Position	Deposit
Identification number	Sequence	of the cDNA with reference to Genbank	number at the CNCM of the corresponding plasmid
		AY274119.3	

FIG. 33 illustrates the use of a synthetic gene for the expression of the SARS-CoV S. Cellular extracts prepared 48 hours after transfection of VeroE6 cells (A) or 293T cells (B) with the plasmids pCI-S, pCI-S-CTE, pCI-S-WPRE and pCI-Ssynth were separated on 8% SDS acrylamide gel and analyzed by Western blotting with the aid of an anti-S rabbit polyclonal antibody and an anti-rabbit IgG(H + L) polyclonal antibody coupled to peroxidase (NA934V, Amersham). The Western blot is visualized by luminescence (ECL+, Amersham) and acquisition on a digital imaging device (FluorS, BioRad). The levels of expression of the S protein were measured by quantifying the 2 predominant bands identified on the image.

FIG. 34 presents a diagram for the construction of recombinant vaccinia viruses VV-TG-S, VV-TG-Ssol, VV-TN-S and VV-TN-Ssol

A. The cDNAs for the S protein and the Ssol polypeptide of SARS-CoV were inserted between the BamHI and SmaI sites of the transfer plasmid pTG186 in order to obtain the plasmids pTG-S and pTG-Ssol.

B. The sequences of the synthetic promoter 480 were then substituted for those of the 7.5 promoter by exchange of the NdeI-PstI fragments of the plasmids pTG186poly, pTG-S and pTG-Ssol in order to obtain the transfer plasmids pTN480, pTN-S and pTN-Ssol.

C. Sequence of the synthetic promoter 480 as contained between the NdeI and PstI sites of the transfer plasmids of the pTN series. An AscI site was inserted in order to facilitate subsequent handling. The restriction sites and the promoter sequence are underlined.

D. The recombinant vaccinia viruses are obtained by double homologous recombination in vivo between the TK cassette of the transfer plasmids of the pTG and pTN series and the TK gene of the Copenhagen strain of the vaccinia virus.

SP: signal peptide predicted (aa 1-13) with the software signalP v2.0 (Nielsen et al., 1997, Protein Engineering, 10: 1-6)

TM: transmembrane region predicted (aa 1196-1218) with the software TMHMM v2.0 (Sonnhammer et al., 1998, Proc. of Sixth Int. Conf. on Intelligent Systems for Molecular Biology, pp. 175-182, AAAI Press). It should be noted that the amino acids W1194 and P1195 possibly form part of the transmembrane region with respective probabilities of 0.13 and 0.42.

TK-L, TK-R: left- and right-hand parts of the vaccinia virus thymidine kinase gene

MCS: multiple cloning site

PE: early promoter

PL: late promoter

PL synth: synthetic late promoter 480

FIG. 35 illustrates the expression of the S protein by recombinant vaccinia viruses, analyzed by Western blotting. Cellular extracts were prepared 18 hours after infection of CV1 cells with the recombinant vaccinia viruses VV-TG, VV-TG-S and VV-TN-S at an M.O.I. of 2 (A). As a control, extracts of VeroE6 cells were prepared 8 hours after infection with SARS-CoV at a multiplicity of infection of 2. Cellular extracts were also prepared 18 hours after infection of CV1 cells with the recombinant vaccinia viruses VV-TG-S, VV-TG-Ssol, VV-TN, VV-TN-S and VV-TN-Ssol (B). They were separated on 8% SDS acrylamide gels and analyzed by Western blotting with the aid of an anti-S rabbit polyclonal antibody and an anti-rabbit IgG(H + L) polyclonal antibody coupled to peroxidase (NA934V, Amersham). "1 μl" and "10 μl" indicates the quantities of cellular extracts deposited on the gel. A molecular mass ladder (kDa) is presented in the figure.

SARS-CoV: extract of VeroE6 cells infected with SARS-CoV

Mock: control extract of noninfected cells

FIG. 36 shows the result of a Western-blot analysis of the secretion of the Ssol polypeptide by the recombinant vaccinia viruses.

A. Supernatants of CV1 cells infected with the recombinant vaccinia virus VV-TN, various clones of the VV-TN-Ssol virus and with the viruses VV-TG-Ssol or VV-TN-Sflag were harvested 18 hours after infection of CV1 cells at an M.O.I. of 2.

B. Supernatants of 293T, FRhK-4, BHK-21 and CV1 cells infected in duplicate (1.2) with the recombinant vaccinia virus VV-TN-Ssol at an M.O.I. of 2 were harvested 18 hours after infection. The supernatant of CV1 cells infected with the virus VV-TN was also harvested as a control (M).

All the supernatants were separated on 8% SDS acrylamide gel according to Laemmli and analyzed by Western blotting with the aid of an anti-FLAG mouse monoclonal antibody and an anti-mouse IgG(H + L) polyclonal antibody coupled to peroxidase (NA931V, Amersham) (A) or with the aid of an anti-S rabbit polyclonal antibody and an anti-rabbit IgG(H + L) polyclonal antibody coupled to peroxidase (NA934V, Amersham) (B). A molecular mass ladder (kDa) is presented in the figure.

FIG. 37 shows the analysis of the Ssol polypeptide, purified on SDS polyacrylamide gel 10, 5 and 2 μl of recombinant Ssol polypeptide purified by anti-FLAG affinity chromatography were separated on 4 to 15% gradient SDS polyacrylamide gel. The Ssol polypeptide and variable quantities of molecular mass markers (MM) were visualized by staining with silver nitrate (Gelcode SilverSNAP stain kit II, Pierce).

FIG. 38 illustrates the immunoreactivity of the recombinant Ssol polypeptide produced by the recombinant vaccinia virus VV-TN-Ssol toward sera of patients suffering from SARS. The reactivity of sera from patients was analyzed by indirect ELISA test against solid phases prepared with the aid of the purified recombinant Ssol polypeptide. The antibodies from patients reacting with the solid phase at a dilution of 1/100 and 1/400 are visualized with a human anti-IgG(H + L) polyclonal antibody coupled to peroxidase (Amersham NA933V) and TMB plus H202 (KPL). The sera of probable SARS cases are identified by a National Reference Center for Influenza Virus serial number and by the initials of the patient and the number of days elapsed since the onset of symptoms, where appropriate. The TV sera are control sera from subjects which were collected in France before the SARS epidemic which occurred in 2003.

FIG. 39 shows the anti-SARS-CoV antibody response in mice after immunization with the recombinant vaccinia viruses. Groups of 7 BALB/c mice were immunized by the i.v. route twice at 4 weeks' interval with 106 pfu of recombinant vaccinia viruses VV-TG, VV-TG-HA, VV-TG-S, VV-TG-Ssol, VV-TN, VV-TN-S, VV-TN-Ssol.

A. Pools of immune sera collected 3 weeks after each of the two immunizations were prepared for each of the groups and were analyzed by indirect ELISA using a lysate of VeroE6 cells infected with SARS-CoV as antigen. The anti-SARS-CoV antibody titers are calculated as the reciprocal of the dilution producing a specific OD of 0.5 after visualization with an anti-mouse IgG polyclonal antibody coupled to peroxidase (NA931V, Amersham) and TMB (KPL).

TABLE I-continued

Sequence listing		Position	Deposit
Identification number	Sequence	of the cDNA with reference to Genbank	number at the CNCM of the corresponding plasmid
		AY274119.3	

10 B. The pools of immune sera were evaluated for their capacity to seroneutralize the infectivity of 100 TCID₅₀ of SARS-CoV on FRhK-4 cells. 4 points are produced for each of the 2-fold dilutions tested from 1/20. The seroneutralizing titer is calculated according to the Reed and Munch method as the reciprocal of the dilution neutralizing the infectivity of 2 wells out of 4.

FIG. 40 describes the construction of the recombinant viruses MVSchw2-SARS-S and MVSchw2-SARS-Ssol.

A. The measles vector is a complete genome of the Schwarz vaccine strain of the measles virus (MV) into which an additional transcription unit has been introduced (Combedet, 2003, Journal of Virology, 77: 11546-11554). The expression of the additional open reading frames (ORF) is controlled by cis-acting elements necessary for the transcription, for the formation of the cap and for the polyadenylation of the transgene which were copied from the elements present at the N/P junction. 2 different vectors allow the insertion between the P (phosphoprotein) and M (matrix) genes on the one hand and the H (hemagglutinin) and L (polymerase) genes on the other hand.

B. The recombinant genomes MVSchw2-SARS-S and MVSchw2-SARS-Ssol of the measles virus were constructed by inserting the ORFs of the S protein and of the Ssol polypeptide into an additional transcription unit located between the P and M genes of the vector.

The various genes of the measles virus (MV) are indicated: N (nucleoprotein), PVC (V/C phosphoprotein and protein), M (matrix), F (fusion), H (hemagglutinin), L (polymerase), T7 = T7 RNA polymerase promoter, hh = hammerhead ribozyme, T7t = T7 phage RNA polymerase terminator sequence, δ = ribozyme of the hepatitis δ virus, (2), (3) = additional transcription units (ATU).

Size of the MV genome: 15 894 nt.

25 SP: signal peptide

TM: transmembrane region

FLAG: FLAG tag

FIG. 41 illustrates the expression of the S protein by the recombinant measles viruses, analyzed by Western blotting.

Cytoplasmic extracts were prepared after infection of Vero cells by different passages of the viruses MVSchw2-SARS-S and MVSchw2-SARS-Ssol and the wild-type virus MWSchw as control. Cellular extracts in loading buffer according to Laemmli were also prepared 8 hours after infection of VeroE6 cells with SARS-CoV at a multiplicity of infection of 3. They were separated on 8% SDS acrylamide gel and analyzed by Western blotting with the aid of an anti-S rabbit polyclonal antibody and an anti-rabbit IgG(H + L) polyclonal antibody coupled to peroxidase (NA934V, Amersham).

A molecular mass ladder (kDa) is presented in the figure.

Pn: nth passage of the virus after coculture of 293-3-46 and Vero cells

SARS-CoV: extract of VeroE6 cells infected with SARS-CoV

35 Mock: control extract of noninfected VeroE6 cells

FIG. 42 shows the expression of the S protein by the recombinant measles viruses, analyzed by immunofluorescence

Vero cells in monolayers on glass slides were infected with the wild-type virus MWSchw (A) or the viruses MVSchw2-SARS-S (B) and MVSchw2-SARS-Ssol (C). When the syncytia have reached 30 to 40% confluence (A, B) or 90-100% (C), the cells were fixed, permeabilized and labeled with anti-SARS-CoV rabbit polyclonal antibodies and an anti-rabbit IgG(H + L) conjugate coupled to FITC (Jackson).

FIG. 43 illustrates the Western-blot analysis of the immunoreactivity of rabbit sera directed against the peptides E1-12, E53-76 and M2-14. The rabbit 20047 was immunized with the peptide E1-12 coupled to KLH. The rabbits 22234 and 22240 were immunized with the peptide E53-76 coupled to KLH. The rabbits 20013 and 20080 were immunized with the peptide M2-14 coupled to KLH. The immune sera were analyzed by Western blotting with the aid of extracts of cells infected with SARS-CoV (B) or with the aid of extracts of cells infected with a recombinant vaccinia virus expressing the protein E (A) or M (C) of the SARS-CoV 031589 isolate. The immunoblots were visualized with the aid of an anti-rabbit IgG(H + L) conjugate coupled to peroxidase (NA934V, Amersham).

The position of the E and M proteins is indicated by an arrow.

A molecular mass ladder (kDa) is presented in the figure.

It should be understood, however, that these examples are given solely by way of illustration of the subject of the invention, and do not constitute in any manner a limitation thereto.

EXAMPLE 1

Cloning and Sequencing of the Genome of the SARS-CoV Strain Derived from the Sample Recorded Under the Number 031589

55 The RNA of the SARS-CoV strain was extracted from the sample of bronchoalveolar washing recorded under the number 031589, performed on a patient at the Hanoi (Vietnam) French hospital suffering from SARS.

60 The isolated RNA was used as template to amplify the cDNAs corresponding to the various open reading frames of the genome (ORF1a, ORF1b, ORF-S, ORF-E, ORF-M, ORF-N (including ORF-13 and ORF-14), ORF3, ORF4, ORF7 to ORF11), and at the noncoding 5' and 3' ends. The sequences of the primers and of the probes used for the amplification/detection were defined based on the available SARS-CoV nucleotide sequence.

In the text which follows, the primers and the probes are identified by: the letter S, followed by a letter which indicates the corresponding region of the genome (L for the 5' end including ORF1a and ORF1b; S, N and N for ORF-S, ORF-M, ORF-N, SE and MN for the corresponding intergene regions), and then optionally by Fn, Rn, with n between 1 and 6 corresponding to the primers used for the nested PCR (F1+R1 pair for the first amplification, F2+R2 pair for the second amplification, and the like), and then by +/- or -/- corresponding to a sense or antisense primer and finally by the positions of the primers with reference to the Genbank sequence AY27411.3; for the sense and antisense S and N primers and the other sense primers only, when a single position is indicated, it corresponds to that of the 5' end of a probe or of a primer of about 20 bases; for the antisense primers other than the S and N primers, when a single position is indicated, it corresponds to that of the 3' end of a probe or of a primer of about 20 bases.

The amplification products thus generated were sequenced with the aid of specific primers in order to determine the complete sequence of the genome of the SARS-CoV strain derived from the sample recorded under the number 031589. These amplification products, with the exception of those corresponding to ORF1a and ORF1b, were then cloned into expression vectors in order to produce the corresponding viral proteins and the antibodies directed against these proteins, in particular by DNA-based immunization.

1. Extraction of the RNAs

The RNAs were extracted with the aid of the QIamp viral RNA extraction mini kit (QIAGEN) according to the manufacturer's recommendations. More specifically: 140 µl of the sample and 560 µl of AVL buffer were vigorously mixed for 15 seconds, incubated for 10 minutes at room temperature and then briefly centrifuged at maximum speed. 560 µl of 100% ethanol were added to the supernatant and the mixture thus obtained was very vigorously stirred for 15 sec. 630 µl of the mixture were then deposited on the column.

The column was placed on a 2 ml tube, centrifuged for 1 min at 8000 rpm, and then the remainder of the preceding mixture was deposited on the same column, centrifuged again, for 1 min at 8000 rpm, and the column was transferred over a clean 2 ml tube. Next, 500 µl of AW1 buffer were added to the column, and then the column was centrifuged for 1 min at 8000 rpm and the eluate was discarded. 500 µl of AW2 buffer were added to the column which was then centrifuged for 3 min at 14 000 rpm and transferred onto a 1.5 ml tube. Finally, 60 µl of AVE buffer were added to the column which was incubated for 1 to 2 min at room temperature and then centrifuged for 1 min at 8000 rpm. The eluate corresponding to the purified RNA was recovered and frozen at -20° C.

2. Amplification, Sequencing and Cloning of the cDNAs

2.1) cDNA Encoding the S Protein

The RNAs extracted from the sample were subjected to reverse transcription with the aid of random sequence hexameric oligonucleotides (pdN6), so as to produce cDNA fragments.

The sequence encoding the SARS-CoV S glycoprotein was amplified in the form of two overlapping DNA fragments: 5' fragment (SARS-Sa, SEQ ID NO: 5) and 3' fragment (SARS-Sb, SEQ ID NO: 6), by carrying out two successive amplifications with the aid of nested primers. The amplicons thus obtained were sequenced, cloned into the PCR plasmid vector 2.1-TOPO™ (INVITROGEN), and then the sequence of the cloned cDNAs was determined.

a) Cloning and Sequencing of the Sa and Sb Fragments

a.1) Synthesis of the cDNA

The reaction mixture containing: RNA (5 µl), H₂O for injection (3.5 µl), 5× reverse transcriptase buffer (4 µl), 5 mM dNTP (2 µl), pdN6 100 µg/ml (4 µl), RNasin 40 IU/µl (0.5 µl) and reverse transcriptase AMV-RT, 10 IU/µl, PROMEGA (1 µl) was incubated in a thermocycler under the following conditions: 45 min at 42° C., 15 min at 55° C., 5 min at 95° C., and then the cDNA obtained was kept at +4° C.

a.2) First PCR Amplification

The 5' and 3' ends of the S gene were respectively amplified with the pairs of primers S/F1+/21350-21372 and S/R1/-/23518-23498, S/F3+/23258-23277 and S/R3/-/25382-25363. The 50 µl reaction mixture containing: cDNA (2 µl), 50 µM primers (0.5 µl), 10× buffer (5 µl), 5 mM dNTP (2 µl), Taq Expand High Fidelity, Roche (0.75 µl) and H₂O (39, 75 µl) was amplified in a thermocycler, under the following conditions: an initial step of denaturation at 94° C. for 2 min was followed by 40 cycles comprising: a step of denaturation at 94° C. for 30 sec, a step of annealing at 55° C. for 30 sec and then a step of extension at 72° C. for 2 min 30 sec, with 10 sec of additional extension at each cycle, and then a final step of extension at 72° C. for 5 min.

a.3) Second PCR Amplification

The products of the first PCR amplification (5' and 3' amplicons) were subjected to a second PCR amplification step (nested PCR) under conditions identical to those of the first amplification, with the pairs of primers S/F2+/21406-21426 and S/R2/-/23454-23435 and S/F4+/23322-23341 and S/R4/-/25348-25329, respectively for the 5' amplicon and the 3' amplicon.

a.4) Cloning and Sequencing of the Sa and Sb Fragments

The Sa (5' end) and Sb (3' end) amplicons thus obtained were purified with the aid of the QIAquick PCR purification kit (QIAGEN), following the manufacturer's instructions, and then they were cloned into the vector PCR2.1-TOPO (Invitrogen kit), to give the plasmids called SARS-S1 and SARS-S2.

The DNA of the Sa and Sb clones was isolated and then the corresponding insert was sequenced with the aid of the Big Dye kit, Applied Biosystem® and universal primers M13 forward and M13 reverse, and primers: S/S+/21867, S/S+/22353, S/S+/22811, S/S+/23754, S/S+/24207, S/S+/24699, S/S+/24348, S/S-/24209, S/S-/23630, S/S-/23038, S/S-/22454, S/S-/21815, S/S-/24784, S/S+/21556, S/S+/23130 and S/S+/24465 following the manufacturer's instructions; the sequences of the Sa and Sb fragments thus obtained correspond to the sequences SEQ ID NO: 5 and SEQ ID NO: 6 in the sequence listing appended as an annex.

The plasmid, called SARS-S1, was deposited under the No. I-3020, on May 12, 2003, at the Collection Nationale de Cultures de Microorganismes, 25 rue du Docteur Roux, 75724 Paris Cedex 15; it contains a 5' fragment of the sequence of the S gene of the SARS-CoV strain derived from the sample recorded under the No. 031589, as defined above, said fragment called Sa corresponding to the nucleotides at positions 21406 to 23454 (SEQ ID NO: 5), with reference to the Genbank sequence AY274119.3 Tor2.

The plasmid, called TOP10F⁺-SARS-S2, was deposited under the No. I-3019, on May 12, 2003, at the Collection Nationale de Cultures de Microorganismes, 25 rue du Docteur Roux, 75724 Paris Cedex 15; it contains a 3' fragment of the sequence of the S gene of the SARS-CoV strain derived from the sample recorded under the No. 031589, as defined above, said fragment called Sb corresponding to the nucleotides at positions 23322 to 25348 (SEQ ID NO: 6), with reference to the Genbank sequence accession No. AY274119.3.

b) Cloning and Sequencing of the Complete cDNA (SARS-S Clone of 4 kb)

The complete S cDNA was obtained from the abovementioned clones SARS-S1 and SARS-S2, in the following manner:

1) A PCR amplification reaction was carried out on a SARS-S2 clone in the presence of the abovementioned primer S/R4/-/25348-25329 and of the primer S/S+/24696-24715: an amplicon of 633 bp was obtained,

2) Another PCR amplification reaction was carried out on another SARS-S2 clone, in the presence of the primers S/F4+/23322-23341 mentioned above and S/S-/24803-24784: an amplicon of 1481 bp was obtained.

The amplification reaction was carried out under the conditions as defined above for the amplification of the Sa and Sb fragments, with the exception that 30 amplification cycles comprising a step of denaturation at 94° C. for 20 sec and a step of extension at 72° C. for 2 min 30 sec were carried out.

3) The 2 amplicons (633 bp and 1481 bp) were purified under the conditions as defined above for the Sa and Sb fragments.

4) Another PCR amplification reaction with the aid of the abovementioned primers S/F4+/23322-23341 and S/R4/-/25348-25329 was carried out on the purified amplicons obtained in 3). The amplification reaction was carried out under the conditions as defined above for the amplification of the Sa and Sb fragments, except that 30 amplification cycles were performed.

The 2026 bp amplicon thus obtained was purified, cloned into the vector PCR2.1-TOPO and then sequenced as above, with the aid of the primers as defined above for the Sa and Sb fragments. The clone thus obtained was called clone 3'.

5) The clone SARS-S1 obtained above and the clone 3' were digested with EcoR I, the bands of about 2 kb thus obtained were gel purified and then amplified by PCR with the abovementioned primers S/F2+/21406-21426 and S/R4/-/25348-25329. The amplification reaction was carried out under the conditions as defined above for the amplification of the Sa and Sb fragments, except that 30 amplification cycles were performed. The amplicon of about 4 kb was purified and sequenced. It was then cloned into the vector PCR2.1-TOPO in order to give the plasmid, called SARS-S, and the insert obtained in this plasmid was sequenced as above, with the aid of the primers as defined above for the Sa and Sb fragments. The cDNA sequences of the insert and of the amplicon encoding the S protein correspond respectively to the sequences SEQ ID NO: 4 and SEQ ID NO: 2 in the sequence listing appended as an annex, they encode the S protein (SEQ ID NO: 3).

The sequence of the amplicon corresponding to the cDNA encoding the S protein of the SARS-CoV strain derived from the sample No. 031589 has the following two mutations compared with the corresponding sequences of respectively the Tor2 and Urbani isolates, the positions of the mutations being indicated with reference to the complete sequence of the genome of the Tor2 isolate (Genbank AY274119.3):

g/t in position 23220; the alanine codon (gct) in position 577 of the amino acid sequence of the S protein of Tor2 is replaced with a serine codon (tct),

c/t in position 24872: this mutation does not modify the amino acid sequence of the S protein, and

the plasmid, called SARS-S, was deposited under the No. I-3059, on Jun. 20, 2003, at the Collection Nationale de Cultures de Microorganismes, 25 rue du Docteur Roux, 75724 Paris Cedex 15; it contains the cDNA sequence encoding the S protein of the SARS-CoV strain derived from the sample recorded under the No. 031589, said sequence corre-

sponding to the nucleotides at positions 21406 to 25348 (SEQ ID NO: 4), with reference to the Genbank sequence AY274119.3.

2.2) cDNA Encoding the M and E Proteins

The RNAs derived from the sample 031589, extracted as above, were subjected to a reverse transcription, combined, during the same step (Titan One Step RT-PCR® kit, Roche), with a PCR amplification reaction, with the aid of the pairs of primers:

S/E/F1+/26051-26070 and S/E/R1-/26455-26436 in order to amplify ORF-E, and

S/M/F1+/26225-26244 and S/M/R1-/27148-27129 in order to amplify ORF-M.

A first reaction mixture containing: 8.6 µl of H₂O for injection, 1 µl of dNTP (5 mM), 0.2 µl of each of the primers (50 µM), 1.25 µl of DTT (100 mM) and 0.25 µl of RNAsin (40 IU/µl) was combined with a second reaction mixture containing: 1 µl of RNA, 7 µl of H₂O for injection, 5 µl of 5×RT-PCR buffer and 0.5 µl of enzyme mixture and the combined mixtures were incubated in a thermocycler under the following conditions: 30 min at 42° C., 10 min at 55° C., 2 min at 94° C. followed by 40 cycles comprising a step of denaturation at 94° C. for 10 sec, a step of annealing at 55° C. for 30 sec and a step of extension at 68° C. for 45 sec, with 3 sec increment per cycle and finally a step of terminal extension at 68° C. for 7 min.

The amplification products thus obtained (M and E amplicons) were subjected to a second PCR amplification (nested PCR) using the Expand High-Fi® kit, Roche), with the aid of the pairs of primers:

S/E/F2+/26082-26101 and S/E/R2-/26413-26394 for the amplicon E, and

S/M/F2+/26330-26350 and S/M/R2-/27098-27078 for the amplicon M.

The reaction mixture containing: 2 µl of the product of the first PCR, 39.25 µl of H₂O for injection, 5 µl of 10× buffer containing MgCl₂, 2 µl of dNTP (5 mM), 0.5 µl of each of the primers (50 µM) and 0.75 µl of enzyme mixture was incubated in a thermocycler under the following conditions: a step of denaturation at 94° C. for 2 min was followed by 30 cycles comprising a step of denaturation at 94° C. for 15 sec, a step of annealing at 60° C. for 30 sec and a step of extension at 72° C. for 45 sec, with 3 sec increment per cycle, and finally a step of terminal extension at 72° C. for 7 min. The amplification products obtained corresponding to the cDNAs encoding the E and M proteins were sequenced as above, with the aid of the primers: S/E/F2+/26082 and S/E/R2-/126394, S/M/F2+/26330, S/M/R2-/27078 cited above and the primers S/M+/26636-26655 and S/M-/26567-26548. They were then cloned, as above, in order to give the plasmids called SARS-E and SARS-M. The DNA of these clones was then isolated and sequenced with the aid of the universal primers M13 forward and M13 reverse and the primers S/M+/26636 and S/M-/26548 mentioned above.

The sequence of the amplicon representing the cDNA encoding the E protein (SEQ ID NO: 13) of the SARS-CoV strain derived from the sample No. 031589 does not contain differences in relation to the corresponding sequences of the isolates AY274119.3-Tor2 and AY278741-Urbani. The sequence of the E protein of the SARS-CoV 031589 strain corresponds to the sequence SEQ ID NO: 14 in the sequence listing appended as an annex.

The plasmid, called SARS-E, was deposited under the No. I-3046, on May 28, 2003, at the Collection Nationale de Cultures de Microorganismes, 25 rue du Docteur Roux, 75724 Paris Cedex 15; it contains the cDNA sequence encoding the E protein of the SARS-CoV strain derived from the

sample recorded under the No. 031589, as defined above, said sequence corresponding to the nucleotides at positions 26082 to 26413 (SEQ ID NO: 15), with reference to the Genbank sequence accession No. AY274119.3.

The sequence of the amplicon representing the cDNA encoding M (SEQ ID NO: 16) from the SARS-CoV strain derived from the sample No. 031589 does not contain differences in relation to the corresponding sequence of the isolate AY274119.3-Tor2. By contrast, at position 26857, the isolate AY278741-Urbani contains a c and the sequence of the SARS-CoV strain derived from the sample recorded under the No. 031589 contains a t. This mutation results in a modification of the amino acid sequence of the corresponding protein: at position 154, a proline (AY278741-Urbani) is changed to serine in the SARS-CoV strain derived from the sample recorded under the No. 031589. The sequence of the M protein of the SARS-CoV strain derived from the sample recorded under the No. 031589 corresponds to the sequence SEQ ID NO: 17 in the sequence listing appended as an annex.

The plasmid, called SARS-M, was deposited under the No. I-3047, on May 28, 2003, at the Collection Nationale de Cultures de Microorganismes, 25 rue du Docteur Roux, 75724 Paris Cedex 15; it contains the cDNA sequence encoding the M protein of the SARS-CoV strain derived from the sample recorded under the No. 031589, as defined above; said sequence corresponding to the nucleotides at positions 26330 to 27098 (SEQ ID NO: 18), with reference to the Genbank sequence accession No. AY274119.3.

2.3) cDNA Corresponding to ORF3, ORF4, ORF7 to ORF11

The same amplification, cloning and sequencing strategy was used to obtain the cDNA fragments corresponding respectively to the following ORFs: ORF3, ORF4, ORF7, ORF8, ORF9, ORF10 and ORF11. The pairs of primers used for the first amplification are:

ORF3 and ORF4: S/SE/F1/+25069-25088 and S/SE/R1/-/26300-26281

ORF7 to ORF11: S/MN/F1/+26898-26917 and S/MN/R1/-/28287-28266

The pairs of primers used for the second amplification are: ORF3 and ORF4: S/SE/F2/+25110-25129 and S/SE/R2/-/26244-26225

ORF7 to ORF11: S/NN/F2/+26977-26996 and S/MN/R2/-/28218-28199

The conditions for the first amplification (RT-PCR) are the following: 45 min at 42° C., 10 min at 55° C., 2 min at 94° C. followed by 40 cycles comprising a step of denaturation at 94° C. for 15 sec, a step of annealing at 58° C. for 30 sec and a step of extension at 68° C. for 1 min, with 5 sec increment per cycle and finally a step of terminal extension at 68° C. for 7 min.

The conditions for the nested PCR are the following: a step of denaturation at 94° C. for 2 min was followed by 40 cycles comprising a step of denaturation at 94° C. for 20 sec, a step of annealing at 58° C. for 30 sec and a step of extension at 72° C. for 50 sec, with 4 sec increment per cycle and finally a step of terminal extension at 72° C. for 7 min.

The amplification products obtained corresponding to the cDNAs containing respectively ORF3 and 4 and ORF7 to 11 were sequenced with the aid of the primers: S/SE/+25363, S/SE/+25835, S/SE/-/25494, S/SE/-/25875, S/MN/+27839, S/MN/+27409, S/MN/-/27836, S/MN/-/27799 and cloned as above for the other ORFs, to give the plasmids called SARS-SE and SARS-MN. The DNA of these clones was isolated and sequenced with the aid of these same primers and of the universal primers M13 sense and M13 antisense.

The sequence of the amplicon representing the cDNA of the region containing ORF3 and ORF4 (SEQ ID NO: 7) of the SARS-CoV strain derived from the sample No. 031589 contains a nucleotide difference in relation to the corresponding sequence of the isolate AY274119-Tor2. This mutation at position 25298 results in a modification of the amino acid sequence of the corresponding protein (ORF3): at position 11, an arginine (AY274119-Tor2) is changed to glycine in the SARS-CoV strain derived from the sample No. 031589. By contrast, no mutation was identified in relation to the corresponding sequence of the isolate AY278741-Urbani. The sequences of ORF3 and 4 of the SARS-COV strain derived from the sample No. 031589 correspond respectively to the sequences SEQ ID NO: 10 and 12 in the sequence listing appended as an annex.

The plasmid, called SARS-SE, was deposited under the No. I-3126, on Nov. 13, 2003, at the Collection Nationale de Cultures de Microorganismes, 25 rue du Docteur Roux, 75724 Paris Cedex 15; it contains the cDNA corresponding to the region situated between ORF-S and ORF-E and overlapping ORF-E of the SARS-CoV strain derived from the sample recorded under the No. 031589, as defined above, said region corresponding to the nucleotides at positions 25110 to 26244 (SEQ ID NO: 8), with reference to the Genbank sequence accession No. AY274119.3.

The sequence of the amplicon representing the cDNA corresponding to the region containing ORF7 to ORF11 (SEQ ID NO: 19) of the SARS-CoV strain derived from the sample No. 031589 does not contain differences in relation to the corresponding sequences of the isolates AY274119-Tor2 and AY278741-Urbani. The sequences of ORF7 to 11 of the SARS-CoV strain derived from the sample No. 031589 correspond respectively to the sequences SEQ ID NO: 22, 24, 26, 28 and 30 in the sequence listing appended as an annex.

The plasmid, called SARS-MN, was deposited under the No. I-3125, on Nov. 13, 2003, at the Collection Nationale de Cultures de Microorganismes, 25 rue du Docteur Roux, 75724 Paris Cedex 15; it contains the cDNA sequence corresponding to the region situated between ORF-M and ORF-N of the SARS-CoV strain derived from the sample recorded under the No. 031589 and collected in Hanoi, as defined above, said sequence corresponding to the nucleotides at positions 26977 to 28218 (SEQ ID NO: 20), with reference to the Genbank sequence accession No. AY274119.3.

The sequence of the amplicon representing the cDNA corresponding to the region containing ORF7 to ORF11 (SEQ ID NO: 19) of the SARS-CoV strain derived from the sample No. 031589 does not contain differences in relation to the corresponding sequences of the isolates AY274119-Tor2 and AY278741-Urbani. The sequences of ORF7 to 11 of the SARS-CoV strain derived from the sample No. 031589 correspond respectively to the sequences SEQ ID NO: 22, 24, 26, 28 and 30 in the sequence listing appended as an annex.

2.4) cDNA Encoding the N Protein and Including ORF13 and ORF14

The cDNA was synthesized and amplified as described above for the fragments Sa and Sb. More specifically, the reaction mixture containing: 5 µl of RNA, 5 µl of H₂O for injection, 4 µl of 5× reverse transcriptase buffer, 2 µl of dNTP (5 mM), 2 µl of oligo 20 T (5 µM), 0.5 µl of rNasin (40 IU/µl) and 1.5 µl of AMV-RT (10 IU/µl Promega) was incubated in a thermocycler under the following conditions: 45 min at 42° C., 15 min at 55° C., 5 min at 95° C., and it was then kept at 4° C.

A first PCR amplification was performed with the pair of primers S/N/F3/+28023 and S/N/R3/-/29460.

The reaction mixture as above for the amplification of the S1 and S2 fragments was incubated in a thermo-cycler, under the following conditions: an initial step of denaturation at 94° C. for 2 min was followed by 40 cycles comprising a step of denaturation at 94° C. for 20 sec, a step of annealing at 55° C. for 30 sec and then a step of extension at 72° C. for 1 min 30 sec with 10 sec of additional extension at each cycle, and then a final step of extension at 72° C. for 5 min.

The amplicon obtained at the first PCR amplification was subjected to a second PCR amplification step (nested PCR) with the pairs of primer S/N/F4+/28054 and S/N/R4-/29430 under conditions identical to those of the first amplification.

The amplification product obtained, corresponding to the cDNA encoding the N protein of the SARS-CoV strain derived from the sample No. 031589, was sequenced with the aid of the primers: S/N/F4+/28054, S/N/R4-/29430, S/N/+28468, S/N/+28918 and S/N/-28607 and cloned as above for the other ORFs, to give the plasmid called SARS-N. The DNA of these clones was isolated and sequenced with the aid of the universal primers M13 sense and M13 antisense, and the primers S/N/+28468, S/N/+28918 and S/N/-28607.

The sequence of the amplicon representing the cDNA corresponding to ORF-N and including ORF13 and ORF14 (SEQ ID NO: 36) of the SARS-CoV strain derived from the sample No. 031589 does not contain differences in relation to the corresponding sequences of the isolates AY274119.3-Tor2 and AY278741-Urbani. The sequence of the N protein of the SARS-CoV strain derived from the sample No. 031589 corresponds to the sequence SEQ ID NO: 37 in the sequence listing appended as an annex.

The sequences of ORF13 and 14 of the SARS-CoV strain derived from the sample No. 031589 correspond respectively to the sequences SEQ ID NO: 32 and 34 in the sequence listing appended as an annex.

The plasmid, called SARS-N, was deposited under the No. I-3048, on Jun. 5, 2003, at the Collection Nationale de Cultures de Microorganismes, 25 rue du Docteur Roux, 75724 Paris Cedex 15; it contains the cDNA encoding the N protein of the SARS-CoV strain derived from the sample recorded under the No. 031589, as defined above, said sequence corresponding to the nucleotides at positions 28054 to 29430 (SEQ ID NO: 38), with reference to the Genbank sequence accession No. AY274119.3.

2.5) Noncoding 5' and 3' Ends

a) Noncoding end (5'NC)

a₁) Synthesis of the cDNA

The RNAs derived from the sample 031589, extracted as above, were subjected to reverse transcription under the following conditions:

The RNA (15 µl) and the primer S/L-/443 (3 µl at the concentration of 5 µM) were incubated for 10 min at 75° C.

Next, the 5× reverse transcriptase buffer (6 µl, INVITROGEN), 10 Mm dNTP (1 µl), 0.1 M DTT (3 µl) were added and the mixture was incubated at, 50° C. for 3 min.

Finally, the reverse transcriptase (3 µl of Superscript®, INVITROGEN) was added to the preceding mixture which was incubated at 50° C. for 1 h 30 min and then at 90° C. for 2 min.

The cDNA thus obtained was purified with the aid of the QIAquick PCR purification kit (QIAGEN), according to the manufacturer's recommendations.

b₁) Terminal Transferase Reaction (TdT)

The cDNA (10 µl) is incubated for 2 min at 100° C., stored in ice, and the following are then added: H₂O (2.5 µl), 5× TdT buffer (4 µl, AMERSHAM), 5 mM dATP (2 µl) and TdT (1.5 µl, AMERSHAM). The mixture thus obtained is incubated for 45 min at 37° C. and then for 2 min at 65° C.

The product obtained is amplified by a first PCR reaction with the aid of the primers: S/L/-225-206 and anchor 14T: 5'-AGATGAATTCGGTACCTTTTTTTTTTTTTTTT-3' (SEQ ID NO: 68). The amplification conditions are the following: an initial step of denaturation at 94° C. for 2 min is followed by 10 cycles comprising a step of denaturation at 94° C. for 10 sec, a step of annealing at 45° C. for 30 sec and then a step of extension at 72° C. for 30 sec and then by 30 cycles comprising a step of denaturation at 94° C. for 10 sec, a step of annealing at 50° C. for 30 sec and then a step of extension at 72° C. for 30 sec, and then a final step of extension at 72° C. for 5 min.

The product of the first PCR amplification was subjected to a second amplification step with the aid of the primers: S/L/-204-185 and anchor 14 T mentioned above under conditions identical to those of the first amplification. The amplicon thus obtained was purified, sequenced with the aid of the primer S/L/-182-163 and it was then cloned as above for the different ORFs, to give the plasmid called SARS-5'NC. The DNA of this clone was isolated and sequenced with the aid of the universal primers M13 sense and M13 antisense and the primer S/L/-182-163 mentioned above.

The amplicon representing the cDNA corresponding to the 5'NC end of the SARS-CoV strain derived from the sample recorded under the No. 031589 corresponds to the sequence SEQ ID NO: 72 in the sequence listing appended as an annex; this sequence does not contain differences in relation to the corresponding sequences of the isolates AY274119.3-Tor2 and AY278741-Urbani.

The plasmid, called SARS-5'NC, was deposited under the No. I-3124, on Nov. 7, 2003, at the Collection Nationale de Cultures de Microorganismes, 25 rue du Docteur Roux, 75724 Paris Cedex 15; it contains the cDNA corresponding to the noncoding 5' end of the genome of the SARS-CoV strain derived from the sample recorded under the No. 031589, as defined above, said sequence corresponding to the nucleotides at positions 1 to 204 (SEQ ID NO: 39), with reference to the Genbank sequence accession No. AY274119.3.

b) Noncoding 3' End (3'NC)

a₁) Synthesis of the cDNA

The RNAs derived from the sample 031589, extracted as above, were subjected to reverse transcription, according to the following protocol: the reaction mixture containing: RNA (5 µl), H₂O (5 µl), 5× reverse transcriptase buffer (4 µl), 5 mM dNTP (2 µl), 5 µM Oligo 20 T (2 µl), 40 U/µl RNasin (0.5 µl) and 10 IU/µl RT-AMV (1.5 µl, PROMEGA) was incubated in a thermo-cycler, under the following conditions: 45 min at 42° C., 15 min at 55° C., 5 min at 95° C., and it was then kept at +4° C.

The cDNA obtained was amplified by a first PCR reaction with the aid of the primers S/N/+28468-28487 and anchor 14 T mentioned above. The amplification conditions are the following: an initial step of denaturation at 94° C. for 2 min is followed by 10 cycles comprising a step of denaturation at 94° C. for 20 sec, a step of annealing at 45° C. for 30 sec and then a step of extension at 72° C. for 50 sec and then 30 cycles comprising a step of denaturation at 94° C. for 20 sec, a step of annealing at 50° C. for 30 sec and then a step of extension at 72° C. for 50 sec, and then a final step of extension at 72° C. for 5 min.

The product of the first PCR amplification was subjected to a second amplification step with the aid of the primers S/N/+28933-28952 and anchor 14 T mentioned above, under conditions identical to those of the first amplification. The amplicon thus obtained was purified, sequenced with the aid of the primer S/N/+29257-29278 and cloned as above for the different ORFs, to give the plasmid called SARS-3'NC. The DNA of this clone was isolated and sequenced with the aid of the universal primers M13 sense and M13 antisense and the primer S/N/+29257-29278 mentioned above.

031589 was performed by carrying out RT-PCR reactions followed by nested PCRs according to the same principles as those described above for the other ORFs. The amplified fragments overlap over several tenths of bases, thus allowing computer reconstruction of the complete sequence of this part of the genome. On average, the amplified fragments are of two kilobases.

14 overlapping fragments, called L0 to L12, were thus amplified with the aid of the following primers:

TABLE II

Primers used for the amplification of the 5' region (ORF1a and ORF1b)				
REGION AMPLIFIED AND SEQUENCED (does not include the primers)	RT-PCR sense primer	RT-PCR antisense primer	Nested PCR sense primer	Nested PCR antisense primer
L0 50-480	S/L0/F1/+30	S/L0/R1/-481		
L1 231-2240	S/L1/F1/+147	S/L1/R1/-2336	S/L1/F2/+211	S/L1/R2/-2241
L2 2156-4167	S/L2/F1/+2033	S/L2/R1/-4192	S/L2/F2/+2136	S/L2/R2/-4168
L3 3913-5324	S/L3bis/F1/+3850	S/L3bis/R1/-5365	S/L3bis/F2/+3892	S/L3bis/R2/-5325
L4b 4952-6023	S/L4b/F1/+4878	S/L4b/R1/-6061	S/L4b/F2/+4932	S/L4b/R2/-6024
L4 5325-7318	S/L4/F1/+5272	S/L4/R1/-7392	S/L4/F2/+5305	S/L4/R2/-7323
L5 7296-9156	S/L5/F1/+7111	S/L5/R1/-9253	S/L5/F2/+7275	S/L5/R2/-9157
L6 9053-11066	S/L6/F1/+8975	S/L6/R1/-11151	S/L6/F2/+9032	S/L6/R2/-11067
L7 10928-12962	S/L7/F1/+10883	S/L7/R1/-13050	S/L7/F2/+10928	S/L7/R2/-12963
L8 12835-14834	S/L8/F1/+12690	S/L8/R1/-14857	S/L8/F2/+12815	S/L8/R2/-14835
L9 14765-16624	S/L9/F1/+14688	S/L9/R1/-16678	S/L9/F2/+14745	S/L9/R2/-16625
L10 16534-18570	S/L10/F1/+16451	S/L10/R1/-18594	S/L10/F2/+16514	S/L10/R2/-18571
L11 18521-20582	S/L11/F1/+18441	S/L11/R1/-20612	S/L11/F2/+18500	S/L11/R2/-20583
L12 20338-22205.	S/L12/F1/+20279	S/L12/R1/-22229	S/L12/F2/+20319	S/L12/R2/-22206

The amplicon representing the cDNA corresponding to the 3'NC end of the SARS-CoV strain derived from the sample recorded under the No. 031589 corresponds to the sequence SEQ ID NO: 73 in the sequence listing appended as an annex; this sequence does not contain differences in relation to the corresponding sequences of the isolates AY274119.3-Tor2 and AY278741-Urbani.

The plasmid called SARS-3'NC was deposited under the No. I-3123 on Nov. 7, 2003, at the Collection Nationale de Cultures de Microorganismes, 25 rue du Docteur Roux, 75724 Paris Cedex 15; it contains the cDNA sequence corresponding to the noncoding 3' end of the genome of the SARS-CoV strain derived from the sample recorded under the No. 031589, as defined above, said sequence corresponding to that situated between the nucleotide at positions 28933 to 29727 (SEQ ID NO: 40), with reference to the Genbank sequence accession No. AY274119.3, ends with a series of nucleotides a.

2.6) ORF1a and ORF1b

The amplification of the 5' region containing ORF1a and ORF1b of the SARS-CoV genome derived from the sample

All the fragments were amplified under the following conditions, except fragment L0 which was amplified as described above for ORF-M:

RT-PCR: 30 min at 42° C., 15 min at 55° C., 2 min at 94° C., and then the cDNA obtained is amplified under the following conditions: 40 cycles comprising: a step of denaturation at 94° C. for 15 sec, a step of annealing at 58° C. for 30 sec and then a step of extension at 68° C. for 1 min 30 sec, with 5 sec additional extension at each cycle, and then a final step of extension at 68° C. for 7 min.

Nested PCR: An initial step of denaturation at 94° C. for 2 min is followed by 35 cycles comprising: a step of denaturation at 94° C. for 15 sec, a step of annealing at 60° C. for 30 sec and then a step of extension at 72° C. for 1 min 30 sec, with 5 sec of additional extension at each cycle, and then a final step of extension at 72° C. for 7 min.

The amplification products were sequenced with the aid of the primers defined in table III below:

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TABLE III

Primers used for the sequencing of the 5' region (ORF1a and ORF1b)	
Names	Sequences (SEQ ID NO: 76 to 139)
S/L3/+4932	5'-CCACACACAGCTTGTGGATA-3'
S/L4/+6401	5'-CCGAAGTTGTAGGCAATGTC-3'
S/L4/+6964	5'-TTTGGTCTCCTTCTTATTG -3'
S/L4/-6817	5'-CCGGCATCCAAACATAATTT-3'
S/L5/-7633	5'-TGGTCAGTAGGGTTGATTGG-3'
S/L5/-8127	5'-CATCCTTTGTGTCAACATCG-3'
S/L5/-8633	5'-GTCACGAGTGACACCATCCT-3'
S/L5/+7839	5'-ATCGCAGAGTCTGCTTCTA-3'
S/L5/+8785	5'-TTCATAGTGCCTGGCTTACC-3'
S/L5/+8255	5'-ATCTTGGCGCATGTATTGAC-3'
S/L6/-9422	5'-TGCATTAGCAGCAACAACAT-3'
S/L6/-9966	5'-TCTGCAGAACAGCAGAAGTG-3'
S/L6/-10542	5'-CCTGTGCAGTTTGTCTGTCA-3'
S/L6/+10677	5'-CCTTGTGGCAATGAAGTACA-3'
S/L6/+10106	5'-ATGTCATTGACAGCAGAA-3'
S/L6/+9571	5'-CTTCAATGGTTTGCCATGTT-3'
S/L7/-11271	5'-TGCGAGCTGTCTATGAGAATA-3'
S/L7/-11801	5'-AACCGAGAGCAGTACCACAG-3'
S/L7/-12383	5'-TTTGGCTGCTGTAGTCAATG-3'
S/L7/+12640	5'-CTACGACAGATGTCTGTGC-3'
S/L7/+12088	5'-GAGCAGGCTGTAGCTAATGG-3'
S/L7/+11551	5'-TTAGGCTATTGTTGCTGCTG-3'
S/L8/-13160	5'-CAGACAACATGAAGCACCAC-3'
S/L8/-13704	5'-CGCTGACGTGATATATGTGG-3'
S/L8/-14284	5'-TGCACAATGAAGGATACACC-3'
S/L8/+14453	5'-ACATAGCTCGCGTCTCAGTT-3'
S/L8/+13968	5'-GGCATTGTAGGCGTACTGAC-3'
S/L8/+13401	5'-GTTTGGCGGTGTAAGTGCAG-3'
S/L9/-15099	5'-TAGTGGCGGCTATTGACTTC-3'
S/L9/-15677	5'-CTAAACCTTGAGCCGCATAG-3'
S/L9/-16247	5'-CATGGTCATAGCAGCACTTG-3'
S/L9/+16323	5'-CCAGGTTGTGATGTCACTGAT-3'
S/L9/+15858	5'-CCTTACCAGATCCATCAAG-3'
S/L9/+15288	5'-CGCAAACATAAACACTTGCTG-3'
S/L10/-16914	5'-AGTGTGGGTACAAGCCAGT-3'
S/L10/-17466	5'-GTTCCAAGGAACATGTCTGG-3'
S/L10/-18022	5'-AGGTGCCTGTGTAGGATGAA-3'
S/L10/+18245	5'-GGGCTGTCTGCAACTAGAG-3'

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TABLE III-continued

Primers used for the sequencing of the 5' region (ORF1a and ORF1b)	
Names	Sequences (SEQ ID NO: 76 to 139)
S/L10/+17663	5'-TCTTACACGCAATCTGCTT-3'
S/L10/+17061	5'-TACCCATCTGCTCGCATAGT-3'
S/L11/-18877	5'-GCAAGCAGAATTAACCCCTCA-3'
S/L11/-19396	5'-AGCACCACCTAAATTGCATC-3'
S/L11/-20002	5'-TGGTCCCTTTGAAGGTGTTA-3'
S/L11/+20245	5'-TCGAACACATCGTTTATGGA-3'
S/L11/+19611	5'-GAAGCACCTGTTTCCATCAT-3'
S/L11/+19021	5'-ACGATGCTCAGCCATGTAGT-3'
SARS/L1/F3/+800	5'-GAGGTGCAGTCACTCGCTAT-3'
SARS/L1/F4/+1391	5'-CAGAGATTGGACCTGAGCAT-3'
SARS/L1/F5/+1925	5'-CAGCAAACCACTCAATTCCT-3'
SARS/L1/R3/-1674	5'-AAATGATGGCAACCTCTTCA-3'
SARS/L1/R4/-1107	5'-CACGTGGTTGAATGACTTTG-3'
SARS/L1/R5/-520	5'-ATTTCTGCAACCAGCTCAAC-3'
SARS/L2/F3/+2664	5'-CGCATTGTCTCTCGTTTAC-3'
SARS/L2/F4/+3232	5'-GAGATTGAGCCAGAACCAGA-3'
SARS/L2/F5/+3746	5'-ATGAGCAGGTTGTCTGGAT-3'
SARS/L2/R3/-3579	5'-CTGCCTTAAGAAGCTGGATG-3'
SARS/L2/R4/-2991	5'-TTTCTTACCAGCATCATCA-3'
SARS/L2/R5/-2529	5'-CACCGTTCTTGAGAACAACC-3'
SARS/L3/F3/+4708	5'-TCTTTGGCTGGCTCTTACAG-3'
SARS/L3/F4/+5305	5'-GCTGGTGATGCTGCTAACTT-3'
SARS/L3/F5/+5822	5'-CCATCAAGCCTGTGTCTGAT-3'
SARS/L3/R3/-5610	5'-CAGGTGGTGCAGACATCATA-3'
SARS/L3/R4/-4988	5'-AACATCAGCACCATCCAAGT-3'
SARS/L3/R5/-4437	5'-ATCGGACACCATAGTCAACG-3'

The sequences of the fragments L0 to L12 of the SARS-CoV strain derived from the sample recorded under the No. 031589 correspond respectively to the sequences SEQ ID NO: 41 to SEQ ID NO: 54 in the sequence listing appended as an annex. Among these sequences, only that corresponding to the fragments L5 contains a nucleotide difference in relation to the corresponding sequence of the isolate AY278741-Urbani. This t/c mutation at position 7919 results in a modification of the amino acid sequence of the corresponding protein, encoded by ORF1a: at position 2552, a valine (gtt codon; AY278741) is changed to alanine (gct codon) in the SARS-CoV strain 031589. By contrast, no mutation was identified in relation to the corresponding sequence of the isolate AY274119.3-Urbani. The other fragments do not exhibit differences in relation to the corresponding sequences of the isolates Tor2 and Urbani.

Production and Purification of the Recombinant N and S Proteins of the SARS-CoV Strain Derived from the Sample Recorded Under the Number 031589

The entire N protein and two polypeptide fragments of the S protein of the SARS-CoV strain derived from the sample recorded under the number 031589 were produced in *E. coli*, in the form of fusion proteins comprising an N- or C-terminal polyhistidine tag. In the two S polypeptides, the N- and C-terminal, hydrophobic sequences of the S protein (signal peptide: positions 1 to 13 and transmembrane helix: positions 1196 to 1218) were deleted whereas the β helix (positions 565 to 687) and the two motifs of the coiled-coil type (positions 895 to 980 and 1155 to 1186) of the S protein were preserved. These two polypeptides consist of: a long fragment (S_L) corresponding to positions 14 to 1193 of the amino acid sequence of the S protein and a short fragment (S_C) corresponding to positions 475 to 1193 of the amino acid sequence of the S protein.

1) Cloning of the cDNAs N, S_L and S_C into the Expression Vectors pIVEX2.3 and pIVEX2.4

The cDNAs corresponding to the N protein and to the S_L and S_C fragments were amplified by PCR under standard conditions, with the aid of the DNA polymerase Platinum Pfx® (INVITROGEN). The plasmids SRAS-N and SRAS-S were used as template and the following oligo-nucleotides as primers:

5'-CCCATATGTCTGATAATGGACCCCAATCAAAC-3' (N sense,

SEQ ID NO: 55)

5'-CCCCCGGGTGCCTGAGTTGAATCAGCAGAAGC-3' (N

antisense, SEQ ID NO: 56)

5'-CCCATATGAGTGACCTTGACCGGTGCACCAC-3' (S_C sense,

SEQ ID NO: 57)

5'-CCCATATGAAACCTTGCACCCACCTGCTC-3' (S_L sense,

SEQ ID NO: 58)

5'-CCCCCGGGTTTAATATATGCTCATATTTTCCC-3' (S_C and

S_L antisense, SEQ ID NO: 29).

The sense primers introduce an NdeI site (underlined) while the antisense primers introduce an XmaI or SmaI site (underlined). The 3 amplification products were column purified (QIAquick PCR Purification kit, QIAGEN) and cloned into an appropriate vector. The plasmid DNA purified from the 3 constructs (QIAfilter Midi Plasmid kit, QIAGEN) was verified by sequencing and digested with the enzymes NdeI and XmaI. The 3 fragments corresponding to the cDNAs N, S_L and S_C were purified on agarose gel and then inserted into the plasmids pIVEX2.3MCS (C-terminal polyhistidine tag) and pIVEX2.4d (N-terminal polyhistidine tag) digested beforehand with the same enzymes. After verification of the constructs, the 6 expression vectors thus obtained (pIV2.3N, pIV2.3 S_C , pIV2.3 S_L , pIV2.4N, pIV2.4 S_C also called pIV2.4 S_1 , pIV2.4 S_2) were then used, on the one hand to test the expression of the proteins in vitro, and on the other hand to transform the bacterial strain BL21(DE3)pDIA17 (NOVAGEN). These constructs encode proteins whose expected molecular mass is the following: pIV2.3N (47174

Da), pIV2.3 S_C (82897 Da), pIV2.3 S_L (132056 Da), pIV2.4N (48996 Da), pIV2.4 S_1 (81076 Da) and pIV2.4 S_2 (133877 Da). Bacteria transformed with pIV2.3N were deposited at the CNCM on Oct. 23, 2003, under the number I-3117, and bacteria transformed with pIV2.4 S_1 were deposited at the CNCM on Oct. 23, 2003, under the number I-3118.

2) Analysis of the Expression of the Recombinant Proteins In Vitro and In Vivo

The expression of recombinant proteins from the 6 recombinant vectors was tested, in a first instance, in a system in vitro (RTS100, Roche). The proteins produced in vitro, after incubation of the recombinant vectors pIVEX for 4 h at 30° C., in the RTS100 system, were analyzed by Western blotting with the aid of an anti-(his)₆ antibody coupled to peroxidase. The result of expression in vitro (FIG. 1) shows that only the N protein is expressed in large quantities, regardless of the position, N- or C-terminal, of the polyhistidine tag. In a second step, the expression of the N and S proteins was tested in vivo at 30° C. in LB medium in the presence or in the absence of inducer (1 mM IPTG). The N protein is very well produced in this bacterial system (FIG. 2) and is found mainly in a soluble fraction after lysis of the bacteria. By contrast, the long version of S (S_L) is very weakly produced and is completely insoluble (FIG. 3). The short version (S_C) also exhibits a very weak solubility, but an expression level that is much higher than that of the long version. Moreover, the construct S_C fused with a polyhistidine tag at the C-terminal position has a smaller size than that expected. An immunodetection experiment with an anti-polyhistidine antibody has shown that this construct was incomplete. In conclusion, the two constructs, pIV2.3N and pIV2.4 S_1 , which express respectively the entire N protein fused with the C-terminal polyhistidine tag and the short S protein fused with the N-terminal polyhistidine tag, were selected in order to produce the two proteins in a large quantity so as to purify them. The plasmids pIV2.3N and pIV2.4 S_1 were deposited respectively under the No. I-3117 and I-3118 at the CNCM, 25 rue du Docteur Roux, 75724 PARIS 15, on Oct. 23, 2003.

3) Analysis of the Antigenic Activity of the Recombinant Proteins

The antigenic activity of the N, S_L and S_C proteins was tested by Western blotting with the aid of two serum samples, obtained from the same patient infected with SARS-CoV, collected 8 days (M12) and 29 days (M13) after the onset of the SARS symptoms. The experimental protocol is as described in example 3. The results illustrated by FIG. 4 show (i) the seroconversion of the patient, and (ii) that the N protein possesses a higher antigenic reactivity than the short S protein.

4) Purification of the N protein from pIV2.3N

Several experiments for purifying the N protein, produced from the vector pIV2.3N, were carried out according to the following protocol. The bacteria BL21(DE3)pDIA17, transformed with the expression vector pIV2.3N, were cultured at 30° C. in 1 liter of culture medium containing 0.1 mg/ml of ampicillin, and induced with 1 mM IPTG when the cell density equivalent to $A_{600}=0.8$ is reached (about 3 hours). After 2 hours of culture in the presence of inducer, the cells were recovered by centrifugation (10 min at 5000 rpm), resuspended in the lysis buffer (50 mM NaH₂PO₄, 0.3 M NaCl, 20 mM imidazole, pH 8, containing the mixture of protease inhibitors Complete®, Roche), and lysed with the French press (12 000 psi). After centrifugation of the bacterial lysate (15 min at 12 000 rpm), the supernatant (50 ml) was deposited at a flow rate of 1 ml/min on a metal chelation column (15 ml) (Ni-NTA superflow, Qiagen), equilibrated with the lysis buffer. After washing the column with 200 ml of lysis buffer,

the N protein was eluted with an imidazole gradient (20→250 mM) in 10 column volumes. The fractions containing the N protein were assembled and analyzed by polyacrylamide gel electrophoresis under denaturing conditions followed by staining with Coomassie blue. The results illustrated by FIG. 5 show that the protocol used makes it possible to purify the N protein with a very satisfactory homogeneity (95%) and a mean yield of 15 mg of protein per liter of culture.

5) Purification of the S_c Protein from pIV2.4S_c (pIV2.4S₁)

The protocol followed for purifying the short S protein is very different from that described above because the protein is highly aggregated in the bacterial system (inclusion bodies). The bacteria BL21(DE3)pDIA17, transformed with the expression vector pIV2.4S₁, were cultured at 30° C. in 1 liter of culture medium containing 0.1 mg/ml of ampicillin, and induced with 1 mM IPTG when the cell density equivalent to A₆₀₀=0.8 is reached (about 3 hours). After 2 hours of culture in the presence of inducer, the cells were recovered by centrifugation (10 min at 5000 rpm), resuspended in the lysis buffer (0.1 M Tris-HCl, 1 mM EDTA, pH 7.5), and lysed with the French press (1200 psi). After centrifugation of the bacterial lysate (15 min at 12 000 rpm), the pellet was resuspended in 25 ml of lysis buffer containing 2% Triton X100 and 10 mM β-mercaptoethanol, and then centrifuged for 20 min at 12 000 rpm. The pellet was resuspended in 10 mM Tris-HCl buffer containing 7 M urea, and gently stirred for 30 min at room temperature. This final washing of the inclusion bodies with 7 M urea is necessary in order to remove most of the *E. coli* membrane proteins which co-sediment with the aggregated S_c protein. After a final centrifugation for 20 min at 12 000 rpm, the final pellet is resuspended in the 10 mM Tris-HCl buffer. The electrophoretic analysis of this preparation (FIG. 6) shows that the short S protein may be purified with a satisfactory homogeneity (about 90%) from the inclusion bodies (insoluble extract).

EXAMPLE 3

Immunodominance of the N Protein

The reactivity of the antibodies present in the serum of patients suffering from atypical pneumopathy caused by the SARS-associated coronavirus (SARS-CoV), toward the various proteins of this virus, was analyzed by Western blotting under the conditions described below.

1) Materials

a) Lysate of Cells Infected with SARS-CoV

Vero E6 cells (2×10⁶) were infected with SARS-CoV (isolate recorded under the number FFM/MA104) at a multiplicity of infection (M.O.I.) of 10⁻¹ or 10⁻² and then incubated in DMEM medium containing 2% FCS, at 35° C. in an atmosphere containing 5% CO₂. 48 hours later, the cellular lawn was washed with PBS and then lysed with 500 μl of loading buffer prepared according to Laemmli and containing β-mercaptoethanol. The samples were then boiled for 10 minutes and then sonicated for 3 times 20 seconds.

b) Antibodies

b₁) Serum from a Patient Suffering from Atypical Pneumopathy

The serum designated by a reference at the National Reference Center for Influenza Viruses (Northern region) under the No. 20033168 is that from a French patient suffering from atypical pneumopathy caused by SARS-CoV collected on day 38 after the onset of the symptoms; the diagnosis of SARS-CoV infection was performed by nested RT-PCR and quantitative PCR.

b₂) Monospecific Rabbit Polyclonal sera Directed Against the N Protein or the S Protein

The sera are those produced from the recombinant N and S_c proteins (example 2), according to the immunization protocol described in example 4; they are the rabbit P13097 serum (anti-N serum) and the rabbit P11135 serum (anti-S serum).

2) Method

20 μl of lysate of cells infected with SARS-CoV at M.O.I. values of 10⁻¹ and 10⁻² and, as a control, 20 μl of a lysate of noninfected cells (mock) were separated on 10% SDS polyacrylamide gel and then transferred onto a nitrocellulose membrane. After blocking in a solution of PBS/5% milk/0.1% Tween and washing in PBS/0.1% Tween, this membrane was hybridized overnight at 4° C. with: (1) the immune serum No. 20033168 diluted 1/300, 1/1000 and 1/3000 in the buffer PBS/1% BSA/0.1% Tween, (ii) the rabbit P13097 serum (anti-N serum) diluted 1/50 000 in the same buffer and (iii) the rabbit P11135 serum (anti-S serum) diluted 1/10 000 in the same buffer. After washing in PBS/Tween, a secondary hybridization was performed with the aid of either sheep polyclonal antibodies directed against the heavy and light chains of human G immunoglobulins and coupled with peroxidase (NA933V, Amersham), or of donkey polyclonal antibodies directed against the heavy and light chains of the rabbit G immunoglobulins and coupled with peroxidase (NA934V, Amersham). The bound antibodies were visualized with the aid of the ECL+ kit (Amersham) and of Hyperfilm MP autoradiography films (Amersham). A molecular mass ladder (kDa) is presented in the figure.

3) Results

FIG. 7 shows that three polypeptides of apparent molecular mass 35, 55 and 200 kDa are specifically detected in the extracts of cells infected with SARS-CoV.

In order to identify these polypeptides, two other immunoblots (FIG. 8) were prepared on the same samples and under the same conditions with rabbit polyclonal antibodies specific for the nucleoprotein N (rabbit P13097, FIG. 8A) and for the spicule protein S (rabbit P11135, FIG. 8B). This experiment shows that the 200 kDa polypeptide corresponds to the SARS-CoV spicule glycoprotein S, that the 55 kDa polypeptide corresponds to the nucleoprotein N while the 35 kDa polypeptide probably represents a truncated or degraded form of N.

The data presented in FIG. 7 therefore show that the serum 20033168 strongly reacts with N and a lot more weakly with the SARS-CoV S since the 35 and 55 kDa polypeptides are visualized in the form of intense bands for 1/300, 1/1000 and 1/3000 dilutions of the immunoserum whereas the 200 kDa polypeptide is only weakly visualized for a dilution of 1/300. It is also possible to note that no other SARS-CoV polypeptide is detected for dilutions greater than 1/300 of the serum 20033168.

This experiment indicates that the antibody response specific for the SARS-CoV N dominates the antibody responses specific for the other SARS-CoV polypeptides and in particular the antibody response directed against the S glycoprotein. It indicates an immunodominance of the nucleoprotein N during human infections with SARS-CoV.

Preparation of Monospecific Polyclonal Antibodies
Directed Against the SRAS-Associated Coronavirus
(SARS-CoV) N and S Proteins

1) Materials and Method

Three rabbits (P13097, P13081, P13031) were immunized with the purified recombinant polypeptide corresponding to the entire nucleoprotein (N), prepared according to the protocol described in example 2. After a first injection of 0.35 mg per rabbit of protein emulsified in complete Freund's adjuvant (intradermal route), the animals received 3 booster injections at 3 and then 4 weeks' interval, of 0.35 mg of recombinant protein emulsified in incomplete Freund's adjuvant.

Three rabbits (P11135, P13042, P14001) were immunized with the recombinant polypeptide corresponding to the short fragment of the S protein (S_c) produced as described in example 2. As this polypeptide is found mainly in the form of inclusion bodies in the bacterial cytoplasm, the animals received 4 intradermal injections at 3-4 weeks' interval of a preparation of inclusion bodies corresponding to 0.5 mg of recombinant protein emulsified in incomplete Freund's adjuvant. The first 3 injections were made with a preparation of inclusion bodies prepared according to the protocol described in example 2, while the fourth injection was made with a preparation of inclusion bodies which were prepared according to the protocol described in example 2 and then purified on sucrose gradient and washed in 2% Triton X100.

For each rabbit, a preimmune (p.i.) serum was prepared before the first immunization and an immune serum (I.S.) 5 weeks after the fourth immunization.

In a first instance, the reactivity of the sera was analyzed by ELISA test on preparations of recombinant proteins similar to those used for the immunizations; the ELISA tests were carried out according to the protocol and with the reagents as described in example 6.

In a second instance, the reactivity of the sera was analyzed by preparing an immunoblot (Western blot) of a lysate of cells infected with SARS-CoV, according to the protocol as described in example 3.

2) Results

The ELISA tests (FIG. 9) demonstrate that the preparations of recombinant N protein and of inclusion bodies of the short fragment of the S protein (S_c) are immunogenic in animals and that the titer of the immune sera is high (more than 1/25 000).

The immunoblot (FIG. 8) shows that the rabbit P13097 immune serum recognizes two polypeptides present in the lysates of cells infected with SARS-CoV: a polypeptide whose apparent molecular mass (50-55 kDa based on experiments) is compatible with that of the nucleoprotein N (422 residues, predicted molecular mass of 46 kDa) and a polypeptide of 35 kDa, which probably represents a truncated or degraded form of N.

This experiment also shows that the rabbit P11135 serum mainly recognizes a polypeptide whose apparent molecular mass (180-220 kDa based on experiments) is compatible with a glycosylated form of S (1255 residues, nonglycosylated polypeptide chain of 139 kDa), as well as lighter polypeptides, which probably represent truncated and/or nonglycosylated forms of S.

In conclusion, all these experiments demonstrate that the recombinant polypeptides expressed in *E. coli* and corresponding to the SARS-CoV N and S proteins make it possible to induce, in animals, polyclonal antibodies capable of recognizing the native forms of these proteins.

Preparation of Monospecific Polyclonal Antibodies
Directed Against the SARS-Associated Coronavirus
(SARS-CoV) M and E Proteins

1) Analysis of the Structure of the M and E Proteins

a) E Protein

The structure of the SARS-CoV E protein (76 amino acids) was analyzed in silico, with the aid of various software packages such as signalP v1.1, NetNGlyc 1.0, THMM 1.0 and 2.0 (Krogh et al., 2001, J. Mol. Biol., 305(3):567-580) or alternatively TOPPED (von Heijne, 1992, J. Mol. Biol. 225, 487-494). The analysis shows that this nonglycosylated polypeptide is a type 1 membrane protein, containing a single transmembrane helix (aa 12-34 according to THMM), and in which the majority of the hydrophilic domain (42 residues) is located at the C-terminal end and probably inside the viral particle (endodomain). It is possible to note an inversion in the topology predicted by versions 1.0 (N-ter is external) and 2.0 (N-ter is internal) of the THMM software, but that other algorithms, in particular TOPPED and THUMBUP (Zhou et Zhou, 2003, Protein Science 12:1547-1555) confirm an external location of the N-terminal end of E.

b) M Protein

A similar analysis carried out on the SARS-CoV M protein (221 amino acids) shows that this polypeptide does not possess a signal peptide (according to the software signalP v1.1) but three transmembrane domains (residues 15-37, 50-72, 77-99 according to THMM2.0) and a large hydrophilic domain (aa 100-221) located inside the viral particle (endodomain). It is probably glycosylated on the asparagine at position 4 (according to NetNGlyc 1.0).

Thus, in agreement with the experimental data known for the other coronaviruses, it is remarkable that the two M and E proteins exhibit endodomains corresponding to the majority of the polypeptides and of the ectodomains that are very small in size.

The ectodomain of E probably corresponds to residues 1 to 11 or 1 to 12 of the protein: MYSFVSEETGT(L), SEQ ID NO: 70. Indeed, the probability associated with the transmembrane location of residue 12 is intermediate (0.56 according to THMM 2.0).

The ectodomain of M probably corresponds to residues 2 to 14 of the protein: ADNGTITVEELKQ, SEQ ID NO: 69. Indeed, the N-terminal methionine of M is very probably cleaved from the mature polypeptide because the residue at position 2 is an alanine (Varshaysky, 1996, 93:12142-12149).

Moreover, the analysis of the hydrophobicity (Kyte & Doolittle Hopp & Woods) of the E protein demonstrates that the C-terminal end of the endodomain of E is hydrophilic and therefore probably exposed at the surface of this domain. Thus, a synthetic peptide corresponding to this end is a good immunogenic candidate for inducing, in animals, antibodies directed against the endodomain of E. Consequently, a peptide corresponding to 24 C-terminal residues of E was synthesized.

2) Preparation of Antibodies Directed Against the Ectodomain of the M and E Proteins and the Endodomain of the E Protein

The peptides M2-14 (ADNGTITVEELKQ, SEQ. ID NO: 69), E1-12 (MYSFVSEETGTL, SEQ ID NO: 70) and E53-76 (KPTVYVYSRV KNLNSSEGVP DLLV, SEQ ID NO: 71) were synthesized by Neosystem. They were coupled with KLH (Keyhole Limpet Hemocyanin) with the aid of MBS (m-maleimido-benzoyl-N-hydroxysuccinimide ester) via a

cysteine added during the synthesis either at the N-terminus of the peptide (case for E53-76) or at the C-terminus (case of M2-14 and E1-12).

Two rabbits were immunized with each of the conjugates, according to the following immunization protocol: after a first injection of 0.5 mg of peptide coupled with KLH and emulsified in complete Freund's adjuvant (intradermal route), the animals receive 2 to 4 booster injections at 3 or 4 weeks' interval of 0.25 mg of peptide coupled to KLH and emulsified in incomplete Freund's adjuvant.

For each rabbit, a preimmune (p.i.) serum was prepared before the first immunization and an immune serum (I.S.) is prepared 3 to 5 weeks after the booster injections.

The reactivity of the sera was analyzed by Western blotting with the aid of extracts of cells infected with SARS-CoV (FIG. 43B) or with the aid of extracts of cells infected with a recombinant vaccinia virus expressing the protein E (VV-TG-E, FIG. 43A) or M (VV-TN-M, FIG. 43C) of the SARS-CoV 031589 isolate.

The immune sera of the rabbits 22234 and 22240, immunized with the conjugate KLH-E53-76, recognize a polypeptide of about 9 to 10 kD, which is present in the extracts of cells infected with SARS-CoV but absent from the extracts, of noninfected cells (FIG. 43B). The apparent mass of this polypeptide is compatible with the predicted mass of the E protein, which is 8.4 kD. Similarly, the immune serum of the rabbit 20047, immunized with the conjugate KLH-E1-12, recognizes a polypeptide present in the extracts of cells infected with the VV-TG-E virus, whose apparent molar mass is compatible with that of the E protein (FIG. 43A).

The immune serum of the rabbits 20013 and 20080, immunized with the conjugate KLH-M2-14, recognizes a polypeptide present in the extracts of cells infected with the VV-TN-M virus (FIG. 43C), whose apparent molar mass (about 18 kD) is compatible with that of the glycoprotein M, which is 25.1 kD and has a high iso-electric point (9.1 for the naked polypeptide).

These results demonstrate that the peptides E1-12 and E53-76, on the one hand, and the peptide M2-14, on the other hand, make it possible to induce, in animals, polyclonal antibodies capable of recognizing the native forms of the SARS-CoV E and M proteins, respectively.

EXAMPLE 6

Analysis of the ELISA Reactivity of the Recombinant N Protein Toward Sera from Patients Suffering from SARS

1) Materials

The antigen used to prepare the solid phases is the purified recombinant nucleoprotein N prepared according to the protocol described in example 2.

The sera to be tested (table IV) were chosen on the basis of the results of analysis of their reactivity by immunofluorescence (IF-SARS titer), toward cells infected with SARS-CoV.

TABLE IV

Sera tested by ELISA					
Reference	Serum No.	Type of serum	Date of the serum***	IF-SARS titer	
	3050	A	Control	na*	nt**
	3048	B	Control	na	nt
	033168	D	Patient 1-SARS	Apr. 27, 2003 (D38)	320
	033397	E	Patient-1 SARS	May 11, 2005 (D52)	320

TABLE IV-continued

Sera tested by ELISA					
Reference	Serum No.	Type of serum	Date of the serum***	IF-SARS titer	
	032632	F	Patient-2 SARS	Mar. 21, 2003 (D17)	2500
	032791	G	Patient-3 SARS	Apr. 04, 2003 (D3)	<40
	033258	H	Patient-3 SARS	Apr. 28, 2003 (D27)	160

*na: not applicable.

**nt: not tested.

***the dates indicated correspond to the number of days after the onset of the SARS symptoms.

2) Method

The N protein (100 µl) diluted at various concentrations in 0.1 M carbonate buffer, pH 9.6 (1, 2 or 4 µg/ml) is distributed into the wells of ELISA plates, and then the plates are incubated overnight at laboratory temperature. The plates are washed with PBS-Tween buffer saturated with PBS-skimmed milk-sucrose (5%) buffer. The test sera (100 µl), diluted beforehand (1/50, 1/100, 1/200, 1/400, 1/800, 1/1600 and 1/3200) are added and then the plates are incubated for 1 h at 37° C. After 3 washings, the peroxidase-labeled anti-human IgG conjugate (reference 209-035-098, JACKSON) diluted 1/18 000 is added and then the plates are incubated for 1 h at 37° C. After 4 washings, the chromogen (TMB) and the substrate (H₂O₂) are added and the plates are incubated for 30 min at room temperature, protected from light. The reaction is then stopped and then the absorbance at 450 nm is measured with the aid of an automated reader.

3) Results

The ELISA tests (FIG. 10) demonstrate that the recombinant N protein preparation is specifically recognized by the antibodies of sera from patients suffering from SARS collected in the late phase of the infection (≥17 days after the onset of the symptoms) whereas it is not significantly recognized by the antibodies of a patient's serum collected in the early phase of the infection (3 days after the onset of the symptoms) or by control sera from subjects not suffering from SARS.

EXAMPLE 7

ELISA Tests Prepared for a Very Specific and Sensitive Detection of a SARS-Associated Coronavirus Infection, from Sera of Patients

1) Indirect ELISA IgG Test

a) Reagents

Preparation of the Plates

The plates are sensitized with a solution of N protein at 2 µg/ml in a 10 mM PBS buffer, pH 7.2, phenol red at 0.25 ml/l. 100 µl of solution are deposited in the wells and left to incubate at room temperature overnight. Saturation is obtained by prewashing in 10 mM PBS/0.1% Tween buffer, followed by washing with a saturation solution PBS, 25% milk/sucrose.

Diluent Sera

Buffer 0.48 g/l TRIS, 10 mM PBS, 3.7 g/l EDTA, 15% v/v milk, pH 6.7

Diluent Conjugate

Citrate buffer (15 g/l), 0.5% Tween, 25% bovine serum, 12% NaCl, 6% v/v skimmed milk pH 6.5

Conjugate

50× anti-human IgG conjugate, marketed by Bio-Rad: Platelia *H. pylori* kit ref 72778

Other Solutions:
Washing solution R2, solutions for visualizing with TMB R8 diluent, R9 chromogen, R10 stopping solution: reagents marketed by Bio-Rad (e.g.: Platelia *pylori* kit, ref 72778)

b) Procedure

Dilute the sera 1/200 in the sample diluent
Distribute 100 μ l/well
Incubation 1 h at 37° C.
3 washings in 10 \times WASHING solution R2 diluted beforehand 10-fold in demineralized water (i.e., 1 \times washing solution)

Distribute 100 μ l of conjugate (50 \times conjugate to be diluted immediately before use in the diluent conjugate provided)
Incubation 1 h at 37° C.

4 washings in 1 \times washing solution
Distribute 200 μ l/well of visualization solution (to be diluted immediately before use e.g.: 1 ml of R9 in 10 ml of R8)

Incubation for 30 min at room temperature in the dark
Stop the reaction with 100 μ l/well of R10
READING at 450/620 nm

The results can be interpreted by taking a THRESHOLD serum giving a response above which the sera tested would be considered as positive. This serum is chosen and diluted so as to give a significantly higher signal than the background noise.

2) Double Epitope Elisa Test

Reagents

Preparation of the Plates

The plates are sensitized with a solution of N protein at 1 μ g/ml in a 10 mM PBS buffer, pH 7.2, phenol red at 0.25 ml/l. 100 μ l of solution are deposited in the wells and left to incubate at room temperature overnight. Saturation is obtained by prewashing in 10 mM PBS/0.1% Tween buffer, followed by washing with a saturation solution 10 mM PBS, 25% (V/V) milk.

Diluent sera and Conjugate

Buffer 50 mM TRIS saline, pH 8, 2% milk

Conjugate

This is the purified recombinant N protein coupled with peroxidase according to the Nakane protocol (Nakane P. K. and Kawaoi A.; (1974): *Peroxydase-labeled antibody, a new method of conjugation. The Journal of Histochemistry and Cytochemistry* Vol. 22, N) 23, pp. 1084-1091), in respective molar ratios 1/2. This ProtN POD conjugate is used at a concentration of 2 μ g/ml in serum/conjugate diluent.

Other Solutions:

Washing solution R2, solutions for visualization with TMB R8, diluent, R9 chromogen, R10 stopping solution: reagents marketed by Bio-Rad (e.g. Platelia *pylori* kit, ref 72778).

b) Procedure

1st step in "predilution" plate

Dilute each serum 1/5 in the predilution plate (48 μ l of diluent+12 μ l of serum).

After having diluted all the sera, distribute 60 μ l of conjugate.

Where appropriate, the serum+conjugate mix is left to incubate.

2nd step in "reaction" plate

Transfer 100 μ l of mixture/well into the reaction plate

Incubation 1 h 37° C.

5 washings in 10 \times WASHING solution R2 diluted 10-fold beforehand in demineralized water (\rightarrow 1 \times washing solution)

Distribute 200 μ l/well of visualization solution (to be diluted immediately before use e.g.: 1 ml of R9 in 10 ml of R8)

Incubation 30 min at room temperature and protected from light

Stop the reaction with 100 μ l/well of R10

READING at 450/620 nm

Likewise as for the indirect ELISA test, the results can be interpreted using a "threshold value" serum. Any serum having a response greater than the threshold value serum will be considered as positive.

2) Results

The sera of patients classified as probable cases of SARS from the French hospital of Hanoi, Vietnam or in relation with the French hospital of Hanoi (JYK) were analyzed using the indirect IgG-N test and the double epitope N test.

The results of the indirect IgG-N test (FIGS. 14 and 15) and double epitope N test (FIGS. 16 and 17) show an excellent correlation between them and with an indirect ELISA test comparing the reactivity of the sera toward a lysate of VeroE6 cells infected or not infected with SARS-CoV (ELISA-SARS-CoV lysate; see table V below). All the sera collected 12 days or more after the onset of the symptoms were found to be positive, including in patients for whom it had not been possible to document the SARS-CoV virus infection by analyzing respiratory samples by RT-PCR, probably because of a sample being collected too late during the infection (\cong D12). In the case of the patient TTH for whom a nasal sample collected on D7 was found to be negative by RT-PCR, the quality of the sample may be in question.

Some sera were found to be negative whereas the presence of SARS-CoV was detected by RT-PCR. They are in all cases early sera collected less than 10 days after the onset of the symptoms (e.g.: serum #032637). In the case of a patient PTTH (serum #032673), only a suspicion of SARS was raised at the time the samples were collected.

In conclusion, the indirect IgG-N and N-double epitope serological tests make it possible to document the SARS-CoV infection in all the patients for the sera collected 12 days or more after the infection.

TABLE V

Results of the ELISA tests						
Sample Num	Patient	Day	PCR-SARS (1)	ELISA SARS-CoV lysate (2)	IgG-N (2nd series)	2Xepitope (2nd series)
033168	JYK	38	POS	+++	>5000	NT
033597	JK	74	POS	NT	\approx 5000	NT
032552	VTT	8	NEG-D3&D8&D12	NEG	<200	<5
032544	CTP	16	NEG-D16&D20	++	>5000	>>20
032546	CJF	15	NEG-D15&D19	++	>5000	>>20

TABLE V-continued

Results of the ELISA tests						
Sample Num	Patient	Day	PCR-SARS (1)	ELISA SARS-CoV lysate (2)	IgG-N (2nd series)	2Xepitope (2nd series)
032548	PTL	17	NEG D17&D21	++	>5000	>>20
032550	NTH	17	NEG- D17&D21	++	>5000	>>20
032553	VTT	8	NEG- D3&D8&D12	NEG	<200	<5
032554	NTBV	4	POS	NEG	<200	<5
032555	NTBV	4	POS	NEG	<200	<5
032564	NTP	15	POS	++	>5000	>>20
032629	NVH	4	POS	NEG	<200	<5
032631	BTTX	9	POS	NEG	<200	<5
032635	NHH	4	POS	NEG	<200	<5
032637	NHB	10	POS	NEG	<200	<5
032642	BTTX	9	POS	NEG	<200	<5
032643	LTDH	1	POS	NEG	<200	<5
032644	NTBV	4	POS	NEG	<200	<5
032646	TTH	12	NEG D7&D12&D16	++	>5000	>>20
032647	DTH	17	NEG D17&D21	++	>5000	>>20
032648	NNT	15	NEG D15&D19	++	>5000	>>20
032649	PTH	17	NEG D17&D21	++	>5000	>>20
032672	LVV	16	NEG D16&D20	+	>5000	>>20
032673	PTTH	NA	NEG	NEG	<200	<5
032674	PNB	17	NEG D17&D21	++	>5000	>>20
032682	VTH	12	NEG D12&D16	++	>5000	>>20
032683	DTV	17	NEG D17&D21	+	>1000	>>20

Remarks:

(1): The RT-PCR analyses were carried out by nested RT-PCR BNL, LC Artus and LC-N on nasal or pharyngeal swabs; POS means that at least one sample was found to be positive in this patient.

(2): The reactivity of the sera in the ELISA test using a lysate of cells infected with SARS-CoV was classified as very highly reactive (+++), highly reactive (++), reactive (+) and negative according to the OD value obtained at the dilutions tested.

EXAMPLE 8

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-continued

Detection of SARS-Associated Coronavirus
(SARS-CoV) by RT-PCR

1) Real Time Development of RT-PCR Conditions with the Aid of Primers Specific for the Gene for the Nucleocapsid Protein—"Light Cycler N" Test

a) Design of the Primers and Probes

The primers and probes were designed from the sequence of the genome of the SARS-CoV strain derived from the sample recorded under the number 031589, with the aid of the programme "Light Cycler Probe Design (Roche)". Thus, the following two series of primers and probes were selected:

series 1 (SEQ ID NO: 60, 61, 64, 65):

sense primer: N+/28507:
5'-GGC ATC GTA TGG GTT G-3' [28507-28522]

antisense primer: N-/28774:
5'-CAG TTT CAC CAC CTC C-3' [28774-28759]

probe 1:
5'-GGC ACC CGC AAT CCT AAT AAC AAT GC-fluorescein
3' [28561-28586]

probe 2:

5' Red705-GCC ACC GTG CTA CAA CTT CCT-phosphate

45 [28588-28608]

series 2 (SEQ ID NO: 62, 63, 66, 67)

50 sense primer: N+/28375:
5'-GGC TAC TAC CGA AGA G-3' [28375-28390]

antisense primer: N-/28702:
5'-AAT TAC CGC GAC TAC G-3' [28702-28687]

55 probe 1: SARS/N/FL:
5'-ATA CAC CCA AAG ACC ACA TTG GC-fluorescein 3'
[28541-28563]

probe 2: SARS/N/LC705:
60 5' Red705-CCC GCA ATC CTA ATA ACA ATG CTG C-
phosphate 3' [28565-28589]

b) Analysis of the Efficacy of the Two Primer Pairs

65 In order to test the respective efficacy of the two pairs of primers, an RT-PCR amplification was carried out on a synthetic RNA corresponding to nucleotides 28054-29430 of the

genome of the SARS-CoV strain derived from the sample recorded under the number 031589 and containing the sequence of the N gene.

More specifically:

This synthetic RNA was prepared by in vitro transcription with the aid of the T7 phage RNA polymerase, of a DNA template obtained by linearization of the plasmid SRAS-N with the enzyme Bam HI. After eliminating the DNA template by digestion with the aid of DNase I, the synthetic RNAs are purified by a phenol-chloroform extraction, followed by two successive precipitations in ammonium acetate and isopropanol. They are then quantified by measuring the absorbance at 260 nm and their quality is checked by the ratio of the absorbances at 260 and 280 nm and by agarose gel electrophoresis. Thus, the concentration of the synthetic RNA preparation used for these studies is 1.6 mg/ml, which corresponds to 2.1×10^{15} copies/ml of RNA.

Decreasing quantities of synthetic RNA were amplified by RT-PCR with the aid of the "Superscript™ One-Step RT-PCR with Platinum® Taq" kit and the pairs of primers No. 1 (N+/28507, N-/28774) (FIG. 1A) and No. 2 (N+/28375, N-/28702) (FIG. 1B), according to the supplier's instructions. The amplification conditions used are the following: the cDNA was synthesized by incubation for 30 min at 45° C., 15 min at 55° C. and then 2 min at 94° C. and it was then amplified by 5 cycles comprising: a step of denaturation at 94° C. for 15 sec, a step of annealing at 45° C. for 30 sec and, then a step of extension at 72° C. for 30 sec, followed by 35 cycles comprising: a step of denaturation at 94° C. for 15 sec, a step of annealing at 55° C. for 30 sec and then a step of extension at 72° C. for 30 sec, with 2 sec of additional extension at each cycle, and a final step of extension at 72° C. for 5 min. The amplification products obtained were then kept at 10° C.

The results presented in FIG. 11 show that the pair of primers No. 2 (N+/28375, N-/28702) makes it possible to detect up to 10 copies of RNA (band of weak intensity) or 10^2 copies (band of good intensity) against 10^4 copies for the pair of primers No. 1 (N+/28507, N-/28774). The amplicons are respectively 268 bp (pair 1) and 328 bp (pair 2).

c) Development of Real Time RT-PCR

A real time RT-PCR was developed with the aid of the pair of primers No. 2 and of the pair of probes consisting of SRAS/N/FL and SRAS/N/LC705 (FIG. 2).

The amplification was carried out on a LightCycler™ (Roche) with the aid of the "Light Cycler RNA Amplification Kit Hybridization Probes" kit (reference 2 015 145, Roche) under the following optimized conditions. A reaction mixture containing: H₂O (6.8 µl), 25 mM MgCl₂ (0.8 µl, 4 µM Mg²⁺ final), 5× reaction mixture (4 µl), 3 µM probe SRAS/N/FL (0.5 µl, 0.075 µM final), 3 µM probe SRAS/N/LC705 (0.5 µl, 0.075 µM final), 10 µM primer N+/28375 (1 µl, 0.5 µM final), 10 µM primer N-/28702 (1 µl, 0.5 µM final), enzyme mixture (0.4 µl) and sample (viral RNA, 5 µl) was amplified according to the following program:

Reverse transcription: 50° C. 10:00 min analysis mode: none

Denaturation: 95° C. 30 sec×1 analysis mode: none

Amplification: 95° C. 2 sec

50° C. 15 sec analysis mode: quantification*{×45

72° C. 13 sec thermal ramp 2.0° C./sec}

* The fluorescence is measured at the end of the annealing and at each cycle (in SINGLE mode).

Annealing: 40° C. 30 sec×1 analysis mode: none

The results presented in FIG. 12 show that this real time RT-PCR is very sensitive since it makes it possible to detect 10^2 copies of synthetic RNA in 100% of the 5 samples ana-

lyzed (29/29 samples in 8 experiments) and up to 10 copies of RNA in 100% of the 5 samples analyzed (40/45 samples in 8 experiments). It also shows that this RT-PCR makes it possible to detect the presence of the SARS-CoV genome in a sample and to quantify the number of genomes present. By way of example, the viral RNA of a SARS-CoV stock cultured on Vero E6 cells was extracted with the aid of the "Qiaamp viral RNA extraction" kit (Qiagen), diluted to 0.05×10^{-14} and analyzed by real time RT-PCR according to the protocol described above; the analysis presented in FIG. 12 shows that this virus stock contains 6.5×10^9 genome-equivalents/ml (geq/ml), which is entirely similar to the 1.0×10^{10} geq/ml value measured with the aid of the "RealArt™ HPA-Coronavirus LC RT PCR Reagents" kit marketed by Artus.

2) Development of Nested RT-PCR Conditions Targeting the Gene for RNA Polymerase—"CDC (Centers for Disease Control and Prevention)/IP Nested RT-PCR" Test

a) Extraction of the Viral RNA

Clinical sample: QIAamp viral RNA Mini Kit (QIAGEN) according to the manufacturer's instructions, or an equivalent technique. The RNA is eluted in a volume of 60 µl.

b) "SNE/SAR" Nested RT-PCR

First step: "SNE" coupled RT-PCR

The Invitrogen "Superscript™ One-Step RT-PCR with Platinum® Taq" kit was used, but the "Titan" kit from Roche Boehringer can be used in its place with similar results.

Oligonucleotides:

SNE-S1 5' GGT TGG GAT TAT CCA AAA TGT GA 3'

SNE-AS1 5' GCA TCA TCA GAA AGA ATC ATC ATG 3'

→Expected size: 440 bp

1. Prepare a mix:

H ₂ O	6.5 µl
Reaction mix 2X	12.5 µl
Oligo SNE-S1 50 µM	0.2 µl
Oligo SNE-AS1 50 µM	0.2 µl
RNAsin 40 U/µl	0.12 µl
RT/Platinum Taq mix	0.5 µl

2. To 20 µl of the mix, add 5 µl of RNA and carry out the amplification on a thermocycler (ABI 9600 conditions):

2.1	45° C.	30 min.	
	55° C.	15 min.	
	94° C.	2 min.	
2.2.	94° C.	15 sec.	} × 5 cycles
	45° C.	30 sec.	
	72° C.	30 sec.	
2.3.	94° C.	15 sec.	} × 35 cycles
	55° C.	30 sec.	
	72° C.	30 sec. + 2 sec./cycle	
2.4.	72° C.	5 min.	
2.5	10° C.	∞	

Storage at 44° C.

The RNAsin (N2511/N2515) from Promega was used as RNase inhibitors.

Synthetic RNAs served as positive control. As the control, 10^3 , 10^2 and 10 copies of synthetic RNA R_{SNE} were amplified in each experiment.

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Second step: "SAR" nested PCR
Oligonucleotides:

SAR1-S 5' CCT CTC TTG TTC TTG CTC GCA 3' 5
SAR1-AS 5' TAT AGT GAG CCG CCA CAC ATG 3'

→Expected size: 121 bp

1. Prepare a mix:

H2O	35.8 µl
Taq buffer 10X	5 µl
MgCl ₂ 25 mM	4 µl
Mix dNTPs 5 mM	2 µl
Oligo SAR1-S 50 µM	0.5 µl
Oligo SAR1-AS 50 µM	0.5 µl
Taq DNA pol 5 U/µl	0.25 µl

AmpliTaq DNA Pot from Applied Biosystems was used (10x buffer without MgCl₂, ref 27216601).

2. To 48 µl of the mix, add 2 µl of the product from the first PCR and carry out the amplification (ABI 9600 conditions):

2.1.	94° C.	2 min.	} × 5 cycles
2.2.	94° C.	30 sec.	
	45° C.	45 sec.	
	72° C.	30 sec.	} × 35 cycles
2.3.	94° C.	30 sec.	
	55° C.	30 sec.	
	72° C.	30 sec. + 1 sec./cycle	}
2.4.	72° C.	5 min.	
2.5	10° C.	∞ min.	

3. Analyze 10 µl of the reaction product on "low-melting" gel (Seakem GTG type) containing 3% agarose.

The sensitivity of the nested test is routinely, under the conditions described, 10 copies of RNA.

4. The fragments can then be purified on QIAquick PCR kit (QIAGEN) and sequenced with the oligos SAR1-S and SAR1-AS.

3) Detection of the SARS-CoV RNA by PCR from Respiratory Samples

a) First Comparative Study

A comparative study was carried out on a series of respiratory samples received by the National Reference Center for the Influenza Virus (Northern region) and likely to contain SARS-CoV. To do this, the RNA was extracted from the samples with the aid of the "Qiam viral RNA extraction" kit (Qiagen) and analyzed by real time RT-PCR, on the one hand with the aid of the pairs of primers and probes of the No. 2 series under the conditions described above on the one hand, and on the other hand with the aid of the kit "LightCycler SARS-CoV quantification kit" marketed by Roche (reference 03 604 438). The results are summarized in table VI below. They show that 18 of the 26 samples are negative and 5 of the 26 samples are positive for the two kits, while one sample is positive for the Roche kit alone and two for the "series 2" N reagents alone. Additionally, for 3 samples (20032701, 20032712, 20032714) the quantities of RNA detected are markedly higher with the reagents (probes and primers) of the No. 2 series. These results indicate that the "series 2" N primers and probes are more sensitive for the detection of the SARS-CoV genome in biological samples than those of the kit currently available.

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TABLE VI

Real time RT-PCR analysis of the RNAs extracted from a series of samples from 5 patients with the aid of the pairs of primers and probes of the No. 2 series ("series 2" N) or of the kit "LightCycler SARS-CoV quantification kit" (Roche). The type of sample is indicated as well as the number of copies of viral genome measured in each of the two tests. NEG: negative RT-PCR.

10	Sample No.	Patient	Type of sample	ROCHE KIT	"Series 2" N
	20033082	K	nasal	NEG	NEG
	20033083	K	pharyngeal	NEG	NEG
	20033086	K	nasal	NEG	NEG
	20033087	K	pharyngeal	NEG	NEG
	20032802	M	nasal	NEG	NEG
	20032803	M	expectoration	NEG	NEG
	20032806	M	nasal or pharyngeal	NEG	NEG
	20031746ARN2	C	pharyngeal	NEG	NEG
	20032711	C	nasal or pharyngeal	39	NEG
	20032910	B	nasal	NEG	NEG
	20032911	B	pharyngeal	NEG	NEG
	20033356	V	expectoration	NEG	NEG
	20033357	V	expectoration	NEG	NEG
	20031725	K	endotracheal asp.	NEG	150
	20032657	K	endotracheal asp.	NEG	NEG
	20032698	K	endotracheal asp.	NEG	NEG
	20032720	K	endotracheal asp.	3	5
	20033074	K	stools	115	257
	20032701	M	pharyngeal	443	1676
	20032702	M	expectoration	NEG	249
	20031747ARN2	C	pharyngeal	NEG	NEG
	20032712	C	unknown	634	6914
	20032714	C	pharyngeal	17	223
	20032800	B	nasal	NEG	NEG
	20033353	V	nasal	NEG	NEG
	20033384	V	nasal	NEG	NEG

b) Second Comparative Study

The performance of various nested RT-PCR and real time RT-PCR methods were then compared for 121 respiratory samples from possible cases of SARS at the French hospital in Hanoi, Vietnam, taken between the 4th and the 17th day after the onset of the symptoms. Among these samples, 14 were found to be positive during a first test using the nested RT-PCR method targeting ORF1b (encoding replicase) as described initially by Bernhard Nocht Institute (BNI nested RT-PCR). Information relating to this test is available on the internet, at the address www.15.bni-hamburg.de/bni2/neu2/getfile.acgi?area_engl=diagnostics&pid=4112.

The various tests compared in this study are:

the quantitative RT-PCR method according to the invention, with the "series 2" N primers and probes described above (LightCycler N column),

the nested RT-PCR test targeting the RNA polymerase gene described above, developed by the CDC, BNI and Institut Pasteur (CDC/IP nested RT-PCR),

the ARTUS kit with the reference "HPA Corona LC RT-PCR Kit #5601-02", which is a real time RT-PCR test targeting the ORF1b gene,

the BNI nested RT-PCR test, also targeting the RNA polymerase gene mentioned above.

The inventors observed:

1) an inter-test variability for the same technique, linked to the degradation of the RNA preparation during repeated thawing, in particular for the samples containing the lowest quantities of RNA,

2) a reduced sensitivity of the CDC/IP nested RT-PCR compared with the BNI nested RT-PCR, and

3) a comparable sensitivity of the quantitative RT-PCR test according to the invention (LightCycler N) compared with the Artus LightCycler (LC) test.

These results, which are presented in table VII below, show that the quantitative RT-PCR test according to the invention constitutes an excellent addition—or an alternative—to the tests currently available. Indeed, the SARS-linked coronavirus is an emergent virus which is capable of changing rapidly. In particular, the gene for the RNA polymerase of the SARS-linked coronavirus, which is targeted in most of the tests currently available, can recombine with that of other coronaviruses not linked to SARS. The use of a test targeting this gene exclusively could then lead to the production of false-negatives.

The quantitative RT-PCR test according to the invention does not target the same genomic region as the ARTUS kit since it targets the gene encoding the N protein. By carrying out a diagnostic test targeting two different genes of the SARS-linked coronavirus, it can therefore be hoped to avoid false-negative type results which could be due to the genetic evolution of the virus.

Furthermore, it appears particularly advantageous to target the gene for the nucleocapsid protein because it is very stable because of the high selection pressure linked to the high structural constraints regarding this protein.

TABLE VII

Comparison of various methods of analysis by gene amplification, from 121 samples of probable cases of SARS at the French hospital in Hanoi, Vietnam (epidemic 2003)

NRC No.	Sample type (1)	Sample collection day	Patient	CDC/IP nested RT-PCR	BNI nested RT-PCR	Artus Light Cyclor kit	Light Cyclor N (IP)
107	N and P			Negative	Negative	Negative	Negative
samples							
032529	P	10	NHB	Negative	Positive	Negative	Negative
032530	N	10	NHB	Positive	Positive	3.10E+01	4.20E+01
032531	P	7	LP	Positive	Positive	7.70E+00	3.10E+00
032534	N	15	BND	Positive	Positive	1.60E+00	Negative
032600	P	4	NHH	Negative	Positive	Negative	1.30E+02
032612	P	17	NTS	Negative	Positive	Negative	Negative
032688	P	9	BTX	Positive	Positive	Negative	Negative
032689	N	4	NVH	Positive	Positive	1.20E+01	2.30E+02
032690	P	4	NVH	Negative	Positive	1.60E+00	Negative
032727	P	8	NVH	Positive	Positive	2.30E+02	4.00E+02
032728	N	8	NVH	Positive	Positive	1.10E+03	1.60E+04
032729	P	14	NHB	Positive	Positive	5.90E+00	3.40E+01
032730	N	14	NHB	Positive	Positive	1.30E+02	4.80E+02
032741	P	8	NHH	Positive	Positive	2.10E+02	1.30E+02
positives				10	14	10	9
fraction detected from the 14 positives				71.4%	100.0%	71.4%	64.3%

(1) P = pharyngeal swab
N = nasal swab

EXAMPLE 9

Production and Characterization of Monoclonal Antibodies Directed Against the N Protein

Balb C mice were immunized with the purified recombinant N protein and their spleen cells fused with an appropriate murine myeloma according to the Köhler and Milstein techniques.

Nineteen anti-N antibody secreting hybridomas were pre-selected and their immunoreactivities determined. These

antibodies do indeed recognize the recombinant N protein (in ELISA) with variable intensities, and the natural viral N protein in ELISA and/or in Western blotting. FIGS. 18 to 20 show the results of these tests for 15 of these 19 monoclonal antibodies.

The highly reactive clones 12, 17, 28, 57, 72, 76, 86, 87, 98, 103, 146, 156, 166, 170, 199, 212, 218, 219 and 222 were subcloned. Specificity studies were carried out with the appropriate tools in order to determine the epitopes recognized and verify the absence of reactivity toward other human coronaviruses and certain respiratory viruses.

Epitope mapping studies (performed on spot membrane with the aid of overlapping peptides of 15 aa) and additional studies performed on the natural N protein in Western blotting revealed the existence of 4 groups of monoclonal antibodies:

1. Monoclonal antibodies specific for a major linear epitope at the N-ter position (75-81, sequence: INTNSVP).

The representative of this group is antibody 156. The hybridoma producing this antibody was deposited at the Collection Nationale de Cultures de Microorganismes (CNCM) of the Institut Pasteur (Paris, France) on Dec. 1, 2004, under the number I-3331. This same epitope is also recognized by a rabbit serum (anti-N polyclonal) obtained by conventional immunization with the aid of this same N protein.

2. Monoclonal antibodies specific for a major linear epitope located in a central position (position 217-224, sequence: ETALALL); the representatives of this group are

the monoclonal antibodies 87 and 166. The hybridoma producing antibody 87 was deposited at the CNCM on Dec. 1, 2004, under the number I-3328.

3. Monoclonal antibodies specific for a major linear epitope located at the C-terminal position (position 403-408, sequence: DFFRQL), the representatives of this group are the antibodies 28, 57 and 143. The hybridoma producing antibody 57 was deposited at the CNCM on Dec. 1, 2004, under the number I-3330.

4. Monoclonal antibodies specific for a discontinuous conformational epitope. This group of antibodies does not rec-

ognize any of the peptides spanning the sequence of the N protein, but react strongly on the non-denatured natural protein. The representative of this final group is the antibody 86. The hybridoma producing this antibody was deposited at the CNCM on Dec. 1, 2004, under the number I-3329.

Table VIII below summarizes the epitope mapping results obtained:

TABLE VIII

Epitope mapping of the monoclonal antibodies			
Antibody	Epitope	Position	Region
28	DFSRQL Q	403 . . . 408	C - Ter.
143	DFSRQL Q		
76	DFSRQL Q		
57	DFSRQL Q		
	FFGMS RI	315 . . . 319	
146	LPQRQ	383 . . . 387	
166	ETALALLLL	217 . . . 224	central
87	ETALALL	217 . . . 224	
156	INTNSGP	75 . . . 81	N-Ter.
86	Conformational		
212	Conformational		
1170	Conformational		

In addition, as illustrated in particular in FIGS. 18 and 19, these antibodies exhibit no reactivity in ELISA and/or in WB toward the N protein of the human corona-virus 229 E.

EXAMPLE 10

Combinations of the Monoclonal Antibodies for the Development of a Sensitive Immunocapture Test Specific for the Viral N Antigen in the Serum or Biological Fluids of Patients Infected with the SARS-CoV Virus

The antibodies listed below were selected because of their very specific properties for an additional capture and detection study of the viral N protein, in the serum of the subjects or patients.

These antibodies were produced in ascites on mice, purified by affinity chromatography and used alone or in combination, as capture antibodies and as signal antibodies.

List of the antibodies selected:

Ab anti-C-ter region (No. 28, 57, 143)

Ab anti-central region (No. 87, 166)

Ab anti-N-ter region (No. 156)

Ab anti-discontinuous conformational epitope (86)

1) Preparation of the Reagents:

a) Immunocapture ELISA Plates

The plates are sensitized with the antibody solutions at 5 µg/ml in 0.1 M carbonate buffer, pH 9.6. The (monovalent or plurivalent) solutions are deposited in a volume of 100 µl in the wells and incubated overnight at room temperature. These plates are then washed with PBS buffer (10 mM pH 7.4 supplemented with 0.1% Tween 20) and then saturated with a PBS solution supplemented with 0.3% BSA and 5% sucrose).

The plates are then dried and then packaged in a bag in the presence of a desiccant. They are ready to use.

b) Conjugates

The purified antibodies were coupled with peroxidase according to the Nakane protocol (Nakane et al.—1974, J. of Histo and cytochemistry, vol. 22, pp. 1084-1091) in a ratio of one molecule of IgG per 3 molecules of peroxidase. These conjugates were purified by exclusion chromatography and stored concentrated (concentration between 1 and 2 mg/ml) in the presence of 50% glycerol and at -20° C. They are diluted for their use in the assays at the final concentration of 1 or 2 µg/ml in PBS buffer (pH 7.4) supplemented with 1% BSA.

c) Other Reagents

Human sera negative for all the serum markers for the HIV, HBV, HCV and THLV viruses

Pool of negative human sera supplemented with 0.5% Triton X 100

Inactivated viral Ag: viral culture supernatant inactivated by irradiation and inactivation verified after placing in culture on sensitive cells—titer of the suspension before inactivation about 10⁷ infectious particles per ml or alternatively about 5×10⁹ physical viral particles per ml of antigen

The Ag samples diluted in negative human serum: these samples were prepared by diluting 1:100 and then by 5-fold serial dilution.

These noninfectious samples mimic human samples thought to contain low to very low concentrations of viral nucleoprotein N. Such samples are not available for routine work.

Washing solution R2, solution for visualization TMB R8, chromogen R9 and stop solution R10, are the generic reagents marketed by Bio-Rad in its ELISA kits (e.g.: *Platelia pylori* kit ref. 72778).

2) Procedure

The samples of human sera overloaded with inactivated viral Ag are distributed in an amount of 100 µl per well, directly in the ready-to-use sensitized plates, and then incubated for 1 hour at 37° C. (Bio-Rad IPS incubation).

The material not bound to the solid phase is removed by 3 washings (washing with dilute R2 solution, automatic LP 35 washer).

The appropriate conjugates, diluted to the final concentration of 1 or 2 µg/ml, are distributed in an amount of 100 µl per well and the plates are again incubated for one hour at 37° C. (IPS incubation).

The excess conjugate is removed by 4 successive washings (dilute R2 solution—LP 35 washer).

The presence of conjugate attached to the plates is visualized after adding 100 µl of visualization solution prepared before use (1 ml of R9 and 10 ml of R8) and after incubation for 30 minutes, at room temperature and protected from light.

The enzymatic reaction is finally blocked by adding 100 µl of R10 reagent (1 N H₂SO₄) to all the wells.

The reading is carried out with the aid of an appropriate microplate reader at double wavelength (450/620 nm).

The results can be interpreted by using, as provisional threshold value, the mean of at least two negative controls multiplied by a factor of 2 or alternatively the mean of 100 negative sera supplemented with an increment corresponding to 6 SD (standard deviation calculated on the 100 individual measurements).

3) Results

Various capture antibody and signal antibody combinations were tested based on the properties of the antibodies

selected, and avoiding the combinations of antibodies specific for the same epitopes in solid phase and as conjugates.

The best results were obtained with the 4 combinations listed below. These results are reproduced in table IX below.

1. Combination F/28

Solid phase (Ab 166+87 central region): conjugate antibody 28 (C-ter)

2. Combination G/28

Solid phase (Ab 86—conformational epitope): conjugate antibody 28 (C-ter)

3. Combination H/28

Solid phase (Ab 86, 166 and 87 central region and conformational epitope): conjugate antibody 28 C-ter)

4. Combination H/28+87

Solid phase (Ab 86, 166 and 87 central region and conformational epitope): mixed conjugate antibodies 28 (C-ter) and 87 (central)

5. Combination G/87

Solid phase (Ab 86—conformational epitope): conjugate antibody 87 (central region)

The first 4 combinations exhibit equivalent and reproduced performance levels, greater than the other combinations used such as for example the combination G/87). Of course, in these combinations, a monoclonal antibody may be replaced with another antibody recognizing the same epitope. Thus, the following variants may be mentioned:

6. Variant of the combination F/28

Solid phase (Ab 87 only): conjugate antibody 57 (C-ter)

7. Variant of the combination G/28

Solid phase (Ab 86—conformational epitope): conjugate antibody 57 (C-ter)

8. Variant of the combination H/28

Solid phase (Ab 86 and 87 central region and conformational epitope): conjugate antibody 57 (C-ter)

9. Variant of the combination H/28+87

Solid phase (Ab 86 and 87 central region and conformational epitope): mixed conjugate antibodies 57 (C-ter) and 87 (central)

TABLE IX

Test of immunoreactivity of the anti-SARS-CoV nucleoprotein Abs: optical densities measured with each combination of antibodies according to the dilutions of the inactivated viral antigen.						
No.	Dilution	F/28	G/28	G/87	H/28	H/28 + 87
0	1/100	5	5	3.495	3.900	5
1	1/500	3.795	3.814	1.379	3.702	3.804
2	1/2 500	2.815	2.950	0.275	3.268	2.680
3	1/12 500	0.987	1.038	0.135	1.374	0.865
4	1/62 500	0.404	0.348	0.125	0.480	0.328
5	1/312 500	0.285	0.211	0.123	0.240	0.215
6	Control	0.210	0.200	0.098	0.186	0.156
7	Control	0.269	0.153	0.104	0.193	0.202

The detection limit for these 4 experimental trials corresponds to the antigen dilution in negative serum 1:62 500. A rapid extrapolation suggests the detection of less than 10^3 infectious particles per ml of sera.

From this study, it is evident that the most appropriate antibodies for the capture of the native viral nucleoprotein are the antibodies specific for the central region and/or for a conformational epitope, both being antibodies also selected for their high affinity for the native antigen.

Having determined the best antibodies for the composition of the solid phase, the antibodies to be selected as a priority for the detection of the antigens attached to the solid phase are the complementary antibodies specific for a dominant epitope

in the C-ter region. The use of any other complementary antibody specific for epitopes located in the N-ter region of the protein leads to average or poor results.

EXAMPLE 11

Eukaryotic Expression Systems for the SARS-Associated Coronavirus (SARS-CoV) Spicule (S) Protein

1) Optimization of the Conditions for Expression of the SARS-CoV S in Mammalian Cells

The conditions for transient expression of the SARS-CoV spicule (S) protein were optimized in mammalian cells (293T, VeroE6).

For that, a DNA fragment containing the cDNA for SARS-CoV S was amplified by PCR with the aid of the oligonucleotides 5'-ATAGGATCCA CCATGTTTAT TTTCT-TATTA TTTCTTACTC TCACT-3' and 5'-ATACTCGAGTT ATGTGTAATG TAATTTGACA CCCTTG-3' from the plasmid pSARS-S (C.N.C.M. No. I-3059) and then inserted between the BamHI and XhoI sites of the plasmid pTRIPΔU3-CMV containing a lentiviral vector TRIP (Sirven, 2001, Mol. Ther., 3, 438-448) in order to obtain the plasmid pTRIP-S. The BamHI and XhoI fragment containing the cDNA for S was then subcloned between BamHI and XhoI of the eukaryotic expression plasmid pcDNA3.1(+) (Clontech) in order to obtain the plasmid pcDNA-S. The NheI and XhoI fragment containing the cDNA for S was then subcloned between the corresponding sites of the expression plasmid pCI (Promega) in order to obtain the plasmid pCI-S. The WPRE sequences of the woodchuck hepatitis virus ("Woodchuck Hepatitis Virus posttranscriptional regulatory element") and the CTE sequences ("constitutive transport element") of the simian retro-virus from Mason-Pfizer were inserted into each of the two plasmids pcDNA-S and pCI-S between the XhoI and XbaI sites in order to obtain respectively the plasmids pcDNA-S-CTE, pcDNA-S-WPRE, pCI-S-CTE and pCI-S-WPRE (FIG. 21). The plasmid pCI-S-WPRE was deposited at the CNCM, on Nov. 22, 2004, under the number I-3323. All the inserts were sequenced with the aid of a BigDye Terminator v1.1 kit (Applied Biosystems) and an automated sequencer ABI377.

The capacity of the plasmid constructs to direct the expression of SARS-CoV S in mammalian cells was assessed after transfection of VeroE6 cells (FIG. 22). In this experiment, monolayers of 5×10^5 VeroE6 cells in 35 mm Petri dishes were transfected with 2 μg of plasmids pcDNA as control), pcDNA-S, pCI and pCI-S and 6 μl of Fugene6 reagent according to the manufacturer's instructions (Roche). After 48 hours of incubation at 37° C. and under 5% CO₂, cellular extracts were prepared in loading buffer according to Laemmli, separated on 8% SDS polyacrylamide gel, and then transferred onto a PVDF membrane (BioRad). The detection of this immunoblot (Western blot) was carried out with the aid of an anti-S rabbit polyclonal serum (immune serum from the rabbit P11135; cf. example 4 above) and donkey polyclonal antibodies directed against rabbit IgGs and coupled with peroxidase (NA934V, Amersham). The bound antibodies were visualized by luminescence with the aid of the ECL+ kit (Amersham) and autoradiography films Hyperfilm MP (Amersham).

This experiment (FIG. 22) shows that the plasmid pcDNA-S does not make it possible to direct the expression of SARS-CoV S at detectable levels whereas the plasmid pCI-S allows a weak expression, close to the limit of detection, which may be detected when the film is overexposed. Similar

results were obtained when the expression of S was sought by immunofluorescence (data not shown). This impossibility to detect effective expression of S cannot be attributed to the detection techniques used since the S protein can be detected at the expected size (180 kDa) in an extract of cells infected with SARS-CoV or in an extract of VeroE6 cells infected with the recombinant vaccinia virus VV-TF7.3 and transfected with the plasmid pcDNA-S. In this latter experiment, the virus VV-TF7.3 expresses the RNA polymerase of the T7 phage and allows the cytoplasmic transcription of an uncapped RNA capable of being efficiently translated. This experiment suggests that the expression defects described above are due to an intrinsic inability of the cDNA for S to be efficiently expressed when the step for transcription to messenger RNA is carried out at the nuclear level.

In a second experiment, the effect of the CTE and WPRE signals on the expression of S was assessed after transfection of VeroE6 (FIG. 23A) and 293T (FIG. 23B) cells and according to a protocol similar to that described above. Whereas the expression of S cannot be detected after transfection of the plasmids pcDNA-S-CTE and pcDNA-S-WPRE derived from pcDNA-S, the insertion of the WPRE and CTE signals greatly improves the expression of S in the context of the expression plasmid pCI-S.

To specify this result, a second series of experiments were carried out where the immunoblot is quantitatively visualized by luminescence and acquisition on a digital imaging device (FluorS, BioRad). The analysis of the results obtained with the QuantityOne v4.2.3 software (BioRad) shows that the WPRE and CTE sequences increase respectively the expression of S by a factor of 20 to 42 and 10 to 26 in Vero E6 cells (table X). In 293T cells (table X), the effect of the CTE sequence is more moderate (4 to 5 times) whereas that of the WPRE sequence remains high (13 to 22 times).

TABLE X

Quantitative analysis of the effect of the CTE and WPRE signals on the expression of SARS-CoV S:			
Plasmid	cell	exp. 1	exp. 2
PCI	VeroE6	0.0	0.0
pCI-S	VeroE6	1.0 ± 0.1	1.0
pCI-S-CTE	VeroE6	9.8 ± 0.9	26.4
pCI-S-WPRE	VeroE6	20.1 ± 2.0	42.3
PCI	293T	0.0	0.0
PCI-S	293T	1.0	1.0
PCI-S-CTE	293T	4.6	4.0
PCI-S-WPRE	293T	27.6	12.8

Cellular extracts were prepared 48 hours after transfection of VeroE6 or 293T cells with the plasmid pCI, pCI-S, pCI-S-CTE and pCI-S-WPRE and analyzed by Western blotting as described in the legend to FIG. 22. The Western blot is visualized by luminescence (ECL+, Amersham) and acquisition on a digital imaging device (FluorS, BioRad). The expression levels are indicated according to an arbitrary scale where the value of 1 represents the level measured after transfection of the plasmid pCI-S. Two independent experiments were carried out for each of the two cell types. In experiment 1 on VeroE6 cells, the transfections were carried out in duplicate and the results are indicated in the form of the mean and standard deviation values for the expression levels measured.

In summary, all these results show that the expression, in mammalian cells, of the cDNA for the SARS-CoV S under the control of the RNA polymerase II promoter sequences requires, to be efficient, the expression of a splice signal and of either of the sequences WPRE and CTE.

2) Production of Stable Lines Allowing the Expression of SARS-CoV S

The cDNA for the SARS-CoV S protein was cloned in the form of a BamHI-XhoI fragment into the plasmid pTRIPAU3-CMV containing a defective lentiviral vector TRIP with central DNA flap (Sirven et al., 2001, Mol. Ther., 3: 438-448) in order to obtain the plasmid pTRIP-S (FIG. 24).

Transient cotransfection according to Zennou et al. (2000, Cell, 101: 173-185) of this plasmid, of an encapsidation plasmid (p8.2) and of a plasmid for expression of the VSV envelope glycoprotein, G (pHCMV-G) in 293T cells allowed the preparation of retroviral pseudoparticles containing the vector TRIP-S and pseudotyped with the envelope protein G. These pseudotyped TRIP-S vectors were used to transduce 293T and FRhK-4 cells: no expression of the S protein could be detected by Western blotting and immunofluorescence in the transduced cells (data not presented).

The optimum expression cassettes consisting of the CMV virus immediate/early promoter, a splice signal, cDNA for S and either of the posttranscriptional signals WPRE or CTE described above were then substituted for the EF1 α -EGFP cassette of the defective lentiviral expression vector with central DNA flap TRIPAU3-EF1 α (Sirven et al., 2001, Mol. Ther., 3: 438-448) (FIG. 25). These substitutions were carried out by a series of successive subclonings of the S expression cassettes which were excised from the plasmids pCT-S-CTE (BglII-ApaI) or respectively pCI-S-WPRE (BglII-SalI) and then inserted between the MluI and KpnI sites or respectively MluI or XhoI sites of the plasmid TRIPAU3-EF1 α in order to obtain the plasmids pTRIP-SD/SA-S-CTE and pTRIP-SD/SA-S-WPRE, deposited at the CNM, on Dec. 1, 2004, under the numbers I-3336 and I-3334, respectively. Pseudotyped vectors were produced according to Zennou et al. (2000, Cell, 101: 173-185) and used to transduce 293T cells (10 000 cells) and FRhK-4 cells (15 000 cells) according to a series of 5 successive transduction cycles with a quantity of vectors corresponding to 25 ng (TRIP-SD/SA-S-CTE) or 22 ng TRIP-SD/SA-S-WPRE) of p24 per cycle.

The transduced cells were cloned by limiting dilution and a series of clones were qualitatively analyzed for the expression of SARS-CoV S by immunofluorescence (data not shown), and then quantitatively by Western blotting (FIG. 25) with the aid of an anti-S rabbit polyclonal serum. The results presented in FIG. 25 show that clones 2 and 15 of FRhK4-S-CTE cells transduced with TRIP-SD/SA-S-CTE and clones 4, 9 and 12 of FRhK4-S-WPRE cells transduced with TRIP-SD/SA-S-WPRE allow the expression of the SARS-CoV S at respectively low, or moderate levels if they are compared to those which can be observed during infection with SARS-CoV.

In summary, the vectors TRIP-SD/SA-S-CTE and TRIP-SD/SA-S-WPRE allow the production of stable clones of FRhK-4 cells and similarly 293T cells expressing SARS-CoV S, whereas the assays carried out with the "parent" vector TRIP-S remained unsuccessful, which demonstrates the need for a splice signal and for either of the sequences CTE and WPRE for the production of stable cell clones expressing the S protein.

In addition, these modifications of the vector TRIP (insertion of a splice signal and of a post-transcriptional signal like CTE and WPRE) could prove advantageous for improving the expression of other cDNAs than that for S.

3) Production of Stable Lines Allowing the Expression of a Soluble Form of SARS-CoV S. Purification of this Recombinant Antigen.

A cDNA encoding a soluble form of the S protein (Ssol) was obtained by fusing the sequences encoding the ectodomain of the protein (amino acids 1 to 1193) with those of a tag (FLAG: DYKDDDDK) via a BspEI linker encoding the SG dipeptide. Practically, in order to obtain the plasmid pcDNA-Ssol, a DNA fragment encoding the ectodomain of SARS-CoV S was amplified by PCR with the aid of the oligonucleotides 5'-ATAGGATCCA CCAIGTTTAT TTTCTTATTA TTTCTTACTC TCACT-3' and 5'-ACCTC-

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CGGAT TTAATATATT GCTCATATTT TCCCAA-3' from the plasmid pcDNA-S, and then inserted between the unique BamHI and BspE1 sites of a modified eukaryotic expression plasmid pcDNA3.1(+) (Clontech) containing the tag sequence FLAG between its BamHI and Xho1 sites:

```
// GGATCC ...nnn... TCC GGA GAT TAT AAA GAT GAC
   BamHI           S  G  D  Y  K  D  D

GAC GAT AAA TAA CTCGAG //
  D  D  K  ter Xho1
```

The Nhe1-Xho1 and BamHI-Xho1 fragments, containing the cDNA for S, were then excised from the plasmid pcDNA-Ssol, and subcloned between the corresponding sites of the plasmid pTRIP-SD/SA-S-CTE and of the plasmid pTRIP-SD/SA-S-WPRE, respectively, in order to obtain the plasmids pTRIP-SD/SA-Ssol-CTE and pTRIP-SD/SA-Ssol-WPRE, deposited at the CNCM, on Dec. 1, 2004, under the numbers I-3337 and I-3335, respectively.

Pseudotyped vectors were produced according to Zennou et al. (2000, Cell, 101:173-185) and used to transduce FRhK-4 cells (15 000 cells) according to a series of 5 successive transduction cycles (15 000 cells) with a quantity of vector corresponding to 24 ng (TRIP-SD/SA-Ssol-CTE) or 40 ng (TRIP-SD/SA-Ssol-WPRE) of p24 per cycle. The transduced cells were cloned by limiting dilution and a series of 16 clones transduced with TRIP-SD/SA-Ssol-CTE and of 15 clones with TRIP-SD/SA-Ssol-WPRE were analyzed for the expression of the Ssol polypeptide by Western blotting visualized with an anti-FLAG monoclonal antibody (FIG. 26 and data not presented), and by capture ELISA specific for the Ssol polypeptide which was developed for this purpose (table XI and data not presented). Part of the process for selecting the best secretory clones is shown in FIG. 26. Capture ELISA is based on the use of solid phases coated with polyclonal antibodies of rabbits immunized with purified and inactivated SARS-CoV. These solid phases allow the capture of the Ssol polypeptide secreted into the cellular supernatants, whose presence is then visualized with a series of steps successively involving the attachment of an anti-FLAG monoclonal antibody (M2, SIGMA), of anti-mouse IgG(H+L) biotinylated rabbit polyclonal antibodies (Jackson) and of a streptavidin-peroxidase conjugate (Amersham) and then the addition of chromogen and substrate (TMB+H₂O₂, KPL).

TABLE XI

Analysis of the expression of the Ssol polypeptide by cell lines transduced with the lentiviral vectors TRIP-SD/SA-Ssol-WPRE and TRIP-SD/SA-Ssol-CTE.		
Vector	Clone	OD (450 nm)
Control	—	0.031
TRIP-SD/SA-Ssol-CTE	CTE2	0.547
	CTE3	0.668
	CTE9	0.171
	CTE12	0.208
	CTE13	0.133
TRIP-SD/SA-Ssol-WPRE	WPRE1	0.061
	WPRE10	0.134

The secretion of the Ssol polypeptide was assessed in the supernatant of a series of cell clones isolated after transduction of FRhK-4 cells with the lentiviral vectors TRIP-SD/SA-Ssol-WPRE and TRIP-SD/SA-Ssol-CTE. The supernatants diluted 1/50 were analyzed by a capture ELISA test specific for SARS-CoV S.

The cell line secreting the highest quantities of Ssol polypeptide in the culture supernatant is the FRhK4-Ssol-CTE3 line. It was subjected to a second series of 5 cycles of

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transduction with the vector TRIP-SD/SA-Ssol-CTE under conditions similar to those described above and then cloned. The subclone secreting the highest quantities of Ssol was selected by a combination of Western blot and capture ELISA analysis: it is the subclone FRhK4-Ssol-30, which was deposited at the CNCM, on Nov. 22, 2004, under the name I-3325.

The FRhK4-Ssol-30 line allows the quantitative production and purification of the recombinant Ssol polypeptide. In a typical experiment where the experimental conditions for growth, production and purification were optimized, the cells of the FRhK4-Ssol-30 line are inoculated in standard culture medium (pyruvate-free DMEM containing 4.5 g/l of glucose and supplemented with 5% FCS, 100 U/ml of penicillin and 100 µg/ml of streptomycin) in the form of a subconfluent monolayer (1 million cells per each 100 cm² in 20 ml of medium). At confluence, the standard medium is replaced with the secretion medium where the quantity of FCS is reduced to 0.5% and the quantity of medium reduced to 16 ml per each 100 cm². The culture supernatant is removed after 4 to 5 days of incubation at 35° C. and under 5% CO₂. The recombinant polypeptide Ssol is purified from the supernatant by the succession of steps of filtration on 0.1 µm polyethersulfone (PES) membrane, concentration by ultrafiltration on a PES membrane with a 50 kD cut-off, affinity chromatography on anti-FLAG matrix with elution with a solution of FLAG peptide (DYKDDDDK) at 100 µg/ml in TBS (50 mM tris, pH 7.4, 150 mM NaCl) and then gel filtration chromatography in TBS on sephadex G-75 beads (Pharmacia). The concentration of the purified recombinant Ssol polypeptide was determined by micro-BCA test (Pierce) and then its biochemical characteristics analyzed.

Analysis by 8% SDS acrylamide gel stained with silver nitrate demonstrates a predominant polypeptide whose molecular mass is about 180 kD and whose degree of purity may be evaluated at 98% (FIG. 27A). Two main peaks are detected by SELDI-TOF mass spectrometry (Cypherger): they correspond to single and double charged forms of a predominant polypeptide whose molecular mass is thus determined at 182.6±3.7 kD (FIGS. 27B and C). After transfer onto Prosorb membrane and rinsing in 0.1% TFA, the N-terminal end of the Ssol polypeptide was sequenced in liquid phase by Edman degradation on 5 residues (ABI494, Applied Biosystems) and determined as being SDLDR (FIG. 27D). This demonstrates that the signal peptide located at the N-terminal end of the SARS-CoV S protein, composed of aa 1 to 13 (MFIFLLFLTLTSG) according to an analysis carried out with the software signalP v2.0 (Nielsen et al., 1997, Protein Engineering, 10:1-6), is cleaved from the mature Ssol polypeptide. The recombinant Ssol polypeptide therefore consists of amino acids 14 to 1193 of the SARS-CoV S protein fused at the C-terminals with a sequence SG DYKDDDDK containing the sequence of the FLAG tag (underlined). The difference between the theoretical molar mass of the naked Ssol polypeptide (132.0 kD) and the real molar mass of the mature polypeptide (182.6 kD) suggests that the Ssol polypeptide is glycosylated.

A preparation of purified Ssol polypeptide, whose protein concentration was determined by micro-BCA test, makes it possible to prepare a calibration series in order to measure, with the aid of the capture ELISA test described above, the concentrations of Ssol present in the culture supernatants and to review the characteristics of the secretory lines. According to this test, the FRhK4-Ssol-CT3 line secretes 4 to 6 µg/ml of polypeptide Ssol while the FRhK4-Ssol-30 line secretes 9 to 13 µg/ml of Ssol after 4 to 5 days of culture at confluence. In addition, the purification scheme presented above makes it

possible routinely to purify from 1 to 2 mg of Ssol polypeptide per liter of culture supernatant.

EXAMPLE 12

Gene Immunization Involving the SARS-Associated Corona Virus (SARS-CoV) Spicule (S) Protein

The effect of a splice signal and of the posttranscriptional signals WPRE and CTE was analyzed after gene immunization of BALB/c mice (FIG. 28).

For that, BALB/c mice were immunized at intervals of 4 weeks by injecting into the tibialis anterior a saline solution of 50 μ g of plasmid DNA of pcDNA-S and pCI-S and, as a control, 50 μ g of plasmid DNA of pcDNA-N (directing the expression of SARS-CoV N) or of pCI-HA (directing the expression of the HA of the influenza virus A/PR/8/34) and the immune sera collected 3 weeks after the 2nd injection. The presence of antibodies directed against the SARS-CoV S was assessed by indirect ELISA using as antigen a lysate of VeroE6 cells infected with SARS-CoV and, as a control, a lysate of noninfected VeroE6 cells. The anti-SARS-CoV antibody titers (TI) are calculated as the reciprocal of the dilution producing a specific OD of 0.5 (difference between OD measured on a lysate of infected cells and OD measured on a lysate of noninfected cells) after visualization with an anti-mouse IgG polyclonal antibody coupled with peroxidase (NA931V, Amersham) and TMB supplemented with H₂O₂ (KPL) (FIG. 28A).

Under these conditions, the expression plasmid pcDNA-S only allows the induction of low antibody titers directed against SARS-CoV S in 3 mice out of 6 ($\text{LOG}_{10}(\text{TI})=1.9\pm 0.6$) whereas the plasmid pcDNA-N allows the induction of anti-N antibodies at high titers ($\text{LOG}_{10}(\text{TI})=3.9\pm 0.3$) in all the animals, and the control plasmids (pCI, pCI-HA) do not result in any detectable antibody ($\text{LOG}_{10}(\text{TI})<1.7$). The plasmid pCI-S equipped with a splice signal allows the induction of antibodies at high titers ($\text{LOG}_{10}(\text{TI})=3.7\pm 0.2$), which are approximately 60 times higher than those observed after injection of the plasmid pcDNA-S ($p<10^{-5}$).

The efficiency of the posttranscriptional signals was studied by carrying out a dose-response study of the anti-S antibody titers induced in the BALB/c mouse as a function of the quantity of plasmid DNA used as immunogen (2 μ g, 10 μ g and 50 μ g). This study (FIG. 28B) demonstrates that the posttranscriptional signal WPRE greatly improves the efficiency of gene immunization when small doses of DNA are used ($p<10^{-5}$ for a dose of 2 μ g of DNA and $p<10^{-2}$ for a dose of 10 μ g), whereas the effect of the CTE signal remains marginal ($p=0.34$ for a dose of 2 μ g of DNA).

Finally, the antibodies induced in mice after gene immunization neutralize the infectivity of SARS-CoV in vitro (FIGS. 29A and 29B) at titers which are consistent with the titers measured by ELISA.

In summary, the use of a splice signal and of the posttranscriptional signal WPRE of the woodchuck hepatitis virus considerably improves the induction of neutralizing antibodies directed against SARS-CoV after gene immunization with the aid of plasmid DNA directing the expression of the cDNA for SARS-CoV S.

EXAMPLE 13

Diagnostic Applications of the S Protein

The ELISA reactivity of the recombinant Ssol polypeptide was analyzed with respect to sera from patients suffering from SARS.

The sera from probable cases of SARS tested were chosen on the basis of the results (positive or negative) of analysis of their specific reactivity toward the native antigens of SARS-CoV by immunofluorescence test on VeroE6 cells infected with SARS-CoV and/or by indirect ELISA test using as antigen a lysate of VeroE6 cells infected with SARS-CoV. The sera of these patients are identified by a serial number of the National Reference Center for Influenza Viruses and by the initials of the patient and the number of days elapsed since the onset of the symptoms. All the sera of probable cases (cf. Table XII) recognize the native antigens of SARS-CoV, with the exception of the serum 032552 of the patient VTT for whom infection with SARS-CoV could not be confirmed by RT-PCR performed on respiratory samples of days 3, 8 and 12. A panel of control sera was used as control (TV sera): they are sera collected in France before the SARS epidemic that occurred in 2003.

TABLE XII

Sera of probable cases of SARS		
Serum	Patient	Sample collection day
031724	JYK	7
033168	JYK	38
033597	JYK	74
032632	NTM	17
032634	THA	15
032541	PHV	10
032542	NIH	17
032552	VTT	8
032633	PTU	16
032791	JLB	3
033258	JLB	27
032703	JCM	8
033153	JCM	29

Solid phases sensitized with the recombinant Ssol polypeptide were prepared by adsorption of a solution of purified Ssol polypeptide at 2 μ g/ml in PBS in the wells of an ELISA plate, and then the plates are incubated overnight at 4° C. and washed with PBS-Tween buffer (PBS, 0.1% Tween 20). After saturating the ELISA plates with a solution of PBS-10% skimmed milk (weight/volume) and washing in PBS-Tween, the sera to be tested (100 μ l) are diluted 1/400 in PBS skimmed milk-Tween buffer (PBS, 3% skimmed milk, 0.1% Tween) and then added to the wells of the sensitized ELISA plate. The plates are incubated for 1 h at 37° C. After 3 washings with PBS-Tween buffer, the anti-human IgG conjugate labeled with peroxidase (ref. NA933V, Amersham) diluted 1/4000 in PBS-skimmed milk-Tween buffer is added, and then the plates are incubated for 1 hour at 37° C. After 6 washings with PBS-Tween buffer, the chromogen (TMB) and the substrate (H₂O₂) are added and the plates are incubated for 10 minutes protected from light. The reaction is stopped by adding a 1 N H₃PO₄ solution, and then the absorbance is measured at 450 nm with a reference at 620 nm.

The ELISA tests (FIG. 30) demonstrate that the recombinant Ssol polypeptide is specifically recognized by the serum antibodies of patients suffering from SARS collected at the medium or late phase of infection (≥ 10 days after the onset of the symptoms) whereas it is not significantly recognized by the serum antibodies of 2 patients (JLB and JCM) collected in the early phase of infection (3 to 8 days after the onset of the symptoms) or by control sera of subjects not suffering from SARS. The serum antibodies of patients JLB and JCM show a seroconversion between days 3 and 27 for the first and 8 and

29 for the second after the onset of the symptoms, which confirms the specificity of the reactivity of these sera toward the Ssol polypeptide.

In conclusion, these results demonstrate that the recombinant Ssol polypeptide may be used as an antigen for the development of an ELISA test for serological diagnosis of infection with SARS-CoV.

EXAMPLE 14

Vaccine Applications of the Recombinant Soluble S Protein

The immunogenicity of the recombinant Ssol polypeptide was studied in mice.

For that, a group of 6 mice was immunized at 3 weeks' interval with 10 µg of recombinant Ssol polypeptide adjuvanted with 1 mg of aluminum hydroxide (Alu-gel-S, Serva) diluted in PBS. Three successive immunizations were performed and the immune sera were collected 3 weeks after each of the immunizations (IS1, IS2, IS3). As a control, a group of mice (mock group) received aluminum hydroxide alone according to the same protocol.

The immune sera were analyzed per pool for each of the 2 groups by indirect ELISA using a lysate of VeroE6 cells infected with SARS-CoV as antigen and as a control a lysate of noninfected VeroE6 cells. The anti-SARS-CoV antibody titers are calculated as the reciprocal of the dilution producing a specific OD of 0.5 after visualization with an anti-mouse IgG(H+L) polyclonal antibody coupled with peroxidase (NA931V, Amersham) and TMB supplemented with H₂O₂ (KPL). This analysis (FIG. 31) shows that the immunization with the Ssol polypeptide induces in mice, from the first immunization, antibodies directed against the native form of the SARS-CoV spicule protein present in the lysate of infected VeroE6 cells. After 2 then 3 immunizations, the anti-S antibody titers become very high.

The immune sera were analyzed per pool for each of the two groups for their capacity to seroneutralize the infectivity of SARS-CoV. 4 points of seroneutralization on FRhK-4 cells (100 TCID₅₀ of SARS-CoV) are produced for each of the 2-fold dilutions tested from 1/20. The seroneutralizing titer is calculated according to the Reed and Munsch method as the reciprocal of the dilution neutralizing the infectivity of 2 wells out of 4. This analysis shows that the antibodies induced in mice by the Ssol polypeptide are neutralizing: the titers observed are very high after 2 and then 3 immunizations (greater than 2560 and 5120 respectively, table XIII).

TABLE XIII

Induction of antibodies directed against SARS-CoV after immunization with the recombinant Ssol polypeptide.		
Group	Sera	Neutralizing Ab
Mock	pi	<20
	IS1	<20
	IS2	<20
	IS3	<20
Ssol	pi	<20
	IS1	57
	IS2	>2560
	IS3	>5120

The immune sera were analyzed per pool for each of the two groups for their capacity to seroneutralize the infectivity of 100 TCID₅₀ of SARS-CoV on FRhK-4 cells. 4 points are produced for each of the 2-fold dilutions tested from 1/20. The seroneutralizing titer is calculated according to the Reed and Munsch method as the reciprocal of the dilution neutralizing the infectivity of 2 wells out of 4.

The neutralizing titers observed in mice immunized with the Ssol polypeptide reach levels far greater than the titers observed by Yang et al. in mice (2004, Nature, 428:561-564) and those observed by Buchholz in the hamster (2004, PNAS 101:9804-9809) which protect respectively mice and hamsters from infection with SARS-CoV. It is therefore probable that the neutralizing antibodies induced in mice after immunization with the Ssol polypeptide protect these animals against infection with SARS-CoV.

EXAMPLE 15

Optimized Synthetic Gene for the Expression in Mammalian Cells of the SARS-Associated Coronavirus (SARS-CoV) Spicule (S) Protein

1) Design of the Synthetic Gene

A synthetic gene encoding the SARS-CoV spicule protein was designed from the gene of the isolate 031589 (plasmid pSARS-S, C.N.C.M. No. I-3059) so as to allow high levels of expression in mammalian cells and in particular in cells of human origin.

For that:

the use of codons of the wild-type gene of the isolate 031589 was modified so as to become close to the bias observed in humans and to improve the efficiency of translation of the corresponding mRNA

the overall GC content of the gene was increased so as to extend the half-life of the corresponding mRNA

the optionally cryptic motifs capable of interfering with an efficient expression of the gene were deleted (splice donor and acceptor sites, polyadenylation signals, sequences very rich (>80%) or very low (<30%) in GC, repeat sequences, sequences involved in the formation of secondary RNA structures, TATA boxes)

a second STOP codon was added to allow efficient termination of translation.

In addition, CpG motifs were introduced into the gene so as to increase its immunogenicity as DNA vaccine. In order to facilitate the manipulation of the synthetic gene, two BamHI and XhoI restriction sites were placed on either side of the open reading frame of the S protein, and the BamHI, XhoI, NheI, KpnI, BspEI and SalI restriction sites were avoided in the synthetic gene.

The sequence of the synthetic gene designed (gene 040530) is given in SEQ ID No: 140.

An alignment of the synthetic gene 040530 with the sequence of the wild-type gene of the isolate 031589 of SARS-CoV deposited at the C.N.C.M. under the number I-3059 (SEQ ID No: 4, plasmid pSRAS-S) is presented in FIG. 32.

2) Plasmid Constructs

The synthetic gene SEQ ID No: 140 was assembled from synthetic oligonucleotides and cloned between the KpnI and SacI sites of the plasmid pUC-Kana in order to give the plasmid 040530pUC-Kana. The nucleotide sequence of the insert of the plasmid 040530pUC-Kana was verified by automated sequencing (Applied).

A KpnI-XhoI fragment containing the synthetic gene 040530 was excised from the plasmid 040530pUC-Kana and subcloned between the NheI and XhoI sites of the expression plasmid pCI (Promega) in order to obtain the plasmid pCI-SSYNTH, deposited at the CNCM on Dec. 1, 2004, under the number I-3333.

A synthetic gene encoding the soluble form of the S protein was then obtained by fusing the synthetic sequences encoding the ectodomain of the S protein (amino acids 1 to 1193) with

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those of the tag (FLAG:DYKDDDDK) via a linker BspE1 encoding the dipeptide SG. Practically, a DNA fragment encoding the ectodomain of the SARS-CoV S was amplified by PCR with the aid of the oligonucleotides 5'-ACTA GCTAGC GGATCCACCATGTTTCATCTT CCTG-3' and 5'-AGTATCCGGAC TTG ATGTACT GCTCGTACTTGC-3' from the plasmid 040530pUC-Kana, digested with Nhe1 and BspE1 and then inserted between the unique Nhe1 and BspE1 sites of the plasmid pCI-Ssol, to give the plasmid pCI-SCUBE, deposited at the CNCM on Dec. 1, 2004, under the number I-3332. The plasmids pCI-Ssol, pCI-Ssol-CTE, and pCI-Ssol-WPRE (deposited at the CNCM, on Nov. 22, 2004, under the number I-3324) had been previously obtained by subcloning the Kpn1-Xho1 fragment excised from the plasmid pcDNA-Ssol (see technical note of DI 2004-106) between the Nhe1 and Xho1 sites of the plasmids pCI, pCI-S-CTE and pCI-S-WPRE respectively.)

The plasmids pCI-Scube and pCI-Ssol encode the same recombinant Ssol polypeptide.

3) Results

The capacity of the synthetic gene encoding the S protein to efficiently direct the expression of the SARS-CoV S in mammalian cells was compared with that of the wild-type gene after transient transfection of primate cells (VeroE6) and of human cells (293T).

In the experiment presented in FIG. 33 and in table XIV, monolayers of 5×10^5 VeroE6 cells or 7×10^5 293T cells in 35 mm Petri dishes were transfected with 2 μ g of plasmids pCI (as control), pCI-S, pCI-S-CTE, pCI-S-WPRE and pCI-S-Ssynth and 6 μ l of Fugene6 reagent according to the manufacturer's instructions (Roche). After 48 hours of incubation at 37° C. and under 5% CO₂, cell extracts were prepared in loading buffer according to Laemmli, separated on 8% SDS polyacrylamide gel and then transferred onto a PVDF membrane (BioRad). The detection of this immunoblot (Western blot) was carried out with the aid of an anti-S rabbit polyclonal serum (immune serum of the rabbit P11135: cf example 4 above) and of donkey polyclonal antibodies directed against rabbit IgGs and coupled with peroxidase (NA934V, Amersham). The immunoblot was quantitatively visualized by luminescence with the aid of the ECL+ kit (Amersham) and acquisition on a digital imaging device (FluorS, BioRad).

The analysis of the results obtained with the software QuantityOne v4.2.3 (BioRad) shows that in this experiment, the plasmid pCI-Synth allows the transient expression of the S protein at high levels in the VeroE6 and 293T cells, whereas the plasmid pCI-S does not make it possible to induce expression at sufficient levels to be detected. The expression. Levels observed are of the order of twice as high as those observed with the plasmid pCI-S-WPRE.

TABLE XIV

Use of a synthetic gene for the expression of the SARS-CoV S.		
Plasmid	VeroE6	293T
pCI	0.0	0.0
pCI-S	≤ 0.1	≤ 0.1
pCI-S-CTE	0.5	≤ 0.1

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TABLE XIV-continued

Use of a synthetic gene for the expression of the SARS-CoV S.		
Plasmid	VeroE6	293T
pCI-S-WPRE	1.0	1.0
pCI-Ssynth	1.8	1.9

Cell extracts prepared 48 hours after transfection of VeroE6 or 293T cells with the plasmids pCI, pCI-S, pCI-S-CTE, pCI-S-WPRE and pCI-S-Ssynth were separated on 8% SDS acrylamide gel and analyzed by Western blotting with the aid of an anti-S rabbit polyclonal antibody and an anti-rabbit IgG(H + L) polyclonal antibody coupled with peroxidase (NA934V, Amersham). The Western blot is visualized by luminescence (ECL+, Amersham) and acquisition on a digital imaging device (FluorS, BioRad). The expression levels of the S protein were measured by quantifying the two predominant bands identified on the image (see FIG. 33) and are indicated according to an arbitrary scale where the value 1 represents the level measured after transfection of the plasmid pCI-S-WPRE.

In a second instance, the capacity of the synthetic gene Scube to efficiently direct the synthesis and the secretion of the Ssol polypeptide by mammalian cells was compared with that of the wild-type gene after transient transfection of hamster cells (BHK-21) and of human cells (293T).

In the experiment presented in table XV, monolayers of 6×10^5 BHK-21 cells and 7×10^5 293T cells in 35 mm Petri dishes were transfected with 2 μ g of plasmids pCI (as control), pCI-Ssol, pCI-Ssol-CTE, pCI-Ssol-WPRE and pCI-Scube and 6 μ l of Fugene6 reagent according to the manufacturer's instructions (Roche). After 48 hours of incubation at 37° C. and under 5% CO₂, the cellular supernatants were collected and quantitatively analyzed for the secretion of the Ssol polypeptide by a capture ELISA test specific for the Ssol polypeptide.

Analysis of the results shows that, in this experiment, the plasmid pCI-Scube allows the expression of the Ssol polypeptide at levels 8 times (BHK-21 cells) to 20 times (293T cells) higher than the plasmid pCI-Ssol. The levels of expression observed are of the order of twice (293T cells) to 5 times (BHK-21 cells) as high as those observed with the plasmid pCI-Ssol-WPRE.

TABLE XV

Use of a synthetic gene for the expression of the Ssol polypeptide.		
Plasmid	BHK	293T
pCI	<20	<20
pCI-Ssol	<20	56 \pm 10
pCI-Ssol-CTE	<20	63 \pm 8
pCI-Ssol-WPRE	28 \pm 1	531 \pm 15
pCI-Scube	152 \pm 6	1140 \pm 20

The supernatants were harvested 48 hours after transfection of BHK or 293T cells with the plasmids pCI, pCI-Ssol, pCI-Ssol-CTE, pCI-Ssol-WPRE and pCI-Scube and quantitatively analyzed for the secretion of the Ssol polypeptide by an ELISA test specific for the Ssol polypeptide. The transfections were carried out in duplicate and the results are presented in the form of means and standard deviations of the concentrations of Ssol polypeptide (ng/ml) measured in the supernatants.

In summary, these results show that the expression, in mammalian cells, of the synthetic gene 040530 encoding SARS-CoV S under the control of RNA polymerase II promoter sequences is much more efficient than that of the wild-type gene of the 031589 isolate. This expression is even more efficient than that directed by the wild-type gene in the presence of the WPRE sequences of the woodchuck hepatitis virus.

4) Applications

The use of the synthetic gene 040530 encoding SARS-CoV S or its Scube variant encoding the polypeptide Ssol is capable of advantageously replacing the wild-type gene in numerous applications where the expression of S is necessary at high levels. In particular in order to:

improve the efficiency of gene immunization with plasmids of the pCI-Ssynth or even pCI-Ssynth-CTE or pCI-Ssynth-WPRE type
 establish novel cell lines expressing higher quantities of the S protein or of the Ssol polypeptide with the aid of recombinant lentiviral vectors carrying the Ssynth gene or the Scube gene respectively
 improve the immunogenicity of the recombinant lentiviral vectors allowing the expression of the S protein or of the Ssol polypeptide
 improve the immunogenicity of live vectors allowing the expression of the S protein or of the Ssol polypeptide like recombinant vaccinia viruses or recombinant measles viruses (see examples 16 and 17 below)

EXAMPLE 16

Expression of the SARS-Associated Coronavirus (SARS-CoV) Spicule (S) Protein with the Aid of Recombinant Vaccinia Viruses

Vaccine Application

Application to the Production of a Soluble Form of the Spicule (S) Protein and Design of a Serological Test for SARS

1) Introduction

The aim of this example is to evaluate the capacity of recombinant vaccinia viruses (VV) expressing various SARS-associated coronavirus (SARS-CoV) antigens to constitute novel vaccine candidates against SARS and a means of producing recombinant antigens in mammalian cells.

For that, the inventors focused on the SARS-CoV spicule (S) protein which makes it possible to induce, after gene immunization in animals, antibodies neutralizing the infectivity of SARS-CoV, and a soluble and secreted form of this protein, the Ssol polypeptide, which is composed of the ectodomain (aa 1-1193) of S fused at its C-ter end with a tag FLAG (DYKDDDDK) via a BspE1 linker encoding the SG dipeptide. This Ssol polypeptide exhibits an antigenicity similar to that of the S protein and allows, after injection into mice in the form of a purified protein adjuvanted with aluminum hydroxide, the induction of high neutralizing antibody titers against SARS-CoV.

The various forms of the S gene were placed under the control of the promoter of the 7.5K gene and then introduced into the thymidine kinase (TK) locus of the Copenhagen strain of the vaccinia virus by double homologous recombination in vivo. In order to improve the immunogenicity of the recombinant vaccinia viruses, a synthetic late promoter was chosen in place of the 7.5K promoter, in order to increase the production of S and Ssol during the late phases of the viral cycle.

After having isolated the recombinant vaccinia viruses and verified their capacity to express the SARS-CoV S antigen, their capacity to induce in mice an immune response against SARS was tested. After having purified the Ssol antigen from the supernatant of infected cells, an ELISA test for serodiagnosis of SARS was designed, and its efficiency was evaluated with the aid of sera from probable cases of SARS.

2) Construction of the Recombinant Viruses

Recombinant vaccinia viruses directing the expression of the S glycoprotein of the 031589 isolate of SARS-CoV and of a soluble and secreted form of this protein, the Ssol polypeptide, under the control of the 7.5K promoter were obtained.

With the aim of increasing the levels of expression of S and Ssol, recombinant viruses in which the cDNAs for S and for Ssol are placed under the control of a late synthetic promoter were also obtained.

The plasmid pTG186poly is a transfer plasmid for the construction of recombinant vaccinia viruses (Kieny, 1986, *Biotechnology*, 4:790-795). As such, it contains the VV thymidine kinase gene into which the promoter of the 7.5K gene has been inserted followed by a multiple cloning site allowing the insertion of heterologous genes (FIG. 34A). The promoter of the 7.5K gene in fact contains a tandem of two promoter sequences that are respectively active during the early (P_E) and late (P_L) phases of the vaccinia virus replication cycle. The BamHI-XhoI fragments were excised from the plasmids pTRIP-S and pcDNA-Ssol respectively and inserted between the BamHI and SmaI sites of the plasmid pTG186poly in order to give the plasmids pTG-S and pTG-Ssol (FIG. 34A). The plasmids pTG-S and pTG-Ssol were deposited at the CNCM, on Dec. 2, 2004, under the numbers I-3338 and I-3339, respectively.

The plasmids pTN480, pTN-S and pTN-Ssol were obtained from the plasmids pTG186poly, pTG-S and pTG-Ssol respectively, by substituting the NdeI-PstI fragment containing the 7.5K promoter by a DNA fragment containing the synthetic late promoter 480, which was obtained by hybridization of the oligonucleotides 5'-TATGAGCTTT TTTTTTTTTT TTTTTTTGGC ATATAAATAG ACTCG-CGCGC CCATCTGCA-3' and 5'-GATGCGCGCGC-CGAGTCTATT TATATGCCAA AAAAAAAAAA AAAAAAAAAAGC TCA-3' (FIG. 34B). The insert was sequenced with the aid of a BigDye Terminator v1.1 kit (Applied Biosystems) and an automated sequencer ABI377. The sequence of the late synthetic promoter 480 as cloned into the transfer plasmids of the pTN series is indicated in FIG. 34C. The plasmids pTN-S and pTN-Ssol were deposited at the CNCM, on Dec. 2, 2004, under the numbers I-3340 and I-3341, respectively.

The recombinant vaccinia viruses were obtained, by double homologous recombination in vivo between the TK cassette of the transfer plasmids of the series pTG and pTN and the TK gene of the Copenhagen strain of the vaccinia virus according to a procedure described by Kieny et al. (1984, *Nature*, 312:163-166). Briefly, CV-1 cells are transfected with the aid of DOTAP (Roche) with genomic DNA of the Copenhagen strain of the vaccinia virus and each of the transfer plasmids of the pTG and pTN series described above, and then superinfected with the helper vaccinia virus VV-ts7 for 24 hours at 33° C. The helper virus is counter-selected by incubation at 40° C. for 2 days and then the recombinant viruses (TK- phenotype) selected by two cloning cycles under agar medium on 143Btk- cells in the presence of BuDr (25 µg/ml). The 6 viruses VV-TG, VV-TG-S, VV-TG-Ssol, VV-TN, VV-TN-S, and VV-TN-Ssol are respectively obtained with the aid of the transfer plasmids pTG186poly, pTG-S, pTG-Ssol, pTN480, pTN-Ssol. The viruses VV-TG and VV-TN do not express any heterologous gene and were used as TK- control in the experiments. The preparations of recombinant viruses were performed on monolayers of CV-1 or BHK-21 cells and the titer in plaque forming units (p.f.u) determined on CV-1 cells according to Earl and Moss (1998, *Current Protocols in Molecular Biology*, 16.16.1-16.16.13).

3) Characterization of the Recombinant Viruses

The expression of the transgenes encoding the S protein and the Ssol polypeptide was assessed by Western blotting.

Monolayers of CV-1 cells were infected at a multiplicity of 2 with various recombinant vaccinia viruses VV-TG, VV-TG-S, VV-TG-Ssol, VV-TN, VV-TN-S and VV-TN-Ssol. After 18 hours of incubation at 37° C. and under 5% CO₂, cellular

extracts were prepared in loading buffer according to Laemmli, separated on 8% SDS polyacrylamide gel and then transferred onto a PVDF membrane (BioRad). The detection of this immunoblot (Western blot) was performed with the aid of an anti-S rabbit polyclonal serum (immune serum from the rabbit P11135: cf. example 4) and donkey polyclonal antibodies directed against rabbit IgGs and coupled with peroxidase (NA934V, Amersham). The bound antibodies were visualized by luminescence with the aid of the ECL+ kit (Amersham) and autoradiography films Hyperfilm MP (Amersham).

As shown in FIG. 35A, the recombinant virus VV-TN-S directs the expression of the S protein at levels which are comparable to those which can be observed 8 h after infection with SARS-CoV but which are much higher than those which can be observed after infection with VV-TG-S. In a second experiment (FIG. 35B), the analysis of variable quantities of cellular extracts shows that the levels of expression observed after infection with viruses of the TN series (VV-TN-S and VV-TN-Ssol) are about 10 times as high as those observed with the viruses of the TG series (VV-TG-S and VV-TG-Ssol, respectively). In addition, the Ssol polypeptide is secreted into the supernatant of CV-1 cells infected with the VV-TN-Ssol virus more efficiently than in the supernatant of cells infected with VV-TG-Ssol (FIG. 36A). In this experiment, the VV-TN-Sflag virus was used as a control because it expresses the membrane form of the S protein fused at its C-ter end with the FLAG tag. The Sflag protein is not detected in the supernatant of cells infected with VV-TN-Sflag, demonstrating that the Ssol polypeptide is indeed actively secreted after infection with VV-TN-Ssol.

These results demonstrate that the recombinant vaccinia viruses are indeed carriers of the transgenes and allow the expression of the SRAS glycoprotein in its membrane form (S) or in a soluble or secreted form (Ssol). The vaccinia viruses carrying the synthetic promoter 480 allow the expression of S and the secretion of Ssol at levels much higher than the viruses carrying the promoter of the 7.5K gene.

4) Application to the Production of a Soluble Form of SARS-CoV S. Purification of this Recombinant Antigen and Diagnostic Applications

The BHK-21 line is the cell line which secretes the highest quantities of Ssol polypeptide after infection with the VV-TN-Ssol virus among the lines tested (BHK-21, CV1, 293T and FrhK-4, FIG. 36B); it allows the quantitative production and purification of the recombinant Ssol polypeptide. In a typical experiment where the experimental conditions for infection, production and purification were optimized, the BHK-21 cells are inoculated in standard culture medium (pyruvate-free DMEM containing 4.5 g/l of glucose and supplemented with 5% TPB, 5% FCS, 100 U/ml of penicillin and 100 µg/ml of streptomycin) in the form of a subconfluent monolayer (10 million cells for each 100 cm² in 25 ml of medium). After 24 h of incubation at 37° C. under 5% CO₂, the cells are infected at an M.O.I. of 0.03 and the standard medium replaced with the secretion medium where the quantity of FCS is reduced to 0.5% and the TPB eliminated. The culture supernatant is removed after 2.5 days of incubation at 35° C. and under 5% CO₂ and the vaccinia virus inactivated by addition of Triton X-100 (0.1%). After filtration on 0.1 µm polyethersulfone (PES) membrane, the recombinant Ssol polypeptide is purified by affinity chromatography on an anti-FLAG matrix with elution with a solution of FLAG peptide (DYKDDDDK) at 100 µg/ml in TBS (50 mM Tris, pH 7.4, 150 mM NaCl).

The analysis by 8% SDS acrylamide gel stained with silver nitrate identified a predominant polypeptide whose molecu-

lar mass is about 180 kD and whose degree of purity is greater than 90% (FIG. 37). The concentration of the purified Ssol recombinant polypeptide was determined by comparison with molecular mass markers and estimated at 24 ng/µl.

This purified Ssol polypeptide preparation makes it possible to produce a calibration series in order to measure, with the aid of a capture ELISA test, the Ssol concentrations present in the culture supernatants. According to this test, the BHK-21 line secretes about 1 µg/ml of Ssol polypeptide under the production conditions described above. In addition, the purification scheme presented makes it possible to purify of the order of 160 µg of Ssol polypeptide per liter of culture supernatant.

The ELISA reactivity of the recombinant Ssol polypeptide was analyzed toward sera from patients suffering from SARS.

The sera of probable cases of SARS tested were chosen on the basis of the results (positive or negative) of analysis of their specific reactivity toward the native antigens of SARS-CoV by immunofluorescence test on VeroE6 cells infected with SARS-CoV and/or by indirect ELISA test using, as antigen, a lysate of VeroE6 cells infected with SARS-CoV. The sera of these patients are identified by a serial number of the National Reference Center for Influenza Viruses and by the patient's initials and the number of days elapsed since the onset of the symptoms. All the sera of probable cases (cf. table XVI) recognize the native antigens of SARS-CoV with the exception of the serum 032552 of the patient VTT, for which infection with SARS-CoV could not be confirmed by RT-PCR performed on respiratory samples of days 3, 8 and 12. A panel of control sera was used as control (TV sera): they are sera collected in France before the SARS epidemic which occurred in 2003.

TABLE XVI

Sera of probable cases of SARS			
Serum	Patient	Sample collection day	
033168	JYK	38	
033597	JYK	74	
032632	NTM	17	
032634	THA	15	
032541	PHV	10	
032542	NIH	17	
032552	VTT	8	
032633	PTU	16	

Solid phases sensitized with the recombinant Ssol polypeptide were prepared by adsorption of a solution of purified Ssol polypeptide at 4 µg/ml in PBS in the wells of an ELISA plate. The plates are incubated overnight at 4° C. and then washed with PES-Tween buffer (PBS, 0.1% Tween 20). After washing with PBS-Tween, the sera to be tested (100 µl) are diluted 1/100 and 1/400 in PBS-skimmed milk-Tween buffer (PBS, 3% skimmed milk, 0.1% Tween) and then added to the wells of the sensitized ELISA plate. The plates are then incubated for 1 h at 37° C. After 3 washings with PBS-Tween buffer, the anti-human IgG conjugate labeled with peroxidase (ref. NA933V, Amersham) diluted 1/4000 in PBS-skimmed milk-Tween buffer is added and then the plates are incubated for one hour at 37° C. After 6 washings with PBS-Tween buffer, the chromogen (TMB) and the substrate (H₂O₂) are added and the plates are incubated for 10 minutes protected from light. The reaction is stopped by adding a 1M solution of H₃PO₄ and then the absorbance is measured at 450 nm with a reference at 620 nm.

The ELISA tests (FIG. 38) demonstrate that the recombinant Ssol polypeptide is specifically recognized by the serum

antibodies of patients suffering from SARS, collected at the middle or late phase of infection (≥ 10 days after the onset of the symptoms), whereas it is not significantly recognized by the serum antibodies of the control sera of subjects not suffering from SARS.

In conclusion, these results demonstrate that the recombinant Ssol polypeptide can be purified from the supernatant of mammalian cells infected with the recombinant vaccinia virus VV-TN-Ssol and can be used as antigen for developing an ELISA test for serological diagnosis of infection with SARS-CoV.

5. Vaccine Applications

The immunogenicity of the recombinant vaccinia viruses was studied in mice.

For that, groups of 7 BALB/c mice were immunized by the i.v. route twice at 4 weeks' interval with 10^6 p.f.u. of recombinant vaccinia viruses VV-TG, VV-TG-S, VV-TG-Ssol, VV-TN, VV-TN-S and VV-TN-Ssol and, as a control, VV-TG-HA which directs the expression of hemagglutinin of the A/PR/8/34 strain of the influenza virus. The immune sera were collected 3 weeks after each of the immunizations (IS1, IS2).

The immune sera were analyzed per pool for each of the groups by indirect ELISA using a lysate of VeroE6 cells infected with SARS-CoV as antigen and, as control, a lysate of noninfected VeroE6 cells. The anti-SARS-CoV antibody titers (TI) are calculated as the reciprocal of the dilution producing a specific OD of 0.5 after visualization with an anti-mouse IgG(H+L) polyclonal antibody coupled with peroxidase (NA931V, Amersham) and TMB supplemented with H_2O_2 (KPL). This analysis (FIG. 39A) shows that immunization with the virus VV-TG-S and VV-TN-S induces in mice, from the first immunization, antibodies directed against the native form of the SARS-CoV spicule protein present in the lysate of infected VeroE6 cells. The responses induced by the VV-TN-S virus are higher than those induced by the VV-TG-S virus after the first (TI=740 and TI=270 respectively) and the second (TI=3230 and TI=600 respectively) immunization. The VV-TN-Ssol virus induces high anti-SARS-CoV antibody titers after two immunizations (TI=640), whereas the virus VV-TG-Ssol induces a response at the detection limit (TI=40).

The immune sera were analyzed per pool for each of the groups for their capacity to seroneutralize the infectivity of SARS-CoV. 4 seroneutralization points on FRhK-4 cells (100 TCID₅₀ of SARS-CoV) are produced for each of the 2-fold dilutions tested from 1/20. The seroneutralizing titer is calculated according to the Reed, and Munsch method as the reciprocal of the dilution neutralizing the infectivity of 2 wells out of 4. This analysis shows that the antibodies induced in mice by the vaccinia viruses expressing the S protein or the Ssol polypeptide are neutralizing and that the viruses with synthetic promoters are more efficient immunogens than the viruses carrying the 7.5K promoter: the highest titers (640) are observed after 2 immunizations with the virus VV-TN-S (FIG. 39B).

The protective power of the neutralizing antibodies induced in mice after immunization with the recombinant vaccinia viruses is evaluated with the aid of a challenge infection with SARS-CoV.

6) Other Applications

Third generation recombinant vaccinia viruses are constructed by substituting the wild-type sequences of the S and Ssol genes by synthetic genes optimized for the expression in mammalian cells, described above. These recombinant vaccinia viruses are capable of expressing larger quantities of S and Ssol antigens and therefore of exhibiting increased immunogenicity.

The recombinant vaccinia virus VV-TN-Ssol can be used for the quantitative production and purification of the Ssol antigen for diagnostic (serology by ELISA) and vaccine (sub-unit vaccine) applications.

EXAMPLE 17

Recombinant Measles Virus Expressing the SARS-Associated Coronavirus (SARS-CoV) Spicule (S) Protein. Vaccine Applications

1) Introduction

The measles vaccine (MV) induces a lasting protective immunity in humans after a single injection (Hilleman, 2002, *Vaccine*, 20: 651-665). The protection conferred is very robust and is based on the induction of an antibody response and of a CD4 and CD8 cell response. The MV genome is very stable and no reversion of the vaccine strains to virulence has ever been observed. The measles virus belongs to the genus *Morbillivirus* of the Paramyxoviridae family; it is an enveloped virus whose genome is a 16 kb single-stranded RNA of negative polarity (FIG. 40A) and whose exclusively cytoplasmic replication cycle excludes any possibility of integration into the genome of the host. The measles vaccine is thus one of the most effective and one of the safest live vaccines used in the human population. Frédéric Tangy's team recently developed an expression vector on the basis of the Schwarz strain of the measles virus, which is the safest attenuated strain and the most widely used in humans as vaccine against measles. This vaccine strain may be isolated from an infectious molecular clone while preserving its immunogenicity in primates and in mice that are sensitive to the infection. It constitutes, after insertion of additional transcription units, a vector for the expression of heterologous sequences (Combretet, 2003, *J. Virol.* 77: 11546-11554). In addition, a recombinant MV Schwarz expressing the envelope glycoprotein of the West Nile virus (WNV) induces an effective and lasting antibody response which protects mice from a lethal challenge infection with WNV (Despres et al., 2004, *J. Infect. Dis.*, in press). All these characteristics make the attenuated Schwarz strain of the measles virus an extremely promising candidate vector for the construction of novel recombinant live vaccines.

The aim of this example is to evaluate the capacity of recombinant measles viruses (MV) expressing various SARS-associated coronavirus (SARS-CoV) antigens to constitute novel candidate vaccines against SARS.

The inventors focused on the SARS-CoV spicule (S) protein, which makes it possible to induce, after gene immunization in animals, antibodies neutralizing the infectivity of SARS-CoV, and on a soluble and secreted form of this protein, the Ssol polypeptide, which is composed of the ectodomain (aa 1-1193) of S fused at its C-ter end with a FLAG tag (DYKDDDDK) via a BspE1 linker encoding the SG dipeptide. This Ssol polypeptide exhibits a similar antigenicity to that of the S protein and allows, after injection into mice in the form of a purified protein adjuvanted with aluminum hydroxide, the induction of high neutralizing antibody titers against SARS-CoV.

The various forms of the S gene were introduced in the form of an additional transcription unit between the P (phosphoprotein) and M (matrix) genes into the cDNA of the Schwarz strain of MV previously described (Combretet, 2003, *J. Virol.* 77: 11546-11554; EP application No. 02291551.6 of Jun. 20, 2002, and EP application No. 02291550.8 of Jun. 20, 2002). After having isolated the recombinant viruses MVSchw2-SARS-S and MVSchw2-

SARS-Ssol and checked their capacity to express the SARS-CoV S antigen, their capacity to induce a protective immune response against SARS in mice and then in monkeys was tested.

2) Construction of the Recombinant Viruses

The plasmid pTM-MVSchw-ATU2 (FIG. 40B) contains an infectious cDNA corresponding to the antigenome of the Schwarz vaccine strain of the measles virus (MV) into which an additional transcription unit (ATU) has been introduced between the P (phosphoprotein) and M (matrix) genes (Combrede, 2003, *Journal of Virology*, 77: 11546-11554). Recombinant genomes MVSchw2-SARS-S and MVSchw2-SARS-Ssol of the measles virus were constructed by inserting ORFs of the S protein and of the Ssol polypeptide into the additional transcription unit of the MVSchw-ATU2 vector.

For that, a DNA fragment containing the SARS-CoV S cDNA was amplified by PCR with the aid of the oligonucleotides 5'-ATACGTACGA CCATGTTTAT TTTCTTATTA TTTCTTACTC TCACT-3' and 5'-ATAGCGCGCT CATTATGTGT AATGTAATTT GACACCCTTG-3' using the plasmid pcDNA-S as template and then inserted into the plasmid pCR®2.1-TOPO (Invitrogen) in order to obtain the plasmid pTOPO-S-MV. The two oligonucleotides used contain restriction sites BsiW1 and BssHII, so as to allow subsequent insertion into the measles vector, and were designed so as to generate a sequence of 3774 nt including the codons for initiation and termination, so as to observe the rule of 6 which stipulates that the length of the genome of a measles virus must be divisible by 6 (Calain & Roux, 1993, *J. Virol.*, 67: 4822-4830; Schneider et al., 1997, *Virology*, 227: 314-322). The insert was sequenced with the aid of a BigDye Terminator v1.1 kit (Applied Biosystems) and an automated sequencer ABI377.

To express a soluble and secreted form of SARS-CoV S, a plasmid containing the cDNA of the Ssol polypeptide corresponding to the ectodomain (aa 1-1193) of SARS-CoV S fused at its C-ter end with the sequence of a FLAG tag (DYKDDDDK) via a BspE1 linker encoding the SG dipeptide was then obtained. For that, a DNA fragment was amplified with the aid of the oligonucleotides 5'-CCATTCAAC AATTTGGCCG-3' and 5'-ATAGGATCCGCGCGCTCATT ATTTATCGTC GTCATCTTTA TAATC-3' from the plasmid pCDNA-Ssol and then inserted into the plasmid pTOPO-S-MV between the SalI and BamHI sites in order to obtain the plasmid pTOPO-S-MV-SF. The sequence generated is 3618 nt long between the BsiW1 and BssHII sites and observes the rule of 6. The insert was sequenced as indicated above.

The BsiW1-BssHII fragments containing the cDNAs for the S protein and the Ssol polypeptide were then excised by digestion of the plasmids pTOPO-S-MV and pTOPO-S-MV-SF and then subcloned between the corresponding sites of the plasmid pTM-MVSchw-ATU2 in order to give the plasmids pTM-MVSchw2-SARS-S and pTM-MVSchw2-SARS-Ssol (FIG. 40B). These two plasmids were deposited at the C.N.C.M. on Dec. 1, 2004, under the numbers I-3326 and I-3327, respectively.

The recombinant measles viruses corresponding to the plasmids pTM-MVSchw2-SARS-S and pTM-MVSchw2-SARS-Ssol were obtained by reverse genetics according to the system based on the use of a helper cell line, described by Radecke et al. (1995, *Embo J.*, 14: 5773-5784) and modified by Parks et al. (1999, *J. Virol.*, 73: 3560-3566). Briefly, the helper cells 293-3-46 are transfected according to the calcium phosphate method with 5 µg of the plasmids pTM-MVSchw2-SARS-S or pTM-MVSchw2-SARS-Ssol and 0.02 µg of the plasmid pEMC-La directing the expression of the MV L polymerase (gift from M. A. Billeter). After incu-

bating, overnight at 37° C., a heat shock is produced for 2 hours at 43° C. and the transfected cells are transferred onto a monolayer of Vero cells. For each of the two plasmids, syncytia appeared after 2 to 3 days of coculture and were transferred successively onto monolayers of Vero cells at 70% confluence in 35 mm Petri dishes and then in, 25 and 75 cm² flasks. When the syncytia have reached 80-90% confluence, the cells are recovered with the aid of a scraper and then frozen and thawed once. After low-speed centrifugation, the supernatant containing the virus is stored in aliquots at -80° C. The titers of the recombinant viruses MVSchw2-SARS-S and MVSchw2-SARS-Ssol were determined by limiting dilution on Vero cells and the titer as dose infecting 50% of the wells (TCID₅₀) calculated according to the Kärber method.

3) Characterization of the Recombinant Viruses

The expression of the transgenes encoding the S protein and the Ssol polypeptide was assessed by Western blotting and immunofluorescence.

Monolayers of Vero cells in T-25 flasks were infected at a multiplicity of 0.05 by various passages of the two viruses MVSchw2-SARS-S and MVSchw2-SARS-Ssol and the wild-type virus MWSchw as a control. When the syncytia had reached 80 to 90% confluence, cytoplasmic extracts were prepared in an extraction buffer (150 mM NaCl, 50 mM Tris-HCl, pH 7.2, 1% Triton X-100, 0.1% SDS, 1% DOC) and then diluted in loading buffer according to Laemmli, separated on 8% SDS polyacrylamide gel and transferred onto a PVDF membrane (BioRad). The detection of this immunoblot (Western blot) was carried out with the aid of an anti-S rabbit polyclonal serum (immune serum of the rabbit P11135: cf. example 4 above) and donkey polyclonal antibodies directed against rabbit IgGs and coupled with peroxidase (NA934V, Amersham). The bound antibodies were visualized by luminescence with the aid of the ECL+ kit (Amersham) and Hyperfilm MP autoradiography films (Amersham).

Vero cells in monolayers on glass slides were infected with the two viruses MVSchw2-SARS-S and MVSchw2-SARS-Ssol and the wild-type virus MWSchw as a control at multiplicities of infection of 0.05. When the syncytia had reached 90 to 100% (MVSchw2-SARS-Ssol virus) or 30 to 40% (MVSchw2-SARS-S, MWSchw) confluence, the cells were fixed in a 4% PBS-PFA solution, permeabilized with a PBS solution containing 0.2% Triton and then labeled with rabbit polyclonal antibodies hyperimmunized with purified and inactivated SARS-CoV virions and with an anti-rabbit IgG (H+L) goat antibody conjugate coupled with FITC (Jackson).

As shown in FIGS. 41 and 42, the recombinant viruses MVSchw2-SARS-S and MVSchw2-SARS-Ssol direct the expression of the S protein and the Ssol polypeptide respectively at levels comparable to those which can be observed 8 h after infection with SARS-CoV. The expression of these polypeptides is stable after 3 passages of the recombinant viruses in cell culture. These results demonstrate that the recombinant measles viruses are indeed carriers of the transgenes and allow the expression of the SARS glycoprotein in its membrane form (S) or in a soluble form (Ssol). The Ssol polypeptide is expected to be secreted by cells infected with the MVSchw2-SARS-Ssol virus as is the case when this same polypeptide is expressed in mammalian cells after transient transfection of the corresponding sequences (cf. example 11 above).

4) Applications

Having shown that the viruses, MVSchw2-SARS-S and MVSchw2-SARS-Ssol allow the expression of the SARS-CoV S, their capacity to induce a protective immune response against SARS-CoV in CD46^{+/+} IFN- α βR^{-/-} mice, which is

sensitive to infection by MV, is evaluated. The antibody response of the immunized mice is evaluated by ELISA test against the native antigens of SARS-CoV and for their capacity to neutralize the infectivity of SARS-CoV in vitro, using the methodologies described above. The protective power of the response will be evaluated by measuring the reduction in the pulmonary viral load 2 days after a nonlethal challenge infection with SARS-CoV.

Second generation recombinant measles viruses are constructed by substituting the wild-type sequences of the S and Sol genes by synthetic genes optimized for expression in mammalian cells, described in example 15 above. These recombinant measles viruses are capable of expressing larger quantities of the S and Ssol antigens and therefore of exhibiting increased immunogenicity.

Alternatively, the wild-type or synthetic genes encoding the S protein or the Ssol polypeptide may be inserted into the measles vector MVSchw-ATU3 in the form of an additional transcription unit located between the H and L genes, and then the recombinant viruses produced and characterized in a similar manner. This insertion is capable of generating recombinant viruses possessing different characteristics (multiplication of the virus, level of expression of the transgene) and possibly an improved immunogenicity compared with those obtained after insertion of the transgenes between the P and N genes.

The recombinant measles virus MVSchw2-SARS-Ssol may be used for the quantitative production and the purification of the Ssol antigen for diagnostic and vaccine applications.

EXAMPLE 18

Other Applications Linked to the S Protein

a) The lentiviral vectors allowing the expression of S or Ssol (or even of fragments of S) can constitute a recombinant vaccine against SARS-CoV, to be used in human or veterinary prophylaxis. In order to demonstrate the feasibility of such a vaccine, the immunogenicity of the recombinant lentiviral vectors TRIP-SD/SA-S-WPRE and TRIP-SD/SA-Ssol-WPRE is studied in mice.

b) Monoclonal antibodies are produced with the aid of the recombinant Ssol polypeptide. According to the results presented in example 14 above, these antibodies or at least the majority of them will recognize the native form of the SARS-CoV S and will be capable of diagnostic and/or prophylactic applications.

c) A serological test for SARS is developed with the Ssol polypeptide used as antigen and the double epitope methodology.

SEQUENCE LISTING

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<211> LENGTH: 29746

<212> TYPE: DNA

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Met Phe Ile Phe Leu Leu Phe Leu
1 5

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Thr Leu Thr Ser Gly Ser Asp Leu Asp Arg Cys Thr Thr Phe Asp Asp
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Val Gln Ala Pro Asn Tyr Thr Gln His Thr Ser Ser Met Arg Gly Val
25 30 35 40

tac tat cct gat gaa att ttt aga tca gac act ctt tat tta act cag      256
Tyr Tyr Pro Asp Glu Ile Phe Arg Ser Asp Thr Leu Tyr Leu Thr Gln
45 50 55

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Asp Leu Phe Leu Pro Phe Tyr Ser Asn Val Thr Gly Phe His Thr Ile
60 65 70

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Asn His Thr Phe Gly Asn Pro Val Ile Pro Phe Lys Asp Gly Ile Tyr
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Phe Ala Ala Thr Glu Lys Ser Asn Val Val Arg Gly Trp Val Phe Gly
90 95 100

tct acc atg aac aac aag tca cag tcg gtg att att att aac aat tct      448
Ser Thr Met Asn Asn Lys Ser Gln Ser Val Ile Ile Ile Asn Asn Ser
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Thr Asn Val Val Ile Arg Ala Cys Asn Phe Glu Leu Cys Asp Asn Pro
125 130 135

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Phe Phe Ala Val Ser Lys Pro Met Gly Thr Gln Thr His Thr Met Ile
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Phe Asp Asn Ala Phe Asn Cys Thr Phe Glu Tyr Ile Ser Asp Ala Phe
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Ser Leu Asp Val Ser Glu Lys Ser Gly Asn Phe Lys His Leu Arg Glu
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Phe Val Phe Lys Asn Lys Asp Gly Phe Leu Tyr Val Tyr Lys Gly Tyr
185 190 195 200

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Gln Pro Ile Asp Val Val Arg Asp Leu Pro Ser Gly Phe Asn Thr Leu
205 210 215

aaa cct att ttt aag ttg cct ctt ggt att aac att aca aat ttt aga      784
Lys Pro Ile Phe Lys Leu Pro Leu Gly Ile Asn Ile Thr Asn Phe Arg
220 225 230

gcc att ctt aca gcc ttt tca cct gct caa gac att tgg ggc acg tca      832
Ala Ile Leu Thr Ala Phe Ser Pro Ala Gln Asp Ile Trp Gly Thr Ser
235 240 245

gct gca gcc tat ttt gtt ggc tat tta aag cca act aca ttt atg ctc      880
Ala Ala Ala Tyr Phe Val Gly Tyr Leu Lys Pro Thr Thr Phe Met Leu
250 255 260

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Asn	Pro	Leu	Ala	Glu	Leu	Lys	Cys	Ser	Val	Lys	Ser	Phe	Glu	Ile	Asp 295	
aaa	gga	att	tac	cag	acc	tct	aat	ttc	agg	gtt	gtt	ccc	tca	gga	gat	1024
Lys	Gly	Ile	Tyr	Gln	Thr	Ser	Asn	Phe	Arg	Val	Val	Pro	Ser	Gly	Asp 310	
gtt	gtg	aga	ttc	cct	aat	att	aca	aac	ttg	tgt	cct	ttt	gga	gag	gtt	1072
Val	Val	Arg	Phe	Pro	Asn	Ile	Thr	Asn	Leu	Cys	Pro	Phe	Gly	Glu	Val 325	
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Phe	Asn	Ala	Thr	Lys	Phe	Pro	Ser	Val	Tyr	Ala	Trp	Glu	Arg	Lys	Lys 340	
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Ile	Ser	Asn	Cys	Val	Ala	Asp	Tyr	Ser	Val	Leu	Tyr	Asn	Ser	Thr	Phe 360	
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Phe	Ser	Thr	Phe	Lys	Cys	Tyr	Gly	Val	Ser	Ala	Thr	Lys	Leu	Asn	Asp 375	
ctt	tgc	ttc	tcc	aat	gtc	tat	gca	gat	tct	ttt	gta	gtc	aag	gga	gat	1264
Leu	Cys	Phe	Ser	Asn	Val	Tyr	Ala	Asp	Ser	Phe	Val	Val	Lys	Gly	Asp 390	
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Asp	Val	Arg	Gln	Ile	Ala	Pro	Gly	Gln	Thr	Gly	Val	Ile	Ala	Asp	Tyr 405	
aat	tat	aaa	ttg	cca	gat	gat	ttc	atg	gg	t	gtc	ctt	gct	tg	aat	1360
Asn	Tyr	Lys	Leu	Pro	Asp	Asp	Phe	Met	Gly	Cys	Val	Leu	Ala	Trp	Asn 420	
act	agg	aac	att	gat	gct	act	tca	act	gg	aat	tat	aat	tat	aaa	tat	1408
Thr	Arg	Asn	Ile	Asp	Ala	Thr	Ser	Thr	Gly	Asn	Tyr	Asn	Tyr	Lys	Tyr 440	
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Asn	Val	Pro	Phe	Ser	Pro	Asp	Gly	Lys	Pro	Cys	Thr	Pro	Pro	Ala	Leu 470	
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Ile	Gly	Tyr	Gln	Pro	Tyr	Arg	Val	Val	Val	Leu	Ser	Phe	Glu	Leu	Leu 500	
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Lys	Asn	Gln	Cys	Val	Asn	Phe	Asn	Phe	Asn	Gly	Leu	Thr	Gly	Thr	Gly 535	
gtg	tta	act	cct	tct	tca	aag	aga	ttt	caa	cca	ttt	caa	caa	ttt	ggc	1744
Val	Leu	Thr	Pro	Ser	Ser	Lys	Arg	Phe	Gln	Pro	Phe	Gln	Gln	Phe	Gly 550	
cgt	gat	g	tct	gat	ttc	act	gat	tcc	g	gat	cct	aaa	aca	tct		1792
Arg	Asp	Val	Ser	Asp	Phe	Thr	Asp	Ser	Val	Arg	Asp	Pro	Lys	Thr	Ser 565	
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Glu	Ile	Leu	Asp	Ile	Ser	Pro	Cys	Ser	Phe	Gly	Gly	Val	Ser	Val	Ile 580	
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Val	Asn	Cys	Thr	Asp	Val	Ser	Thr	Ala	Ile	His	Ala	Asp	Gln	Leu	Thr		
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Pro	Ala	Trp	Arg	Ile	Tyr	Ser	Thr	Gly	Asn	Asn	Val	Phe	Gln	Thr	Gln		
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Ala	Gly	Cys	Leu	Ile	Gly	Ala	Glu	His	Val	Asp	Thr	Ser	Tyr	Glu	Cys		
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Asp	Ile	Pro	Ile	Gly	Ala	Gly	Ile	Cys	Ala	Ser	Tyr	His	Thr	Val	Ser		
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Leu	Leu	Arg	Ser	Thr	Ser	Gln	Lys	Ser	Ile	Val	Ala	Tyr	Thr	Met	Ser		
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Leu	Gly	Ala	Asp	Ser	Ser	Ile	Ala	Tyr	Ser	Asn	Asn	Thr	Ile	Ala	Ile		
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cct	act	aac	ttt	tca	att	agc	att	act	aca	gaa	gta	atg	cct	gtt	tct		2224
Pro	Thr	Asn	Phe	Ser	Ile	Ser	Ile	Thr	Thr	Glu	Val	Met	Pro	Val	Ser		
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Met	Ala	Lys	Thr	Ser	Val	Asp	Cys	Asn	Met	Tyr	Ile	Cys	Gly	Asp	Ser		
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Thr	Glu	Cys	Ala	Asn	Leu	Leu	Gln	Tyr	Gly	Ser	Phe	Cys	Thr	Gln			
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Leu	Asn	Arg	Ala	Leu	Ser	Gly	Ile	Ala	Ala	Glu	Gln	Asp	Arg	Asn	Thr		
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Arg	Glu	Val	Phe	Ala	Gln	Val	Lys	Gln	Met	Tyr	Lys	Thr	Pro	Thr	Leu		
				765					770					775			
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Lys	Tyr	Phe	Gly	Gly	Phe	Asn	Phe	Ser	Gln	Ile	Leu	Pro	Asp	Pro	Leu		
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Lys	Pro	Thr	Lys	Arg	Ser	Phe	Ile	Glu	Asp	Leu	Leu	Phe	Asn	Lys	Val		
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Thr	Leu	Ala	Asp	Ala	Gly	Phe	Met	Lys	Gln	Tyr	Gly	Glu	Cys	Leu	Gly		
						815						820					
gat	att	aat	gct	aga	gat	ctc	att	tgt	gcg	cag	aag	ttc	aat	gga	ctt		2608
Asp	Ile	Asn	Ala	Arg	Asp	Leu	Ile	Cys	Ala	Gln	Lys	Phe	Asn	Gly	Leu		
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Thr	Val	Leu	Pro	Pro	Leu	Leu	Thr	Asp	Asp	Met	Ile	Ala	Ala	Tyr	Thr		
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Ala	Ala	Leu	Val	Ser	Gly	Thr	Ala	Thr	Ala	Gly	Trp	Thr	Phe	Gly	Ala		
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Gly	Ala	Ala	Leu	Gln	Ile	Pro	Phe	Ala	Met	Gln	Met	Ala	Tyr	Arg	Phe		
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Asn	Gly	Ile	Gly	Val	Thr	Gln	Asn	Val	Leu	Tyr	Glu	Asn	Gln	Lys	Gln		
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Thr Thr Thr Ser Thr	Ala Leu Gly Lys Leu	Gln Asp Val Val Asn Gln	
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aat gct caa gca tta	aac aca ctt gtt aaa	caa ctt agc tct aat ttt	2944
Asn Ala Gln Ala Leu	Asn Thr Leu Val Lys	Gln Leu Ser Ser Asn Phe	
	940	945	950
ggc gca att tca agt	gtg cta aat gat atc	ctt tcg cga ctt gat aaa	2992
Gly Ala Ile Ser Ser	Val Leu Asn Asp Ile	Leu Ser Arg Leu Asp Lys	
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gtc gag gcg gag gta	caa att gac agg tta	att aca ggc aga ctt caa	3040
Val Glu Ala Glu Val	Gln Ile Asp Arg Leu	Ile Thr Gly Arg Leu Gln	
	970	975	980
agc ctt caa acc tat	gta aca caa caa cta	atc agg gct gct gaa atc	3088
Ser Leu Gln Thr Tyr	Val Thr Gln Gln Leu	Ile Arg Ala Ala Glu Ile	
985	990	995	1000
agg gct tct gct aat	ctt gct gct act aaa	atg tct gag tgt gtt	3133
Arg Ala Ser Ala Asn	Leu Ala Ala Thr Lys	Met Ser Glu Cys Val	
	1005	1010	1015
ctt gga caa tca aaa	aga gtt gac ttt tgt	gga aag ggc tac cac	3178
Leu Gly Gln Ser Lys	Arg Val Asp Phe Cys	Gly Lys Gly Tyr His	
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ctt atg tcc ttc cca	caa gca gcc ccg cat	ggc gtt gtc ttc cta	3223
Leu Met Ser Phe Pro	Gln Ala Ala Pro His	Gly Val Val Phe Leu	
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cat gtc acg tat gtg	cca tcc cag gag agg	aac ttc acc aca gcg	3268
His Val Thr Tyr Val	Pro Ser Gln Glu Arg	Asn Phe Thr Thr Ala	
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cca gca att tgt cat	gaa ggc aaa gca tac	ttc cct cgt gaa ggt	3313
Pro Ala Ile Cys His	Glu Gly Lys Ala Tyr	Phe Pro Arg Glu Gly	
	1065	1070	1075
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Val Phe Val Phe Asn	Gly Thr Ser Trp Phe	Ile Thr Gln Arg Asn	
	1080	1085	1090
ttc ttt tct cca caa	ata att act aca gac	aat aca ttt gtc tca	3403
Phe Phe Ser Pro Gln	Ile Ile Thr Thr Asp	Asn Thr Phe Val Ser	
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Gly Asn Cys Asp Val	Val Ile Gly Ile Ile	Asn Asn Thr Val Tyr	
	1110	1115	1120
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Asp Pro Leu Gln Pro	Glu Leu Asp Ser Phe	Lys Glu Glu Leu Asp	
	1125	1130	1135
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Lys Tyr Phe Lys Asn	His Thr Ser Pro Asp	Val Asp Leu Gly Asp	
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att tca ggc att aac	gct tct gtc gtc aac	att caa aaa gaa att	3583
Ile Ser Gly Ile Asn	Ala Ser Val Val Asn	Ile Gln Lys Glu Ile	
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Asp Arg Leu Asn Glu	Val Ala Lys Asn Leu	Asn Glu Ser Leu Ile	
	1170	1175	1180
gac ctt caa gaa ttg	gga aaa tat gag caa	tat att aaa tgg cct	3673
Asp Leu Gln Glu Leu	Gly Lys Tyr Glu Gln	Tyr Ile Lys Trp Pro	
	1185	1190	1195
tgg tat gtt tgg ctc	ggc ttc att gct gga	cta att gcc atc gtc	3718
Trp Tyr Val Trp Leu	Gly Phe Ile Ala Gly	Leu Ile Ala Ile Val	
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Leu Lys Gly Ala Cys Ser Cys Gly Ser Cys Cys Lys Phe Asp Glu	
1230	1235 1240
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Asp Asp Ser Glu Pro Val Leu Lys Gly Val Lys Leu His Tyr Thr	
1245	1250 1255
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His Thr Ser Ser Met Arg Gly Val Tyr Tyr Pro Asp Glu Ile Phe Arg	
35 40 45	
Ser Asp Thr Leu Tyr Leu Thr Gln Asp Leu Phe Leu Pro Phe Tyr Ser	
50 55 60	
Asn Val Thr Gly Phe His Thr Ile Asn His Thr Phe Gly Asn Pro Val	
65 70 75 80	
Ile Pro Phe Lys Asp Gly Ile Tyr Phe Ala Ala Thr Glu Lys Ser Asn	
85 90 95	
Val Val Arg Gly Trp Val Phe Gly Ser Thr Met Asn Asn Lys Ser Gln	
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Ser Val Ile Ile Ile Asn Asn Ser Thr Asn Val Val Ile Arg Ala Cys	
115 120 125	
Asn Phe Glu Leu Cys Asp Asn Pro Phe Phe Ala Val Ser Lys Pro Met	
130 135 140	
Gly Thr Gln Thr His Thr Met Ile Phe Asp Asn Ala Phe Asn Cys Thr	
145 150 155 160	
Phe Glu Tyr Ile Ser Asp Ala Phe Ser Leu Asp Val Ser Glu Lys Ser	
165 170 175	
Gly Asn Phe Lys His Leu Arg Glu Phe Val Phe Lys Asn Lys Asp Gly	
180 185 190	
Phe Leu Tyr Val Tyr Lys Gly Tyr Gln Pro Ile Asp Val Val Arg Asp	
195 200 205	
Leu Pro Ser Gly Phe Asn Thr Leu Lys Pro Ile Phe Lys Leu Pro Leu	
210 215 220	
Gly Ile Asn Ile Thr Asn Phe Arg Ala Ile Leu Thr Ala Phe Ser Pro	
225 230 235 240	
Ala Gln Asp Ile Trp Gly Thr Ser Ala Ala Ala Tyr Phe Val Gly Tyr	
245 250 255	
Leu Lys Pro Thr Thr Phe Met Leu Lys Tyr Asp Glu Asn Gly Thr Ile	
260 265 270	
Thr Asp Ala Val Asp Cys Ser Gln Asn Pro Leu Ala Glu Leu Lys Cys	
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Ser Val Lys Ser Phe Glu Ile Asp Lys Gly Ile Tyr Gln Thr Ser Asn	
290 295 300	

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Val	Tyr	Ala	Trp	Glu	Arg	Lys	Lys	Ile	Ser	Asn	Cys	Val	Ala	Asp	Tyr				340							345							350											
Ser	Val	Leu	Tyr	Asn	Ser	Thr	Phe	Phe	Ser	Thr	Phe	Lys	Cys	Tyr	Gly				355							360							365											
Val	Ser	Ala	Thr	Lys	Leu	Asn	Asp	Leu	Cys	Phe	Ser	Asn	Val	Tyr	Ala				370							375							380											
Asp	Ser	Phe	Val	Val	Lys	Gly	Asp	Asp	Val	Arg	Gln	Ile	Ala	Pro	Gly											385							390			395				400				
Gln	Thr	Gly	Val	Ile	Ala	Asp	Tyr	Asn	Tyr	Lys	Leu	Pro	Asp	Asp	Phe											405							410						415					
Met	Gly	Cys	Val	Leu	Ala	Trp	Asn	Thr	Arg	Asn	Ile	Asp	Ala	Thr	Ser											420							425							430				
Thr	Gly	Asn	Tyr	Asn	Tyr	Lys	Tyr	Arg	Tyr	Leu	Arg	His	Gly	Lys	Leu											435							440						445					
Arg	Pro	Phe	Glu	Arg	Asp	Ile	Ser	Asn	Val	Pro	Phe	Ser	Pro	Asp	Gly											450							455							460				
Lys	Pro	Cys	Thr	Pro	Pro	Ala	Leu	Asn	Cys	Tyr	Trp	Pro	Leu	Asn	Asp											465							470				475				480			
Tyr	Gly	Phe	Tyr	Thr	Thr	Thr	Gly	Ile	Gly	Tyr	Gln	Pro	Tyr	Arg	Val											485							490						495					
Val	Val	Leu	Ser	Phe	Glu	Leu	Leu	Asn	Ala	Pro	Ala	Thr	Val	Cys	Gly											500							505							510				
Pro	Lys	Leu	Ser	Thr	Asp	Leu	Ile	Lys	Asn	Gln	Cys	Val	Asn	Phe	Asn											515							520						525					
Phe	Asn	Gly	Leu	Thr	Gly	Thr	Gly	Val	Leu	Thr	Pro	Ser	Ser	Lys	Arg											530							535						540					
Phe	Gln	Pro	Phe	Gln	Gln	Phe	Gly	Arg	Asp	Val	Ser	Asp	Phe	Thr	Asp											545							550						555				560	
Ser	Val	Arg	Asp	Pro	Lys	Thr	Ser	Glu	Ile	Leu	Asp	Ile	Ser	Pro	Cys											565							570							575				
Ser	Phe	Gly	Gly	Val	Ser	Val	Ile	Thr	Pro	Gly	Thr	Asn	Ala	Ser	Ser											580							585							590				
Glu	Val	Ala	Val	Leu	Tyr	Gln	Asp	Val	Asn	Cys	Thr	Asp	Val	Ser	Thr											595							600							605				
Ala	Ile	His	Ala	Asp	Gln	Leu	Thr	Pro	Ala	Trp	Arg	Ile	Tyr	Ser	Thr											610							615							620				
Gly	Asn	Asn	Val	Phe	Gln	Thr	Gln	Ala	Gly	Cys	Leu	Ile	Gly	Ala	Glu											625							630							635				640
His	Val	Asp	Thr	Ser	Tyr	Glu	Cys	Asp	Ile	Pro	Ile	Gly	Ala	Gly	Ile											645							650							655				
Cys	Ala	Ser	Tyr	His	Thr	Val	Ser	Leu	Leu	Arg	Ser	Thr	Ser	Gln	Lys											660							665							670				
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Tyr	Ser	Asn	Asn	Thr	Ile	Ala	Ile	Pro	Thr	Asn	Phe	Ser	Ile	Ser	Ile											690							695							700				
Thr	Thr	Glu	Val	Met	Pro	Val	Ser	Met	Ala	Lys	Thr	Ser	Val	Asp	Cys											705							710							715				720
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 770 775 780
 Ser Gln Ile Leu Pro Asp Pro Leu Lys Pro Thr Lys Arg Ser Phe Ile
 785 790 795 800
 Glu Asp Leu Leu Phe Asn Lys Val Thr Leu Ala Asp Ala Gly Phe Met
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 Lys Gln Tyr Gly Glu Cys Leu Gly Asp Ile Asn Ala Arg Asp Leu Ile
 820 825 830
 Cys Ala Gln Lys Phe Asn Gly Leu Thr Val Leu Pro Pro Leu Leu Thr
 835 840 845
 Asp Asp Met Ile Ala Ala Tyr Thr Ala Ala Leu Val Ser Gly Thr Ala
 850 855 860
 Thr Ala Gly Trp Thr Phe Gly Ala Gly Ala Ala Leu Gln Ile Pro Phe
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 Ala Met Gln Met Ala Tyr Arg Phe Asn Gly Ile Gly Val Thr Gln Asn
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 Val Leu Tyr Glu Asn Gln Lys Gln Ile Ala Asn Gln Phe Asn Lys Ala
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 915 920 925
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 930 935 940
 Val Lys Gln Leu Ser Ser Asn Phe Gly Ala Ile Ser Ser Val Leu Asn
 945 950 955 960
 Asp Ile Leu Ser Arg Leu Asp Lys Val Glu Ala Glu Val Gln Ile Asp
 965 970 975
 Arg Leu Ile Thr Gly Arg Leu Gln Ser Leu Gln Thr Tyr Val Thr Gln
 980 985 990
 Gln Leu Ile Arg Ala Ala Glu Ile Arg Ala Ser Ala Asn Leu Ala Ala
 995 1000 1005
 Thr Lys Met Ser Glu Cys Val Leu Gly Gln Ser Lys Arg Val Asp
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 Pro His Gly Val Val Phe Leu His Val Thr Tyr Val Pro Ser Gln
 1040 1045 1050
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 1055 1060 1065
 Ala Tyr Phe Pro Arg Glu Gly Val Phe Val Phe Asn Gly Thr Ser
 1070 1075 1080
 Trp Phe Ile Thr Gln Arg Asn Phe Phe Ser Pro Gln Ile Ile Thr
 1085 1090 1095
 Thr Asp Asn Thr Phe Val Ser Gly Asn Cys Asp Val Val Ile Gly
 1100 1105 1110
 Ile Ile Asn Asn Thr Val Tyr Asp Pro Leu Gln Pro Glu Leu Asp
 1115 1120 1125
 Ser Phe Lys Glu Glu Leu Asp Lys Tyr Phe Lys Asn His Thr Ser
 1130 1135 1140
 Pro Asp Val Asp Leu Gly Asp Ile Ser Gly Ile Asn Ala Ser Val

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1145	1150	1155
Val Asn Ile Gln Lys Glu Ile Asp Arg Leu Asn Glu Val Ala Lys		
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Asn Leu Asn Glu Ser Leu Ile Asp Leu Gln Glu Leu Gly Lys Tyr		
1175	1180	1185
Glu Gln Tyr Ile Lys Trp Pro Trp Tyr Val Trp Leu Gly Phe Ile		
1190	1195	1200
Ala Gly Leu Ile Ala Ile Val Met Val Thr Ile Leu Leu Cys Cys		
1205	1210	1215
Met Thr Ser Cys Cys Ser Cys Leu Lys Gly Ala Cys Ser Cys Gly		
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<210> SEQ ID NO 4

<211> LENGTH: 3943

<212> TYPE: DNA

<213> ORGANISM: CORONAVIRUS

<400> SEQUENCE: 4

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aacatacttc atctatgagg ggggtttact atcctgatga aatttttaga tcagacactc    240
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caactggtaa ttataattat aaatataggt atcttagaca tggcaagcctt aggccctttg   1440
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<210> SEQ ID NO 5
 <211> LENGTH: 2049
 <212> TYPE: DNA
 <213> ORGANISM: CORONAVIRUS

<400> SEQUENCE: 5

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<211> LENGTH: 2027
<212> TYPE: DNA
<213> ORGANISM: CORONAVIRUS

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<210> SEQ ID NO 7
<211> LENGTH: 1096

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<212> TYPE: DNA

<213> ORGANISM: CORONAVIRUS

<400> SEQUENCE: 7

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gggcttccag ttcatttga atttactgct gctatttgtt accatctatt cacatctttt	420
gcttgcctgc gcaggtatgg aggcgcaatt tttgtacctc tatgcctga tatattttct	480
acaatgcatc aacgcatgta gaattattat gagatggttg ctttgggtga agtgcaaatc	540
caagaaccca ttactttatg atgccaacta ctttgggtgc tggcacacac ataactatga	600
ctactgtata ccatataaca gtgtcacaga tacaattgtc gttactgaag gtgacggcat	660
ttcaacacca aaactcaaag aagactacca aattgggtgt tattctgagg ataggcactc	720
agggtttaa gactatgtcg ttgtacatgg ctatttcacc gaagtctact accagcttga	780
gtctacacaa attactacag aactggtat tgaaaatgct acattcttca tctttaacaa	840
gcttgtttaa gaccaccgca atgtgcaaat acacacaatc gacggctctt caggagtgc	900
taatccagca atggatccaa tttatgatga gccgacgacg actactagcg tgcctttgta	960
agcacaagaa agtgagtacg aacttatgta ctcatctggt tcggaagaaa caggtagctt	1020
aatagttaat agcgtacttc ttttcttgc tttcgtggtta tcttggctag tcacactagc	1080
catccttact gcgctt	1096

<210> SEQ ID NO 8

<211> LENGTH: 1135

<212> TYPE: DNA

<213> ORGANISM: CORONAVIRUS

<400> SEQUENCE: 8

attgccatcg tcatggttac aatcttgctt tgttgcata ctagtgttg cagttgcctc	60
aagggtgcat gctcttggg tcttggctgc aagtttgatg aggatgactc tgagccagtt	120
ctcaagggtg tcaaattaca ttacacataa acgaacttat ggatttgttt atgagatttt	180
ttactcttgg atcaattact gcacagccag taaaaattga caatgcttct cctgcaagta	240
ctgttcatgc tacagcaacg ataccgctac aagcctcact ccctttcgga tggcttgtta	300
ttggcgttgc atttcttgcg gtttttcaga gcgctaccaa aataattgcg ctcaataaaa	360
gatggcagct agccctttat aagggttcc agttcatttg caatttactg ctgctatttg	420
ttaccatcta ttcacatctt ttgcttgcg ctgcaggat ggaggcga tttttgtacc	480
tctatgcctt gatataattt ctacaatgca tcaacgcatg tagaattatt atgagatggt	540
ggcttgggtg gaagtgcaaa tccaagaacc cactacttta tgatgccaac tactttgttt	600
gctggcacac acataactat gactactgta taccatataa cagtgtcaca gatacaattg	660
tcgttactga aggtgacgac atttcaaac caaaactcaa agaagactac caaattggtg	720
gttattctga ggataggcac tcagggtgta aagactatgt cgttgatcat ggctatttca	780
ccgaagtta ctaccagctt gactctacac aaattactac agacactggt attgaaatg	840
ctacattctt catctttaac aagcttgtta aagaccacc gaatgtgcaa atacacaaa	900

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tcgacggctc ttcaggagtt gctaataccag caatggatcc aatttatgat gagccgacga 960
cgactactag cgtgcctttg taagcacaag aaagtgagta cgaacttatg tactcattcg 1020
tttgggaaga aacaggtagc ttaatagtta atagegtact tctttttctt gctttcgtgg 1080
tattcttgct agtcacacta gccatcctta ctgcgcttcg attgtgtgcg tactg 1135

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<210> SEQ ID NO 9
<211> LENGTH: 1096
<212> TYPE: DNA
<213> ORGANISM: CORONAVIRUS
<220> FEATURE:
<221> NAME/KEY: CDS
<222> LOCATION: (137)..(958)
<223> OTHER INFORMATION:

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<400> SEQUENCE: 9

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tcttgctttg ttgcatgact agttgttgca gttgcctcaa ggggtgcatgc tcttgtggtt 60
cttggctgcaa gtttgatgag gatgactctg agccagttct caaggggtgc aaattacatt 120
acacataaac gaactt atg gat ttg ttt atg aga ttt ttt act ctt gga tca 172
      Met Asp Leu Phe Met Arg Phe Phe Thr Leu Gly Ser
      1             5             10
att act gca cag cca gta aaa att gac aat gct tct cct gca agt act 220
Ile Thr Ala Gln Pro Val Lys Ile Asp Asn Ala Ser Pro Ala Ser Thr
      15             20             25
gtt cat gct aca gca acg ata ccg cta caa gcc tca ctc cct ttc gga 268
Val His Ala Thr Ala Thr Ile Pro Leu Gln Ala Ser Leu Pro Phe Gly
      30             35             40
tgg ctt gtt att ggc gtt gca ttt ctt gct gtt ttt cag agc gct acc 316
Trp Leu Val Ile Gly Val Ala Phe Leu Ala Val Phe Gln Ser Ala Thr
      45             50             55             60
aaa ata att gcg ctc aat aaa aga tgg cag cta gcc ctt tat aag ggc 364
Lys Ile Ile Ala Leu Asn Lys Arg Trp Gln Leu Ala Leu Tyr Lys Gly
      65             70             75
ttc cag ttc att tgc aat tta ctg ctg cta ttt gtt acc atc tat tca 412
Phe Gln Phe Ile Cys Asn Leu Leu Leu Leu Phe Val Thr Ile Tyr Ser
      80             85             90
cat ctt ttg ctt gtc gct gca ggt atg gag gcg caa ttt ttg tac ctc 460
His Leu Leu Leu Val Ala Ala Gly Met Glu Ala Gln Phe Leu Tyr Leu
      95             100             105
tat gcc ttg ata tat ttt cta caa tgc atc aac gca tgt aga att att 508
Tyr Ala Leu Ile Tyr Phe Leu Gln Cys Ile Asn Ala Cys Arg Ile Ile
      110             115             120
atg aga tgt tgg ctt tgt tgg aag tgc aaa tcc aag aac cca tta ctt 556
Met Arg Cys Trp Leu Cys Trp Lys Cys Lys Ser Lys Asn Pro Leu Leu
      125             130             135             140
tat gat gcc aac tac ttt gtt tgc tgg cac aca cat aac tat gac tac 604
Tyr Asp Ala Asn Tyr Phe Val Cys Trp His Thr His Asn Tyr Asp Tyr
      145             150             155
tgt ata cca tat aac agt gtc aca gat aca att gtc gtt act gaa ggt 652
Cys Ile Pro Tyr Asn Ser Val Thr Asp Thr Ile Val Val Thr Glu Gly
      160             165             170
gac ggc att tca aca cca aaa ctc aaa gaa gac tac caa att ggt ggt 700
Asp Gly Ile Ser Thr Pro Lys Leu Lys Glu Asp Tyr Gln Ile Gly Gly
      175             180             185
tat tct gag gat agg cac tca ggt gtt aaa gac tat gtc gtt gta cat 748
Tyr Ser Glu Asp Arg His Ser Gly Val Lys Asp Tyr Val Val Val His
      190             195             200
ggc tat ttc acc gaa gtt tac tac cag ctt gag tct aca caa att act 796
Gly Tyr Phe Thr Glu Val Tyr Tyr Gln Leu Glu Ser Thr Gln Ile Thr
      205             210             215             220

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aca gac act ggt att gaa aat gct aca ttc ttc atc ttt aac aag ctt      844
Thr Asp Thr Gly Ile Glu Asn Ala Thr Phe Phe Ile Phe Asn Lys Leu
      225                      230                      235

ggt aaa gac cca ccg aat gtg caa ata cac aca atc gac ggc tct tca      892
Val Lys Asp Pro Pro Asn Val Gln Ile His Thr Ile Asp Gly Ser Ser
      240                      245                      250

gga gtt gct aat cca gca atg gat cca att tat gat gag ccg acg acg      940
Gly Val Ala Asn Pro Ala Met Asp Pro Ile Tyr Asp Glu Pro Thr Thr
      255                      260                      265

act act agc gtg cct ttg taagcacaag aaagtgagta cgaacttatg      988
Thr Thr Ser Val Pro Leu
      270

tactcattcg ttccggaaga aacaggtacg ttaatagtta atagcgtact tctttttctt 1048

gctttcgtgg tattcttctgt agtcacacta gccatcctta ctgcgctt      1096

<210> SEQ ID NO 10
<211> LENGTH: 274
<212> TYPE: PRT
<213> ORGANISM: CORONAVIRUS

<400> SEQUENCE: 10

Met Asp Leu Phe Met Arg Phe Phe Thr Leu Gly Ser Ile Thr Ala Gln
 1      5      10      15
Pro Val Lys Ile Asp Asn Ala Ser Pro Ala Ser Thr Val His Ala Thr
 20      25      30
Ala Thr Ile Pro Leu Gln Ala Ser Leu Pro Phe Gly Trp Leu Val Ile
 35      40      45
Gly Val Ala Phe Leu Ala Val Phe Gln Ser Ala Thr Lys Ile Ile Ala
 50      55      60
Leu Asn Lys Arg Trp Gln Leu Ala Leu Tyr Lys Gly Phe Gln Phe Ile
 65      70      75      80
Cys Asn Leu Leu Leu Phe Val Thr Ile Tyr Ser His Leu Leu Leu
 85      90      95
Val Ala Ala Gly Met Glu Ala Gln Phe Leu Tyr Leu Tyr Ala Leu Ile
100     105     110
Tyr Phe Leu Gln Cys Ile Asn Ala Cys Arg Ile Ile Met Arg Cys Trp
115     120     125
Leu Cys Trp Lys Cys Lys Ser Lys Asn Pro Leu Leu Tyr Asp Ala Asn
130     135     140
Tyr Phe Val Cys Trp His Thr His Asn Tyr Asp Tyr Cys Ile Pro Tyr
145     150     155     160
Asn Ser Val Thr Asp Thr Ile Val Val Thr Glu Gly Asp Gly Ile Ser
165     170     175
Thr Pro Lys Leu Lys Glu Asp Tyr Gln Ile Gly Gly Tyr Ser Glu Asp
180     185     190
Arg His Ser Gly Val Lys Asp Tyr Val Val Val His Gly Tyr Phe Thr
195     200     205
Glu Val Tyr Tyr Gln Leu Glu Ser Thr Gln Ile Thr Thr Asp Thr Gly
210     215     220
Ile Glu Asn Ala Thr Phe Phe Ile Phe Asn Lys Leu Val Lys Asp Pro
225     230     235     240
Pro Asn Val Gln Ile His Thr Ile Asp Gly Ser Ser Gly Val Ala Asn
245     250     255
Pro Ala Met Asp Pro Ile Tyr Asp Glu Pro Thr Thr Thr Thr Ser Val
260     265     270

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Pro Leu

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<210> SEQ ID NO 11
<211> LENGTH: 1096
<212> TYPE: DNA
<213> ORGANISM: CORONAVIRUS
<220> FEATURE:
<221> NAME/KEY: CDS
<222> LOCATION: (558)..(1019)
<223> OTHER INFORMATION:

<400> SEQUENCE: 11

tcttgctttg ttgcatgact agttggttga gttgectcaa gggtgcatgc tcttgggtt      60
cttgtgcaaa gtttgatgag gatgactctg agccagttct caagggtgtc aaattacatt      120
acacataaac gaacttatgg atttgtttat gagatttttt actcttggat caattactgc      180
acagccagta aaaattgaca atgcttctcc tgcaagtact gttcatgcta cagcaacgat      240
accgctacaa gcctcactcc ctttcggatg gcttgattatt ggcgttgcat ttcttgctgt      300
ttttcagagc gctaccacaaa taattgctgc caataaaaga tggcagctag ccctttataa      360
gggcttccag ttcatttgca atttactgct gctatttggc accatctatt cacatctttt      420
gcttgctgct gcaggtatgg aggcgcaatt tttgtacctc tatgccttga tatattttct      480
acaatgcatc aacgcatgta gaattattat gagatggtgg ctttgggtga agtgcaaatc      540
caagaaccca ttactttt atg atg oca act act ttg ttt gct ggc aca cac      590
                Met Met Pro Thr Thr Leu Phe Ala Gly Thr His
                1                    5                      10

ata act atg act act gta tac cat ata aca gtg tca cag ata caa ttg      638
Ile Thr Met Thr Thr Val Tyr His Ile Thr Val Ser Gln Ile Gln Leu
                15                    20                      25

tcg tta ctg aag gtg acg gca ttt caa cac caa aac tca aag aag act      686
Ser Leu Leu Lys Val Thr Ala Phe Gln His Gln Asn Ser Lys Lys Thr
                30                    35                      40

acc aaa ttg gtg gtt att ctg agg ata ggc act cag gtg tta aag act      734
Thr Lys Leu Val Val Ile Leu Arg Ile Gly Thr Gln Val Leu Lys Thr
                45                    50                      55

atg tcg ttg tac atg gct att tca ccg aag ttt act acc agc ttg agt      782
Met Ser Leu Tyr Met Ala Ile Ser Pro Lys Phe Thr Thr Ser Leu Ser
                60                    65                      70                      75

cta cac aaa tta cta cag aca ctg gta ttg aaa atg cta cat tct tca      830
Leu His Lys Leu Leu Gln Thr Leu Val Leu Lys Met Leu His Ser Ser
                80                    85                      90

tct tta aca agc ttg tta aag acc cac cga atg tgc aaa tac aca caa      878
Ser Leu Thr Ser Leu Leu Lys Thr His Arg Met Cys Lys Tyr Thr Gln
                95                    100                   105

tcg acg gct ctt cag gag ttg cta atc cag caa tgg atc caa ttt atg      926
Ser Thr Ala Leu Gln Glu Leu Leu Ile Gln Gln Trp Ile Gln Phe Met
                110                   115                   120

atg agc cga cga cga cta cta cgc tgc ctt tgt aag cac aag aaa gtg      974
Met Ser Arg Arg Arg Leu Leu Ala Cys Leu Cys Lys His Lys Lys Val
                125                   130                   135

agt acg aac tta tgt act cat tcg ttt cgg aag aaa cag gta cgt      1019
Ser Thr Asn Leu Cys Thr His Ser Phe Arg Lys Lys Gln Val Arg
                140                   145                   150

taatagttaa tagcgtactt cttttctctg ctttcgtggg attcttgcta gtcacactag      1079
ccatccttac tgcgctt      1096

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<210> SEQ ID NO 12
<211> LENGTH: 154
<212> TYPE: PRT

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<213> ORGANISM: CORONAVIRUS

<400> SEQUENCE: 12

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Met Met Pro Thr Thr Leu Phe Ala Gly Thr His Ile Thr Met Thr Thr
 1           5           10           15
Val Tyr His Ile Thr Val Ser Gln Ile Gln Leu Ser Leu Leu Lys Val
          20           25           30
Thr Ala Phe Gln His Gln Asn Ser Lys Lys Thr Thr Lys Leu Val Val
          35           40           45
Ile Leu Arg Ile Gly Thr Gln Val Leu Lys Thr Met Ser Leu Tyr Met
          50           55           60
Ala Ile Ser Pro Lys Phe Thr Thr Ser Leu Ser Leu His Lys Leu Leu
 65           70           75           80
Gln Thr Leu Val Leu Lys Met Leu His Ser Ser Ser Leu Thr Ser Leu
          85           90           95
Leu Lys Thr His Arg Met Cys Lys Tyr Thr Gln Ser Thr Ala Leu Gln
          100          105          110
Glu Leu Leu Ile Gln Gln Trp Ile Gln Phe Met Met Ser Arg Arg Arg
          115          120          125
Leu Leu Ala Cys Leu Cys Lys His Lys Lys Val Ser Thr Asn Leu Cys
          130          135          140
Thr His Ser Phe Arg Lys Lys Gln Val Arg
145           150

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<210> SEQ ID NO 13

<211> LENGTH: 332

<212> TYPE: DNA

<213> ORGANISM: CORONAVIRUS

<220> FEATURE:

<221> NAME/KEY: CDS

<222> LOCATION: (36)..(263)

<223> OTHER INFORMATION:

<400> SEQUENCE: 13

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tgcccttgta agcacaagaa agtgagtacg aactt atg tac tca ttc gtt tcg      53
                Met Tyr Ser Phe Val Ser
                1           5
gaa gaa aca ggt acg tta ata gtt aat agc gta ctt ctt ttt ctt gct      101
Glu Glu Thr Gly Thr Leu Ile Val Asn Ser Val Leu Leu Phe Leu Ala
          10           15           20
ttc gtg gta ttc ttg cta gtc aca cta gcc atc ctt act gcg ctt cga      149
Phe Val Val Phe Leu Leu Val Thr Leu Ala Ile Leu Thr Ala Leu Arg
          25           30           35
ttg tgt gcg tac tgc tgc aat att gtt aac gtg agt tta gta aaa cca      197
Leu Cys Ala Tyr Cys Cys Asn Ile Val Asn Val Ser Leu Val Lys Pro
          40           45           50
acg gtt tac gtc tac tcg cgt gtt aaa aat ctg aac tct tct gaa gga      245
Thr Val Tyr Val Tyr Ser Arg Val Lys Asn Leu Asn Ser Ser Glu Gly
          55           60           65           70
gtt cct gat ctt ctg gtc taaacgaact aactattatt attattctgt      293
Val Pro Asp Leu Leu Val
          75
ttggaacttt aacattgctt atcatggcag acaacggta      332

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<210> SEQ ID NO 14

<211> LENGTH: 76

<212> TYPE: PRT

<213> ORGANISM: CORONAVIRUS

<400> SEQUENCE: 14

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Met Tyr Ser Phe Val Ser Glu Glu Thr Gly Thr Leu Ile Val Asn Ser
 1 5 10 15
 Val Leu Leu Phe Leu Ala Phe Val Val Phe Leu Leu Val Thr Leu Ala
 20 25 30
 Ile Leu Thr Ala Leu Arg Leu Cys Ala Tyr Cys Cys Asn Ile Val Asn
 35 40 45
 Val Ser Leu Val Lys Pro Thr Val Tyr Val Tyr Ser Arg Val Lys Asn
 50 55 60
 Leu Asn Ser Ser Glu Gly Val Pro Asp Leu Leu Val
 65 70 75

<210> SEQ ID NO 15
 <211> LENGTH: 332
 <212> TYPE: DNA
 <213> ORGANISM: CORONAVIRUS

<400> SEQUENCE: 15

tgcccttgta agcacaagaa agtgagtagc aacttatgta ctcattcggt tcggaagaaa 60
 caggtagcgtt aatagttaat agcgtacttc tttttcttgc tttcgtggta ttcttgctag 120
 tcacactagc catccttact gcgcttcgat tgtgtgcgta ctgctgcaat attgttaacg 180
 tgagtttagt aaaaccaacg gtttacgtct actcgcgtgt taaaaatctg aactcttctg 240
 aaggagtcc tgatcttctg gtctaaca ga actaactatt attattattc tgtttggaac 300
 ttttaacattg cttatcatgg cagacaacgg ta 332

<210> SEQ ID NO 16
 <211> LENGTH: 708
 <212> TYPE: DNA
 <213> ORGANISM: CORONAVIRUS
 <220> FEATURE:
 <221> NAME/KEY: CDS
 <222> LOCATION: (41)..(703)
 <223> OTHER INFORMATION:

<400> SEQUENCE: 16

tattattatt attctgtttg gaactttaac attgcttacc atg gca gac aac ggt 55
 Met Ala Asp Asn Gly
 1 5
 act att acc gtt gag gag ctt aaa caa ctc ctg gaa caa tgg aac cta 103
 Thr Ile Thr Val Glu Glu Leu Lys Gln Leu Leu Glu Gln Trp Asn Leu
 10 15 20
 gta ata ggt ttc cta ttc cta gcc tgg att atg tta cta caa ttt gcc 151
 Val Ile Gly Phe Leu Phe Leu Ala Trp Ile Met Leu Leu Gln Phe Ala
 25 30 35
 tat tct aat cgg aac agg ttt ttg tac ata ata aag ctt gtt ttc ctc 199
 Tyr Ser Asn Arg Asn Arg Phe Leu Tyr Ile Ile Lys Leu Val Phe Leu
 40 45 50
 tgg ctc ttg tgg cca gta aca ctt gct tgt ttt gtg ctt gct gct gtc 247
 Trp Leu Leu Trp Pro Val Thr Leu Ala Cys Phe Val Leu Ala Ala Val
 55 60 65
 tac aga att aat tgg gtg act ggc ggg att gcg att gca atg gct tgt 295
 Tyr Arg Ile Asn Trp Val Thr Gly Gly Ile Ala Ile Ala Met Ala Cys
 70 75 80 85
 att gta ggc ttg atg tgg ctt agc tac ttc gtt gct tcc ttc agg ctg 343
 Ile Val Gly Leu Met Trp Leu Ser Tyr Phe Val Ala Ser Phe Arg Leu
 90 95 100
 ttt gct cgt acc cgc tca atg tgg tca ttc aac cca gaa aca aac att 391
 Phe Ala Arg Thr Arg Ser Met Trp Ser Phe Asn Pro Glu Thr Asn Ile
 105 110 115
 ctt ctc aat gtg cct ctc cgg ggg aca att gtg acc aga ccg ctc atg 439

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Leu Leu Asn Val Pro Leu Arg Gly Thr Ile Val Thr Arg Pro Leu Met
    120                125                130

gaa agt gaa ctt gtc att ggt gct gtg atc att cgt ggt cac ttg cga    487
Glu Ser Glu Leu Val Ile Gly Ala Val Ile Ile Arg Gly His Leu Arg
    135                140                145

atg gcc gga cac tcc cta ggg cgc tgt gac att aag gac ctg cca aaa    535
Met Ala Gly His Ser Leu Gly Arg Cys Asp Ile Lys Asp Leu Pro Lys
    150                155                160                165

gag atc act gtg gct aca tca cga acg ctt tct tat tac aaa tta gga    583
Glu Ile Thr Val Ala Thr Ser Arg Thr Leu Ser Tyr Tyr Lys Leu Gly
    170                175                180

gcg tcg cag cgt gta ggc act gat tca ggt ttt gct gca tac aac cgc    631
Ala Ser Gln Arg Val Gly Thr Asp Ser Gly Phe Ala Ala Tyr Asn Arg
    185                190                195

tac cgt att gga aac tat aaa tta aat aca gac cac gcc ggt agc aac    679
Tyr Arg Ile Gly Asn Tyr Lys Leu Asn Thr Asp His Ala Gly Ser Asn
    200                205                210

gac aat att gct ttg cta gta cag taagt    708
Asp Asn Ile Ala Leu Leu Val Gln
    215                220

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<210> SEQ ID NO 17
<211> LENGTH: 221
<212> TYPE: PRT
<213> ORGANISM: CORONAVIRUS

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<400> SEQUENCE: 17

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Met Ala Asp Asn Gly Thr Ile Thr Val Glu Glu Leu Lys Gln Leu Leu
 1                5                10                15

Glu Gln Trp Asn Leu Val Ile Gly Phe Leu Phe Leu Ala Trp Ile Met
    20                25                30

Leu Leu Gln Phe Ala Tyr Ser Asn Arg Asn Arg Phe Leu Tyr Ile Ile
    35                40                45

Lys Leu Val Phe Leu Trp Leu Leu Trp Pro Val Thr Leu Ala Cys Phe
    50                55                60

Val Leu Ala Ala Val Tyr Arg Ile Asn Trp Val Thr Gly Gly Ile Ala
    65                70                75                80

Ile Ala Met Ala Cys Ile Val Gly Leu Met Trp Leu Ser Tyr Phe Val
    85                90                95

Ala Ser Phe Arg Leu Phe Ala Arg Thr Arg Ser Met Trp Ser Phe Asn
    100                105                110

Pro Glu Thr Asn Ile Leu Leu Asn Val Pro Leu Arg Gly Thr Ile Val
    115                120                125

Thr Arg Pro Leu Met Glu Ser Glu Leu Val Ile Gly Ala Val Ile Ile
    130                135                140

Arg Gly His Leu Arg Met Ala Gly His Ser Leu Gly Arg Cys Asp Ile
    145                150                155                160

Lys Asp Leu Pro Lys Glu Ile Thr Val Ala Thr Ser Arg Thr Leu Ser
    165                170                175

Tyr Tyr Lys Leu Gly Ala Ser Gln Arg Val Gly Thr Asp Ser Gly Phe
    180                185                190

Ala Ala Tyr Asn Arg Tyr Arg Ile Gly Asn Tyr Lys Leu Asn Thr Asp
    195                200                205

His Ala Gly Ser Asn Asp Asn Ile Ala Leu Leu Val Gln
    210                215                220

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<210> SEQ ID NO 18
<211> LENGTH: 769

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<212> TYPE: DNA

<213> ORGANISM: CORONAVIRUS

<400> SEQUENCE: 18

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cctgatcttc tggctctaac gaactaacta ttattattat tctgtttga actttaacat    60
tgcttatcat ggcagacaac ggtactatta cgttgagga gcttaacaa ctctggaac    120
aatggaacct agtaataggt ttcctattcc tagcctggat tatgttacta caatttgcc    180
attctaateg gaacagggtt ttgtacataa taaagcttgt tttcctctgg ctcttggtgc    240
cagtaaacct tgcttgtttt gtgcttctg ctgtctacag aattaattgg gtgactggcg    300
ggattgcatg tgcaatggct tgtattgtag gcttgatgtg gcttagctac ttcggtgctt    360
ccttcagget gtttgctcgt acccgctcaa tgtggtcatt caaccagaa acaaacattc    420
ttctcaatgt gcctctccgg gggacaattg tgaccagacc gctcatggaa agtgaacttg    480
tcattggtgc tgtgatcatt cgtggtcact tgcgaatggc cggacactcc ctagggcgct    540
gtgacattaa ggacctgcca aaagagatca ctgtggctac atcacgaacg ctttcttatt    600
acaattagg agcgtcgcag cgtgtaggca ctgattcagg ttttctgca tacaaccgct    660
accgtattgg aaactataaa ttaaatacag accacgcccg tagcaacgac aatattgctt    720
tgctagtaca gtaagtgaca acagatggtt catcttgttg acttccagg                769

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<210> SEQ ID NO 19

<211> LENGTH: 1231

<212> TYPE: DNA

<213> ORGANISM: CORONAVIRUS

<400> SEQUENCE: 19

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taccgtattg gaaactataa attaaataca gaccacgccg gtagcaacga caatattgct    60
ttgctagtac agtaagtgac aacagatggt tcatcttgtt gacttccagg ttacaatagc    120
agagatattg attatcatta tgaggacttt caggattgct atttggaaac ttgacgttat    180
aataagtcca atagtgagac aattatttaa gcctctaact aagaagaatt attcggagtt    240
agatgatgaa gaacctatgg agttagatta tocataaac gaacatgaaa attattctct    300
tcctgacatt gattgtatct acatcttgcg agctatatca ctatcaggag tgtgtagag    360
gtacgactgt actactaaaa gaaccttgcc catcaggaac atacgagggc aattcaccat    420
ttcaccctct tgctgacaat aaatttgcac taacttgcac tagcacacac tttgcttttg    480
cttgctgta cggtactcga catacctatc agctgcgtgc aagatcagtt tcacaaaaac    540
ttttcatcag acaagaggag gttcaacaag agctctactc gccacttttt ctctattgtg    600
ctgctctagt atttttaata ctttgcttca ccattaagag aaagacagaa tgaatgagct    660
cactttaatt gacttctatt tgtgcttttt agcctttctg ctattccttg ttttaataat    720
gcttattata ttttggtttt cactcgaat ccaggatcta gaagaacctt gtaccaaggt    780
ctaaacgaac atgaaacttc tcattgtttt gacttgtatt tctctatgca gttgcatatg    840
cactgtagta cagcgtgtg catctaataa acctcatgtg cttgaagatc cttgtaaggt    900
acaacactag gggtaatact tatagcactg cttggctttg tgctctagga aagggtttac    960
cttttcatag atggcacact atggttcaaa catgcacacc taatgttact atcaactgtc    1020
aagatccagc tgggtgtgct cttatageta ggtgttgta ccttcatgaa ggtcaccaaa    1080
ctgctgcatt tagagacgta cttgtgtttt taaataaacg aacaaattaa aatgtctgat    1140
aatggacccc aatcaaacca acgtagtgcc ccccgatta catttgggtg acccacagat    1200
tcaactgaca ataaccagaa tggaggacgc a                1231

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atg gag tta gat tat cca taaaacgaac atgaaaatta ttctcttctct      304
Met Glu Leu Asp Tyr Pro
  60

gacattgatt gtatttaccat cttgcgagct atatcactat caggagtgtg ttagaggtag      364
gactgtacta ctaaaagaac cttgcccatc aggaacatac gagggcaatt caccatttca      424
ccctcttgct gacaataaat ttgcactaac ttgcactagc acacactttg cttttgcttg      484
tgctgacggg actcgacata cctatcagct gcgtgcaaga tcagtttcac caaaaacttt      544
catcagacaa gaggaggttc aacaagagct ctactcgcca ctttttctca ttgttgctgc      604
tctagtattt ttaactcttt gcttcacatc taagagaaag acagaatgaa tgagctcact      664
ttaattgact tctatttggt ctttttagcc tttctgctat tccttgtttt aataatgctt      724
attatatttt ggttttctc cgaatccag gatctagaag aaccttgtag caaagtctaa      784
acgaacatga aacttctcat tgttttgact tgtatttctc tatgcagttg catatgcact      844
gtagtacagc gctgtgcatc taataaacct catgtgcttg aagatccttg taaggtaaaa      904
cactaggggt aatacttata gcaactgctg gctttgtgct ctaggaaaagg ttttaccttt      964
tcatagatgg cacactatgg ttcaaacatg cacacctaact gttactatca actgtcaaga     1024
tccagctggt ggtgcgctta tagctagggt ttggtacctt catgaagggt accaaaactgc     1084
tgcatttaga gacgtacttg ttgttttaaa taaacgaaca aattaaaatg tctgataatg     1144
gacccaatc aaaccaacgt agtgcccccc gcattacatt tgggtggaccc acagattcaa     1204
ctgacaataa ccagaatgga ggacgca                                           1231

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<210> SEQ ID NO 22
<211> LENGTH: 63
<212> TYPE: PRT
<213> ORGANISM: CORONAVIRUS

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<400> SEQUENCE: 22

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Met Phe His Leu Val Asp Phe Gln Val Thr Ile Ala Glu Ile Leu Ile
 1             5             10             15

Ile Ile Met Arg Thr Phe Arg Ile Ala Ile Trp Asn Leu Asp Val Ile
          20             25             30

Ile Ser Ser Ile Val Arg Gln Leu Phe Lys Pro Leu Thr Lys Lys Asn
          35             40             45

Tyr Ser Glu Leu Asp Asp Glu Glu Pro Met Glu Leu Asp Tyr Pro
          50             55             60

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<210> SEQ ID NO 23
<211> LENGTH: 1231
<212> TYPE: DNA
<213> ORGANISM: CORONAVIRUS
<220> FEATURE:
<221> NAME/KEY: CDS
<222> LOCATION: (285)..(650)
<223> OTHER INFORMATION:

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<400> SEQUENCE: 23

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ttgctagtac agtaagttag aacagatggt tcatcttggt gacttccagg ttacaatagc     120
agagatattg attatcatta tgaggacttt caggattgct atttggaaac ttgacgttat     180
aataagttca atagttagac aattatntaa gcctctaact aagaagaatt attcggagtt     240
agatgatgaa gaacctatgg agttagatta tccataaaac gaac atg aaa att att     296
Met Lys Ile Ile
 1

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ctc ttc ctg aca ttg att gta ttt aca tct tgc gag cta tat cac tat    344
Leu Phe Leu Thr Leu Ile Val Phe Thr Ser Cys Glu Leu Tyr His Tyr
5          10          15          20

cag gag tgt gtt aga ggt acg act gta cta cta aaa gaa cct tgc cca    392
Gln Glu Cys Val Arg Gly Thr Thr Val Leu Leu Lys Glu Pro Cys Pro
          25          30          35

tca gga aca tac gag ggc aat tca cca ttt cac cct ctt gct gac aat    440
Ser Gly Thr Tyr Glu Gly Asn Ser Pro Phe His Pro Leu Ala Asp Asn
          40          45          50

aaa ttt gca cta act tgc act agc aca cac ttt gct ttt gct tgt gct    488
Lys Phe Ala Leu Thr Cys Thr Ser Thr His Phe Ala Phe Ala Cys Ala
          55          60          65

gac ggt act cga cat acc tat cag ctg cgt gca aga tca gtt tca cca    536
Asp Gly Thr Arg His Thr Tyr Gln Leu Arg Ala Arg Ser Val Ser Pro
          70          75          80

aaa ctt ttc atc aga caa gag gag gtt caa caa gag ctc tac tcg cca    584
Lys Leu Phe Ile Arg Gln Glu Glu Val Gln Gln Glu Leu Tyr Ser Pro
85          90          95          100

ctt ttt ctc att gtt gct gct cta gta ttt tta ata ctt tgc ttc acc    632
Leu Phe Leu Ile Val Ala Ala Leu Val Phe Leu Ile Leu Cys Phe Thr
          105          110          115

att aag aga aag aca gaa tgaatgagct cactttaatt gacttctatt    680
Ile Lys Arg Lys Thr Glu
          120

tgtgtctttt agcctttctg ctattccttg ttttaataat gcttattata ttttggtttt    740

cactcgaaat ccaggateta gaagaacctt gtaccaaagt ctaaacgaac atgaaacttc    800

tcattgtttt gacttgatt tctctatgca gttgcatatg cactgtagta cagcgtgtg    860

catctaataa acctcatgtg cttgaagatc cttgtaagggt acaacactag gggtaatact    920

tatagcactg cttgctttg tgctctagga aaggttttac cttttcatag atggcacact    980

atggttcaaa catgcacacc taatgttact atcaactgtc aagatccagc tgggtgtgcg    1040

cttatagcta ggtgttgta ccttcatgaa ggtcacaaa ctgctgcatt tagagacgta    1100

cttggtgttt taataaacg aacaaattaa aatgtctgat aatggacccc aatcaaacca    1160

acgtagtgcc ccccgatta ctttggtgg acccacagat tcaactgaca ataaccagaa    1220

tggaggacgc a    1231

<210> SEQ ID NO 24
<211> LENGTH: 122
<212> TYPE: PRT
<213> ORGANISM: CORONAVIRUS

<400> SEQUENCE: 24
Met Lys Ile Ile Leu Phe Leu Thr Leu Ile Val Phe Thr Ser Cys Glu
1          5          10          15

Leu Tyr His Tyr Gln Glu Cys Val Arg Gly Thr Thr Val Leu Leu Lys
20          25          30

Glu Pro Cys Pro Ser Gly Thr Tyr Glu Gly Asn Ser Pro Phe His Pro
35          40          45

Leu Ala Asp Asn Lys Phe Ala Leu Thr Cys Thr Ser Thr His Phe Ala
50          55          60

Phe Ala Cys Ala Asp Gly Thr Arg His Thr Tyr Gln Leu Arg Ala Arg
65          70          75          80

Ser Val Ser Pro Lys Leu Phe Ile Arg Gln Glu Glu Val Gln Gln Glu
85          90          95

Leu Tyr Ser Pro Leu Phe Leu Ile Val Ala Ala Leu Val Phe Leu Ile

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100	105	110	
Leu Cys Phe Thr Ile Lys Arg Lys Thr Glu			
115	120		
<210> SEQ ID NO 25			
<211> LENGTH: 1231			
<212> TYPE: DNA			
<213> ORGANISM: CORONAVIRUS			
<220> FEATURE:			
<221> NAME/KEY: CDS			
<222> LOCATION: (650)..(781)			
<223> OTHER INFORMATION:			
<400> SEQUENCE: 25			
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agagatattg attatcatta tgaggacttt caggattgct atttggaaac ttgacgttat			180
aataagttca atagtggagac aattatntaa gcctctaact aagaagaatt attcggagtt			240
agatgatgaa gaacctatgg agttagatta tccataaaac gaacatgaaa attattctct			300
tcctgacatt gattgtatnt acatcttgcg agctatatca ctatcaggag tgtgtagag			360
gtacgactgt actactaaaa gaaccttgcc catcaggaac atacgagggc aattcaccat			420
ttcacctct tgctgacaat aaatttgac taacttgac tagcacacac tttgcttttg			480
cttgctga cggctactga catacctatc agctgcctgc aagatcagtt tcacaaaaac			540
ttttcatcag acaagaggag gtccaacaag agctctactc gccacttttt ctctattgtg			600
ctgctctagt atttttaata ctttgcttca ccattaagag aaagacaga atg aat gag			658
		Met Asn Glu	
		1	
ctc act tta att gac ttc tat ttg tgc ttt tta gcc ttt ctg cta ttc			706
Leu Thr Leu Ile Asp Phe Tyr Leu Cys Phe Leu Ala Phe Leu Leu Phe			
5	10	15	
ctt gtt tta ata atg ctt att ata ttt tgg ttt tca ctc gaa atc cag			754
Leu Val Leu Ile Met Leu Ile Ile Phe Trp Phe Ser Leu Glu Ile Gln			
20	25	30	35
gat cta gaa gaa cct tgt acc aaa gtc taaacgaaca tgaaacttct			801
Asp Leu Glu Glu Pro Cys Thr Lys Val			
40			
cattgttttg acttgtatnt ctctatgcag ttgcatatgc actgtagtac agcgtgtgc			861
atctaataaa cctcatgtgc ttgaagatcc ttgtaaggta caacactagg ggtaatactt			921
atagcactgc ttggctttgt gctctaggaa aggttttacc ttttcataga tggcacacta			981
tggttcaaac atgcacacct aatgttacta tcaactgtca agatccagct ggtgggtgcgc			1041
ttatagctag gtggttggtac cttcatgaaag gtcaccaaac tgctgcattt agagacgtac			1101
ttggtgtttt aaataaacga acaaatataa atgtctgata atggacccca atcaaaccaa			1161
cgtagtgccc cccgcattac atttgggtgga cccacagatt caactgacaa taaccagaat			1221
ggaggacgca			1231
<210> SEQ ID NO 26			
<211> LENGTH: 44			
<212> TYPE: PRT			
<213> ORGANISM: CORONAVIRUS			
<400> SEQUENCE: 26			
Met Asn Glu Leu Thr Leu Ile Asp Phe Tyr Leu Cys Phe Leu Ala Phe			
1	5	10	15

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Leu Leu Phe Leu Val Leu Ile Met Leu Ile Ile Phe Trp Phe Ser Leu
 20 25 30

Glu Ile Gln Asp Leu Glu Glu Pro Cys Thr Lys Val
 35 40

<210> SEQ ID NO 27
 <211> LENGTH: 1231
 <212> TYPE: DNA
 <213> ORGANISM: CORONAVIRUS
 <220> FEATURE:
 <221> NAME/KEY: CDS
 <222> LOCATION: (791) .. (907)
 <223> OTHER INFORMATION:

<400> SEQUENCE: 27

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 agagatattg attatcatta tgaggacttt caggattgct atttggaaac ttgacgttat 180
 aataagttca atagtgcagac aattatntaa gcctctaact aagaagaatt attcggagtt 240
 agatgatgaa gaacctatgg agttagatta tccataaaac gaacatgaaa attattctct 300
 tcctgacatt gattgtatnt acatcttgcg agctatatca ctatcaggag tgtgtagtag 360
 gtacgactgt actactaaaa gaaccttgcc catcaggaac atacgagggc aattcaccat 420
 ttcaccctct tgctgacaat aaatttgcac taacttgcac tagcacacac tttgcttttg 480
 cttgtgctga cggctactcga catacctatc agctgcgtgc aagatcagtt tcacaaaaac 540
 ttttcatcag acaagaggag gttcaacaag agctctactc gccacttttt ctctattggtg 600
 ctgctctagt atttttaata ctttgcttca ccattaagag aaagacagaa tgaatgagct 660
 cactttaatt gacttctatt tgtgcttttt agcctttctg ctattccttg ttttaataat 720
 gcttattata ttttggtttt cactcgaat ccaggatcta gaagaacctt gtaccaaagt 780
 ctaaacgaac atg aaa ctt ctc att gtt ttg act tgt att tct cta tgc 829
 Met Lys Leu Leu Ile Val Leu Thr Cys Ile Ser Leu Cys
 1 5 10
 agt tgc ata tgc act gta gta cag cgc tgt gca tct aat aaa cct cat 877
 Ser Cys Ile Cys Thr Val Val Gln Arg Cys Ala Ser Asn Lys Pro His
 15 20 25
 gtg ctt gaa gat cct tgt aag gta caa cac taggggtaatt acttatagca 927
 Val Leu Glu Asp Pro Cys Lys Val Gln His
 30 35
 ctgcttggtt ttgtgctcta ggaaagggtt taccttttca tagatggcac actatgggtc 987
 aaacatgcac acctaagtgt actatcaact gtcaagatcc agctggtggt gcgcttatag 1047
 ctagggtgtg gtaccttcat gaaggtcacc aaactgctgc atttagagac gtacttggtg 1107
 ttttaataaa acgaacaaat taaaatgtct gataatggac cccaatcaaa ccaacgtagt 1167
 gccccccgca ttacatttgg tggacccaca gattcaactg acaataacca gaatggagga 1227
 cgca 1231

<210> SEQ ID NO 28
 <211> LENGTH: 39
 <212> TYPE: PRT
 <213> ORGANISM: CORONAVIRUS

<400> SEQUENCE: 28

Met Lys Leu Leu Ile Val Leu Thr Cys Ile Ser Leu Cys Ser Cys Ile
 1 5 10 15

Cys Thr Val Val Gln Arg Cys Ala Ser Asn Lys Pro His Val Leu Glu

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20	25	30	
Asp Pro Cys Lys Val Gln His			
35			
<210> SEQ ID NO 29			
<211> LENGTH: 1231			
<212> TYPE: DNA			
<213> ORGANISM: CORONAVIRUS			
<220> FEATURE:			
<221> NAME/KEY: CDS			
<222> LOCATION: (876)..(1127)			
<223> OTHER INFORMATION:			
<400> SEQUENCE: 29			
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agagatattg attatcatta tgaggacttt caggattgct atttggaaac ttgacgttat			180
aataagttca atagtggagac aattatntaa gcctctaact aagaagaatt attcggagtt			240
agatgatgaa gaacctatgg agttagatta tccataaaac gaacatgaaa attattctct			300
tcctgacatt gattgtatnt acatcttgcg agctatatca ctatcaggag tgtgtagag			360
gtacgactgt actactaaaa gaaccttgcc catcaggaac atacgagggc aattcaccat			420
ttcacctct tgctgacaat aaatttgac taacttgac tagcacacac tttgcttttg			480
cttgctga cggctactga catacctatc agctgcgtgc aagatcagtt tcacaaaaac			540
ttttcatcag acaagaggag gttcaacaag agctctactc gccacttttt ctctattgtg			600
ctgctctagt atttttaata ctttgcttca ccattaagag aaagacagaa tgaatgagct			660
cactttaatt gacttctatt tgtgcttttt agcctttctg ctattccttg ttttaataat			720
gcttattata ttttggtttt cactcgaaat ccaggatcta gaagaacctt gtaccaaagt			780
ctaaacgaac atgaaacttc tcattgcttt gacttgatt tctctatgca gttgcatatg			840
cactgtagta cagcgtgtg catctaataa acctc atg tgc ttg aag atc ctt			893
		Met Cys Leu Lys Ile Leu	
		1 5	
gta agg tac aac act agg ggt aat act tat agc act gct tgg ctt tgt			941
Val Arg Tyr Asn Thr Arg Gly Asn Thr Tyr Ser Thr Ala Trp Leu Cys			
	10	15	20
gct cta gga aag gtt tta cct ttt cat aga tgg cac act atg gtt caa			989
Ala Leu Gly Lys Val Leu Pro Phe His Arg Trp His Thr Met Val Gln			
	25	30	35
aca tgc aca cct aat gtt act atc aac tgt caa gat cca gct ggt ggt			1037
Thr Cys Thr Pro Asn Val Thr Ile Asn Cys Gln Asp Pro Ala Gly Gly			
	40	45	50
gcg ctt ata gct agg tgt tgg tac ctt cat gaa ggt cac caa act gct			1085
Ala Leu Ile Ala Arg Cys Trp Tyr Leu His Glu Gly His Gln Thr Ala			
	55	60	65
			70
gca ttt aga gac gta ctt gtt gtt tta aat aaa cga aca aat			1127
Ala Phe Arg Asp Val Leu Val Val Leu Asn Lys Arg Thr Asn			
	75	80	
taaaatgtct gataatggac cccaatcaaa ccaacgtagt gcccccgca ttacatttgg			1187
tggaccacaca gattcaactg acaataacca gaatggagga cgca			1231

<210> SEQ ID NO 30

<211> LENGTH: 84

<212> TYPE: PRT

<213> ORGANISM: CORONAVIRUS

<400> SEQUENCE: 30

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Met Cys Leu Lys Ile Leu Val Arg Tyr Asn Thr Arg Gly Asn Thr Tyr
 1 5 10 15
 Ser Thr Ala Trp Leu Cys Ala Leu Gly Lys Val Leu Pro Phe His Arg
 20 25 30
 Trp His Thr Met Val Gln Thr Cys Thr Pro Asn Val Thr Ile Asn Cys
 35 40 45
 Gln Asp Pro Ala Gly Gly Ala Leu Ile Ala Arg Cys Trp Tyr Leu His
 50 55 60
 Glu Gly His Gln Thr Ala Ala Phe Arg Asp Val Leu Val Val Leu Asn
 65 70 75 80
 Lys Arg Thr Asn

<210> SEQ ID NO 31

<211> LENGTH: 21221

<212> TYPE: DNA

<213> ORGANISM: CORONAVIRUS

<400> SEQUENCE: 31

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 gaggcacgtg aacacctcaa aaatggcact tgtggtctag tagagctgga aaaaggcgta 180
 ctgccccage ttgaacagcc ctatgtgttc attaaacggt ctgatgcctt aagcaccaat 240
 cacggccaca aggtcgttga gctggttgca gaaatggacg gcattcagta cggtcgtagc 300
 ggtataaacac tgggagtact cgtgccacat gtgggcgaaa cccaattgc ataccgcaat 360
 gttcttcttc gtaagaacgg taataaggga gccggtggtc atagctatgg catcgatcta 420
 aagtcttatg acttagtgta cgagcttggc actgatccca ttgaagatta tgaacaaaac 480
 tggaaacta agcatggcag tggtgcactc cgtgaactca ctctgagct caatggaggt 540
 gcagtcactc gctatgtcga caacaatttc tgtggcccag atgggtaccc tcttgattgc 600
 atcaaagatt ttctcgcacg cgcgggcaag tcaatgtgca ctctttccga acaacttgat 660
 tacatcgagt cgaagagagg tgtctactgc tgccgtgacc atgagcatga aattgcctgg 720
 ttactgagc gctctgataa gagctacgag caccagacac ccttcgaaat taagagtgcc 780
 aagaaatttg acactttcaa aggggaatgc ccaaagtttg tgtttcctct taactcaaaa 840
 gtcaaagtca ttcaaccacg tgttgaaaag aaaaagactg agggtttcat ggggcgtata 900
 cgtctgtgtg accctgttgc atctccacag gagtgtaaca atatgcactt gtctaccttg 960
 atgaaatgta atcattgcga tgaagtttca tggcagacgt gcgactttct gaaagccact 1020
 tgtgaaacatt gtggcactga aaatttagtt attgaaggac ctactacatg tgggtaccta 1080
 cctactaatg ctgtagttaa aatgccatgt cctgcctgtc aagaccaga gattggacct 1140
 gagcatagtg ttgcagatta tcacaaccac tcaaacattg aaactcgact ccgcaaggga 1200
 ggtaggacta gatgttttgg aggctgtgtg tttgcctatg ttggctgcta taataagcgt 1260
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 tctgcttcta caagtgcctt tattgacct ataaagagtc ttgattacaa gtctttcaaa 1500
 accattgttg agtctcgcgg taactataaa gttaccaagg gaaagcccgt aaaagtgct 1560
 tggaaacattg gacaacagag atcagtttta acaccactgt gtggttttcc ctacaggct 1620

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gctgggtgta	tcagatcaat	ttttgogcgc	acacttgatg	cagcaaacca	ctcaattcct	1680
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ttctcaagg	atgcttggga	gattctcaaa	tttctcatta	cagggtgttt	tgacatcgtc	1980
aagggtcaaa	tacaggttgc	ttcagataac	atcaaggatt	gtgtaaaatg	cttcattgat	2040
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cgatcactca	acttagtgta	agtcttcac	gctcaaagca	agggacttta	ccgtcagtgt	2160
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acaccagtct	gtgtaaatgg	cctcatgctc	ttagagatta	aggacaaaga	acaactctgc	2400
gcattgtctc	ctggtttact	ggctacaaac	aatgtcttcc	gcttaaaagg	gggtgcacca	2460
attaaggtg	taacctttgg	agaagatact	gtttgggaag	ttcaaggtta	caagaatgtg	2520
agaatcacat	ttgagcttga	tgaacgtgtt	gacaaagtgc	ttaatgaaaa	gtgctctgtc	2580
tacactgtyt	aatccggtag	cgaagttact	gagtttgc	gtgtttagc	agaggctggt	2640
gtgaagactt	tacaaccagt	ttctgatctc	cttaccaaca	tgggtatgta	tcttgatgag	2700
tggagtgtag	ctacattcta	cttatttgat	gatgctggty	aagaaaactt	ttcatcacgt	2760
atgtattgty	ccttttacc	tccagatgag	gaagaagagg	acgatgcaga	gtgtgaggaa	2820
gaagaaattg	atgaaacctg	tgaacatgag	tacggtacag	aggatgatta	tcaaggtctc	2880
cctctggaat	ttggtycctc	agctgaaaca	gttcgagtyt	aggaaaga	agaggaagac	2940
tggctggatg	atactactga	gcaatcagag	attgagccag	aaccagaacc	tacacctgaa	3000
gaaccagtta	atcagtttac	tggttattta	aaacttactg	acaatgttgc	cattaaatgt	3060
gttgacatcg	ttaaggagcc	acaaagtctc	aatcctatgg	tgattgtaaa	tgctgctaac	3120
atacacctga	aacatggtgg	tggtytagca	ggtgcactca	acaaggcaac	caatggtgcc	3180
atgcaaaagg	agagtgatga	ttacattaag	ctaaatggcc	ctcttacagt	aggaggtctt	3240
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aattagtgtt gtctgttaat ccctatgttt gcaatgcccc aggttgtgat gtcactgatg 16080
tgacacaact gtatctagga ggtatgagct attattgcaa gtcacataag cctcccatta 16140
gttttccatt atgtgtcaat ggtcaggttt ttggtttata caaaaacaca tgtgtaggca 16200
gtgacaatgt cactgacttc aatgcatag caacatgtga ttggactaat gctggcgatt 16260
acatacttgc caacacttgt actgagagac tcaagctttt cgcagcagaa acgctcaaag 16320
ccactgagga aacatttaag ctgtcatatg gtattgccac tgtacgcgaa gtactctctg 16380
acagagaatt gcatctttca tgggaggttg gaaaacctag accaccattg aacagaaact 16440
atgtctttac tggttaccgt gtaactaaaa atagttaaagt acagattgga gagtacacct 16500
ttgaaaaagg tgactatggt gatgtgttg tgtacagagg tactacgaca tacaagtga 16560
atgttggtga ttactttgtg ttgacatctc aactgttaat gccacttagt gcacctactc 16620
tagtgccaca agagcactat gtgagaatta ctggcttcta ccaacactc aacatctcag 16680
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tcaagattgg acctgaaaga acgtgtgtgc tgtgtgacaa acgtgcaact tgcttttcta 18360
cttcacaga tacttatgcc tgctggaatc attctgtggg ttttgaactat gtctataacc 18420

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catttatgat	tgatgttcag	cagtggggct	ttacgggtaa	ccttcagagt	aaccatgacc	18480
aacattgcca	ggtacatgga	aatgcacatg	tggttagttg	tgatgctatc	atgactagat	18540
gtttagcagt	ccatgagtgc	tttgtaagc	gogttgattg	gtctgtttaa	tacctatta	18600
taggagatga	actgaggggt	aattctgctt	gcagaaaagt	acaacacatg	gttgtgaagt	18660
ctgcattgct	tgctgataag	ttccagttc	ttcatgacat	tggaaatcca	aaggctatca	18720
agtggtgcc	tcaggctgaa	gtagaatgga	agttctacga	tgctcagcca	tgtagtgaca	18780
aagcttaca	aatagaggaa	ctctctatt	cttatgctac	acatcacgat	aaattcactg	18840
atggtgtttg	ttgttttgg	aattgtaacg	ttgatcgtta	cccagccaat	gcaattgtgt	18900
gtaggtttga	cacaagatc	ttgtcaaact	tgaacttacc	aggctgtgat	ggtggtagtt	18960
tgatgtgaa	taagcatgca	ttccacactc	cagctttcga	taaaagtgca	tttactaatt	19020
taaagcaatt	gcctttcttt	tactattctg	atagtccttg	tgagtctcat	ggcaacaag	19080
tagtgtcgga	tattgattat	gttccactca	aatctgctac	gtgtattaca	cgatgcaatt	19140
taggtggtgc	tgtttcgaga	caccatgcaa	atgagtaccg	acagtacttg	gatgcatata	19200
atatgatgat	ttctgctgga	tttagcctat	ggatttaca	acaatttgat	acttataacc	19260
tgtggaatac	atttaccagg	ttacagagtt	tagaaaatgt	ggcttataat	gttgttaata	19320
aaggacactt	tgatggacac	gccggcgaag	cacctgttcc	catcattaat	aatgctgttt	19380
acacaaaggt	agatggtatt	gatgtggaga	tctttgaaaa	taagacaaca	cttctgttta	19440
atgttgcat	tgagctttgg	gctaagcgta	acattaaacc	agtgccagag	attaagatac	19500
tcaataattt	gggtgttgat	atcgtcgcta	atactgtaat	ctgggactac	aaaagagaag	19560
ccccagcaca	tgatcttaca	ataggtgtct	gcacaatgac	tgacattgcc	aagaaacctt	19620
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acctttttag	aaacgcccgt	aatggtgttt	taataacaga	aggttcagtc	aaaggtctaa	19740
caccttcaaa	gggaccagca	caagctagcg	tcaatggagt	cacattaatt	ggagaatcag	19800
taaaaacaca	gtttaactac	tttaagaaag	tagacggcat	tattcaacag	ttgctgaaa	19860
cctactttac	tcagagcaga	gacttagagg	attttaagcc	cagatcaca	atggaaactg	19920
actttctcga	gctcgtatg	gatgaattca	tacagcgata	taagctcgag	ggctatgcct	19980
tcgaacacat	cgtttatgga	gatttcagtc	atggacaact	tggcggctct	catttaatga	20040
taggcttagc	caagcgtcca	caagattcac	cacttaaatt	agaggatttt	atcctatgg	20100
acagcacagt	gaaaaattac	ttcataacag	atgcgcaaac	aggttcatca	aaatgtgtgt	20160
gttctgtgat	tgatctttta	cttgatgact	ttgtcgagat	aataaagtca	caagatttgt	20220
cagtgatctc	aaaagtggtc	aaggttacaa	ttgactatgc	tgaaatttca	ttcatgcttt	20280
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aaccaggtgt	tcgcatgcct	aacttgatca	agatgcaaag	aatgcttctt	gaaaagtgtg	20400
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agtatactca	actgtgtcaa	tacttaata	cacttacttt	agctgtaccc	tacaacatga	20520
gagtatttca	ctttggtgct	ggctctgata	aaggagttgc	accaggtaca	gctgtgctca	20580
gacaatgggt	gccaaactgc	acactacttg	tcgattcaga	tcttaatgac	ttcgtctccg	20640
acgcagatc	tactttaatt	ggagactgtg	caacagtaca	tacggctaat	aaatgggacc	20700
ttattattag	cgatattgat	gaccctagga	ccaaacatgt	gacaaaagag	aatgactcta	20760
aagaaggggt	ttcacttat	ctgtgtggat	ttataaagca	aaaactagcc	ctgggtgggt	20820

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ctatagctgt aaagataaca gagcattctt ggaatgctga cctttacaag cttatgggcc 20880
atctctcatg gtggacagct tttgttacia atgtaaatgc atcatcatcg gaagcatttt 20940
taattggggc taactatctt ggcaagccga aggaacaaat tgatggctat accatgcatg 21000
ctaactacat tttctggagg aacacaaaac ctatccagtt gtcttcctat tcaactctttg 21060
acatgagcaa atttctcttt aaattaagag gaactgctgt aatgtctctt aaggagaate 21120
aatcaatga tatgatttat tctctcttgg aaaaaggtag gcttatcatt agagaaaaca 21180
acagagttgt ggtttcaagt gatattcttg ttaacaacta a 21221

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<210> SEQ ID NO 32
<211> LENGTH: 297
<212> TYPE: DNA
<213> ORGANISM: CORONAVIRUS

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<400> SEQUENCE: 32

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atggacccca atcaaaccaa cgtagtgtccc cccgcattac atttgggtgga cccacagatt 60
caactgacaa taaccagaat ggaggacgca atggggcaag gccaaaacag cgccgacccc 120
aagggttacc caataaact gcgtcttgggt tcacagctct cactcagcat ggcaaggagg 180
aacttagatt ccctcgagge cagggogttc caatcaacac caatagtgggt ccagatgacc 240
aaattggcta ctaccgaaga gctaccgcac gagttcgtgg tggtagcggc aaaatga 297

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<210> SEQ ID NO 33
<211> LENGTH: 98
<212> TYPE: PRT
<213> ORGANISM: CORONAVIRUS

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<400> SEQUENCE: 33

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```

Met Asp Pro Asn Gln Thr Asn Val Val Pro Pro Ala Leu His Leu Val
1           5           10           15
Asp Pro Gln Ile Gln Leu Thr Ile Thr Arg Met Glu Asp Ala Met Gly
20           25           30
Gln Gly Gln Asn Ser Ala Asp Pro Lys Val Tyr Pro Ile Ile Leu Arg
35           40           45
Leu Gly Ser Gln Leu Ser Leu Ser Met Ala Arg Arg Asn Leu Asp Ser
50           55           60
Leu Glu Ala Arg Ala Phe Gln Ser Thr Pro Ile Val Val Gln Met Thr
65           70           75           80
Lys Leu Ala Thr Thr Glu Glu Leu Pro Asp Glu Phe Val Val Val Thr
85           90           95

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Ala Lys

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<210> SEQ ID NO 34
<211> LENGTH: 213
<212> TYPE: DNA
<213> ORGANISM: CORONAVIRUS

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<400> SEQUENCE: 34

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```

atgctgccac cgtgctacaa cttcctcaag gaacaacatt gccaaaaggc ttctacgcag 60
agggaaagcag agggcgagcgt caagcctctt ctcgctcttc atcacgtagt cgcggtaatt 120
caagaaattc aactcctggc agcagtaggg gaaattctcc tgctcgaatg gctagcggag 180
gtggtgaaac tgccctcgcg ctattgctgc tag 213

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<210> SEQ ID NO 35
<211> LENGTH: 70
<212> TYPE: PRT

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<213> ORGANISM: CORONAVIRUS

<400> SEQUENCE: 35

```

Met Leu Pro Pro Cys Tyr Asn Phe Leu Lys Glu Gln His Cys Gln Lys
1           5           10           15
Ala Ser Thr Gln Arg Glu Ala Glu Ala Ala Val Lys Pro Leu Leu Ala
20           25           30
Pro His His Val Val Ala Val Ile Gln Glu Ile Gln Leu Leu Ala Ala
35           40           45
Val Gly Glu Ile Leu Leu Leu Glu Trp Leu Ala Glu Val Val Lys Leu
50           55           60
Pro Ser Arg Tyr Cys Cys
65           70

```

<210> SEQ ID NO 36

<211> LENGTH: 1377

<212> TYPE: DNA

<213> ORGANISM: CORONAVIRUS

<220> FEATURE:

<221> NAME/KEY: CDS

<222> LOCATION: (67)..(1335)

<223> OTHER INFORMATION:

<400> SEQUENCE: 36

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atgaagggtca ccaaactgct gcatttagag acgtacttgt tgttttaaat aaacgaacaa      60
attaaa atg tct gat aat gga ccc caa tca aac caa cgt agt gcc ccc      108
  Met Ser Asp Asn Gly Pro Gln Ser Asn Gln Arg Ser Ala Pro
  1           5           10
cgc att aca ttt ggt gga ccc aca gat tca act gac aat aac cag aat      156
Arg Ile Thr Phe Gly Gly Pro Thr Asp Ser Thr Asp Asn Asn Gln Asn
15           20           25           30
gga gga cgc aat ggg gca agg cca aaa cag cgc cga ccc caa ggt tta      204
Gly Gly Arg Asn Gly Ala Arg Pro Lys Gln Arg Arg Pro Gln Gly Leu
35           40           45
ccc aat aat act gcg tct tgg ttc aca gct ctc act cag cat ggc aag      252
Pro Asn Asn Thr Ala Ser Trp Phe Thr Ala Leu Thr Gln His Gly Lys
50           55           60
gag gaa ctt aga ttc cct cga ggc cag ggc gtt cca atc aac acc aat      300
Glu Glu Leu Arg Phe Pro Arg Gly Gln Gly Val Pro Ile Asn Thr Asn
65           70           75
agt ggt cca gat gac caa att ggc tac tac cga aga gct acc cga cga      348
Ser Gly Pro Asp Asp Gln Ile Gly Tyr Tyr Arg Arg Ala Thr Arg Arg
80           85           90
gtt cgt ggt ggt gac ggc aaa atg aaa gag ctc agc ccc aga tgg tac      396
Val Arg Gly Gly Asp Gly Lys Met Lys Glu Leu Ser Pro Arg Trp Tyr
95           100           105           110
ttc tat tac cta gga act ggc cca gaa gct tca ctt ccc tac ggc gct      444
Phe Tyr Tyr Leu Gly Thr Gly Pro Glu Ala Ser Leu Pro Tyr Gly Ala
115           120           125
aac aaa gaa ggc atc gta tgg gtt gca act gag gga gcc ttg aat aca      492
Asn Lys Glu Gly Ile Val Trp Val Ala Thr Glu Gly Ala Leu Asn Thr
130           135           140
ccc aaa gac cac att ggc acc cgc aat cct aat aac aat gct gcc acc      540
Pro Lys Asp His Ile Gly Thr Arg Asn Pro Asn Asn Asn Ala Ala Thr
145           150           155
gtg cta caa ctt cct caa gga aca aca ttg cca aaa ggc ttc tac gca      588
Val Leu Gln Leu Pro Gln Gly Thr Thr Leu Pro Lys Gly Phe Tyr Ala
160           165           170
gag gga agc aga ggc ggc agt caa gcc tct tct cgc tcc tca tca cgt      636
Glu Gly Ser Arg Gly Gly Ser Gln Ala Ser Ser Arg Ser Ser Ser Arg
175           180           185           190

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agt cgc ggt aat tca aga aat tca act cct ggc agc agt agg gga aat      684
Ser Arg Gly Asn Ser Arg Asn Ser Thr Pro Gly Ser Ser Arg Gly Asn
      195                               200                               205

tct cct gct cga atg gct agc gga ggt ggt gaa act gcc ctc gcg cta      732
Ser Pro Ala Arg Met Ala Ser Gly Gly Glu Thr Ala Leu Ala Leu
      210                               215                               220

ttg ctg cta gac aga ttg aac cag ctt gag agc aaa gtt tct ggt aaa      780
Leu Leu Leu Asp Arg Leu Asn Gln Leu Glu Ser Lys Val Ser Gly Lys
      225                               230                               235

ggc caa caa caa caa ggc caa act gtc act aag aaa tct gct gct gag      828
Gly Gln Gln Gln Gln Gly Gln Thr Val Thr Lys Lys Ser Ala Ala Glu
      240                               245                               250

gca tct aaa aag cct cgc caa aaa cgt act gcc aca aaa cag tac aac      876
Ala Ser Lys Lys Pro Arg Gln Lys Arg Thr Ala Thr Lys Gln Tyr Asn
      255                               260                               265                               270

gtc act caa gca ttt ggg aga cgt ggt cca gaa caa acc caa gga aat      924
Val Thr Gln Ala Phe Gly Arg Arg Gly Pro Glu Gln Thr Gln Gly Asn
      275                               280                               285

ttc ggg gac caa gac cta atc aga caa gga act gat tac aaa cat tgg      972
Phe Gly Asp Gln Asp Leu Ile Arg Gln Gly Thr Asp Tyr Lys His Trp
      290                               295                               300

ccg caa att gca caa ttt gct cca agt gcc tct gca ttc ttt gga atg     1020
Pro Gln Ile Ala Gln Phe Ala Pro Ser Ala Ser Ala Phe Phe Gly Met
      305                               310                               315

tca cgc att ggc atg gaa gtc aca cct tcg gga aca tgg ctg act tat     1068
Ser Arg Ile Gly Met Glu Val Thr Pro Ser Gly Thr Trp Leu Thr Tyr
      320                               325                               330

cat gga gcc att aaa ttg gat gac aaa gat cca caa ttc aaa gac aac     1116
His Gly Ala Ile Lys Leu Asp Asp Lys Asp Pro Gln Phe Lys Asp Asn
      335                               340                               345                               350

gtc ata ctg ctg aac aag cac att gac gca tac aaa aca ttc cca cca     1164
Val Ile Leu Leu Asn Lys His Ile Asp Ala Tyr Lys Thr Phe Pro Pro
      355                               360                               365

aca gag cct aaa aag gac aaa aag aaa aag act gat gaa gct cag cct     1212
Thr Glu Pro Lys Lys Asp Lys Lys Lys Lys Thr Asp Glu Ala Gln Pro
      370                               375                               380

ttg ccg cag aga caa aag aag cag ccc act gtg act ctt ctt cct gcg     1260
Leu Pro Gln Arg Gln Lys Lys Gln Pro Thr Val Thr Leu Leu Pro Ala
      385                               390                               395

gct gac atg gat gat ttc tcc aga caa ctt caa aat tcc atg agt gga     1308
Ala Asp Met Asp Asp Phe Ser Arg Gln Leu Gln Asn Ser Met Ser Gly
      400                               405                               410

gct tct gct gat tca act cag gca taa acactcatga tgaccacaca           1355
Ala Ser Ala Asp Ser Thr Gln Ala
      415                               420

aggcagatgg gctatgtaaa cg                                           1377

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<210> SEQ ID NO 37

<211> LENGTH: 422

<212> TYPE: PRT

<213> ORGANISM: CORONAVIRUS

<400> SEQUENCE: 37

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Met Ser Asp Asn Gly Pro Gln Ser Asn Gln Arg Ser Ala Pro Arg Ile
1      5      10      15

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Thr Phe Gly Gly Pro Thr Asp Ser Thr Asp Asn Asn Gln Asn Gly Gly
20     25     30

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Arg Asn Gly Ala Arg Pro Lys Gln Arg Arg Pro Gln Gly Leu Pro Asn
35     40     45

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Asn Thr Ala Ser Trp Phe Thr Ala Leu Thr Gln His Gly Lys Glu Glu
 50                               55                               60

Leu Arg Phe Pro Arg Gly Gln Gly Val Pro Ile Asn Thr Asn Ser Gly
65                               70                               75                               80

Pro Asp Asp Gln Ile Gly Tyr Tyr Arg Arg Ala Thr Arg Arg Val Arg
                               85                               90                               95

Gly Gly Asp Gly Lys Met Lys Glu Leu Ser Pro Arg Trp Tyr Phe Tyr
                               100                               105                               110

Tyr Leu Gly Thr Gly Pro Glu Ala Ser Leu Pro Tyr Gly Ala Asn Lys
                               115                               120                               125

Glu Gly Ile Val Trp Val Ala Thr Glu Gly Ala Leu Asn Thr Pro Lys
                               130                               135                               140

Asp His Ile Gly Thr Arg Asn Pro Asn Asn Asn Ala Ala Thr Val Leu
145                               150                               155                               160

Gln Leu Pro Gln Gly Thr Thr Leu Pro Lys Gly Phe Tyr Ala Glu Gly
                               165                               170                               175

Ser Arg Gly Gly Ser Gln Ala Ser Ser Arg Ser Ser Ser Arg Ser Arg
                               180                               185                               190

Gly Asn Ser Arg Asn Ser Thr Pro Gly Ser Ser Arg Gly Asn Ser Pro
                               195                               200                               205

Ala Arg Met Ala Ser Gly Gly Gly Glu Thr Ala Leu Ala Leu Leu Leu
                               210                               215                               220

Leu Asp Arg Leu Asn Gln Leu Glu Ser Lys Val Ser Gly Lys Gly Gln
225                               230                               235                               240

Gln Gln Gln Gly Gln Thr Val Thr Lys Lys Ser Ala Ala Glu Ala Ser
                               245                               250                               255

Lys Lys Pro Arg Gln Lys Arg Thr Ala Thr Lys Gln Tyr Asn Val Thr
                               260                               265                               270

Gln Ala Phe Gly Arg Arg Gly Pro Glu Gln Thr Gln Gly Asn Phe Gly
                               275                               280                               285

Asp Gln Asp Leu Ile Arg Gln Gly Thr Asp Tyr Lys His Trp Pro Gln
290                               295                               300

Ile Ala Gln Phe Ala Pro Ser Ala Ser Ala Phe Phe Gly Met Ser Arg
305                               310                               315                               320

Ile Gly Met Glu Val Thr Pro Ser Gly Thr Trp Leu Thr Tyr His Gly
                               325                               330                               335

Ala Ile Lys Leu Asp Asp Lys Asp Pro Gln Phe Lys Asp Asn Val Ile
                               340                               345                               350

Leu Leu Asn Lys His Ile Asp Ala Tyr Lys Thr Phe Pro Pro Thr Glu
355                               360                               365

Pro Lys Lys Asp Lys Lys Lys Lys Thr Asp Glu Ala Gln Pro Leu Pro
370                               375                               380

Gln Arg Gln Lys Lys Gln Pro Thr Val Thr Leu Leu Pro Ala Ala Asp
385                               390                               395                               400

Met Asp Asp Phe Ser Arg Gln Leu Gln Asn Ser Met Ser Gly Ala Ser
                               405                               410                               415

Ala Asp Ser Thr Gln Ala
                               420

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<210> SEQ ID NO 38

<211> LENGTH: 1377

<212> TYPE: DNA

<213> ORGANISM: CORONAVIRUS

<400> SEQUENCE: 38

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attaaaaatgt ctgataatgg accccaatca aaccaacgta gtgccccccg cattacattt    120
ggtggaccca cagattcaac tgacaataac cagaatggag gacgcaatgg ggcaaggcca    180
aaacagcgcc gacccaaggt tttacccaat aatactgcgt cttgggtcac agctctcact    240
cagcatggca aggaggaact tagattccct cgaggccagg gcgttccaat caacaccaat    300
agtggccag atgaccaaat tggctactac cgaagagcta cccgacgagt tcgtgggtgt    360
gacggcaaaa tgaagagct cagccccaga tggctactct attacctagg aactggccca    420
gaagcttcac ttccctacgg cgctaacaaa gaaggcatcg tatgggttgc aactgagggg    480
gccttgaata cacccaaaga ccacattggc acccgcaatc ctaataacaa tgctgccacc    540
gtgctacaac ttccctcaag aacaacattg ccaaaaggct tctacgcaga gggaaagcaga    600
ggcggcagtc aagcctcttc tcgctctca tcacgtagtc gcgtaattc aagaaattca    660
actcctggca gcagtgggg aaattctct gctcgaatgg ctacggagg tggtgaaact    720
gccctcgcgc tattgtctgt agacagattg aaccagcttg agagcaaagt ttctggtaaa    780
ggccaacaac aacaaggcca aactgtcact aagaaatctg ctgctgaggc atctaaaaag    840
cctcgccaaa aacgtactgc cacaaaacag tacaacgtca ctcaagcatt tgggagacgt    900
ggtccagaac aaaccaaggt aaatttcggg gaccaagacc taatcagaca aggaactgat    960
tacaaaacatt ggccgcaaat tgcacaattt gctccaagtg cctctgcatt ctttggatg    1020
tcacgcattg gcatggaagt cacaccttcg ggaacatggc tgacttatca tggagccatt    1080
aaattggatg acaaagatcc acaattcaaa gacaacgtca tactgctgaa caagcacatt    1140
gacgcataca aacattccc accaacagag cctaaaaagg acaaaaagaa aaagactgat    1200
gaagctcagc ctttgccgca gagacaaaag aagcagccca ctgtgactct tcttctcgcg    1260
gttgacatgg atgatttctc cagacaactt caaaattcca tgagtggagc ttctgctgat    1320
tcaactcagg cataaacact catgatgacc acacaaggca gatgggetat gtaaacg    1377

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<210> SEQ ID NO 39
<211> LENGTH: 204
<212> TYPE: DNA
<213> ORGANISM: CORONAVIRUS

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<400> SEQUENCE: 39

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atattaggtt ttacctacc caggaaaagc caaccaacct cgatctcttg tagatctgtt    60
ctctaaacga actttaaaat ctgtgtagct gtctctcggc tgcattgcta gtgcacctac    120
gcagtataaa caataataaa ttttactgtc gttgacaaga aacgagtaac tcgtccctct    180
tctgcagact gcttacgggt tcgt                204

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<210> SEQ ID NO 40
<211> LENGTH: 809
<212> TYPE: DNA
<213> ORGANISM: CORONAVIRUS

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<400> SEQUENCE: 40

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actcaagcat tgggagacg tggtcagaa caaacccaag gaaatttcgg ggaccaagac    60
ctaatacagc aaggaactga ttacaacat tggccgcaaa ttgcacaatt tgctccaagt    120
gcctctgcat tctttggaat gtcacgcatt ggcatggaag tcacaccttc gggaaacatgg    180
ctgacttacc atggagccat taaattggat gacaaagatc cacaattcaa agacaacgtc    240
atactgctga acaagcacat tgacgcatac aaaacattcc caccaacaga gcctaaaaag    300

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gacaaaaaga aaaagactga tgaagctcag cctttgccgc agagacaaaa gaagcagccc	360
actgtgactc ttcttctctc ggctgacatg gatgatttct ccagacaact tcaaaattcc	420
atgagtggag cttctgctga ttcaactcag gcataaacac tcatgatgac cacacaaggc	480
agatgggcta tgtaaactgt ttcgcaattc cgtttacgat acatagtcta ctcttgca	540
gaatgaatte tcgtaactaa acagcacaag taggtttagt taactttaat ctccatagc	600
aatctttaat caatgtgtaa cattagggag gacttgaaag agccaccaca ttttcacga	660
ggccacgcgg agtacgatcg agggtagcgt gaataatgct agggagagct gcctatatgg	720
aagagcccta atgtgtaaaa ttaatttttag tagtgctatc cccatgtgat tttaatagct	780
tcttaggaga atgacaaaaa aaaaaaaaa	809

<210> SEQ ID NO 41
 <211> LENGTH: 448
 <212> TYPE: DNA
 <213> ORGANISM: CORONAVIRUS

<400> SEQUENCE: 41

aatgaacaca tagggctgtt caagctgggg cagtacgct tttccagct ctactagacc	60
acaagtgcc a ttttgaggt gttcaogtgc ctccgatagg gcctcttcca cagagtccc	120
gaagccacgc actagcacgt ctctaactg aaggacaggc aaactgagtt ggacgtgtgt	180
tttctcgty acaccaagaa caaggctctc catcttacct ttcggtcaca cccggacgaa	240
acctaggtat gctgatgac gactgcaaca cggacgaaac cgtaacgagt ctgcagaaga	300
gggacgagtt actcgtttct tgtcaacgac agtaaaattt attattgttt atactgcgta	360
ggtgcactag gcatgcagcc gagcgacagc tacacagatt ttaaagtctg tttagagaac	420
agatctacaa gagatcgagg ttggttgg	448

<210> SEQ ID NO 42
 <211> LENGTH: 2033
 <212> TYPE: DNA
 <213> ORGANISM: CORONAVIRUS

<400> SEQUENCE: 42

atacctaggt ttcgtccggg tgtgaccgaa aggtaagatg gagagccttg ttcttggtgt	60
caacgagaaa acacacgtcc aactcagttt gcctgtcctt caggttagag acgtgctagt	120
gcgtggcttc ggggactctg tggaaagagc cctatcgag gcacgtgaac acctcaaaaa	180
tggcacttgt ggtctagtag agctggaaaa aggcgtactg cccagcttg aacagcccta	240
tgtgttcatt aaacgttctg atgccttaag caccaatcac ggccacaagg tcgttgagct	300
ggttgacaaa atggacggca ttcagtacgg tcgtagcggg ataactgag gagtactcgt	360
gccacatgtg ggcaaaacc caattgcata ccgcaatgtt cttcttcgta agaacggtaa	420
taagggagcc ggtggtcata gctatggcat cgatctaaag tcttatgact taggtgacga	480
gcttgccact gatcccattg aagattatga acaaaactgg aactactaagc atggcagtg	540
tgcactccgt gaactcactc gtgagctcaa tggaggtgca gtcactcgt atgtcgacaa	600
caatttctgt ggcccagatg ggtaccctct tgattgcatc aaagatttct tcgcacgcgc	660
gggcaagtca atgtgcactc tttccgaaca acttgattac atcgagtcga agagaggtgt	720
ctactgctgc cgtgaccatg agcatgaaat tgctgggttc actgagcgt ctgataagag	780
ctacgagcac cagacaccct tcgaaattaa gagtgccaag aaatttgaca ctttcaaagg	840
ggaatgcccc aagtttgtgt ttcctcttaa ctcaaaagtc aaagtcattc aaccacgtgt	900

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tgaaaagaaa aagactgagg gtttcatggg gcgtatacgc tctgtgtacc ctgttgcatc 960
tccacaggag tgtaacaata tgcacttgtc taccttgatg aaatgtaatc attgcgatga 1020
agtttcatgg cagacgtgcg actttctgaa agccacttgt gaacattgtg gcaactgaaaa 1080
tttagttatt gaaggaccta ctacatgtgg gtacctacct actaatgctg tagtgaaaaat 1140
gccatgtcct gcctgtcaag acccagagat tggacctgag catagtgttg cagattatca 1200
caaccactca aacattgaaa ctgcactccg caagggaggt aggactagat gttttggagg 1260
ctgtgtgttt gcctatgttg gctgctataa taagcgtgcc tactgggttc ctctgtctag 1320
tgctgatatt ggctcaggcc atactggcat tactgggtgac aatgtggaga ccttgaatga 1380
ggatctcctt gagatactga gtcgtgaacg tgtaaacatt aacattgttg gcgattttca 1440
tttgaatgaa gaggttgcca tcattttggc atctttctct gcttctacaa gtgcctttat 1500
tgacactata aagagtcttg attacaagtc tttcaaaacc attgttgagt cctgcggtaa 1560
ctataaagtt accaagggaa agcccgtaaa aggtgcttgg aacattggac aacagagatc 1620
agttttaaca ccaactgttg gttttccctc acaggctgct ggtgttatca gatcaatttt 1680
tgcgcgcaca ctgtatgcag caaacactc aattcctgat ttgcaaagag cagctgtcac 1740
catacttgat ggtatttctg aacagtcatt acgtcttgtc gacgccatgg tttatacttc 1800
agacctgctc accaacagtg tcattattat ggcatatgta actggtggtc ttgtacaaca 1860
gacttctcag tggttgteta atcttttggg cactactggt gaaaaactca ggctatctt 1920
tgaatggatt gagggcaaac ttagtgcagg agttgaattt ctcaaggatg cttgggagat 1980
tctcaaatct ctcattacag gtgttttga catcgtcaag ggtcaaatac agg 2033

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<210> SEQ ID NO 43

<211> LENGTH: 2018

<212> TYPE: DNA

<213> ORGANISM: CORONAVIRUS

<400> SEQUENCE: 43

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ggattgaggc gaaacttagt gcaggagttg aatttctcaa ggatgcttgg gagattctca 60
aatttctcat tacagggtgt tttgacatcg tcaagggtca aatacagggt gcttcagata 120
acatcaagga ttgtgtaaaa tgcttcattg atgttggtta caaggcactc gaaatgtgca 180
ttgatcaagt cactatcgct gccgcaaagt tgcgatcact caacttaggt gaagtcttca 240
tcgctcaaag caagggactt tacctgcagt gtatacgtgg caaggagcag ctgcaactac 300
tcatgcctct taaggcacca aaagaagtaa cctttcttga aggtgattca catgacacag 360
tacttacctc tgaggaggtt gttctcaaga acggtgaact cgaagcactc gagacgcccg 420
ttgatagctt cacaaatgga gctatcgttg gcacaccagt ctgtgtaaat ggccctcatg 480
tcttagagat taaggacaaa gaacaatact gcgcattgtc tcctgggtta ctggctacaa 540
acaatgtcct tcgcttaaaa gggggtgcac caattaaagg tgtaaccttt ggagaagata 600
ctgtttggga agttcaagg tacaagaatg tgagaatcac atttgagctt gatgaacgtg 660
ttgacaaaag gcttaatgaa aagtgtctct tctacactgt tgaatccggt accgaagtta 720
ctgagtttgc atgtgttgta gcagaggctg ttgtgaagac tttacaacca gtttctgatc 780
tccttaccaa catgggtatt gatcttgatg agtggagtgt agctacattc tacttatttg 840
atgatgctgg tgaagaaaa ttttcatcac gtatgtattg ttctttttac cctccagatg 900
aggaagaaga ggacgatgca gagtvtgagg aagaagaat tgatgaaacc tgtgaacatg 960
agtacggtac agaggatgat tatcaaggtc tccctctgga atttggtgcc tcagctgaaa 1020

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cagttcgagt tgaggaagaa gaagaggaag actggctgga tgatactact gagcaatcag 1080
agattgagcc agaaccagaa cctacacctg aagaaccagt taatcagttt actggttatt 1140
taaaacttac tgacaatggt gccattaaat gtggtgacat cgttaaggag gcacaaagtg 1200
ctaactctat ggtgattgta aatgctgcta acatacacct gaaacatggt ggtggtgtag 1260
cagggtgcaact caacaaggca accaatggtg ccatgcacaa ggagagtgat gattacatta 1320
agctaaatgg ccctcttaca gtaggagggt cttgtttget ttctggacat aatcttgcta 1380
agaagtgtct gcatgttgtt ggacctaac taaatgcagg tgaggacatc cagcttctta 1440
aggcagcata tgaaaatttc aattcacagg acatcttact tgcaccattg ttgtcagcag 1500
gcatatttgg tgctaaacca cttcagtctt tacaagtgtg cgtgcagacg gttcgtacac 1560
aggtttatat tgcagtcaat gacaaagctc tttatgagca ggttgctcag gattatcttg 1620
ataacctgaa gcctagatggt gaagcaccta aacaagagga gccacacaa acagaagatt 1680
ccaaaactga ggagaaatct gtcgtacaga agcctgtcga tgtgaagcca aaaattaagg 1740
cctgcattga tgaggttacc acaacactgg aagaaactaa gtttcttacc aataagttac 1800
tcttgtttgc tgatatcaat ggtaagcttt accatgattc tcagaacatg cttagagggtg 1860
aagatatgtc tttccttgag aaggatgcac cttacatggt aggtgatggt atcactagtg 1920
gtgatatac ttgtgttgta atacctcca aaaaggctgg tggcactact gagatgctct 1980
caagagcttt gaagaaagtg ccagttgatg agtatata 2018

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<210> SEQ ID NO 44

<211> LENGTH: 1442

<212> TYPE: DNA

<213> ORGANISM: CORONAVIRUS

<400> SEQUENCE: 44

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ttgatgaggt taccacaaca ctggaagaaa ctaagtttct taccaataag ttactcttgt 60
ttgctgatat caatggtaag ctttaacctg attctcagaa catgcttaga ggtgaagata 120
tgtctttcct tgagaaggat gcaccttaca tggtaggtga tgttatcact agtggtgata 180
tcacttgtgt tgtaataccc tccaaaaagg ctggtggcac tactgagatg ctctcaagag 240
ctttgaagaa agtgccagtt gatgagtata taaccacgta ccctggacaa ggatgtgctg 300
ggtatacact tgaggaagct aagactgctc ttaagaaatg caaatctgca ttttatgtac 360
taccttcaga agcacctaata gctaagggaag agattctagg aactgtatcc tggaaattga 420
gagaaatgct tgctcatgct gaagagacaa gaaaattaat gcctatatgc atggatgta 480
gagccataat ggcaaccatc caacgtaagt ataaaggaat taaaattcaa gagggcatcg 540
ttgactatgg tgtccgatcc ttcttttata ctagttaaaga gcctgtagct tctattatta 600
cgaagctgaa ctctctaaat gagccgcttg tcacaatgcc aattggttat gtgacacatg 660
gttttaactc tgaagaggct gcgcgctgta tgcgttctct taaagctcct gccgtagtgt 720
cagtatcatc accagatgct gttactacat ataatggata cctcacttcg tcatcaaaga 780
catctgagga gcactttgta gaaacagttt ctttggtggt ctcttacaga gattggctct 840
attcaggaca gcgtacagag ttaggtgttg aatttcttaa gcgtgggtgac aaaatttgtt 900
accacactct ggagagcccc gtcgagtttc atcttgacgg tgaggttcct tcacttgaca 960
aactaaagag tctcttatcc ctgcgggagg ttaagactat aaaagtgttc acaactgtgg 1020
acaacactaa tctccacaca cagcttgtgg atatgtctat gacatagga cagcagtttg 1080
gtccaacata cttggatggt gctgatgta caaaaattaa acctcatgta aatcatgagg 1140

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gtaagacttt ctttgtacta cctagtgatg acacactacg tagtgaagct ttcgagtact 1200
accatactct tgatgagagt tttcttggtta ggtacatgtc tgctttaaac cacacaaaga 1260
aatggaaatt tcctcaagtt ggtggtttaa cttcaattaa atgggctgat aacaattggt 1320
at ttgtctag tgttttatta gcacttcaac agcttgaagt caaattcaat gcaccagcac 1380
ttcaagagge ttattataga gcccgctgctg gtgatgctgc taacttttgt gcaactcatac 1440
tc 1442

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<210> SEQ ID NO 45
<211> LENGTH: 1050
<212> TYPE: DNA
<213> ORGANISM: CORONAVIRUS

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<400> SEQUENCE: 45

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atatgtctat gacatatgga cagcagtttg gtccaacata cttggatggt gctgatgta 60
caaaaattaa acctcatgta aatcatgagg gtaagacttt ctttgtacta cctagtgatg 120
acacactacg tagtgaagct ttcgagtact accatactct tgatgagagt tttcttggtta 180
ggtacatgtc tgctttaaac cacacaaaga aatggaaatt tcctcaagtt ggtggtttaa 240
cttcaattaa atgggctgat aacaattggt at ttgtctag tgttttatta gcacttcaac 300
agcttgaagt caaattcaat gcaccagcac ttcaagagge ttattataga gcccgctgctg 360
gtgatgctgc taacttttgt gcaactcatac tcgcttacag taataaaact gttggcgagc 420
ttggatgagt cagagaaact atgacccatc ttctacagca tgctaatttg gaatctgcaa 480
agcgagtctt taatgtggtg tgtaaacatt gtggtcagaa aactactacc ttaacgggtg 540
tagaagctgt gatgtatag ggtactctat cttatgataa tcttaagaca ggtgttcca 600
ttccatggtg gtgtggtcgt gatgctacac aatatctagt acaacaagag tcttcttttg 660
ttatgatgtc tgcaccacct gctgagtata aattacagca aggtacattc ttatgtgcca 720
atgagtacac tggtaactat cagtgtggtc attacactca tataactgct aaggagacct 780
tctatcgtat tgacggagct caccttacia agatgtcaga gtacaaagga ccagtgactg 840
atgttttcta caagaaaca tcttacta caaccatcaa gccgtgtgctg tataaactcg 900
atggagtac ttacacagag attgaaccaa aattggatgg gtattataaa aaggataatg 960
cttactatac agagcagcct atagacctg taccaactca accattacca atgcgagtt 1020
ttgataattt caaactcaca tgttctaaca 1050

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<210> SEQ ID NO 46
<211> LENGTH: 1995
<212> TYPE: DNA
<213> ORGANISM: CORONAVIRUS

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<400> SEQUENCE: 46

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tttgtgcact catactcgct tacagtaata aaactggttg cgagcttggg gatgtcagag 60
aaactatgac ccatcttcta cagcatgcta at ttggaatc tgcaaagcga gttcttaatg 120
tgggtgtaaa acattgtggt cagaaaacta ctaccttaac ggggtgtagaa gctgtgatgt 180
atatgggtac tctatcttat gataatctta agacaggtgt ttccattcca tgtgtgtgtg 240
gtcgtgatgc tacacaatat ctagtacaac aagagtcttc tttgttatg atgtctgcac 300
cacctgctga gtataaatta cagcaaggta cattcttatg tgcgaatgag tacactggta 360
actatcagtg tggtcattac actcatataa ctgctaagga gacctctat cgtattgacg 420
gagctcacct tacaaagatg tcagagtaca aaggaccagt gactgatggt ttctacaagg 480

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aaacatctta cactacaacc atcaagcctg tgtcgtataa actcgatgga gttacttaca	540
cagagattga accaaaaattg gatgggtatt ataaaaagga taatgcttac tatacagagc	600
agcctataga ccttgtacca actcaacat taccaaatgc gagttttgat aatttcaaac	660
tcacatgttc taacacaaaa tttgctgatg atttfaatca aatgacaggc ttcacaaagc	720
cagcttcacg agagctatct gtcacattct tcccagactt gaatggcgat gtagtggcta	780
ttgactatag aactattca gcgagtttca agaaagggtgc taaattactg cataagccaa	840
ttgtttggca cattaaccag gctacaacca agacaacggt caaaccaaac acttgggtgt	900
tacgttgtct ttggagtaca aagccagtag atacttcaa ttcattttaa gttctggcag	960
tagaagacac acaaggaatg gacaatcttg cttgtgaaag tcaacaaccc acctctgaag	1020
aagtagtgga aaatcctacc atacagaagg aagtcataga gtgtgacgtg aaaactaccg	1080
aagttgtagg caatgtcata cttaaaccat cagatgaagg tgttaaagta acacaagagt	1140
taggtcatga ggatcttatg gctgcttatg tggaaaacac aagcattacc attaagaaac	1200
ctaattgagct ttcactagcc ttaggtttaa aaacaattgc cactcatggt attgctgcaa	1260
ttaatagtgt tccttggagt aaaattttgg cttatgtcaa accattctta ggacaagcag	1320
caattacaac atcaaatgc gctaagagat tagcacaacg tgtgtttaa aattatagc	1380
cttatgtgtt tacattattg ttccaattgt gtacttttac taaaagtacc aattctagaa	1440
ttagagcttc actacctaca actattgcta aaaatagtgt taagagtgtt gctaaattat	1500
gtttgatgc cggcattaat tatgtgaagt cacccaaatt ttctaattg ttcacaatcg	1560
ctatgtggct attgttgta agtatttgc taggttctct aatctgtgta actgctgctt	1620
ttggtgact cttatcctaat tttgggtgct cttcttattg taatggcggt agagaattgt	1680
atcttaattc gtctaactg actactatgg atttctgtga aggttctttt ccttcagca	1740
ttgtttaaag tggattagac tcccttgatt cttatccagc tcttgaacc attcaggtga	1800
cgatttcate gtacaageta gacttgacaa ttttaggtct ggccgctgag tgggttttgg	1860
catatatggt gttcacaaaa ttcttttatt tattaggtct ttcagctata atgcaggtgt	1920
tctttggeta ttttctagct catttcatca gcaattcttg gctcatgtgg tttatcatta	1980
gtattgtaca aatgg	1995

<210> SEQ ID NO 47

<211> LENGTH: 1884

<212> TYPE: DNA

<213> ORGANISM: CORONAVIRUS

<400> SEQUENCE: 47

aattcttggc tcatgtgggt tatcattagt attgtacaaa tggcaccggt ttctgcaatg	60
gttaggatgt acatcttctt tgcttcttcc tactacatat ggaagagcta tgttcatatc	120
atggatgggt gcacctcttc gacttgcatg atgtgctata agcgcaatcg tgccacacgc	180
gttgagtgta caactattgt taatggcatg aagagatctt tctatgtcta tgcaaatgga	240
ggccgtggct tctgcaagac tcacaattgg aattgtctca attgtgacac attttgcact	300
ggtagtacat tcattagtga tgaagtgtct cgtgatttgt cactccagtt taaaagacca	360
atcaacccta ctgaccagtc atcgtatatt gttgatagtg ttgctgtgaa aaatggcgcg	420
cttcacctct actttgacaa ggctgggtcaa aagacctatg agagacatcc gctctcccat	480
tttgcatt tagacaattt gagagctaac aacactaaag gttcactgcc tattaatgtc	540
atagtttttg atggcaagtc caaatgcgac gagtctgctt ctaagtctgc ttctgtgtac	600

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tacagtcagc	tgatgtgcc	acctattctg	ttgcttgacc	aagctcttgt	atcagacgtt	660
ggagatagta	ctgaagtttc	cgtaagatg	tttgatgctt	atgtcgacac	cttttcagca	720
acttttagtg	ttcctatgga	aaaacttaag	gcacttgttg	ctacagctca	cagcgagtta	780
gcaaaggggtg	tagctttaga	tggtgtcctt	tctacattcg	tgctagctgc	ccgacaaggt	840
gttgttgata	ccgatgttga	cacaaaggat	gttattgaat	gtctcaaact	ttcacatcac	900
tctgacttag	aagtgcacgg	tgacagttgt	aacaatttca	tgctcaccta	taataaggtt	960
gaaaacatga	cgcccagaga	tcttggcgca	tgtattgact	gtaatgcaag	gcatatcaat	1020
gcccagtag	caaaaagtca	caatgtttca	ctcatctgga	atgtaaaaga	ctacatgtct	1080
ttatctgaac	agctgcgtaa	acaaattctg	agtgtgccca	agaagaacaa	catacctttt	1140
agactaactt	gtgctacaac	tagacaggtt	gtcaatgtca	taactactaa	aatctcactc	1200
aagggtggta	agattgttag	tacttgtttt	aaacttatgc	ttaaggccac	attattgtgc	1260
gttcttgctg	cattggtttg	ttatatcggt	atgccagtac	atacattgtc	aatccatgat	1320
ggttacacaa	atgaaatcat	tggttacaaa	gccattcagg	atgggtgcac	tcgtgacatc	1380
atcttactg	atgattgttt	tgcaaaaaa	catgctgggt	ttgacgcatg	gtttagccag	1440
cggtgtggtt	catacaaaaa	tgacaaaagc	tgccctgtag	tagctgctat	cattacaaga	1500
gagattgggt	tcatagtgcc	tggttacccg	ggactgtgct	tgagagcaat	caatgggtgac	1560
ttcttgcaat	ttctacctcg	tgtttttagt	gctgttgcca	acatttgcta	cacaccttcc	1620
aaactcattg	agtatagtga	ttttgctacc	tctgcttgog	ttcttgctgc	tgagtgtaca	1680
atttttaag	atgctatggg	caaacctgtg	ccatattggt	atgacactaa	tttgctagag	1740
ggttctat	cttatagtga	gcttctgcca	gacactcggt	atgtgcttat	ggatgggtcc	1800
atcatacagt	ttcctaacac	ttacctggag	ggttctgtta	gagtagtaac	aacttttgat	1860
gctgagtact	gtagacatgg	taca				1884

<210> SEQ ID NO 48

<211> LENGTH: 2020

<212> TYPE: DNA

<213> ORGANISM: CORONAVIRUS

<400> SEQUENCE: 48

cactcgttat	gtgcttatgg	atggttccat	catacagttt	cctaacactt	acctggaggg	60
ttctgttaga	gtagtaacaa	cttttgatgc	tgagtactgt	agacatggta	catgcgaaaag	120
gtcagaagta	ggtatttgcc	tatctaccag	tggtagatgg	gttcttaata	atgagcatta	180
cagagctcta	tcaggagttt	tctgtgggtg	tgatgctgatg	aatctcatag	ctaactcctt	240
tactcctcct	gtgcaacctg	tggtgtcctt	agatgtgtct	gcttcagtag	tggtctgggtg	300
tattattgce	atattgggtga	cttgtgctgc	ctactacttt	atgaaattca	gacgtgtttt	360
tggtgagtac	aacctgttg	ttgctgctaa	tgcaactttg	tttttgatgt	ctttcactat	420
actctgtctg	gtaccagctt	acagctttct	gccgggagtc	tactcagtct	tttacttgta	480
cttgacatte	tatttcacca	atgatgtttc	attcttggct	caccttcaat	ggtttgccat	540
gttttctcct	atgtgacctt	tttgataac	agcaatctat	gtattctgta	tttctctgaa	600
gcactgccat	tggttcttta	acaactatct	taggaaaaga	gcatgttta	atggagttac	660
atntagtacc	ttcgaggagg	ctgctttgtg	tacctttttg	ctcaacaagg	aaatgtacct	720
aaaattgcgt	agcgagacac	tggtgacct	tacacagtat	aacaggtatc	ttgctctata	780
taacaagtac	aagtatttca	gtggagcctt	agatactacc	agctatcgtg	aagcagcttg	840

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ctgccactta gcaaaggctc taaatgactt tagcaactca ggtgctgatg ttctctacca	900
accaccacag acatcaatca cttctgctgt tctgcagagt ggttttagga aaatggcatt	960
cccgtcaggc aaagtgaag ggtgcatggt acaagtaacc tgtggaacta caactcttaa	1020
tggattgtgg ttggatgaca cagtatactg tocaagacat gtcatttgca cagcagaaga	1080
catgcttaat cctaactatg aagatctgct cattcgcaaa tccaaccata gctttcttgt	1140
tcaggctggc aatgttcaac ttcgtgttat tggccattct atgcaaaatt gctgcttag	1200
gcttaaagtt gatacttcta accctaagac acccaagtat aaatttgtcc gtatccaacc	1260
tggtaaaaa ttttcagttc tagcatgcta caatggttca ccactctggg tttatcagtg	1320
tgcoatgaga cctaatacata ccattaaagg ttctttcctt aatggatcat gtggtagtgt	1380
tggttttaac attgattatg attgctgtgc tttctgctat atgcatcata tggagcttcc	1440
aacaggagta cacgctggta ctgacttaga aggtaaatcc tatggtccat ttgttgacag	1500
acaaactgca caggctgcag gtacagacac aaccataaca ttaaatgttt tggcatggct	1560
gtatgctgct gttatcaatg gtgatagggt gtttcttaat agattcacca ctactttgaa	1620
tgactttaac cttgtggcaa tgaagtacaa ctatgaacct ttgacacaag atcatgttga	1680
catattggga cctctttctg ctcaaacagg aattgccctc ttagatatgt gtgctgcttt	1740
gaaagagctg ctgcagaatg gtatgaatgg togtactatc cttggtagca ctattttaga	1800
agatgagttt acaccatttg atgttgtagt acaatgctct ggtgttacct tccaaggtaa	1860
gttcaagaaa attgttaagg gcaactcatca ttggatgctt ttaactttct tgacatcact	1920
attgattcct gttcaaaagta cacagtggtc actgttttcc tttgtttacg agaatgcttt	1980
cttgccattt actcttggtg ttatggcaat tgctgcatgt	2020

<210> SEQ ID NO 49

<211> LENGTH: 2040

<212> TYPE: DNA

<213> ORGANISM: CORONAVIRUS

<400> SEQUENCE: 49

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taggttggtt aagcctttga tgaagtacaa gtatttccact ttaggccttt ttggtgtgtc	120
tgtacaaaa ctacaagggt gttccagttc tgtgtaaat gtacctgtac catcactctt	180
agggaaatcta gccattttga gatcttggtg gtctgatagt aatgccagca caaacctacc	240
tcccttcgaa ttgttatagt aggcaagtgc attgtcatca gtacaagctg tttgtgtggt	300
accagccgca caggacatct gtcgtagtgc tactggactc agttcattat tctgtagttt	360
aacagctgag ttggctctta gagctgtaac aataagaggc caagccaaat ttggtgaatt	420
gtccatgtta atttcaactaa gttgaacaat cttgctatcc gcatcaacaa cttgctggat	480
ttcccagagt gcagatgcat atgtaaagggt gttaccatca caagtgttct tgtaggtagc	540
ataatcaggg acaacaacca tgagtttggc tgctgtagtc aatggtaga tgttgagtgg	600
aacacaacca tcacgcgcat tgttgataat gttgttaagt gcatcattat caagcttcct	660
aagcatagtg aagagcattg tttgcatagc actagttact tttgccctct tgcctcaga	720
tcttgctctg ttgtacattt gggctatagc ctgatctgcc atcttttcca acttgctgtg	780
catggcagca tcacgggtcaa actcagattt agccacattc aaagatttct ttaacttttt	840
gagaacgact tcagaatcac cattagctac agcctgctca taggcctcct gggcagtggc	900
ataagcggca tatgatggtg aagaactaaa ttctgaagca atagcctgaa gagtagcagc	960

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gttatcgagc atttcctcgc acaacctatt aatgtctaca gcaccctgca tggatagcaa 1020
aacagacaaa agagaaacca tcttctcgaa agcttcagtt gtgtcttttg caagaagaat 1080
atcattgtgg agttgtacac attgtgccca caatttagaa gatgactcta ctctaagttg 1140
ttgaagaacc gagagcagta ccacagatgt gcactttacg tcagacattt tagactgtac 1200
agtagcaacc ttgatacatg gtttacctcc aatacccaac aacttaatgt taagcttgaa 1260
agcatcaata ctactcttag gaggcaaaag ccctgggag ttcataatcc taaattcttg 1320
tgtagagacc aagtagtcat aaacaccaag agtaagcctg aagtaacggg tgagtaaaca 1380
gaaaaggcca aagtagcagc agcaacaata gcctaagaaa caataaaca gcatgatata 1440
ctgtaaggtg ttgccagtaa taaataacaa tgggtaatac tcaacacaca caaacactat 1500
agctctagct aaaaacatga tagtcgtaac gacaccagaa tagttagagg ttacagaaat 1560
aactaaggcc cacatggaaa tagcttgatc taaagcatta ccatagtaga ctttgtaaac 1620
aagtgtaatg acattcatca gtgtccaac acgtctagca gcatcatcat aaacagtgcg 1680
agctgtcatg agaataagca aaactaaagc tgaagcatac ataacacaat ccttaagcct 1740
ataaccagac aagctagtgt cagccaatc aagccatgac atgatacga tcaaccagct 1800
agcaggcatg tagaccatat taaagtaagc aactgttgca agagaaggta acagaaacaa 1860
gcacaagaat gcgtgcttat gcttaacaag cagcatagca catgcagcaa ttgccataat 1920
accaagagta aatggcaaga aagcattctc gtaaacaaag aaaaacagtg accactgtgt 1980
actttgaaca agaatcaata gtgatgtcaa gaaagttaa agcatccaat gatgagtgca 2040

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<210> SEQ ID NO 50

<211> LENGTH: 2012

<212> TYPE: DNA

<213> ORGANISM: CORONAVIRUS

<400> SEQUENCE: 50

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cttgtaggtt tgttacagac acacaaaaag ggctaaagt gaaatacttg tacttcatca 60
aaggcttaa caacctaat agaggtagg tgctgggag tttagctgct acagtacgtc 120
ttcaggctgg aaatgtaca gaagtacctg ccaattcaac tgtgctttcc ttctgtgctt 180
ttgcagtaga ccctgctaaa gcatataagg attacctagc aagtggagga caaccaatca 240
ccaactgtgt gaagatgttg tgtacacaca ctggtacagg acaggcaatt actgtaacac 300
cagaagctaa catggaccaa gagtcctttg gtggtgcttc atgttgtctg tattgtagat 360
gccacattga ccatccaaat cctaaaggat tctgtgactt gaaaggtaag tacgtccaaa 420
tacctaccac ttgtgcta at gaccagtggt gttttact tagaaacaca gtctgtaccg 480
tctgcggaat gtggaagggt tatggctgta gttgtgacca actccgcaa cccttgatgc 540
agtctcggga tgcataacg tttttaaacy ggtttgcggt gtaagtgag cccgtcttac 600
accgtgcggc acaggcacta gtactgatgt cgtctacagg gcttttgata tttacaacga 660
aaaagtgtct ggttttgcaa agttcctaaa aactaattgc tgtcgcttcc aggagaagga 720
tgaggaagge aatttattag actcttactt tgtagttaag aggcatacta tgtctaacta 780
ccaacatgaa gagactattt ataacttggt taaagattgt ccagcggttg ctgtccatga 840
ctttttcaag tttagagtag atgggtgacat ggtaccacat atatcacgtc agegtctaac 900
taaatacaca atggctgatt tagtctatgc tctacgtcat tttgatgagg gtaattgtga 960
tacattaaaa gaaatactcg tcacatacaa ttgctgtgat gatgattatt tcaataagaa 1020
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gcggtgacgc caatcattat taaagactgt acaattctgc gatgctatgc gtgatgcagg 1140
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tgatttcgta caagtagcac caggctgcgg agttcctatt gtggattcat attactcatt 1260
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tctcgcaaaa ccacttatta agtgggattt gctgaaatat gattttacgg aagagagact 1380
ttgtctcttc gaccgttatt ttaaattatt ggaccagaca taccatccca attgtattaa 1440
ctgtttggat gataggtgta tccttcattg tgcaaaacttt aatgtgttat tttctactgt 1500
gtttccacct acaagttttg gaccactagt aagaaaaata tttgtagatg gtgttccttt 1560
tgttgtttca actggatacc attttcgtga gttaggagtc gtacataatc aggatgtaaa 1620
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gcatgcagct tctggcaatt tattgctaga taaacgcact acatgctttt cagtagctgc 1740
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tgactttgct gtgtctaaag gtttctttaa ggaaggaagt tctgttgaac taaaacactt 1860
cttctttgct caggatggca acgctgctat cagtgattat gactattatc gttataatct 1920
gccacaatg tgatgatca gacaactcct attcgtagtt gaagtgttg ataataactt 1980
tgattgttac gatgggtgct gtattaatgc ca 2012

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<210> SEQ ID NO 51

<211> LENGTH: 1877

<212> TYPE: DNA

<213> ORGANISM: CORONAVIRUS

<400> SEQUENCE: 51

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gtacttcgcy tacagtggca ataccatag acagcttaaa tgtttcctca gtggctttga 60
gcgtttctgc tgcgaaaagc ttgagtctct cagtacaagt gttggcaagt atgtaategc 120
cagcattagt ccaatcacat gttgctatcg cattgaagtc agtgacattg tcaactgcta 180
cacatgtggt tttgtataaa ccaaaaacct gaccattagc acataatgga aaactaatgg 240
gaggcttatg tgacttgcaa taatagctca tacctcctag atacagtgtg gtcacatcag 300
tgacatcaca acctggggca ttgcaaacat agggattaac agacaacact aatttgtgtg 360
atgttgaat gacatggtca tagcagcact tgcaacatag gaatggtctc ctaatacagg 420
caccgcaacy aagtgaagtc tgtgaattgc acaatacaca agcacctaca gcctgcaaga 480
ctgtatgtgg tgtgtacata gcctcataaa actcaggttc ccagtaccgt gaggtgttat 540
cattagttag cattacggaa tacatgtcca acatgtggcc agtaagctca tcatgtaact 600
ttctaataga ttgtaaatca aagtgaaga catcagcata ctctgatta ggatgttttg 660
taagtgggta agcatcaata gccagtgaca cgaaccttc aatcataagt gtaccatctg 720
ttttgacaat atcatcgaca aacagcctg cgcctaatat tcttgatgga tctgggtaag 780
gcaggtacac gtaatcatct ccttgtttaa cttagcattgt atgctgtgag caaaattcgt 840
gaggtccttt agtaaggta gtctcagtc aacattttgc ctgagacatg aacacattat 900
tttgataata aagaactgcc ttaaagtctc taatgctagc tactaaaact tgagccgcat 960
agttactggt atagcacaca acggcatcat cagaaagaat catcatggag aaatgtttac 1020
gcaggtaaag gtaaaactca tccacgaatt catgatcaac atccctattt ctatagagac 1080
actcatagag cctgtgttgt agattgcgga catactgtgc agctatctta ttaccatcag 1140
ttgaaagaag tgcatttaca ttggtgttaa cagcttgaca aatgttaaag acactattag 1200

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cataagcagt	tgtagcatca	cgggatgatg	ttccacctgg	ttaacatat	agtgagccgc	1260
cacacatgac	catctcactt	aatacttgcg	cacactcggt	agctaacctg	tagaaacggt	1320
gtgataagtt	acagcaagtg	ttatgtttgc	gagcaagaac	aagagaggcc	attatcctaa	1380
gcatgttagg	catggctctg	tcacattttg	gataatccca	acccataagg	tgtggagttt	1440
ctacatcact	gtaaacagtt	ttaacatat	tatgccagcc	accgtaaac	ttgcttgttc	1500
caattaccac	agtagctcct	ctagtggcgg	ctattgactt	caataatttc	tgatgaaact	1560
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cactaatggc	atacttaaga	ttcatttgag	ttatagtagg	gatgacatta	cgcttagtat	1680
acgcgaaaag	tgcactttga	tcctcataac	tcattgagtc	ataataaagt	ctagccttac	1740
cccatttatt	aaatgggaaa	ccagctgatt	tatccagatt	gttaacgatt	acttggttgg	1800
cattaataca	gccaccatcg	taacaatcaa	agtatttata	aacaacttca	actacgaata	1860
ggagttgtct	gatatca					1877

<210> SEQ ID NO 52

<211> LENGTH: 2051

<212> TYPE: DNA

<213> ORGANISM: CORONAVIRUS

<400> SEQUENCE: 52

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acgaacacga	ctctgtctga	caatccttc	agtgtatcac	tgagcatttg	tactatctta	120
atacgacta	cattccaggg	caagccttta	tacatgagtg	gtataagatg	tttaaacctgg	180
tcacctgggtg	gaggttttgc	attaactctg	gtgaattctg	tgttattttc	agtgccaaca	240
taaccagtcg	gtacagctac	taagttaaca	cctgtagaaa	atcctagctg	gagaggtagg	300
ttagtaccca	cagcatctct	agttgcatga	cagccctcta	catcaaagcc	aatccacgca	360
cgaaactgac	gaatagcttc	ttcgcgggtg	ataaacatat	tagggtaacc	attgacttgg	420
taattcattt	tgaaacccat	catagagatg	agtctacggg	aggctcatgct	ctttggtatg	480
cctggtagtg	caacacataa	tccttcagtc	ttgaacttta	tatcaacgct	gagggtgtgta	540
gggtgcctgtg	taggatgaag	accagtaatg	atcttactac	agtccttaaa	aagtccagtt	600
acattttctg	cttgtaatgt	agccacattg	cgacgtggta	tttctagact	tgtaaattgc	660
agtttgtcat	aaagatctct	atcagacatt	atgcacaaaa	tgccaatttt	tgccttctgtg	720
atagccacat	tgaagcgggt	gacattacaa	gagtggtctg	tttcagtagt	ttgtgtgaat	780
atgacatagt	catattcaga	accctgtgat	gaatcaacag	tctgcgtagg	caatcctaag	840
atttttgaag	ctacagcgtt	ctgtgaatta	taagggtgaga	taaaaacagc	ttttctccaa	900
gcaggattgc	gtgtaagaaa	ttctcttaca	acgcctattt	gaggctctgtt	gattgcagat	960
gaaacatcat	gtgtaataac	acctttgtag	aacattttga	agcattgagc	tgacttatcc	1020
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ggacaacggc	gacaagttcc	aaggaacatg	tctggaccta	ttgttttcat	aagtctgcac	1140
actgaattaa	aatattctgg	ttctagtgtg	cctttagcca	gcaatgtgcg	gggggctggt	1200
aattgagcag	gatcgccaat	atagacgtag	tgttttgcac	gaagtctagc	attgacaaca	1260
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tctggcaatg	catttacagt	gcagaaaaca	tactgttcta	gtgttgtaatt	cactttgaa	1380
ttatcaaaac	actctacgcg	cgcaacgcga	ggtagtatcc	tactacattt	atctatgggc	1440

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aaatatttta atgccttttc acatagggca tcaacagctg catgagagca tgcggtatac	1500
actatgcgag cagatgggta atagagagca agtccgatgg caaaatgact cttaccagta	1560
ccagggtgct cttggagtgt agagtacttt tgcagccga ccttttgata atttgaaca	1620
ttgctagaaa actcatctga gatgttgagt gttgggtaca agccagtaat tctcacatag	1680
tgctcttggt gcactagagt aggtgcacta agtggcatta cagtgtgaga tgtcaacaca	1740
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taaccagtaa agacatagtt tctgttcaat ggtggtctag gttttccaac ctcccatgaa	1920
agatgcaatt ctctgtcaga gagtacttcg cgtacagtgg caataccata tgacagctta	1980
aatgtttcct cagtggcttt gagcgtttct gctgcgaaaa gcttgagtct ctccagtaca	2040
gtgttgcaa g	2051

<210> SEQ ID NO 53

<211> LENGTH: 2075

<212> TYPE: DNA

<213> ORGANISM: CORONAVIRUS

<400> SEQUENCE: 53

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aatttcagca tagtcaattg taaccttgac cacttttgaa atcactgaca aatcttgtga	120
ctttattatc tcgacaaagt catcaagtaa aagatcaatc acagaacaca cacattttga	180
tgaacctggt tgcgcatctg ttatgaagta atttttcact gtgctgtcca tagggataaa	240
atcctcta at taaagtgggt aatcttgtga gcgcttggt aagcctatca ttaaatgaag	300
accgccaagt tgtccatgac tgaaatctcc ataaacgatg tgttcgaagg catagccctc	360
gagcttatat cgctgtatga attcatccat agcagctcg agaaagtcag tttccatttg	420
tgatctgggc ttaaaatcct ctaagtctct gctctgagta aagtaggttt caggcaactg	480
ttgaataatg ccgtctactt tcttaaagta gttaaactgt gtttttactg attctccaat	540
taatgtgact ccattgacgc tagcttgtgc tggctccctt gaagggtta gacctttgac	600
tgaaccttct gttattaaaa caccattacg ggcgtttcta aaaaggctta cctgtccttc	660
cactctacca tcaacaaga cagtaagtga agaacaagca ctctcagtag gtttcttggc	720
aatgtcagtc attgtgcaga cacctattgt agatacatgt gctggggctt ctcttttgta	780
gtcccagatt acagtattag cagcgatatc aacacccaaa ttattgagta tcttaatctc	840
tggcactggt ttaatgttac gcttagccca aagctcaaat gcaacattaa caggaagtgt	900
tgtcttattt tcaaagatct ccacatcaat accatctacc tttgtgtaa cagcattatt	960
aatgatggaa acagggtgct cgccggcgtg tccatcaaag tgtcctttat taacaacatt	1020
ataagccaca ttttctaaac tctgtaacct ggtaaatgta ttccacaggt tataagtatc	1080
aaattgtttg taaatccata ggctaaatcc agcagaaatc atcatattat atgcatccaa	1140
gtactgtcgg tactcatttg catggtgtct gcaaacagca ccacctaata tgcacgtgt	1200
aatacacgta gcagatttga gtggaacata atcaatatcc gacactactt gtttgccatg	1260
agactcaca ggactatcag aatagtaaaa gaaaggcaat tgctttaaata tagtaaatgc	1320
acttttatcg aaagctggag tgtggaatgc atgcttattc acatacaaac taccaccatc	1380
acagcctggt aagttcaagt ttgacaagac tcttgtgtca aacctacaca caattgcatt	1440
ggctgggtaa cgatcaacgt tacaattcca aaacaacaa acaccatcag tgaatttatc	1500

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gtgatgtgta	gcataagaat	agaagagttc	ctctattttg	taagctttgt	cactacatgg	1560
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atttccaatg	tcatgaagaa	ctggaaactt	atcagcaagc	aatgcagact	tcacaaccat	1680
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ctgaaggtta	cccgtaaagc	cccactgctg	aacatcaatc	ataaatgggt	tatagacata	1920
gtcaaaaacc	acagaatgat	tccagcaggc	ataagtatct	gatgaagtag	aaaagcaagt	1980
tgacagtttg	tcacacagac	aacacgttct	ttcaggtcca	atcttgacia	agtacttcat	2040
tgatgtaagc	tcaaagccat	gcgcccaaag	gacga			2075

<210> SEQ ID NO 54

<211> LENGTH: 1891

<212> TYPE: DNA

<213> ORGANISM: CORONAVIRUS

<400> SEQUENCE: 54

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tcataacaga	tgcgcaaaca	ggttcatcaa	aatgtgtgtg	ttctgtgatt	gatcttttac	120
ttgatgactt	tgctgagata	ataaagtca	aagatttctc	agtgatttca	aaagtggtea	180
aggttacaat	tgactatgct	gaaatttcat	tcctgctttg	gtgtaaggat	ggacatgttg	240
aaaccttcta	cccaaaaacta	caagcaagtc	aagcgtggca	accaggtggt	gcatgacctc	300
acttgtaaaa	gatgcaaaaga	atgcttcttg	aaaagtgtga	ccttcagaat	tatggtgaaa	360
atgctgttat	acaaaagga	ataatgatga	atgtcgcaaa	gtatactcaa	ctgtgtcaat	420
acttaaatca	acttacttta	gctgtacctc	acaacatgag	agttattcac	tttgggtctg	480
gctctgataa	aggagttgca	ccaggtacag	ctgtgctcag	acaatgggtg	ccaactggca	540
cactacttgt	cgattcagat	cttaatgact	tcgtctccga	cgcagattct	actttaattg	600
gagactgtgc	aacagtagat	acggctaata	aatgggacct	tattattagc	gatatgtatg	660
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ctcttctgga	aaaaggttag	cttatcatta	gagaaaacaa	cagagttgtg	gtttcaagtg	1140
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gtggtagtga	ccttgaccgg	tgcaccactt	ttgatgatgt	tcaagctcct	aattacactc	1260
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ttaatcatac	gtttggcaac	cctgtcatac	cttttaagga	tggattttat	tttctgcca	1440
cagagaaatc	aaatgttgtc	cgtggttggg	tttttggttc	taccatgaac	aacaagtcac	1500
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tgtgtgacaa ccctttcttt gctgtttcta aacctatggg tacacagaca catactatga 1620
tattcgataa tgcatttaat tgcactttcg agtacatatac tgatgccttt tcgcttgatg 1680
tttcagaaaa gtcaggtaat tttaaacact tacgagagtt tgtgtttaaa aataaagatg 1740
ggttttctta tgtttataag ggctatcaac ctatagatgt agttcgtgat ctaccttctg 1800
gttttaacac tttgaaacct atttttaagt tgcctcttgg tattaacatt acaaatttta 1860
gagcattctt tacagccttt tcacctgctc a 1891

```

```

<210> SEQ ID NO 55
<211> LENGTH: 32
<212> TYPE: DNA
<213> ORGANISM: artificial sequence
<220> FEATURE:
<223> OTHER INFORMATION: N sens primer

```

```

<400> SEQUENCE: 55

```

```

cccatatgtc tgataatgga ccccaatcaa ac 32

```

```

<210> SEQ ID NO 56
<211> LENGTH: 32
<212> TYPE: DNA
<213> ORGANISM: artificial sequence
<220> FEATURE:
<223> OTHER INFORMATION: N antisens primer

```

```

<400> SEQUENCE: 56

```

```

cccccggtg cctgagttga atcagcagaa gc 32

```

```

<210> SEQ ID NO 57
<211> LENGTH: 31
<212> TYPE: DNA
<213> ORGANISM: artificial sequence
<220> FEATURE:
<223> OTHER INFORMATION: Sc sens primer

```

```

<400> SEQUENCE: 57

```

```

cccatatgag tgaccttgac cgggtcacca c 31

```

```

<210> SEQ ID NO 58
<211> LENGTH: 30
<212> TYPE: DNA
<213> ORGANISM: artificial sequence
<220> FEATURE:
<223> OTHER INFORMATION: SL sens primer

```

```

<400> SEQUENCE: 58

```

```

cccatatgaa accttgacac ccacctgctc 30

```

```

<210> SEQ ID NO 59
<211> LENGTH: 33
<212> TYPE: DNA
<213> ORGANISM: Sc and SL antisens primer

```

```

<400> SEQUENCE: 59

```

```

cccccggtt taatatattg ctcatatattt ccc 33

```

```

<210> SEQ ID NO 60
<211> LENGTH: 16
<212> TYPE: DNA
<213> ORGANISM: Sens set 1 primer

```

```

<400> SEQUENCE: 60

```

```

ggcctegtat gggttg 16

```


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```

<210> SEQ ID NO 61
<211> LENGTH: 16
<212> TYPE: DNA
<213> ORGANISM: Antisens set 2 (28774-28759) primer

<400> SEQUENCE: 61
cagtttcacc acctcc 16

<210> SEQ ID NO 62
<211> LENGTH: 16
<212> TYPE: DNA
<213> ORGANISM: Sens set 2 (28375-28390) primer

<400> SEQUENCE: 62
ggctactacc gaagag 16

<210> SEQ ID NO 63
<211> LENGTH: 16
<212> TYPE: DNA
<213> ORGANISM: Antisens set 2 (28702-28687)primer

<400> SEQUENCE: 63
aattaccgcg actacg 16

<210> SEQ ID NO 64
<211> LENGTH: 26
<212> TYPE: DNA
<213> ORGANISM: Probe 1/set 1 (28561-28586)

<400> SEQUENCE: 64
ggcaccgca atcctaataa caatgc 26

<210> SEQ ID NO 65
<211> LENGTH: 21
<212> TYPE: DNA
<213> ORGANISM: Probe 2/set 1 (28588-28608)

<400> SEQUENCE: 65
gccaccgtgc tacaacttcc t 21

<210> SEQ ID NO 66
<211> LENGTH: 23
<212> TYPE: DNA
<213> ORGANISM: Probe 1/set 2 /probe N/FL (28541-28563)

<400> SEQUENCE: 66
atacacccaa agaccacatt ggc 23

<210> SEQ ID NO 67
<211> LENGTH: 25
<212> TYPE: DNA
<213> ORGANISM: Probe 2/set 2/probe SARS/N/LC705 (28565-28589)

<400> SEQUENCE: 67
cccgcaatcc taataacaat gctgc 25

<210> SEQ ID NO 68
<211> LENGTH: 30
<212> TYPE: DNA
<213> ORGANISM: artificial sequence
<220> FEATURE:
<223> OTHER INFORMATION: Anchor primer 14T

<400> SEQUENCE: 68

```

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 agatgaattc ggtacctttt tttttttttt 30

<210> SEQ ID NO 69
 <211> LENGTH: 13
 <212> TYPE: PRT
 <213> ORGANISM: artificial sequence
 <220> FEATURE:
 <223> OTHER INFORMATION: M2-14 peptide

<400> SEQUENCE: 69

Ala Asp Asn Gly Thr Ile Thr Val Glu Glu Leu Lys Gln
 1 5 10

<210> SEQ ID NO 70
 <211> LENGTH: 12
 <212> TYPE: PRT
 <213> ORGANISM: artificial sequence
 <220> FEATURE:
 <223> OTHER INFORMATION: E1-12 peptide

<400> SEQUENCE: 70

Met Tyr Ser Phe Val Ser Glu Glu Thr Gly Thr Leu
 1 5 10

<210> SEQ ID NO 71
 <211> LENGTH: 24
 <212> TYPE: PRT
 <213> ORGANISM: artificial sequence
 <220> FEATURE:
 <223> OTHER INFORMATION: E53-72 peptide

<400> SEQUENCE: 71

Lys Pro Thr Val Tyr Val Tyr Ser Arg Val Lys Asn Leu Asn Ser Ser
 1 5 10 15

Glu Gly Val Pro Asp Leu Leu Val
 20

<210> SEQ ID NO 72
 <211> LENGTH: 153
 <212> TYPE: DNA
 <213> ORGANISM: CORONAVIRUS

<400> SEQUENCE: 72

gatattaggt ttttacctac ccaggaaaag ccaaccaacc tcgatctctt gtagatctgt 60

tctctaaaacg aactttaaaa tctgtgtagc tgtegcctcg ctgcatgcct agtgcacct 120

cgcagtataa acaataataa attttactgt cgt 153

<210> SEQ ID NO 73
 <211> LENGTH: 410
 <212> TYPE: DNA
 <213> ORGANISM: CORONAVIRUS

<400> SEQUENCE: 73

ttctccagac aacttcaaaa ttccatgagt ggagcttctg ctgattcaac tcaggcataa 60

acactcatga tgaccacaca aggcagatgg gctatgtaaa cgttttcgca attccgttta 120

cgatacatag tctactcttg tgcagaatga atttctgtaa ctaaacagca caagtagggt 180

tagttaactt taatctcaca tagcaatctt taatcaatgt gtaacattag ggaggacttg 240

aaagagccac cacatthtca tcgaggccac ggggagtacg atcgagggta cagtgaataa 300

tgctaggag agctgcctat atggaagagc cctaatgtgt aaaattaatt ttagtagtgc 360

tatcccacatg tgattttaat agcttcttag gagaatgaca aaaaaaaaaa 410

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<210> SEQ ID NO 74
<211> LENGTH: 4382
<212> TYPE: PRT
<213> ORGANISM: CORONAVIRUS

<400> SEQUENCE: 74

Met Glu Ser Leu Val Leu Gly Val Asn Glu Lys Thr His Val Gln Leu
1      5      10     15
Ser Leu Pro Val Leu Gln Val Arg Asp Val Leu Val Arg Gly Phe Gly
20     25     30
Asp Ser Val Glu Glu Ala Leu Ser Glu Ala Arg Glu His Leu Lys Asn
35     40     45
Gly Thr Cys Gly Leu Val Glu Leu Glu Lys Gly Val Leu Pro Gln Leu
50     55     60
Glu Gln Pro Tyr Val Phe Ile Lys Arg Ser Asp Ala Leu Ser Thr Asn
65     70     75     80
His Gly His Lys Val Val Glu Leu Val Ala Glu Met Asp Gly Ile Gln
85     90     95
Tyr Gly Arg Ser Gly Ile Thr Leu Gly Val Leu Val Pro His Val Gly
100    105   110
Glu Thr Pro Ile Ala Tyr Arg Asn Val Leu Leu Arg Lys Asn Gly Asn
115    120   125
Lys Gly Ala Gly Gly His Ser Tyr Gly Ile Asp Leu Lys Ser Tyr Asp
130    135   140
Leu Gly Asp Glu Leu Gly Thr Asp Pro Ile Glu Asp Tyr Glu Gln Asn
145    150   155   160
Trp Asn Thr Lys His Gly Ser Gly Ala Leu Arg Glu Leu Thr Arg Glu
165    170   175
Leu Asn Gly Gly Ala Val Thr Arg Tyr Val Asp Asn Asn Phe Cys Gly
180    185   190
Pro Asp Gly Tyr Pro Leu Asp Cys Ile Lys Asp Phe Leu Ala Arg Ala
195    200   205
Gly Lys Ser Met Cys Thr Leu Ser Glu Gln Leu Asp Tyr Ile Glu Ser
210    215   220
Lys Arg Gly Val Tyr Cys Cys Arg Asp His Glu His Glu Ile Ala Trp
225    230   235   240
Phe Thr Glu Arg Ser Asp Lys Ser Tyr Glu His Gln Thr Pro Phe Glu
245    250   255
Ile Lys Ser Ala Lys Lys Phe Asp Thr Phe Lys Gly Glu Cys Pro Lys
260    265   270
Phe Val Phe Pro Leu Asn Ser Lys Val Lys Val Ile Gln Pro Arg Val
275    280   285
Glu Lys Lys Lys Thr Glu Gly Phe Met Gly Arg Ile Arg Ser Val Tyr
290    295   300
Pro Val Ala Ser Pro Gln Glu Cys Asn Asn Met His Leu Ser Thr Leu
305    310   315   320
Met Lys Cys Asn His Cys Asp Glu Val Ser Trp Gln Thr Cys Asp Phe
325    330   335
Leu Lys Ala Thr Cys Glu His Cys Gly Thr Glu Asn Leu Val Ile Glu
340    345   350
Gly Pro Thr Thr Cys Gly Tyr Leu Pro Thr Asn Ala Val Val Lys Met
355    360   365
Pro Cys Pro Ala Cys Gln Asp Pro Glu Ile Gly Pro Glu His Ser Val
370    375   380

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Ala	Asp	Tyr	His	Asn	His	Ser	Asn	Ile	Glu	Thr	Arg	Leu	Arg	Lys	Gly	385		390			395					400
Gly	Arg	Thr	Arg	Cys	Phe	Gly	Gly	Cys	Val	Phe	Ala	Tyr	Val	Gly	Cys			405			410					415
Tyr	Asn	Lys	Arg	Ala	Tyr	Trp	Val	Pro	Arg	Ala	Ser	Ala	Asp	Ile	Gly			420			425					430
Ser	Gly	His	Thr	Gly	Ile	Thr	Gly	Asp	Asn	Val	Glu	Thr	Leu	Asn	Glu			435			440					445
Asp	Leu	Leu	Glu	Ile	Leu	Ser	Arg	Glu	Arg	Val	Asn	Ile	Asn	Ile	Val			450			455					460
Gly	Asp	Phe	His	Leu	Asn	Glu	Glu	Val	Ala	Ile	Ile	Leu	Ala	Ser	Phe			465			470					480
Ser	Ala	Ser	Thr	Ser	Ala	Phe	Ile	Asp	Thr	Ile	Lys	Ser	Leu	Asp	Tyr			485			490					495
Lys	Ser	Phe	Lys	Thr	Ile	Val	Glu	Ser	Cys	Gly	Asn	Tyr	Lys	Val	Thr			500			505					510
Lys	Gly	Lys	Pro	Val	Lys	Gly	Ala	Trp	Asn	Ile	Gly	Gln	Gln	Arg	Ser			515			520					525
Val	Leu	Thr	Pro	Leu	Cys	Gly	Phe	Pro	Ser	Gln	Ala	Ala	Gly	Val	Ile			530			535					540
Arg	Ser	Ile	Phe	Ala	Arg	Thr	Leu	Asp	Ala	Ala	Asn	His	Ser	Ile	Pro			545			550					560
Asp	Leu	Gln	Arg	Ala	Ala	Val	Thr	Ile	Leu	Asp	Gly	Ile	Ser	Glu	Gln			565			570					575
Ser	Leu	Arg	Leu	Val	Asp	Ala	Met	Val	Tyr	Thr	Ser	Asp	Leu	Leu	Thr			580			585					590
Asn	Ser	Val	Ile	Ile	Met	Ala	Tyr	Val	Thr	Gly	Gly	Leu	Val	Gln	Gln			595			600					605
Thr	Ser	Gln	Trp	Leu	Ser	Asn	Leu	Leu	Gly	Thr	Thr	Val	Glu	Lys	Leu			610			615					620
Arg	Pro	Ile	Phe	Glu	Trp	Ile	Glu	Ala	Lys	Leu	Ser	Ala	Gly	Val	Glu			625			630					640
Phe	Leu	Lys	Asp	Ala	Trp	Glu	Ile	Leu	Lys	Phe	Leu	Ile	Thr	Gly	Val			645			650					655
Phe	Asp	Ile	Val	Lys	Gly	Gln	Ile	Gln	Val	Ala	Ser	Asp	Asn	Ile	Lys			660			665					670
Asp	Cys	Val	Lys	Cys	Phe	Ile	Asp	Val	Val	Asn	Lys	Ala	Leu	Glu	Met			675			680					685
Cys	Ile	Asp	Gln	Val	Thr	Ile	Ala	Gly	Ala	Lys	Leu	Arg	Ser	Leu	Asn			690			695					700
Leu	Gly	Glu	Val	Phe	Ile	Ala	Gln	Ser	Lys	Gly	Leu	Tyr	Arg	Gln	Cys			705			710					720
Ile	Arg	Gly	Lys	Glu	Gln	Leu	Gln	Leu	Leu	Met	Pro	Leu	Lys	Ala	Pro			725			730					735
Lys	Glu	Val	Thr	Phe	Leu	Glu	Gly	Asp	Ser	His	Asp	Thr	Val	Leu	Thr			740			745					750
Ser	Glu	Glu	Val	Val	Leu	Lys	Asn	Gly	Glu	Leu	Glu	Ala	Leu	Glu	Thr			755			760					765
Pro	Val	Asp	Ser	Phe	Thr	Asn	Gly	Ala	Ile	Val	Gly	Thr	Pro	Val	Cys			770			775					780
Val	Asn	Gly	Leu	Met	Leu	Leu	Glu	Ile	Lys	Asp	Lys	Glu	Gln	Tyr	Cys			785			790					800
Ala	Leu	Ser	Pro	Gly	Leu	Leu	Ala	Thr	Asn	Asn	Val	Phe	Arg	Leu	Lys			805			810					815

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Gly Gly Ala Pro Ile Lys Gly Val Thr Phe Gly Glu Asp Thr Val Trp
 820 825 830
 Glu Val Gln Gly Tyr Lys Asn Val Arg Ile Thr Phe Glu Leu Asp Glu
 835 840 845
 Arg Val Asp Lys Val Leu Asn Glu Lys Cys Ser Val Tyr Thr Val Glu
 850 855 860
 Ser Gly Thr Glu Val Thr Glu Phe Ala Cys Val Val Ala Glu Ala Val
 865 870 875 880
 Val Lys Thr Leu Gln Pro Val Ser Asp Leu Leu Thr Asn Met Gly Ile
 885 890 895
 Asp Leu Asp Glu Trp Ser Val Ala Thr Phe Tyr Leu Phe Asp Asp Ala
 900 905 910
 Gly Glu Glu Asn Phe Ser Ser Arg Met Tyr Cys Ser Phe Tyr Pro Pro
 915 920 925
 Asp Glu Glu Glu Glu Asp Asp Ala Glu Cys Glu Glu Glu Glu Ile Asp
 930 935 940
 Glu Thr Cys Glu His Glu Tyr Gly Thr Glu Asp Asp Tyr Gln Gly Leu
 945 950 955 960
 Pro Leu Glu Phe Gly Ala Ser Ala Glu Thr Val Arg Val Glu Glu Glu
 965 970 975
 Glu Glu Glu Asp Trp Leu Asp Asp Thr Thr Glu Gln Ser Glu Ile Glu
 980 985 990
 Pro Glu Pro Glu Pro Thr Pro Glu Glu Pro Val Asn Gln Phe Thr Gly
 995 1000 1005
 Tyr Leu Lys Leu Thr Asp Asn Val Ala Ile Lys Cys Val Asp Ile
 1010 1015 1020
 Val Lys Glu Ala Gln Ser Ala Asn Pro Met Val Ile Val Asn Ala
 1025 1030 1035
 Ala Asn Ile His Leu Lys His Gly Gly Gly Val Ala Gly Ala Leu
 1040 1045 1050
 Asn Lys Ala Thr Asn Gly Ala Met Gln Lys Glu Ser Asp Asp Tyr
 1055 1060 1065
 Ile Lys Leu Asn Gly Pro Leu Thr Val Gly Gly Ser Cys Leu Leu
 1070 1075 1080
 Ser Gly His Asn Leu Ala Lys Lys Cys Leu His Val Val Gly Pro
 1085 1090 1095
 Asn Leu Asn Ala Gly Glu Asp Ile Gln Leu Leu Lys Ala Ala Tyr
 1100 1105 1110
 Glu Asn Phe Asn Ser Gln Asp Ile Leu Leu Ala Pro Leu Leu Ser
 1115 1120 1125
 Ala Gly Ile Phe Gly Ala Lys Pro Leu Gln Ser Leu Gln Val Cys
 1130 1135 1140
 Val Gln Thr Val Arg Thr Gln Val Tyr Ile Ala Val Asn Asp Lys
 1145 1150 1155
 Ala Leu Tyr Glu Gln Val Val Met Asp Tyr Leu Asp Asn Leu Lys
 1160 1165 1170
 Pro Arg Val Glu Ala Pro Lys Gln Glu Glu Pro Pro Asn Thr Glu
 1175 1180 1185
 Asp Ser Lys Thr Glu Glu Lys Ser Val Val Gln Lys Pro Val Asp
 1190 1195 1200
 Val Lys Pro Lys Ile Lys Ala Cys Ile Asp Glu Val Thr Thr Thr
 1205 1210 1215
 Leu Glu Glu Thr Lys Phe Leu Thr Asn Lys Leu Leu Leu Phe Ala

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1220	1225	1230
Asp Ile Asn Gly Lys Leu Tyr	His Asp Ser Gln Asn Met Leu Arg	
1235	1240	1245
Gly Glu Asp Met Ser Phe Leu	Glu Lys Asp Ala Pro Tyr Met Val	
1250	1255	1260
Gly Asp Val Ile Thr Ser Gly	Asp Ile Thr Cys Val Val Ile Pro	
1265	1270	1275
Ser Lys Lys Ala Gly Gly Thr	Thr Glu Met Leu Ser Arg Ala Leu	
1280	1285	1290
Lys Lys Val Pro Val Asp Glu	Tyr Ile Thr Thr Tyr Pro Gly Gln	
1295	1300	1305
Gly Cys Ala Gly Tyr Thr Leu	Glu Glu Ala Lys Thr Ala Leu Lys	
1310	1315	1320
Lys Cys Lys Ser Ala Phe Tyr	Val Leu Pro Ser Glu Ala Pro Asn	
1325	1330	1335
Ala Lys Glu Glu Ile Leu Gly	Thr Val Ser Trp Asn Leu Arg Glu	
1340	1345	1350
Met Leu Ala His Ala Glu Glu	Thr Arg Lys Leu Met Pro Ile Cys	
1355	1360	1365
Met Asp Val Arg Ala Ile Met	Ala Thr Ile Gln Arg Lys Tyr Lys	
1370	1375	1380
Gly Ile Lys Ile Gln Glu Gly	Ile Val Asp Tyr Gly Val Arg Phe	
1385	1390	1395
Phe Phe Tyr Thr Ser Lys Glu	Pro Val Ala Ser Ile Ile Thr Lys	
1400	1405	1410
Leu Asn Ser Leu Asn Glu Pro	Leu Val Thr Met Pro Ile Gly Tyr	
1415	1420	1425
Val Thr His Gly Phe Asn Leu	Glu Glu Ala Ala Arg Cys Met Arg	
1430	1435	1440
Ser Leu Lys Ala Pro Ala Val	Val Ser Val Ser Ser Pro Asp Ala	
1445	1450	1455
Val Thr Thr Tyr Asn Gly Tyr	Leu Thr Ser Ser Ser Lys Thr Ser	
1460	1465	1470
Glu Glu His Phe Val Glu Thr	Val Ser Leu Ala Gly Ser Tyr Arg	
1475	1480	1485
Asp Trp Ser Tyr Ser Gly Gln	Arg Thr Glu Leu Gly Val Glu Phe	
1490	1495	1500
Leu Lys Arg Gly Asp Lys Ile	Val Tyr His Thr Leu Glu Ser Pro	
1505	1510	1515
Val Glu Phe His Leu Asp Gly	Glu Val Leu Ser Leu Asp Lys Leu	
1520	1525	1530
Lys Ser Leu Leu Ser Leu Arg	Glu Val Lys Thr Ile Lys Val Phe	
1535	1540	1545
Thr Thr Val Asp Asn Thr Asn	Leu His Thr Gln Leu Val Asp Met	
1550	1555	1560
Ser Met Thr Tyr Gly Gln Gln	Phe Gly Pro Thr Tyr Leu Asp Gly	
1565	1570	1575
Ala Asp Val Thr Lys Ile Lys	Pro His Val Asn His Glu Gly Lys	
1580	1585	1590
Thr Phe Phe Val Leu Pro Ser	Asp Asp Thr Leu Arg Ser Glu Ala	
1595	1600	1605
Phe Glu Tyr Tyr His Thr Leu	Asp Glu Ser Phe Leu Gly Arg Tyr	
1610	1615	1620

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Met	Ser	Ala	Leu	Asn	His	Thr	Lys	Lys	Trp	Lys	Phe	Pro	Gln	Val
1625						1630					1635			
Gly	Gly	Leu	Thr	Ser	Ile	Lys	Trp	Ala	Asp	Asn	Asn	Cys	Tyr	Leu
1640						1645					1650			
Ser	Ser	Val	Leu	Leu	Ala	Leu	Gln	Gln	Leu	Glu	Val	Lys	Phe	Asn
1655						1660					1665			
Ala	Pro	Ala	Leu	Gln	Glu	Ala	Tyr	Tyr	Arg	Ala	Arg	Ala	Gly	Asp
1670						1675					1680			
Ala	Ala	Asn	Phe	Cys	Ala	Leu	Ile	Leu	Ala	Tyr	Ser	Asn	Lys	Thr
1685						1690					1695			
Val	Gly	Glu	Leu	Gly	Asp	Val	Arg	Glu	Thr	Met	Thr	His	Leu	Leu
1700						1705					1710			
Gln	His	Ala	Asn	Leu	Glu	Ser	Ala	Lys	Arg	Val	Leu	Asn	Val	Val
1715						1720					1725			
Cys	Lys	His	Cys	Gly	Gln	Lys	Thr	Thr	Thr	Leu	Thr	Gly	Val	Glu
1730						1735					1740			
Ala	Val	Met	Tyr	Met	Gly	Thr	Leu	Ser	Tyr	Asp	Asn	Leu	Lys	Thr
1745						1750					1755			
Gly	Val	Ser	Ile	Pro	Cys	Val	Cys	Gly	Arg	Asp	Ala	Thr	Gln	Tyr
1760						1765					1770			
Leu	Val	Gln	Gln	Glu	Ser	Ser	Phe	Val	Met	Met	Ser	Ala	Pro	Pro
1775						1780					1785			
Ala	Glu	Tyr	Lys	Leu	Gln	Gln	Gly	Thr	Phe	Leu	Cys	Ala	Asn	Glu
1790						1795					1800			
Tyr	Thr	Gly	Asn	Tyr	Gln	Cys	Gly	His	Tyr	Thr	His	Ile	Thr	Ala
1805						1810					1815			
Lys	Glu	Thr	Leu	Tyr	Arg	Ile	Asp	Gly	Ala	His	Leu	Thr	Lys	Met
1820						1825					1830			
Ser	Glu	Tyr	Lys	Gly	Pro	Val	Thr	Asp	Val	Phe	Tyr	Lys	Glu	Thr
1835						1840					1845			
Ser	Tyr	Thr	Thr	Thr	Ile	Lys	Pro	Val	Ser	Tyr	Lys	Leu	Asp	Gly
1850						1855					1860			
Val	Thr	Tyr	Thr	Glu	Ile	Glu	Pro	Lys	Leu	Asp	Gly	Tyr	Tyr	Lys
1865						1870					1875			
Lys	Asp	Asn	Ala	Tyr	Tyr	Thr	Glu	Gln	Pro	Ile	Asp	Leu	Val	Pro
1880						1885					1890			
Thr	Gln	Pro	Leu	Pro	Asn	Ala	Ser	Phe	Asp	Asn	Phe	Lys	Leu	Thr
1895						1900					1905			
Cys	Ser	Asn	Thr	Lys	Phe	Ala	Asp	Asp	Leu	Asn	Gln	Met	Thr	Gly
1910						1915					1920			
Phe	Thr	Lys	Pro	Ala	Ser	Arg	Glu	Leu	Ser	Val	Thr	Phe	Phe	Pro
1925						1930					1935			
Asp	Leu	Asn	Gly	Asp	Val	Val	Ala	Ile	Asp	Tyr	Arg	His	Tyr	Ser
1940						1945					1950			
Ala	Ser	Phe	Lys	Lys	Gly	Ala	Lys	Leu	Leu	His	Lys	Pro	Ile	Val
1955						1960					1965			
Trp	His	Ile	Asn	Gln	Ala	Thr	Thr	Lys	Thr	Thr	Phe	Lys	Pro	Asn
1970						1975					1980			
Thr	Trp	Cys	Leu	Arg	Cys	Leu	Trp	Ser	Thr	Lys	Pro	Val	Asp	Thr
1985						1990					1995			
Ser	Asn	Ser	Phe	Glu	Val	Leu	Ala	Val	Glu	Asp	Thr	Gln	Gly	Met
2000						2005					2010			
Asp	Asn	Leu	Ala	Cys	Glu	Ser	Gln	Gln	Pro	Thr	Ser	Glu	Glu	Val
2015						2020					2025			

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Val	Glu	Asn	Pro	Thr	Ile	Gln	Lys	Glu	Val	Ile	Glu	Cys	Asp	Val
2030						2035					2040			
Lys	Thr	Thr	Glu	Val	Val	Gly	Asn	Val	Ile	Leu	Lys	Pro	Ser	Asp
2045						2050					2055			
Glu	Gly	Val	Lys	Val	Thr	Gln	Glu	Leu	Gly	His	Glu	Asp	Leu	Met
2060						2065					2070			
Ala	Ala	Tyr	Val	Glu	Asn	Thr	Ser	Ile	Thr	Ile	Lys	Lys	Pro	Asn
2075						2080					2085			
Glu	Leu	Ser	Leu	Ala	Leu	Gly	Leu	Lys	Thr	Ile	Ala	Thr	His	Gly
2090						2095					2100			
Ile	Ala	Ala	Ile	Asn	Ser	Val	Pro	Trp	Ser	Lys	Ile	Leu	Ala	Tyr
2105						2110					2115			
Val	Lys	Pro	Phe	Leu	Gly	Gln	Ala	Ala	Ile	Thr	Thr	Ser	Asn	Cys
2120						2125					2130			
Ala	Lys	Arg	Leu	Ala	Gln	Arg	Val	Phe	Asn	Asn	Tyr	Met	Pro	Tyr
2135						2140					2145			
Val	Phe	Thr	Leu	Leu	Phe	Gln	Leu	Cys	Thr	Phe	Thr	Lys	Ser	Thr
2150						2155					2160			
Asn	Ser	Arg	Ile	Arg	Ala	Ser	Leu	Pro	Thr	Thr	Ile	Ala	Lys	Asn
2165						2170					2175			
Ser	Val	Lys	Ser	Val	Ala	Lys	Leu	Cys	Leu	Asp	Ala	Gly	Ile	Asn
2180						2185					2190			
Tyr	Val	Lys	Ser	Pro	Lys	Phe	Ser	Lys	Leu	Phe	Thr	Ile	Ala	Met
2195						2200					2205			
Trp	Leu	Leu	Leu	Leu	Ser	Ile	Cys	Leu	Gly	Ser	Leu	Ile	Cys	Val
2210						2215					2220			
Thr	Ala	Ala	Phe	Gly	Val	Leu	Leu	Ser	Asn	Phe	Gly	Ala	Pro	Ser
2225						2230					2235			
Tyr	Cys	Asn	Gly	Val	Arg	Glu	Leu	Tyr	Leu	Asn	Ser	Ser	Asn	Val
2240						2245					2250			
Thr	Thr	Met	Asp	Phe	Cys	Glu	Gly	Ser	Phe	Pro	Cys	Ser	Ile	Cys
2255						2260					2265			
Leu	Ser	Gly	Leu	Asp	Ser	Leu	Asp	Ser	Tyr	Pro	Ala	Leu	Glu	Thr
2270						2275					2280			
Ile	Gln	Val	Thr	Ile	Ser	Ser	Tyr	Lys	Leu	Asp	Leu	Thr	Ile	Leu
2285						2290					2295			
Gly	Leu	Ala	Ala	Glu	Trp	Val	Leu	Ala	Tyr	Met	Leu	Phe	Thr	Lys
2300						2305					2310			
Phe	Phe	Tyr	Leu	Leu	Gly	Leu	Ser	Ala	Ile	Met	Gln	Val	Phe	Phe
2315						2320					2325			
Gly	Tyr	Phe	Ala	Ser	His	Phe	Ile	Ser	Asn	Ser	Trp	Leu	Met	Trp
2330						2335					2340			
Phe	Ile	Ile	Ser	Ile	Val	Gln	Met	Ala	Pro	Val	Ser	Ala	Met	Val
2345						2350					2355			
Arg	Met	Tyr	Ile	Phe	Phe	Ala	Ser	Phe	Tyr	Tyr	Ile	Trp	Lys	Ser
2360						2365					2370			
Tyr	Val	His	Ile	Met	Asp	Gly	Cys	Thr	Ser	Ser	Thr	Cys	Met	Met
2375						2380					2385			
Cys	Tyr	Lys	Arg	Asn	Arg	Ala	Thr	Arg	Val	Glu	Cys	Thr	Thr	Ile
2390						2395					2400			
Val	Asn	Gly	Met	Lys	Arg	Ser	Phe	Tyr	Val	Tyr	Ala	Asn	Gly	Gly
2405						2410					2415			
Arg	Gly	Phe	Cys	Lys	Thr	His	Asn	Trp	Asn	Cys	Leu	Asn	Cys	Asp

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2420	2425	2430
Thr Phe Cys Thr Gly Ser 2435	Thr Phe Ile Ser Asp 2440	Glu Val Ala Arg 2445
Asp Leu Ser Leu Gln Phe 2450	Lys Arg Pro Ile Asn 2455	Pro Thr Asp Gln 2460
Ser Ser Tyr Ile Val Asp 2465	Ser Val Ala Val Lys 2470	Asn Gly Ala Leu 2475
His Leu Tyr Phe Asp Lys 2480	Ala Gly Gln Lys Thr 2485	Tyr Glu Arg His 2490
Pro Leu Ser His Phe Val 2495	Asn Leu Asp Asn Leu 2500	Arg Ala Asn Asn 2505
Thr Lys Gly Ser Leu Pro 2510	Ile Asn Val Ile Val 2515	Phe Asp Gly Lys 2520
Ser Lys Cys Asp Glu Ser 2525	Ala Ser Lys Ser Ala 2530	Ser Val Tyr Tyr 2535
Ser Gln Leu Met Cys Gln 2540	Pro Ile Leu Leu Leu 2545	Asp Gln Ala Leu 2550
Val Ser Asp Val Gly Asp 2555	Ser Thr Glu Val Ser 2560	Val Lys Met Phe 2565
Asp Ala Tyr Val Asp Thr 2570	Phe Ser Ala Thr Phe 2575	Ser Val Pro Met 2580
Glu Lys Leu Lys Ala Leu 2585	Val Ala Thr Ala His 2590	Ser Glu Leu Ala 2595
Lys Gly Val Ala Leu Asp 2600	Gly Val Leu Ser Thr 2605	Phe Val Ser Ala 2610
Ala Arg Gln Gly Val Val 2615	Asp Thr Asp Val Asp 2620	Thr Lys Asp Val 2625
Ile Glu Cys Leu Lys Leu 2630	Ser His His Ser Asp 2635	Leu Glu Val Thr 2640
Gly Asp Ser Cys Asn Asn 2645	Phe Met Leu Thr Tyr 2650	Asn Lys Val Glu 2655
Asn Met Thr Pro Arg Asp 2660	Leu Gly Ala Cys Ile 2665	Asp Cys Asn Ala 2670
Arg His Ile Asn Ala Gln 2675	Val Ala Lys Ser His 2680	Asn Val Ser Leu 2685
Ile Trp Asn Val Lys Asp 2690	Tyr Met Ser Leu Ser 2695	Glu Gln Leu Arg 2700
Lys Gln Ile Arg Ser Ala 2705	Ala Lys Lys Asn Asn 2710	Ile Pro Phe Arg 2715
Leu Thr Cys Ala Thr Thr 2720	Arg Gln Val Val Asn 2725	Val Ile Thr Thr 2730
Lys Ile Ser Leu Lys Gly 2735	Gly Lys Ile Val Ser 2740	Thr Cys Phe Lys 2745
Leu Met Leu Lys Ala Thr 2750	Leu Leu Cys Val Leu 2755	Ala Ala Leu Val 2760
Cys Tyr Ile Val Met Pro 2765	Val His Thr Leu Ser 2770	Ile His Asp Gly 2775
Tyr Thr Asn Glu Ile Ile 2780	Gly Tyr Lys Ala Ile 2785	Gln Asp Gly Val 2790
Thr Arg Asp Ile Ile Ser 2795	Thr Asp Asp Cys Phe 2800	Ala Asn Lys His 2805
Ala Gly Phe Asp Ala Trp 2810	Phe Ser Gln Arg Gly 2815	Gly Ser Tyr Lys 2820

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Asn Asp	Lys Ser Cys Pro	Val	Val Ala Ala Ile	Ile Thr Arg Glu	
2825		2830		2835	
Ile Gly	Phe Ile Val Pro	Gly	Leu Pro Gly Thr	Val Leu Arg Ala	
2840		2845		2850	
Ile Asn	Gly Asp Phe Leu	His	Phe Leu Pro Arg	Val Phe Ser Ala	
2855		2860		2865	
Val Gly	Asn Ile Cys Tyr	Thr	Pro Ser Lys Leu	Ile Glu Tyr Ser	
2870		2875		2880	
Asp Phe	Ala Thr Ser Ala	Cys	Val Leu Ala Ala	Glu Cys Thr Ile	
2885		2890		2895	
Phe Lys	Asp Ala Met Gly	Lys	Pro Val Pro Tyr	Cys Tyr Asp Thr	
2900		2905		2910	
Asn Leu	Leu Glu Gly Ser	Ile	Ser Tyr Ser Glu	Leu Arg Pro Asp	
2915		2920		2925	
Thr Arg	Tyr Val Leu Met	Asp	Gly Ser Ile Ile	Gln Phe Pro Asn	
2930		2935		2940	
Thr Tyr	Leu Glu Gly Ser	Val	Arg Val Val Thr	Thr Phe Asp Ala	
2945		2950		2955	
Glu Tyr	Cys Arg His Gly	Thr	Cys Glu Arg Ser	Glu Val Gly Ile	
2960		2965		2970	
Cys Leu	Ser Thr Ser Gly	Arg	Trp Val Leu Asn	Asn Glu His Tyr	
2975		2980		2985	
Arg Ala	Leu Ser Gly Val	Phe	Cys Gly Val Asp	Ala Met Asn Leu	
2990		2995		3000	
Ile Ala	Asn Ile Phe Thr	Pro	Leu Val Gln Pro	Val Gly Ala Leu	
3005		3010		3015	
Asp Val	Ser Ala Ser Val	Val	Ala Gly Gly Ile	Ile Ala Ile Leu	
3020		3025		3030	
Val Thr	Cys Ala Ala Tyr	Tyr	Phe Met Lys Phe	Arg Arg Val Phe	
3035		3040		3045	
Gly Glu	Tyr Asn His Val	Val	Ala Ala Asn Ala	Leu Leu Phe Leu	
3050		3055		3060	
Met Ser	Phe Thr Ile Leu	Cys	Leu Val Pro Ala	Tyr Ser Phe Leu	
3065		3070		3075	
Pro Gly	Val Tyr Ser Val	Phe	Tyr Leu Tyr Leu	Thr Phe Tyr Phe	
3080		3085		3090	
Thr Asn	Asp Val Ser Phe	Leu	Ala His Leu Gln	Trp Phe Ala Met	
3095		3100		3105	
Phe Ser	Pro Ile Val Pro	Phe	Trp Ile Thr Ala	Ile Tyr Val Phe	
3110		3115		3120	
Cys Ile	Ser Leu Lys His	Cys	His Trp Phe Phe	Asn Asn Tyr Leu	
3125		3130		3135	
Arg Lys	Arg Val Met Phe	Asn	Gly Val Thr Phe	Ser Thr Phe Glu	
3140		3145		3150	
Glu Ala	Ala Leu Cys Thr	Phe	Leu Leu Asn Lys	Glu Met Tyr Leu	
3155		3160		3165	
Lys Leu	Arg Ser Glu Thr	Leu	Leu Pro Leu Thr	Gln Tyr Asn Arg	
3170		3175		3180	
Tyr Leu	Ala Leu Tyr Asn	Lys	Tyr Lys Tyr Phe	Ser Gly Ala Leu	
3185		3190		3195	
Asp Thr	Thr Ser Tyr Arg	Glu	Ala Ala Cys Cys	His Leu Ala Lys	
3200		3205		3210	
Ala Leu	Asn Asp Phe Ser	Asn	Ser Gly Ala Asp	Val Leu Tyr Gln	
3215		3220		3225	

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Pro	Pro	Gln	Thr	Ser	Ile	Thr	Ser	Ala	Val	Leu	Gln	Ser	Gly	Phe
	3230					3235					3240			
Arg	Lys	Met	Ala	Phe	Pro	Ser	Gly	Lys	Val	Glu	Gly	Cys	Met	Val
	3245					3250					3255			
Gln	Val	Thr	Cys	Gly	Thr	Thr	Thr	Leu	Asn	Gly	Leu	Trp	Leu	Asp
	3260					3265					3270			
Asp	Thr	Val	Tyr	Cys	Pro	Arg	His	Val	Ile	Cys	Thr	Ala	Glu	Asp
	3275					3280					3285			
Met	Leu	Asn	Pro	Asn	Tyr	Glu	Asp	Leu	Leu	Ile	Arg	Lys	Ser	Asn
	3290					3295					3300			
His	Ser	Phe	Leu	Val	Gln	Ala	Gly	Asn	Val	Gln	Leu	Arg	Val	Ile
	3305					3310					3315			
Gly	His	Ser	Met	Gln	Asn	Cys	Leu	Leu	Arg	Leu	Lys	Val	Asp	Thr
	3320					3325					3330			
Ser	Asn	Pro	Lys	Thr	Pro	Lys	Tyr	Lys	Phe	Val	Arg	Ile	Gln	Pro
	3335					3340					3345			
Gly	Gln	Thr	Phe	Ser	Val	Leu	Ala	Cys	Tyr	Asn	Gly	Ser	Pro	Ser
	3350					3355					3360			
Gly	Val	Tyr	Gln	Cys	Ala	Met	Arg	Pro	Asn	His	Thr	Ile	Lys	Gly
	3365					3370					3375			
Ser	Phe	Leu	Asn	Gly	Ser	Cys	Gly	Ser	Val	Gly	Phe	Asn	Ile	Asp
	3380					3385					3390			
Tyr	Asp	Cys	Val	Ser	Phe	Cys	Tyr	Met	His	His	Met	Glu	Leu	Pro
	3395					3400					3405			
Thr	Gly	Val	His	Ala	Gly	Thr	Asp	Leu	Glu	Gly	Lys	Phe	Tyr	Gly
	3410					3415					3420			
Pro	Phe	Val	Asp	Arg	Gln	Thr	Ala	Gln	Ala	Ala	Gly	Thr	Asp	Thr
	3425					3430					3435			
Thr	Ile	Thr	Leu	Asn	Val	Leu	Ala	Trp	Leu	Tyr	Ala	Ala	Val	Ile
	3440					3445					3450			
Asn	Gly	Asp	Arg	Trp	Phe	Leu	Asn	Arg	Phe	Thr	Thr	Thr	Leu	Asn
	3455					3460					3465			
Asp	Phe	Asn	Leu	Val	Ala	Met	Lys	Tyr	Asn	Tyr	Glu	Pro	Leu	Thr
	3470					3475					3480			
Gln	Asp	His	Val	Asp	Ile	Leu	Gly	Pro	Leu	Ser	Ala	Gln	Thr	Gly
	3485					3490					3495			
Ile	Ala	Val	Leu	Asp	Met	Cys	Ala	Ala	Leu	Lys	Glu	Leu	Leu	Gln
	3500					3505					3510			
Asn	Gly	Met	Asn	Gly	Arg	Thr	Ile	Leu	Gly	Ser	Thr	Ile	Leu	Glu
	3515					3520					3525			
Asp	Glu	Phe	Thr	Pro	Phe	Asp	Val	Val	Arg	Gln	Cys	Ser	Gly	Val
	3530					3535					3540			
Thr	Phe	Gln	Gly	Lys	Phe	Lys	Lys	Ile	Val	Lys	Gly	Thr	His	His
	3545					3550					3555			
Trp	Met	Leu	Leu	Thr	Phe	Leu	Thr	Ser	Leu	Leu	Ile	Leu	Val	Gln
	3560					3565					3570			
Ser	Thr	Gln	Trp	Ser	Leu	Phe	Phe	Phe	Val	Tyr	Glu	Asn	Ala	Phe
	3575					3580					3585			
Leu	Pro	Phe	Thr	Leu	Gly	Ile	Met	Ala	Ile	Ala	Ala	Cys	Ala	Met
	3590					3595					3600			
Leu	Leu	Val	Lys	His	Lys	His	Ala	Phe	Leu	Cys	Leu	Phe	Leu	Leu
	3605					3610					3615			
Pro	Ser	Leu	Ala	Thr	Val	Ala	Tyr	Phe	Asn	Met	Val	Tyr	Met	Pro

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3620	3625	3630
Ala Ser Trp Val Met Arg Ile Met Thr Trp Leu Glu Leu Ala Asp 3635 3640 3645		
Thr Ser Leu Ser Gly Tyr Arg Leu Lys Asp Cys Val Met Tyr Ala 3650 3655 3660		
Ser Ala Leu Val Leu Leu Ile Leu Met Thr Ala Arg Thr Val Tyr 3665 3670 3675		
Asp Asp Ala Ala Arg Arg Val Trp Thr Leu Met Asn Val Ile Thr 3680 3685 3690		
Leu Val Tyr Lys Val Tyr Tyr Gly Asn Ala Leu Asp Gln Ala Ile 3695 3700 3705		
Ser Met Trp Ala Leu Val Ile Ser Val Thr Ser Asn Tyr Ser Gly 3710 3715 3720		
Val Val Thr Thr Ile Met Phe Leu Ala Arg Ala Ile Val Phe Val 3725 3730 3735		
Cys Val Glu Tyr Tyr Pro Leu Leu Phe Ile Thr Gly Asn Thr Leu 3740 3745 3750		
Gln Cys Ile Met Leu Val Tyr Cys Phe Leu Gly Tyr Cys Cys Cys 3755 3760 3765		
Cys Tyr Phe Gly Leu Phe Cys Leu Leu Asn Arg Tyr Phe Arg Leu 3770 3775 3780		
Thr Leu Gly Val Tyr Asp Tyr Leu Val Ser Thr Gln Glu Phe Arg 3785 3790 3795		
Tyr Met Asn Ser Gln Gly Leu Leu Pro Pro Lys Ser Ser Ile Asp 3800 3805 3810		
Ala Phe Lys Leu Asn Ile Lys Leu Leu Gly Ile Gly Gly Lys Pro 3815 3820 3825		
Cys Ile Lys Val Ala Thr Val Gln Ser Lys Met Ser Asp Val Lys 3830 3835 3840		
Cys Thr Ser Val Val Leu Leu Ser Val Leu Gln Gln Leu Arg Val 3845 3850 3855		
Glu Ser Ser Ser Lys Leu Trp Ala Gln Cys Val Gln Leu His Asn 3860 3865 3870		
Asp Ile Leu Leu Ala Lys Asp Thr Thr Glu Ala Phe Glu Lys Met 3875 3880 3885		
Val Ser Leu Leu Ser Val Leu Leu Ser Met Gln Gly Ala Val Asp 3890 3895 3900		
Ile Asn Arg Leu Cys Glu Glu Met Leu Asp Asn Arg Ala Thr Leu 3905 3910 3915		
Gln Ala Ile Ala Ser Glu Phe Ser Ser Leu Pro Ser Tyr Ala Ala 3920 3925 3930		
Tyr Ala Thr Ala Gln Glu Ala Tyr Glu Gln Ala Val Ala Asn Gly 3935 3940 3945		
Asp Ser Glu Val Val Leu Lys Lys Leu Lys Lys Ser Leu Asn Val 3950 3955 3960		
Ala Lys Ser Glu Phe Asp Arg Asp Ala Ala Met Gln Arg Lys Leu 3965 3970 3975		
Glu Lys Met Ala Asp Gln Ala Met Thr Gln Met Tyr Lys Gln Ala 3980 3985 3990		
Arg Ser Glu Asp Lys Arg Ala Lys Val Thr Ser Ala Met Gln Thr 3995 4000 4005		
Met Leu Phe Thr Met Leu Arg Lys Leu Asp Asn Asp Ala Leu Asn 4010 4015 4020		

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Asn Ile	Ile Asn Asn Ala Arg	Asp Gly Cys Val Pro	Leu Asn Ile
4025	4030	4035	
Ile Pro	Leu Thr Thr Ala Ala	Lys Leu Met Val Val	Val Pro Asp
4040	4045	4050	
Tyr Gly	Thr Tyr Lys Asn Thr	Cys Asp Gly Asn Thr	Phe Thr Tyr
4055	4060	4065	
Ala Ser	Ala Leu Trp Glu Ile	Gln Gln Val Val Asp	Ala Asp Ser
4070	4075	4080	
Lys Ile	Val Gln Leu Ser Glu	Ile Asn Met Asp Asn	Ser Pro Asn
4085	4090	4095	
Leu Ala	Trp Pro Leu Ile Val	Thr Ala Leu Arg Ala	Asn Ser Ala
4100	4105	4110	
Val Lys	Leu Gln Asn Asn Glu	Leu Ser Pro Val Ala	Leu Arg Gln
4115	4120	4125	
Met Ser	Cys Ala Ala Gly Thr	Thr Gln Thr Ala Cys	Thr Asp Asp
4130	4135	4140	
Asn Ala	Leu Ala Tyr Tyr Asn	Asn Ser Lys Gly Gly	Arg Phe Val
4145	4150	4155	
Leu Ala	Leu Leu Ser Asp His	Gln Asp Leu Lys Trp	Ala Arg Phe
4160	4165	4170	
Pro Lys	Ser Asp Gly Thr Gly	Thr Ile Tyr Thr Glu	Leu Glu Pro
4175	4180	4185	
Pro Cys	Arg Phe Val Thr Asp	Thr Pro Lys Gly Pro	Lys Val Lys
4190	4195	4200	
Tyr Leu	Tyr Phe Ile Lys Gly	Leu Asn Asn Leu Asn	Arg Gly Met
4205	4210	4215	
Val Leu	Gly Ser Leu Ala Ala	Thr Val Arg Leu Gln	Ala Gly Asn
4220	4225	4230	
Ala Thr	Glu Val Pro Ala Asn	Ser Thr Val Leu Ser	Phe Cys Ala
4235	4240	4245	
Phe Ala	Val Asp Pro Ala Lys	Ala Tyr Lys Asp Tyr	Leu Ala Ser
4250	4255	4260	
Gly Gly	Gln Pro Ile Thr Asn	Cys Val Lys Met Leu	Cys Thr His
4265	4270	4275	
Thr Gly	Thr Gly Gln Ala Ile	Thr Val Thr Pro Glu	Ala Asn Met
4280	4285	4290	
Asp Gln	Glu Ser Phe Gly Gly	Ala Ser Cys Cys Leu	Tyr Cys Arg
4295	4300	4305	
Cys His	Ile Asp His Pro Asn	Pro Lys Gly Phe Cys	Asp Leu Lys
4310	4315	4320	
Gly Lys	Tyr Val Gln Ile Pro	Thr Thr Cys Ala Asn	Asp Pro Val
4325	4330	4335	
Gly Phe	Thr Leu Arg Asn Thr	Val Cys Thr Val Cys	Gly Met Trp
4340	4345	4350	
Lys Gly	Tyr Gly Cys Ser Cys	Asp Gln Leu Arg Glu	Pro Leu Met
4355	4360	4365	
Gln Ser	Ala Asp Ala Ser Thr	Phe Leu Asn Gly Phe	Ala Val
4370	4375	4380	

<210> SEQ ID NO 75

<211> LENGTH: 2695

<212> TYPE: PRT

<213> ORGANISM: CORONAVIRUS

<400> SEQUENCE: 75

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Arg	Val	Cys	Gly	Val	Ser	Ala	Ala	Arg	Leu	Thr	Pro	Cys	Gly	Thr	Gly
1				5					10					15	
Thr	Ser	Thr	Asp	Val	Val	Tyr	Arg	Ala	Phe	Asp	Ile	Tyr	Asn	Glu	Lys
			20					25					30		
Val	Ala	Gly	Phe	Ala	Lys	Phe	Leu	Lys	Thr	Asn	Cys	Cys	Arg	Phe	Gln
		35					40					45			
Glu	Lys	Asp	Glu	Glu	Gly	Asn	Leu	Leu	Asp	Ser	Tyr	Phe	Val	Val	Lys
	50					55					60				
Arg	His	Thr	Met	Ser	Asn	Tyr	Gln	His	Glu	Glu	Thr	Ile	Tyr	Asn	Leu
65					70					75					80
Val	Lys	Asp	Cys	Pro	Ala	Val	Ala	Val	His	Asp	Phe	Phe	Lys	Phe	Arg
				85					90					95	
Val	Asp	Gly	Asp	Met	Val	Pro	His	Ile	Ser	Arg	Gln	Arg	Leu	Thr	Lys
			100					105					110		
Tyr	Thr	Met	Ala	Asp	Leu	Val	Tyr	Ala	Leu	Arg	His	Phe	Asp	Glu	Gly
		115					120					125			
Asn	Cys	Asp	Thr	Leu	Lys	Glu	Ile	Leu	Val	Thr	Tyr	Asn	Cys	Cys	Asp
	130					135					140				
Asp	Asp	Tyr	Phe	Asn	Lys	Lys	Asp	Trp	Tyr	Asp	Phe	Val	Glu	Asn	Pro
145					150					155					160
Asp	Ile	Leu	Arg	Val	Tyr	Ala	Asn	Leu	Gly	Glu	Arg	Val	Arg	Gln	Ser
				165					170					175	
Leu	Leu	Lys	Thr	Val	Gln	Phe	Cys	Asp	Ala	Met	Arg	Asp	Ala	Gly	Ile
			180					185					190		
Val	Gly	Val	Leu	Thr	Leu	Asp	Asn	Gln	Asp	Leu	Asn	Gly	Asn	Trp	Tyr
		195					200					205			
Asp	Phe	Gly	Asp	Phe	Val	Gln	Val	Ala	Pro	Gly	Cys	Gly	Val	Pro	Ile
	210					215					220				
Val	Asp	Ser	Tyr	Tyr	Ser	Leu	Leu	Met	Pro	Ile	Leu	Thr	Leu	Thr	Arg
225					230					235					240
Ala	Leu	Ala	Ala	Glu	Ser	His	Met	Asp	Ala	Asp	Leu	Ala	Lys	Pro	Leu
				245					250					255	
Ile	Lys	Trp	Asp	Leu	Leu	Lys	Tyr	Asp	Phe	Thr	Glu	Glu	Arg	Leu	Cys
			260					265						270	
Leu	Phe	Asp	Arg	Tyr	Phe	Lys	Tyr	Trp	Asp	Gln	Thr	Tyr	His	Pro	Asn
		275					280					285			
Cys	Ile	Asn	Cys	Leu	Asp	Asp	Arg	Cys	Ile	Leu	His	Cys	Ala	Asn	Phe
	290					295					300				
Asn	Val	Leu	Phe	Ser	Thr	Val	Phe	Pro	Pro	Thr	Ser	Phe	Gly	Pro	Leu
305					310					315					320
Val	Arg	Lys	Ile	Phe	Val	Asp	Gly	Val	Pro	Phe	Val	Val	Ser	Thr	Gly
				325					330					335	
Tyr	His	Phe	Arg	Glu	Leu	Gly	Val	Val	His	Asn	Gln	Asp	Val	Asn	Leu
			340					345					350		
His	Ser	Ser	Arg	Leu	Ser	Phe	Lys	Glu	Leu	Leu	Val	Tyr	Ala	Ala	Asp
		355					360					365			
Pro	Ala	Met	His	Ala	Ala	Ser	Gly	Asn	Leu	Leu	Leu	Asp	Lys	Arg	Thr
	370					375					380				
Thr	Cys	Phe	Ser	Val	Ala	Ala	Leu	Thr	Asn	Asn	Val	Ala	Phe	Gln	Thr
385					390					395					400
Val	Lys	Pro	Gly	Asn	Phe	Asn	Lys	Asp	Phe	Tyr	Asp	Phe	Ala	Val	Ser
				405					410					415	
Lys	Gly	Phe	Phe	Lys	Glu	Gly	Ser	Ser	Val	Glu	Leu	Lys	His	Phe	Phe
			420					425					430		

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Phe Ala Gln Asp Gly Asn Ala Ala Ile Ser Asp Tyr Asp Tyr Tyr Arg
435 440 445
Tyr Asn Leu Pro Thr Met Cys Asp Ile Arg Gln Leu Leu Phe Val Val
450 455 460
Glu Val Val Asp Lys Tyr Phe Asp Cys Tyr Asp Gly Gly Cys Ile Asn
465 470 475 480
Ala Asn Gln Val Ile Val Asn Asn Leu Asp Lys Ser Ala Gly Phe Pro
485 490 495
Phe Asn Lys Trp Gly Lys Ala Arg Leu Tyr Tyr Asp Ser Met Ser Tyr
500 505 510
Glu Asp Gln Asp Ala Leu Phe Ala Tyr Thr Lys Arg Asn Val Ile Pro
515 520 525
Thr Ile Thr Gln Met Asn Leu Lys Tyr Ala Ile Ser Ala Lys Asn Arg
530 535 540
Ala Arg Thr Val Ala Gly Val Ser Ile Cys Ser Thr Met Thr Asn Arg
545 550 555 560
Gln Phe His Gln Lys Leu Leu Lys Ser Ile Ala Ala Thr Arg Gly Ala
565 570 575
Thr Val Val Ile Gly Thr Ser Lys Phe Tyr Gly Gly Trp His Asn Met
580 585 590
Leu Lys Thr Val Tyr Ser Asp Val Glu Thr Pro His Leu Met Gly Trp
595 600 605
Asp Tyr Pro Lys Cys Asp Arg Ala Met Pro Asn Met Leu Arg Ile Met
610 615 620
Ala Ser Leu Val Leu Ala Arg Lys His Asn Thr Cys Cys Asn Leu Ser
625 630 635 640
His Arg Phe Tyr Arg Leu Ala Asn Glu Cys Ala Gln Val Leu Ser Glu
645 650 655
Met Val Met Cys Gly Gly Ser Leu Tyr Val Lys Pro Gly Gly Thr Ser
660 665 670
Ser Gly Asp Ala Thr Thr Ala Tyr Ala Asn Ser Val Phe Asn Ile Cys
675 680 685
Gln Ala Val Thr Ala Asn Val Asn Ala Leu Leu Ser Thr Asp Gly Asn
690 695 700
Lys Ile Ala Asp Lys Tyr Val Arg Asn Leu Gln His Arg Leu Tyr Glu
705 710 715 720
Cys Leu Tyr Arg Asn Arg Asp Val Asp His Glu Phe Val Asp Glu Phe
725 730 735
Tyr Ala Tyr Leu Arg Lys His Phe Ser Met Met Ile Leu Ser Asp Asp
740 745 750
Ala Val Val Cys Tyr Asn Ser Asn Tyr Ala Ala Gln Gly Leu Val Ala
755 760 765
Ser Ile Lys Asn Phe Lys Ala Val Leu Tyr Tyr Gln Asn Asn Val Phe
770 775 780
Met Ser Glu Ala Lys Cys Trp Thr Glu Thr Asp Leu Thr Lys Gly Pro
785 790 795 800
His Glu Phe Cys Ser Gln His Thr Met Leu Val Lys Gln Gly Asp Asp
805 810 815
Tyr Val Tyr Leu Pro Tyr Pro Asp Pro Ser Arg Ile Leu Gly Ala Gly
820 825 830
Cys Phe Val Asp Asp Ile Val Lys Thr Asp Gly Thr Leu Met Ile Glu
835 840 845
Arg Phe Val Ser Leu Ala Ile Asp Ala Tyr Pro Leu Thr Lys His Pro

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850					855					860					
Asn	Gln	Glu	Tyr	Ala	Asp	Val	Phe	His	Leu	Tyr	Leu	Gln	Tyr	Ile	Arg
865					870					875					880
Lys	Leu	His	Asp	Glu	Leu	Thr	Gly	His	Met	Leu	Asp	Met	Tyr	Ser	Val
			885						890					895	
Met	Leu	Thr	Asn	Asp	Asn	Thr	Ser	Arg	Tyr	Trp	Glu	Pro	Glu	Phe	Tyr
			900					905						910	
Glu	Ala	Met	Tyr	Thr	Pro	His	Thr	Val	Leu	Gln	Ala	Val	Gly	Ala	Cys
		915					920					925			
Val	Leu	Cys	Asn	Ser	Gln	Thr	Ser	Leu	Arg	Cys	Gly	Ala	Cys	Ile	Arg
	930					935					940				
Arg	Pro	Phe	Leu	Cys	Cys	Lys	Cys	Cys	Tyr	Asp	His	Val	Ile	Ser	Thr
945					950					955					960
Ser	His	Lys	Leu	Val	Leu	Ser	Val	Asn	Pro	Tyr	Val	Cys	Asn	Ala	Pro
				965					970					975	
Gly	Cys	Asp	Val	Thr	Asp	Val	Thr	Gln	Leu	Tyr	Leu	Gly	Gly	Met	Ser
			980					985						990	
Tyr	Tyr	Cys	Lys	Ser	His	Lys	Pro	Pro	Ile	Ser	Phe	Pro	Leu	Cys	Ala
			995				1000					1005			
Asn	Gly	Gln	Val	Phe	Gly	Leu	Tyr	Lys	Asn	Thr	Cys	Val	Gly	Ser	
	1010					1015						1020			
Asp	Asn	Val	Thr	Asp	Phe	Asn	Ala	Ile	Ala	Thr	Cys	Asp	Trp	Thr	
	1025					1030						1035			
Asn	Ala	Gly	Asp	Tyr	Ile	Leu	Ala	Asn	Thr	Cys	Thr	Glu	Arg	Leu	
	1040					1045						1050			
Lys	Leu	Phe	Ala	Ala	Glu	Thr	Leu	Lys	Ala	Thr	Glu	Glu	Thr	Phe	
	1055					1060						1065			
Lys	Leu	Ser	Tyr	Gly	Ile	Ala	Thr	Val	Arg	Glu	Val	Leu	Ser	Asp	
	1070					1075						1080			
Arg	Glu	Leu	His	Leu	Ser	Trp	Glu	Val	Gly	Lys	Pro	Arg	Pro	Pro	
	1085					1090						1095			
Leu	Asn	Arg	Asn	Tyr	Val	Phe	Thr	Gly	Tyr	Arg	Val	Thr	Lys	Asn	
	1100					1105						1110			
Ser	Lys	Val	Gln	Ile	Gly	Glu	Tyr	Thr	Phe	Glu	Lys	Gly	Asp	Tyr	
	1115					1120						1125			
Gly	Asp	Ala	Val	Val	Tyr	Arg	Gly	Thr	Thr	Thr	Tyr	Lys	Leu	Asn	
	1130					1135						1140			
Val	Gly	Asp	Tyr	Phe	Val	Leu	Thr	Ser	His	Thr	Val	Met	Pro	Leu	
	1145					1150						1155			
Ser	Ala	Pro	Thr	Leu	Val	Pro	Gln	Glu	His	Tyr	Val	Arg	Ile	Thr	
	1160					1165						1170			
Gly	Leu	Tyr	Pro	Thr	Leu	Asn	Ile	Ser	Asp	Glu	Phe	Ser	Ser	Asn	
	1175					1180						1185			
Val	Ala	Asn	Tyr	Gln	Lys	Val	Gly	Met	Gln	Lys	Tyr	Ser	Thr	Leu	
	1190					1195						1200			
Gln	Gly	Pro	Pro	Gly	Thr	Gly	Lys	Ser	His	Phe	Ala	Ile	Gly	Leu	
	1205					1210						1215			
Ala	Leu	Tyr	Tyr	Pro	Ser	Ala	Arg	Ile	Val	Tyr	Thr	Ala	Cys	Ser	
	1220					1225						1230			
His	Ala	Ala	Val	Asp	Ala	Leu	Cys	Glu	Lys	Ala	Leu	Lys	Tyr	Leu	
	1235					1240						1245			
Pro	Ile	Asp	Lys	Cys	Ser	Arg	Ile	Ile	Pro	Ala	Arg	Ala	Arg	Val	
	1250					1255						1260			

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Glu	Cys	Phe	Asp	Lys	Phe	Lys	Val	Asn	Ser	Thr	Leu	Glu	Gln	Tyr
1265						1270					1275			
Val	Phe	Cys	Thr	Val	Asn	Ala	Leu	Pro	Glu	Thr	Thr	Ala	Asp	Ile
1280						1285					1290			
Val	Val	Phe	Asp	Glu	Ile	Ser	Met	Ala	Thr	Asn	Tyr	Asp	Leu	Ser
1295						1300					1305			
Val	Val	Asn	Ala	Arg	Leu	Arg	Ala	Lys	His	Tyr	Val	Tyr	Ile	Gly
1310						1315					1320			
Asp	Pro	Ala	Gln	Leu	Pro	Ala	Pro	Arg	Thr	Leu	Leu	Thr	Lys	Gly
1325						1330					1335			
Thr	Leu	Glu	Pro	Glu	Tyr	Phe	Asn	Ser	Val	Cys	Arg	Leu	Met	Lys
1340						1345					1350			
Thr	Ile	Gly	Pro	Asp	Met	Phe	Leu	Gly	Thr	Cys	Arg	Arg	Cys	Pro
1355						1360					1365			
Ala	Glu	Ile	Val	Asp	Thr	Val	Ser	Ala	Leu	Val	Tyr	Asp	Asn	Lys
1370						1375					1380			
Leu	Lys	Ala	His	Lys	Asp	Lys	Ser	Ala	Gln	Cys	Phe	Lys	Met	Phe
1385						1390					1395			
Tyr	Lys	Gly	Val	Ile	Thr	His	Asp	Val	Ser	Ser	Ala	Ile	Asn	Arg
1400						1405					1410			
Pro	Gln	Ile	Gly	Val	Val	Arg	Glu	Phe	Leu	Thr	Arg	Asn	Pro	Ala
1415						1420					1425			
Trp	Arg	Lys	Ala	Val	Phe	Ile	Ser	Pro	Tyr	Asn	Ser	Gln	Asn	Ala
1430						1435					1440			
Val	Ala	Ser	Lys	Ile	Leu	Gly	Leu	Pro	Thr	Gln	Thr	Val	Asp	Ser
1445						1450					1455			
Ser	Gln	Gly	Ser	Glu	Tyr	Asp	Tyr	Val	Ile	Phe	Thr	Gln	Thr	Thr
1460						1465					1470			
Glu	Thr	Ala	His	Ser	Cys	Asn	Val	Asn	Arg	Phe	Asn	Val	Ala	Ile
1475						1480					1485			
Thr	Arg	Ala	Lys	Ile	Gly	Ile	Leu	Cys	Ile	Met	Ser	Asp	Arg	Asp
1490						1495					1500			
Leu	Tyr	Asp	Lys	Leu	Gln	Phe	Thr	Ser	Leu	Glu	Ile	Pro	Arg	Arg
1505						1510					1515			
Asn	Val	Ala	Thr	Leu	Gln	Ala	Glu	Asn	Val	Thr	Gly	Leu	Phe	Lys
1520						1525					1530			
Asp	Cys	Ser	Lys	Ile	Ile	Thr	Gly	Leu	His	Pro	Thr	Gln	Ala	Pro
1535						1540					1545			
Thr	His	Leu	Ser	Val	Asp	Ile	Lys	Phe	Lys	Thr	Glu	Gly	Leu	Cys
1550						1555					1560			
Val	Asp	Ile	Pro	Gly	Ile	Pro	Lys	Asp	Met	Thr	Tyr	Arg	Arg	Leu
1565						1570					1575			
Ile	Ser	Met	Met	Gly	Phe	Lys	Met	Asn	Tyr	Gln	Val	Asn	Gly	Tyr
1580						1585					1590			
Pro	Asn	Met	Phe	Ile	Thr	Arg	Glu	Glu	Ala	Ile	Arg	His	Val	Arg
1595						1600					1605			
Ala	Trp	Ile	Gly	Phe	Asp	Val	Glu	Gly	Cys	His	Ala	Thr	Arg	Asp
1610						1615					1620			
Ala	Val	Gly	Thr	Asn	Leu	Pro	Leu	Gln	Leu	Gly	Phe	Ser	Thr	Gly
1625						1630					1635			
Val	Asn	Leu	Val	Ala	Val	Pro	Thr	Gly	Tyr	Val	Asp	Thr	Glu	Asn
1640						1645					1650			
Asn	Thr	Glu	Phe	Thr	Arg	Val	Asn	Ala	Lys	Pro	Pro	Pro	Gly	Asp
1655						1660					1665			

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Gln Phe 1670	Lys His Leu Ile 1675	Pro Leu Met Tyr Lys 1680	Gly Leu Pro Trp 1685
Asn Val 1685	Val Arg Ile Lys 1690	Val Gln Met Leu Ser 1695	Asp Thr Leu
Lys Gly 1700	Leu Ser Asp Arg 1705	Val Phe Val Leu Trp 1710	Ala His Gly
Phe Glu 1715	Leu Thr Ser Met 1720	Lys Tyr Phe Val Lys 1725	Ile Gly Pro Glu
Arg Thr 1730	Cys Cys Leu Cys 1735	Asp Lys Arg Ala Thr 1740	Cys Phe Ser Thr
Ser Ser 1745	Asp Thr Tyr Ala 1750	Cys Trp Asn His Ser 1755	Val Gly Phe Asp
Tyr Val 1760	Tyr Asn Pro Phe 1765	Met Ile Asp Val Gln 1770	Gln Trp Gly Phe
Thr Gly 1775	Asn Leu Gln Ser 1780	Asn His Asp Gln His 1785	Cys Gln Val His
Gly Asn 1790	Ala His Val Ala 1795	Ser Cys Asp Ala Ile 1800	Met Thr Arg Cys
Leu Ala 1805	Val His Glu Cys 1810	Phe Val Lys Arg Val 1815	Asp Trp Ser Val
Glu Tyr 1820	Pro Ile Ile Gly 1825	Asp Glu Leu Arg Val 1830	Asn Ser Ala Cys
Arg Lys 1835	Val Gln His Met 1840	Val Lys Ser Ala Leu 1845	Leu Leu Ala Asp
Lys Phe 1850	Pro Val Leu His 1855	Asp Ile Gly Asn Pro 1860	Lys Ala Ile Lys
Cys Val 1865	Pro Gln Ala Glu 1870	Val Glu Trp Lys Phe 1875	Tyr Asp Ala Gln
Pro Cys 1880	Ser Asp Lys Ala 1885	Tyr Lys Ile Glu Glu 1890	Leu Phe Tyr Ser
Tyr Ala 1895	Thr His His Asp 1900	Lys Phe Thr Asp Gly 1905	Val Cys Leu Phe
Trp Asn 1910	Cys Asn Val Asp 1915	Arg Tyr Pro Ala Asn 1920	Ala Ile Val Cys
Arg Phe 1925	Asp Thr Arg Val 1930	Leu Ser Asn Leu Asn 1935	Leu Pro Gly Cys
Asp Gly 1940	Gly Ser Leu Tyr 1945	Val Asn Lys His Ala 1950	Phe His Thr Pro
Ala Phe 1955	Asp Lys Ser Ala 1960	Phe Thr Asn Leu Lys 1965	Gln Leu Pro Phe
Phe Tyr 1970	Tyr Ser Asp Ser 1975	Pro Cys Glu Ser His 1980	Gly Lys Gln Val
Val Ser 1985	Asp Ile Asp Tyr 1990	Val Pro Leu Lys Ser 1995	Ala Thr Cys Ile
Thr Arg 2000	Cys Asn Leu Gly 2005	Gly Ala Val Cys Arg 2010	His His Ala Asn
Glu Tyr 2015	Arg Gln Tyr Leu 2020	Asp Ala Tyr Asn Met 2025	Met Ile Ser Ala
Gly Phe 2030	Ser Leu Trp Ile 2035	Tyr Lys Gln Phe Asp 2040	Thr Tyr Asn Leu
Trp Asn 2045	Thr Phe Thr Arg 2050	Leu Gln Ser Leu Glu 2055	Asn Val Ala Tyr
Asn Val	Val Asn Lys Gly His	Phe Asp Gly His Ala	Gly Glu Ala

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2060			2065			2070		
Pro Val	Ser Ile	Ile Asn	Asn Ala	Val Tyr	Thr Lys	Val Asp	Gly	
2075			2080			2085		
Ile Asp	Val Glu	Ile Phe	Glu Asn	Lys Thr	Thr Leu	Pro Val	Asn	
2090			2095			2100		
Val Ala	Phe Glu	Leu Trp	Ala Lys	Arg Asn	Ile Lys	Pro Val	Pro	
2105			2110			2115		
Glu Ile	Lys Ile	Leu Asn	Asn Leu	Gly Val	Asp Ile	Ala Ala	Asn	
2120			2125			2130		
Thr Val	Ile Trp	Asp Tyr	Lys Arg	Glu Ala	Pro Ala	His Val	Ser	
2135			2140			2145		
Thr Ile	Gly Val	Cys Thr	Met Thr	Asp Ile	Ala Lys	Lys Pro	Thr	
2150			2155			2160		
Glu Ser	Ala Cys	Ser Ser	Leu Thr	Val Leu	Phe Asp	Gly Arg	Val	
2165			2170			2175		
Glu Gly	Gln Val	Asp Leu	Phe Arg	Asn Ala	Arg Asn	Gly Val	Leu	
2180			2185			2190		
Ile Thr	Glu Gly	Ser Val	Lys Gly	Leu Thr	Pro Ser	Lys Gly	Pro	
2195			2200			2205		
Ala Gln	Ala Ser	Val Asn	Gly Val	Thr Leu	Ile Gly	Glu Ser	Val	
2210			2215			2220		
Lys Thr	Gln Phe	Asn Tyr	Phe Lys	Lys Val	Asp Gly	Ile Ile	Gln	
2225			2230			2235		
Gln Leu	Pro Glu	Thr Tyr	Phe Thr	Gln Ser	Arg Asp	Leu Glu	Asp	
2240			2245			2250		
Phe Lys	Pro Arg	Ser Gln	Met Glu	Thr Asp	Phe Leu	Glu Leu	Ala	
2255			2260			2265		
Met Asp	Glu Phe	Ile Gln	Arg Tyr	Lys Leu	Glu Gly	Tyr Ala	Phe	
2270			2275			2280		
Glu His	Ile Val	Tyr Gly	Asp Phe	Ser His	Gly Gln	Leu Gly	Gly	
2285			2290			2295		
Leu His	Leu Met	Ile Gly	Leu Ala	Lys Arg	Ser Gln	Asp Ser	Pro	
2300			2305			2310		
Leu Lys	Leu Glu	Asp Phe	Ile Pro	Met Asp	Ser Thr	Val Lys	Asn	
2315			2320			2325		
Tyr Phe	Ile Thr	Asp Ala	Gln Thr	Gly Ser	Ser Lys	Cys Val	Cys	
2330			2335			2340		
Ser Val	Ile Asp	Leu Leu	Leu Asp	Asp Phe	Val Glu	Ile Ile	Lys	
2345			2350			2355		
Ser Gln	Asp Leu	Ser Val	Ile Ser	Lys Val	Val Lys	Val Thr	Ile	
2360			2365			2370		
Asp Tyr	Ala Glu	Ile Ser	Phe Met	Leu Trp	Cys Lys	Asp Gly	His	
2375			2380			2385		
Val Glu	Thr Phe	Tyr Pro	Lys Leu	Gln Ala	Ser Gln	Ala Trp	Gln	
2390			2395			2400		
Pro Gly	Val Ala	Met Pro	Asn Leu	Tyr Lys	Met Gln	Arg Met	Leu	
2405			2410			2415		
Leu Glu	Lys Cys	Asp Leu	Gln Asn	Tyr Gly	Glu Asn	Ala Val	Ile	
2420			2425			2430		
Pro Lys	Gly Ile	Met Met	Asn Val	Ala Lys	Tyr Thr	Gln Leu	Cys	
2435			2440			2445		
Gln Tyr	Leu Asn	Thr Leu	Thr Leu	Ala Val	Pro Tyr	Asn Met	Arg	
2450			2455			2460		

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Val	Ile	His	Phe	Gly	Ala	Gly	Ser	Asp	Lys	Gly	Val	Ala	Pro	Gly
	2465					2470					2475			
Thr	Ala	Val	Leu	Arg	Gln	Trp	Leu	Pro	Thr	Gly	Thr	Leu	Leu	Val
	2480					2485					2490			
Asp	Ser	Asp	Leu	Asn	Asp	Phe	Val	Ser	Asp	Ala	Asp	Ser	Thr	Leu
	2495					2500					2505			
Ile	Gly	Asp	Cys	Ala	Thr	Val	His	Thr	Ala	Asn	Lys	Trp	Asp	Leu
	2510					2515					2520			
Ile	Ile	Ser	Asp	Met	Tyr	Asp	Pro	Arg	Thr	Lys	His	Val	Thr	Lys
	2525					2530					2535			
Glu	Asn	Asp	Ser	Lys	Glu	Gly	Phe	Phe	Thr	Tyr	Leu	Cys	Gly	Phe
	2540					2545					2550			
Ile	Lys	Gln	Lys	Leu	Ala	Leu	Gly	Gly	Ser	Ile	Ala	Val	Lys	Ile
	2555					2560					2565			
Thr	Glu	His	Ser	Trp	Asn	Ala	Asp	Leu	Tyr	Lys	Leu	Met	Gly	His
	2570					2575					2580			
Phe	Ser	Trp	Trp	Thr	Ala	Phe	Val	Thr	Asn	Val	Asn	Ala	Ser	Ser
	2585					2590					2595			
Ser	Glu	Ala	Phe	Leu	Ile	Gly	Ala	Asn	Tyr	Leu	Gly	Lys	Pro	Lys
	2600					2605					2610			
Glu	Gln	Ile	Asp	Gly	Tyr	Thr	Met	His	Ala	Asn	Tyr	Ile	Phe	Trp
	2615					2620					2625			
Arg	Asn	Thr	Asn	Pro	Ile	Gln	Leu	Ser	Ser	Tyr	Ser	Leu	Phe	Asp
	2630					2635					2640			
Met	Ser	Lys	Phe	Pro	Leu	Lys	Leu	Arg	Gly	Thr	Ala	Val	Met	Ser
	2645					2650					2655			
Leu	Lys	Glu	Asn	Gln	Ile	Asn	Asp	Met	Ile	Tyr	Ser	Leu	Leu	Glu
	2660					2665					2670			
Lys	Gly	Arg	Leu	Ile	Ile	Arg	Glu	Asn	Asn	Arg	Val	Val	Val	Ser
	2675					2680					2685			
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 <211> LENGTH: 20
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 <213> ORGANISM: Artificial sequence
 <220> FEATURE:
 <223> OTHER INFORMATION: S/L3/+/4932 primer

<400> SEQUENCE: 76

ccacacacag cttgtggata

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<210> SEQ ID NO 77
 <211> LENGTH: 20
 <212> TYPE: DNA
 <213> ORGANISM: Artificial sequence
 <220> FEATURE:
 <223> OTHER INFORMATION: S/L4/+/6401 primer

<400> SEQUENCE: 77

ccgaagttgt aggcaatgtc

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<210> SEQ ID NO 78
 <211> LENGTH: 20
 <212> TYPE: DNA
 <213> ORGANISM: Artificial sequence
 <220> FEATURE:
 <223> OTHER INFORMATION: S/L4/+/6964 primer

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<400> SEQUENCE: 78
 ttggtgctc cttcttattg 20

<210> SEQ ID NO 79
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 <212> TYPE: DNA
 <213> ORGANISM: Artificial sequence
 <220> FEATURE:
 <223> OTHER INFORMATION: S/L4/-/6817 primer

<400> SEQUENCE: 79
 ccggcatcca aacataattt 20

<210> SEQ ID NO 80
 <211> LENGTH: 20
 <212> TYPE: DNA
 <213> ORGANISM: Artificial sequence
 <220> FEATURE:
 <223> OTHER INFORMATION: S/L5/-/7633 primer

<400> SEQUENCE: 80
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<210> SEQ ID NO 81
 <211> LENGTH: 20
 <212> TYPE: DNA
 <213> ORGANISM: Artificial sequence
 <220> FEATURE:
 <223> OTHER INFORMATION: S/L5/-/8127 primer

<400> SEQUENCE: 81
 catcctttgt gtcaacatcg 20

<210> SEQ ID NO 82
 <211> LENGTH: 20
 <212> TYPE: DNA
 <213> ORGANISM: Artificial sequence
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 <223> OTHER INFORMATION: S/L5/-/8633 primer

<400> SEQUENCE: 82
 gtcacgagtg acaccatcct 20

<210> SEQ ID NO 83
 <211> LENGTH: 20
 <212> TYPE: DNA
 <213> ORGANISM: Artificial sequence
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 <223> OTHER INFORMATION: S/L5/+ /7839 primer

<400> SEQUENCE: 83
 atgcgacgag tctgcttcta 20

<210> SEQ ID NO 84
 <211> LENGTH: 20
 <212> TYPE: DNA
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<400> SEQUENCE: 84
 ttcatagtgc ctggcttacc 20

<210> SEQ ID NO 85
 <211> LENGTH: 20
 <212> TYPE: DNA

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<213> ORGANISM: Artificial sequence
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 <223> OTHER INFORMATION: S/L5/+ /8255 primer

 <400> SEQUENCE: 85

 atcttggcgc atgtattgac 20

<210> SEQ ID NO 86
 <211> LENGTH: 20
 <212> TYPE: DNA
 <213> ORGANISM: Artificial sequence
 <220> FEATURE:
 <223> OTHER INFORMATION: S/L6/- /9422 primer

 <400> SEQUENCE: 86

 tgcattagca gcaacaacat 20

<210> SEQ ID NO 87
 <211> LENGTH: 20
 <212> TYPE: DNA
 <213> ORGANISM: Artificial sequence
 <220> FEATURE:
 <223> OTHER INFORMATION: S/L6/- /9966 primer

 <400> SEQUENCE: 87

 tctgcagaac agcagaagtg 20

<210> SEQ ID NO 88
 <211> LENGTH: 20
 <212> TYPE: DNA
 <213> ORGANISM: Artificial sequence
 <220> FEATURE:
 <223> OTHER INFORMATION: S/L6/- /10542 primer

 <400> SEQUENCE: 88

 cctgtgcagt ttgtctgtca 20

<210> SEQ ID NO 89
 <211> LENGTH: 20
 <212> TYPE: DNA
 <213> ORGANISM: Artificial sequence
 <220> FEATURE:
 <223> OTHER INFORMATION: S/L6/+ /10677 primer

 <400> SEQUENCE: 89

 ccttgtggca atgaagtaca 20

<210> SEQ ID NO 90
 <211> LENGTH: 20
 <212> TYPE: DNA
 <213> ORGANISM: Artificial sequence
 <220> FEATURE:
 <223> OTHER INFORMATION: S/L6/+ /10106 primer

 <400> SEQUENCE: 90

 atgtcatttg cacagcagaa 20

<210> SEQ ID NO 91
 <211> LENGTH: 20
 <212> TYPE: DNA
 <213> ORGANISM: Artificial sequence
 <220> FEATURE:
 <223> OTHER INFORMATION: S/L6/+ /9571 primer

 <400> SEQUENCE: 91

 cttcaatggt ttgccatggt 20

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<210> SEQ ID NO 92
<211> LENGTH: 20
<212> TYPE: DNA
<213> ORGANISM: Artificial sequence
<220> FEATURE:
<223> OTHER INFORMATION: S/L7/-/11271 primer

<400> SEQUENCE: 92
tgcgagctgt catgagaata 20

<210> SEQ ID NO 93
<211> LENGTH: 20
<212> TYPE: DNA
<213> ORGANISM: Artificial sequence
<220> FEATURE:
<223> OTHER INFORMATION: S/L7/-/11801 primer

<400> SEQUENCE: 93
aaccgagagc agtaccacag 20

<210> SEQ ID NO 94
<211> LENGTH: 20
<212> TYPE: DNA
<213> ORGANISM: Artificial sequence
<220> FEATURE:
<223> OTHER INFORMATION: S/L7/-/12383 primer

<400> SEQUENCE: 94
tttggtgctgt gtagtcaatg 20

<210> SEQ ID NO 95
<211> LENGTH: 20
<212> TYPE: DNA
<213> ORGANISM: Artificial sequence
<220> FEATURE:
<223> OTHER INFORMATION: S/L7/+12640 primer

<400> SEQUENCE: 95
ctacgacaga tgtcctgtgc 20

<210> SEQ ID NO 96
<211> LENGTH: 20
<212> TYPE: DNA
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<223> OTHER INFORMATION: S/L7/+12088 primer

<400> SEQUENCE: 96
gagcaggctg tagctaattg 20

<210> SEQ ID NO 97
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<213> ORGANISM: Artificial sequence
<220> FEATURE:
<223> OTHER INFORMATION: S/L7/+11551 primer

<400> SEQUENCE: 97
ttaggctatt gttgctgctg 20

<210> SEQ ID NO 98
<211> LENGTH: 20
<212> TYPE: DNA
<213> ORGANISM: Artificial sequence
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<223> OTHER INFORMATION: S/L8/-/13160 primer

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<400> SEQUENCE: 98
cagacaacat gaagcaccac 20

<210> SEQ ID NO 99
<211> LENGTH: 20
<212> TYPE: DNA
<213> ORGANISM: Artificial sequence
<220> FEATURE:
<223> OTHER INFORMATION: S/L8/-/13704 primer

<400> SEQUENCE: 99
cgctgacgtg atatatgtgg 20

<210> SEQ ID NO 100
<211> LENGTH: 20
<212> TYPE: DNA
<213> ORGANISM: Artificial sequence
<220> FEATURE:
<223> OTHER INFORMATION: S/L8/-/14284 primer

<400> SEQUENCE: 100
tgcacaatga aggatacacc 20

<210> SEQ ID NO 101
<211> LENGTH: 20
<212> TYPE: DNA
<213> ORGANISM: Artificial sequence
<220> FEATURE:
<223> OTHER INFORMATION: S/L8/+14453 primer

<400> SEQUENCE: 101
acatagctcg cgtctcagtt 20

<210> SEQ ID NO 102
<211> LENGTH: 20
<212> TYPE: DNA
<213> ORGANISM: Artificial sequence
<220> FEATURE:
<223> OTHER INFORMATION: S/L8/+13968 primer

<400> SEQUENCE: 102
ggcattgtag gcgtactgac 20

<210> SEQ ID NO 103
<211> LENGTH: 19
<212> TYPE: DNA
<213> ORGANISM: Artificial sequence
<220> FEATURE:
<223> OTHER INFORMATION: S/L8/+13401 primer

<400> SEQUENCE: 103
gtttgcggtg taagtgcag 19

<210> SEQ ID NO 104
<211> LENGTH: 20
<212> TYPE: DNA
<213> ORGANISM: Artificial sequence
<220> FEATURE:
<223> OTHER INFORMATION: S/L9/-/15098 primer

<400> SEQUENCE: 104
tagtggcggc tattgacttc 20

<210> SEQ ID NO 105
<211> LENGTH: 20
<212> TYPE: DNA

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<213> ORGANISM: Artificial sequence
 <220> FEATURE:
 <223> OTHER INFORMATION: S/L9/-/15677 primer

 <400> SEQUENCE: 105

 ctaaaccttg agccgcatag 20

 <210> SEQ ID NO 106
 <211> LENGTH: 20
 <212> TYPE: DNA
 <213> ORGANISM: Artificial sequence
 <220> FEATURE:
 <223> OTHER INFORMATION: S/L9/-/16247 primer

 <400> SEQUENCE: 106

 catggtcata gcagcacttg 20

 <210> SEQ ID NO 107
 <211> LENGTH: 21
 <212> TYPE: DNA
 <213> ORGANISM: Artificial sequence
 <220> FEATURE:
 <223> OTHER INFORMATION: S/L9+/16323 primer

 <400> SEQUENCE: 107

 ccaggttgty atgtcactga t 21

 <210> SEQ ID NO 108
 <211> LENGTH: 20
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 <400> SEQUENCE: 108

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 <210> SEQ ID NO 109
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 <223> OTHER INFORMATION: S/L9+/15288 primer

 <400> SEQUENCE: 109

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 <210> SEQ ID NO 110
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 <212> TYPE: DNA
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 <223> OTHER INFORMATION: S/L10/-/16914 primer

 <400> SEQUENCE: 110

 agtggtgggt acaagccagt 20

 <210> SEQ ID NO 111
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 <212> TYPE: DNA
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 <220> FEATURE:
 <223> OTHER INFORMATION: S/L10/-/17466 primer

 <400> SEQUENCE: 111

 gttccaagga acatgtctgg 20

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<210> SEQ ID NO 112
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<213> ORGANISM: Artificial sequence
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<400> SEQUENCE: 112
aggtgcctgt gtaggatgaa 20

<210> SEQ ID NO 113
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<212> TYPE: DNA
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<223> OTHER INFORMATION: S/L10+/18245 primer

<400> SEQUENCE: 113
gggctgtcat gcaactagag 20

<210> SEQ ID NO 114
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<212> TYPE: DNA
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<223> OTHER INFORMATION: S/L10+/17663 primer

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tcttacacgc aatcctgctt 20

<210> SEQ ID NO 115
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<212> TYPE: DNA
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<210> SEQ ID NO 116
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<400> SEQUENCE: 116
gcaagcagaa ttaaccctca 20

<210> SEQ ID NO 117
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<212> TYPE: DNA
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<400> SEQUENCE: 117
agcaccacct aaattgcatc 20

<210> SEQ ID NO 118
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<223> OTHER INFORMATION: S/L11/-/20002 primer

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<210> SEQ ID NO 119
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tcgaacacat cgtttatgga 20

<210> SEQ ID NO 120
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<210> SEQ ID NO 121
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<210> SEQ ID NO 122
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<400> SEQUENCE: 122
gaggtgcagt cactcgctat 20

<210> SEQ ID NO 123
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<400> SEQUENCE: 123
cagagattgg acctgagcat 20

<210> SEQ ID NO 124
<211> LENGTH: 20
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<400> SEQUENCE: 124
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<210> SEQ ID NO 125
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<212> TYPE: DNA

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<213> ORGANISM: Artificial sequence
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 <210> SEQ ID NO 126
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 <210> SEQ ID NO 127
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 <400> SEQUENCE: 127

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 <210> SEQ ID NO 128
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 <210> SEQ ID NO 129
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 <400> SEQUENCE: 129

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 <210> SEQ ID NO 130
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 <210> SEQ ID NO 131
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 <213> ORGANISM: Artificial sequence
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 <400> SEQUENCE: 131

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<210> SEQ ID NO 132
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<213> ORGANISM: Artificial sequence
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<400> SEQUENCE: 132
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<210> SEQ ID NO 133
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<213> ORGANISM: Artificial sequence
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<223> OTHER INFORMATION: SARS/L2/R5/-/2529 primer

<400> SEQUENCE: 133
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<210> SEQ ID NO 134
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<213> ORGANISM: Artificial sequence
<220> FEATURE:
<223> OTHER INFORMATION: SARS/L3/F3+/4708 primer

<400> SEQUENCE: 134
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<210> SEQ ID NO 135
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<213> ORGANISM: Artificial sequence
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<223> OTHER INFORMATION: SRAS/L3/F4+/5305 primer

<400> SEQUENCE: 135
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<210> SEQ ID NO 136
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<213> ORGANISM: Artificial sequence
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<223> OTHER INFORMATION: SARS/L3/F5+/5822 primer

<400> SEQUENCE: 136
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<210> SEQ ID NO 137
<211> LENGTH: 20
<212> TYPE: DNA
<213> ORGANISM: Artificial sequence
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<223> OTHER INFORMATION: SARS/L3/R3/-/5610 primer

<400> SEQUENCE: 137
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<210> SEQ ID NO 138
<211> LENGTH: 20
<212> TYPE: DNA
<213> ORGANISM: Artificial sequence
<220> FEATURE:
<223> OTHER INFORMATION: SARS/L3/R4/-/4988 primer

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<400> SEQUENCE: 138

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<210> SEQ ID NO 139

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<212> TYPE: DNA

<213> ORGANISM: Artificial sequence

<220> FEATURE:

<223> OTHER INFORMATION: SARS/L3/R5/-/4437 primer

<400> SEQUENCE: 139

atcggacacc atagtcaacg 20

<210> SEQ ID NO 140

<211> LENGTH: 7788

<212> TYPE: DNA

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<220> FEATURE:

<223> OTHER INFORMATION: synthetic S gene

<400> SEQUENCE: 140

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<213> ORGANISM: Artificial sequence
<220> FEATURE:
<223> OTHER INFORMATION: SNE-S1 primer

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<212> TYPE: DNA
<213> ORGANISM: Artificial sequence
<220> FEATURE:
<223> OTHER INFORMATION: SNE-AS1 primer

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<210> SEQ ID NO 143
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<212> TYPE: DNA
<213> ORGANISM: Artificial sequence
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 <220> FEATURE:
 <223> OTHER INFORMATION: PCR primer

<400> SEQUENCE: 145

ataggatcca ccatgtttat tttcttatta tttcttactc tcact 45

<210> SEQ ID NO 146
 <211> LENGTH: 37
 <212> TYPE: DNA
 <213> ORGANISM: Artificial sequence
 <220> FEATURE:
 <223> OTHER INFORMATION: PCR primer

<400> SEQUENCE: 146

atactcgagt tatgtgtaat gtaatttgac acccttg 37

<210> SEQ ID NO 147
 <211> LENGTH: 45
 <212> TYPE: DNA
 <213> ORGANISM: Artificial sequence
 <220> FEATURE:
 <223> OTHER INFORMATION: PCR primer

<400> SEQUENCE: 147

ataggatcca ccatgtttat tttcttatta tttcttactc tcact 45

<210> SEQ ID NO 148
 <211> LENGTH: 36
 <212> TYPE: DNA
 <213> ORGANISM: Artificial sequence
 <220> FEATURE:
 <223> OTHER INFORMATION: PCR primer

<400> SEQUENCE: 148

acctccggat ttaatatatt gctcatatt tcccaa 36

<210> SEQ ID NO 149
 <211> LENGTH: 13
 <212> TYPE: PRT
 <213> ORGANISM: Artificial sequence
 <220> FEATURE:
 <223> OTHER INFORMATION: N-terminal end of SRAS-CoV S protein (amino-acids 1 to 13)

<400> SEQUENCE: 149

Met Phe Ile Phe Leu Leu Phe Leu Thr Leu Thr Ser Gly
 1 5 10

<210> SEQ ID NO 150
 <211> LENGTH: 10
 <212> TYPE: PRT

-continued

<213> ORGANISM: Artificial sequence
 <220> FEATURE:
 <223> OTHER INFORMATION: oligopeptide

 <400> SEQUENCE: 150

 Ser Gly Asp Tyr Lys Asp Asp Asp Asp Lys
 1 5 10

<210> SEQ ID NO 151
 <211> LENGTH: 34
 <212> TYPE: DNA
 <213> ORGANISM: Artificial sequence
 <220> FEATURE:
 <223> OTHER INFORMATION: PCR primer

 <400> SEQUENCE: 151

 actagctagc ggatccacca tgttcatctt cctg 34

<210> SEQ ID NO 152
 <211> LENGTH: 33
 <212> TYPE: DNA
 <213> ORGANISM: Artificial sequence
 <220> FEATURE:
 <223> OTHER INFORMATION: PCR primer

 <400> SEQUENCE: 152

 agtatccgga cttgatgtac tgctcgact tgc 33

<210> SEQ ID NO 153
 <211> LENGTH: 59
 <212> TYPE: DNA
 <213> ORGANISM: Artificial sequence
 <220> FEATURE:
 <223> OTHER INFORMATION: oligonucleotid

 <400> SEQUENCE: 153

 tatgagcttt tttttttttt tttttttggc atataaatag actcggcgcg ccatctgca 59

<210> SEQ ID NO 154
 <211> LENGTH: 53
 <212> TYPE: DNA
 <213> ORGANISM: Artificial sequence
 <220> FEATURE:
 <223> OTHER INFORMATION: oligonucleotid

 <400> SEQUENCE: 154

 gatggcgcgc cgagtctatt tatatgccaa aaaaaaaaaa aaaaaaagc tca 53

<210> SEQ ID NO 155
 <211> LENGTH: 45
 <212> TYPE: DNA
 <213> ORGANISM: Artificial sequence
 <220> FEATURE:
 <223> OTHER INFORMATION: PCR primer

 <400> SEQUENCE: 155

 atacgtacga ccatgtttat tttcttatta tttcttactc tcaact 45

<210> SEQ ID NO 156
 <211> LENGTH: 40
 <212> TYPE: DNA
 <213> ORGANISM: Artificial sequence
 <220> FEATURE:
 <223> OTHER INFORMATION: PCR primer

 <400> SEQUENCE: 156

 atagcgcgct cattaatgtgt aatgtaattt gacacccttg 40

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<210> SEQ ID NO 157
 <211> LENGTH: 20
 <212> TYPE: DNA
 <213> ORGANISM: Artificial sequence
 <220> FEATURE:
 <223> OTHER INFORMATION: PCR primer

<400> SEQUENCE: 157

ccatttcaac aatttgccg

20

<210> SEQ ID NO 158
 <211> LENGTH: 45
 <212> TYPE: DNA
 <213> ORGANISM: Artificial sequence
 <220> FEATURE:
 <223> OTHER INFORMATION: PCR primer

<400> SEQUENCE: 158

ataggatccg cgcgctcatt atttatcgtc gtcattctta taatc

45

The invention claimed is:

1. A method for the detection of a SARS-associated coronavirus infection, from a biological sample, by indirect IgG ELISA using the SARS-associated coronavirus N protein, which comprises providing ELISA plates that have been sensitized with a solution consisting of N protein at a concentration of between 0.5 and 4 µg/ml in a 10 mM PBS buffer, pH 7.2, phenol red at 0.25 ml/l.

2. A method for the detection of a SARS-associated coronavirus infection, from a biological sample, by double epitope ELISA, comprising mixing a serum to be tested with the antigen attached to a solid support, wherein said antigen is a SARS-associated coronavirus N protein and wherein said solid support is sensitized with a solution consisting of N protein at a concentration of between 0.5 and 4 µg/ml in a 10 mM PBS buffer, pH 7.2, phenol red at 0.25 ml/l.

25 3. The method as claimed in claim 2, wherein said N protein is at a concentration of 1 µg/ml.

4. The method as claimed in claim 1, wherein said biological sample is collected 12 days or more after said infection.

5. The method as claimed in claim 2, wherein said biological sample is collected 12 days or more after said infection.

30 6. The method as claimed in claim 1, wherein said N protein is at a concentration of 2 µg/ml.

7. The method as claimed in claim 2, wherein said visualizing antigen consists of said SARS-associated coronavirus N protein conjugated to a visualizing molecule selected from the group consisting of a radioactive atom, a dye, a fluorescent molecule, a fluorophore, and an enzyme.

8. The method as claimed in claim 7, wherein said enzyme is a peroxidase.

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